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MULTI-SCALE ANALYSIS OF SKIN HYPER-PIGMENTATION EVOLUTION

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ABSTRACT

In this paper, we use statistical inference and multi-spectral images to quantify the evolution of skin hyper-pigmentation lesions under treatment. We show that statistical inference allows getting change maps of the disease which can be useful for dermatologists to analyze the disease evolution. Indeed, a local change map is obtained by computing the deviation between two multi-spectral images in a region of interest (ROI). Then, we normalize the obtained map and develop a statistical inference framework to quantify the changes. Finally, we propose a criterion that integrates change maps in order to quantify the treatment efficacy on a patient.

Index Terms— multi-scale analysis, statistical inference, multi-spectral image, skin, hyper-pigmentation.

1. INTRODUCTION

In dermatology, spectral information is used to quantify the severity of pigmentation lesions like melasma. To this end, spectro-colorimeters allow the measurement of an average spectrum on a small skin area. Such measurements can be repeated to get an evolution curve per patient, or to do statistical analysis of a treatment efficacy on a group of patients. The lack of spatial information about the lesion is the weak point of spectro-colorimeters. To overcome this limit we propose to use multi-spectral images. Multi-spectral images are series of gray scale frames taken at different wavelengths. For hyper-pigmentation, the range of the wavelengths is from 400 nm to 900 nm.

To characterize hyper-pigmentation lesions, we decompose the image analysis into three steps: the first step is the lesion segmentation. This segmentation can be done manually, with methods based on physical model [1], on CIE L^*a^*b decomposition [1], or on independent component analysis (ICA) [2]. We proposed a method based on support vector machine (SVM) in [2]. The second step is to design a spectral signature of the lesion. Such a signature can be empirical, for example the Luminance of the CIE L^*a^*b decomposition, or a spectrum obtained by physical analysis [1]. We proposed in [3] a statistical approach based on ICA and on

a function optimisation to obtain a spectral signature. The third step is evaluating the lesion spatial changes between two measurements, which is the purpose of this paper. To perform this evaluation, we have a series of multi-spectral images $[HS_0, \dots, HS_k, \dots, HS_n]$ taken at times $[t_0, \dots, t_k, \dots, t_n]$. Moreover, we have a binary classification mask M_{p_0} defining the ROI computed on HS_0 during the first step, and a spectral signature that allows the integration of the multi-spectral images into gray scale images $[I_0, \dots, I_k, \dots, I_n]$. Thereby, we aim to use the Statistical Parametric Mapping (SPM) methodology [4] based on statistical inference to map the changes between I_0 and each of the I_k images inside the ROI.

The paper is organized as follows. Section 2 describes the statistical inference to quantify local significance of changes. Section 3 proposes a homogeneity criterion to integrate a change map into a scalar criterion. Finally, we demonstrate experimental results on patients whose lesions of melasma have a decreasing severity.

2. STATISTICAL INFERENCE

For skin pigmentation analysis, dermatologists can be interested by both height and low intensity changes inside a lesion. In fact, we are interested to evaluate if a treatment affect the pathological area by decreasing the pigmentation in localized spots, or affect large areas. To this end we propose to use statistical inference. Such a methodology was used for fMRI studies on brain activity [4] to detect small intensity but spatially consistent changes and height intensity changes with a multi-scale analysis called SPM. We adapt this framework to our specific detection problem considering a change map defined by $C_k = I_k - I_0$.

2.1. Change map normalisation

Statistical inference is a mathematical model for highly correlated Gaussian random field. Two transformations are then need for C_k to fit the model. First, we smooth C_k with a Gaussian convolution filter to increase its spatial correlation. Second, the ROI of C_k denoted $C_k^{M_{p_0}}$ is defined by a unique

class on a radiometric image. Thus, $C_k^{M_{p_0}}$ histogram can be model as monomodal. We use a histogram specification to transform $C_k^{M_{p_0}}$ histogram into a Gaussian distribution. We center and reduce the obtained distribution to standardize it. In the following we will denote C_k^N the normalized change map.

2.2. Change map thresholding

C_k^N satisfies the conditions to apply the Gaussian field theory [5]. An important characteristic of a Gaussian field F of size S is the DT (Differential Topology) characteristic defined by:

$$\chi(A_u) = (-1)^{(D-1)} \sum_{k=0}^{D-1} (-1)^k \chi_k(A_u), \quad (1)$$

where the excursion set A_u is defined by:

$$A_u = \{s \in S, F(s) \geq u\}, \quad (2)$$

where u is a given threshold, D the dimension of the field and $\chi_k(A_u)$ the number of points $s \in A_u$ satisfying the conditions: (a) $F(s) = u$, (b) $F^{(i)}(s) = 0, \forall i \in [1, D-1]$, (c) $F^{(D)}(s) > 0$ and (d) the $(D-1) \times (D-1)$ matrix of second order partial derivatives of $F(s)$ has exactly k negative eigenvalues. In the above expressions, $F^{(i)}$ denotes the i^{th} derivative of F . In the 2D case (i.e. $D = 2$), for a given threshold u , the expectation of the DT characteristic of a centred and reduced Gaussian field can be written as [5]:

$$\mathbb{E}_{\chi(A_u)} = \mathcal{S}(2\pi)^{\frac{3}{2}} |\Lambda|^{\frac{1}{2}} \sigma^{-3} u e^{-\frac{u^2}{2\sigma^2}} \quad (3)$$

with \mathcal{S} the area of the field, Λ the 2×2 correlation matrix of the field derivatives, and $\sigma^2 = \mathbb{E}(F^2)$.

In order to estimate the likelihood of a pixel of C_k^N and its neighborhood under the Gaussian field assumption, we compare its characteristic at a threshold u to the DT characteristic. Two parameters can be used to compute this statistic: the maximum intensity and the spatial extent of the considered neighborhood above u .

Let x_0 denote the maximum intensity of $R_u^{x_0}$, a region of C_k^N above the threshold u . The likelihood of this region under the Gaussian field assumption is [6]:

$$P(R_u^{x_0} \in F) = \frac{\mathbb{E}_{\chi(A_{x_0})}}{\mathbb{E}_{\chi(A_u)}} = \frac{x_0}{u} e^{\frac{u^2 - x_0^2}{2}}. \quad (4)$$

Let S_0 denote the spatial extent of $R_u^{S_0}$, a region of C_k^N above the threshold u . The expectation for this region under the Gaussian field assumption is [6]

$$\mathbb{E}(R_u^{S_0} \in F) = \frac{\mathbb{E}(N_u)}{\mathbb{E}_{\chi(A_u)}}, \quad (5)$$

where $\mathbb{E}(N_u)$ is the expectation to have N_u pixels above the threshold u in the random field F . As F follows a standard

Algorithm 1 Statistical Map

Require: I_0, I_k, M_{p_0}

- 1: $C_k = I_k - I_0$
 - 2: Smooth and normalize C_k to get C_k^N (see section 2.1)
 - 3: Choose a family $U = u_1, \dots, u_{n_u}$
 - 4: **for** $i = 1$ to n_u **do**
 - 5: Compute the connected components above u_i
 - 6: **for** each connected component **do**
 - 7: Calculate $P(R_u^{x_0, S_0} \in F)$ with eq. 9
 - 8: **end for**
 - 9: **end for**
 - 10: Merge the obtained connected clusters obtained with the family U to get SM_{t_k}
-

normal distribution,

$$\mathbb{E}(N_u) = \mathcal{S} \int_u^\infty (2\pi)^{\frac{1}{2}} e^{-\frac{x^2}{2}} = \mathcal{S}\Phi(-u) \quad (6)$$

then,

$$\mathbb{E}(R_u^{S_0} \in F) = \frac{\Phi(-u)}{(2\pi)^{\frac{3}{2}} |\Lambda|^{\frac{1}{2}} u e^{-\frac{u^2}{2}}}. \quad (7)$$

Thus, the number of pixels in a region above the threshold u has an exponential distribution with the parameter $\lambda_u = 1/\mathbb{E}(R_u^{S_0} \in F)$. We can then write the probability of a region $R_u^{S_0}$ to appear in a realization of the Gaussian field by:

$$P(R_u^{S_0} \in F) = e^{-\lambda_u S_0} = e^{\frac{(2\pi)^{\frac{3}{2}} |\Lambda|^{\frac{1}{2}} S_0 u e^{-\frac{u^2}{2}}}{\Phi(-u)}}. \quad (8)$$

Finally, for a region of C_k^N above the threshold u with the maximum intensity x_0 and the spatial extent S_0 , the probability of C_k^N to be a realisation of F is [6]:

$$P(R_u^{x_0, S_0} \in F) = \min(P(R_u^{x_0} \in F), P(R_u^{S_0} \in F)). \quad (9)$$

Thereby, to detect changes between I_0 and I_k we define a family of n_u thresholds $U = \{u_1, \dots, u_{n_u}\}$. For each u_i , C_k^N is segmented into regions denoted $R_{u_i}^{x_0, S_0}$. This segmentation is obtained by a connected components analysis of the class defined by the pixels of C_k^N above the threshold u_i . Then, we assign a probability $P(R_{u_i}^{x_0, S_0} \in F)$ to each region $R_{u_i}^{x_0, S_0}$. The superposition of the obtained regions for each threshold gives SM_{t_k} , a statistical map of the changes (Figs. 2 and 3).

The algorithm 1 summarizes the procedure for obtaining SM_{t_k} between I_0 and I_k for a given patient.

2.3. Rare event hypothesis

The statistical approach proposed above makes the rare events hypothesis. In studied images, a large region of the ROI can change if the disease decreases significantly. Thus, if we apply the methodology described above, only a small amount of the changes will be quantified and the resulting homogeneity

criterion will be difficult to analyze. We then introduce the normalisation:

$$F_{t_k/0}^{N_{de}} = F_{t_k/0}^{Ne} + \frac{1}{N_r} \sum_{r=t_0}^{t_k} \left(\mu_{C_{t_0}}(F_{t_r/0}^{Ne}) - \mu_{C_{t_0}}(F_{t_1/0}^{Ne}) \right) \quad (10)$$

where N_r is the number of time measurements between t_0 and t_k . $\mu_{C_{t_0}}(F_{t_r/0}^{Ne})$ (respectively $\mu_{C_{t_0}}(F_{t_1/0}^{Ne})$) denotes the average pixel intensity in the interest area of $F_{t_r/0}^{Ne}$ (respectively $F_{t_1/0}^{Ne}$) delimited by C_{t_0} . If the normalised data with Eq. (10) does not respect the rare event hypothesis, the calculated statistics by Eq. (9) will then be biased. Nevertheless, we do not use the absolute values of these statistics but their spatial repartition.

3. CHANGE CRITERION

The homogeneity map SM_{t_k} can be integrated into a scalar criterion in order to get a treatment efficacy quantification. To do so, we first define a function $f_s(p)$ that represents the spatial repartition of the changes by:

$$f_s(p) = \frac{\text{card}\{x : SM_{t_k}(x) \leq p\}}{\text{card}\{SM_{t_k}\}}. \quad (11)$$

$f_s(p)$ is an increasing function from $[0, 1]$ to $[0, 1]$. As the interpretation of $f_s(p)$ is not straightforward, we prefer to work with $f_{t_k}^p(s)$ that is the inverse function of $f_s(p)$. A linear interpolation is used to calculate $f_{t_k}^p(s)$ with a regular sampling along s . $f_{t_k}^p(s)$ is also an increasing function from $[0, 1]$ to $[0, 1]$. Then, an integration of $f_{t_k}^p(s)$ to a scalar H_{t_k} is computed by:

$$H_{t_k} = 2 \int_0^1 |f_{t_k}^p(s) - s| ds. \quad (12)$$

$H_{t_k} \in [0, 1]$ represents the area between the function $f_{t_k}^p(s)$ and the identity function $Id(s) = s$. Id corresponds to the most heterogeneous repartition that can be expected. Then, the higher H_{t_k} is, the more homogeneous changes are. We use the Riemann integral to calculate H_{t_k} with discrete data. The homogeneity measure of Eq. (12) makes sense only when some changes appear in the image between t_0 and t_k . We estimated that, if less than 10% of the ROI change between I_0 and I_k , the criterion H_{t_k} cannot be interpreted.

4. EXPERIMENTAL RESULTS

This section shows results obtained by the proposed method on patients taken from a clinical study involving melasma. We used the method proposed in [3] to obtain the spectral signature that gives I_k , and used a SVM classification to calculate M_{p_0} . Then, the images series I_0, \dots, I_n were co-registered with the algorithm proposed in [7]. To smooth C_k we use a 13 pixel diameter Gaussian kernel. The chosen family of thresholds is $U = [1, 1.5, 2, 2.5, 3]$ for all studied images. This

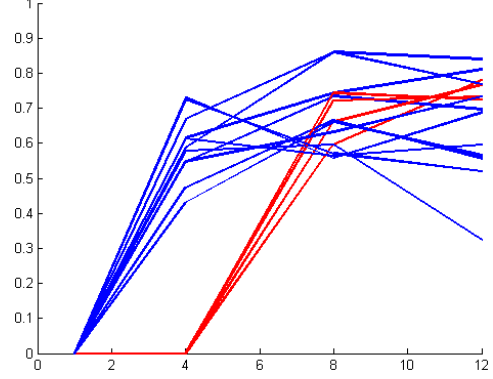


Fig. 1. Time sequences of H for patients whose pathology spatially decrease by more than 10%. **X-axis:** time in weeks ($t_0 = 1, t_1 = 4, t_2 = 8, t_3 = 12$), **Y-axis:** H value.

thresholds family has been manually selected on a subset of images. Figs. 2 and 3 show two examples of obtained maps for two patients' time series. Fig. 2 corresponds to a patient where dermatologist diagnoses a significant change whereas Fig. 3 corresponds to a patient whose diagnosis reveals few changes. As one can see, the proposed method allows us to detect precisely the areas where a change appears. Moreover, we can distinguish at an earlier stage non significant changes (in red) that can become significant later (in blue). We ran the computation of H on a series of 22 patients under a treatment. The clinical analysis of this population concludes that the treatment had globally an effect. In the population of 22 patients, we observed that for 7 patients the criterion H is not interpretable. Indeed, for five patients less than 10% of the ROI changes at time $t_2 = 8$ or $t_3 = 12$ and for two patients there were less than 10% of the ROI that change in all the time sequence. For the others fifteen patients (see Fig. 1) we observe an global growth of H with time. For most patients (in blue) main changes occurred in the second measurement time $t_1 = 4$, and for 4 patients (in red), main changes occurred in time $t_2 = 8$. These results show that the homogeneity criterion gives relevant interpretation of disease progression during treatment.

5. CONCLUSION

In this paper, we adapt the SPM methodology to skin lesion analysis. Associated with a classification and a spectral signature, it gives a precise map of the changes appearing during the treatment phase. For an individual patient, the proposed method gives a spatial analysis of the changes. As the proposed method is automatic, the severity estimation is more robust in a time sequence than a human inspection. For the change detection to succeed, both co-registration between time measurements and classification of the ROI should be accurate.

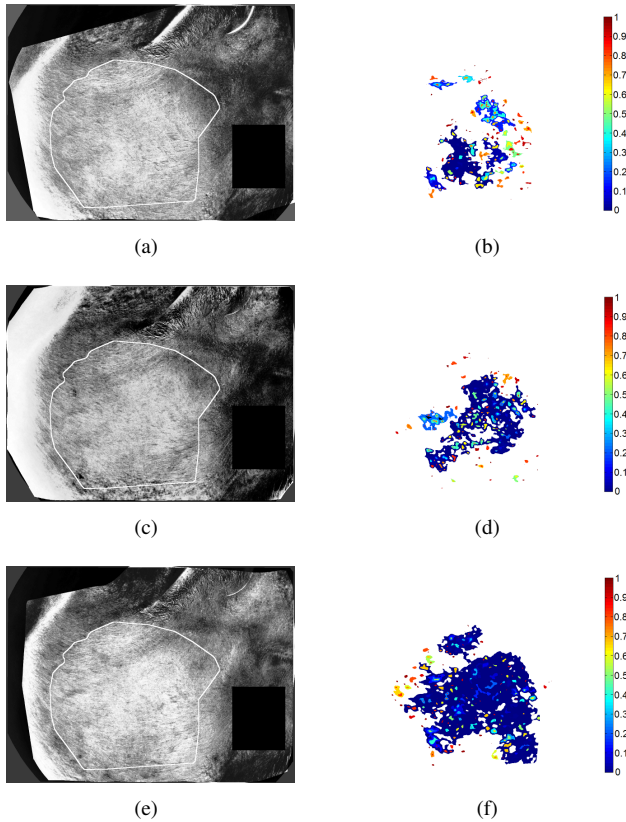


Fig. 2. Change maps obtained for a patient whose pathology decreased during the treatment period. a,c,e) equalised difference images on the spectral feature maps between each time t_1 , t_2 et t_3 and t_0 . b,d,f) homogeneity maps SM_{t_k} for the three time t_1 , t_2 et t_3 using the normalisation of Eq. (10).

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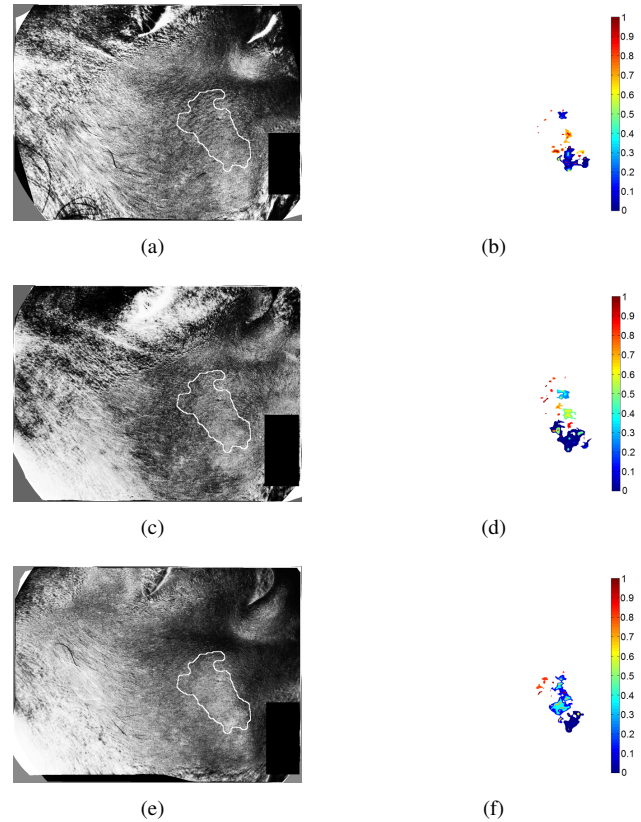


Fig. 3. Change maps obtained for a patient whose pathology decreased during the treatment period. a,c,e) equalised difference image on the spectral feature maps between each time t_1 , t_2 et t_3 and t_0 . b,d,f) homogeneity maps SM_{t_k} for the three time t_1 , t_2 et t_3 using the normalisation of Eq. (10).

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