

Can biomarkers be used to improve diagnosis and prediction of metabolic syndrome in childhood cancer survivors? A systematic review

Pluimakers, V.G.; Santen, S.S. van; Fiocco, M.; Bakker, M.C.E.; Lelij, A.J. van der; Heuveleibrink, M.M. van den; Neggers, S.J.C.M.M.

Citation

Pluimakers, V. G., Santen, S. S. van, Fiocco, M., Bakker, M. C. E., Lelij, A. J. van der, Heuvel-eibrink, M. M. van den, & Neggers, S. J. C. M. M. (2021). Can biomarkers be used to improve diagnosis and prediction of metabolic syndrome in childhood cancer survivors?: A systematic review. *Obesity Reviews*, *22*(11). doi:10.1111/obr.13312

Version:Publisher's VersionLicense:Creative Commons CC BY-NC-ND 4.0 licenseDownloaded from:https://hdl.handle.net/1887/3277372

Note: To cite this publication please use the final published version (if applicable).

PEDIATRIC OBESITY/OBESITY COMORBIDITY

Can biomarkers be used to improve diagnosis and prediction of metabolic syndrome in childhood cancer survivors? A systematic review

Vincent G. Pluimakers ¹ Selveta	S. van Santen ^{1,2} Marta Fiocco ^{1,3,4}
Marie-Christine E. Bakker ^{1,5} Aart	J. van der Lelij ² 💿 🛛
Marry M. van den Heuvel-Eibrink ¹ 💿	Sebastian J. C. M. M. Neggers ^{1,2}

¹Princess Máxima Center for Pediatric Oncology, Utrecht, Netherlands

²Department of Medicine, Endocrinology, Erasmus Medical Center, Rotterdam, Netherlands

³Medical Statistics, Department of Biomedical Data Science, Leiden UMC, Leiden, Netherlands

⁴Mathematical Institute, Leiden University, Leiden, Netherlands

⁵Department of Medicine, University Medical Center Utrecht, Netherlands

Correspondence

V.G. Pluimakers, Princess Máxima Centre for Pediatric Oncology, Heidelberglaan 25, Utrecht 3584 CS, Netherlands. Email: v.g.pluimakers@prinsesmaximacentrum. nl

Funding information Erasmus Medisch Centrum; Princess Máxima Center for Pediatric Oncology

Summary

Childhood cancer survivors (CCS) are at increased risk to develop metabolic syndrome (MetS), diabetes, and cardiovascular disease. Common criteria underestimate adiposity and possibly underdiagnose MetS, particularly after abdominal radiotherapy. A systematic literature review and meta-analysis on the diagnostic and predictive value of nine newer MetS related biomarkers (adiponectin, leptin, uric acid, hsCRP, TNF-alpha, IL-1, IL-6, apolipoprotein B (apoB), and lipoprotein(a) [lp(a)]) in survivors and adult non-cancer survivors was performed by searching PubMed and Embase. Evidence was summarized with GRADE after risk of bias evaluation (QUADAS-2/QUIPS). Eligible studies on promising biomarkers were pooled. We identified 175 general population and five CCS studies. In the general population, valuable predictive biomarkers are uric acid, adiponectin, hsCRP and apoB (high level of evidence), and leptin (moderate level of evidence). Valuable diagnostic biomarkers are hsCRP, adiponectin, uric acid, and leptin (low, low, moderate, and high level of evidence, respectively). Meta-analysis showed OR for hyperuricemia of 2.94 (age-/ sex-adjusted), OR per unit uric acid increase of 1.086 (unadjusted), and AUC for hsCRP of 0.71 (unadjusted). Uric acid, adiponectin, hsCRP, leptin, and apoB can be alternative biomarkers in the screening setting for MetS in survivors, to enhance early identification of those at high risk of subsequent complications.

KEYWORDS

biomarker, childhood cancer survivors, systematic review, the metabolic syndrome

Abbreviations: ALL, acute lymphoblastic leukemia; apoA1, apolipoprotein A1; apoB, apolipoprotein B; AUC, area under the curve; BMI, body mass index; CCS, childhood cancer survivors; CVD, cardiovascular disease; DXA, dual-energy X-ray Absorptiometry; GRADE, Grading of Recommendations Assessment Development and Evaluation; HDL, high density lipoproteins; HOMA-IR, Homeostatic Model Assessment of Insulin Resistance; HR, hazard ratio; hsCRP, high sensitivity C-reactive protein; IL-1, interleukin 1; IL-6, interleukin 6; LDL, low density lipoproteins; Lp(a), lipoprotein(a); MetS, metabolic syndrome; OR, odds ratio; ROC, receiver operating characteristic; T2DM, type 2 diabetes mellitus; TNF-alpha, Tumor Necrosis Factor alpha.

Vincent G. Pluimakers, Selveta S. van Santen, Marry M. van den Heuvel-Eibrink, and Sebastian J.C.M.M. Neggers contributed equally to the content of this manuscript

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Authors. *Obesity Reviews* published by John Wiley & Sons Ltd on behalf of World Obesity Federation.

Wiley

1 | INTRODUCTION

Childhood cancer 5-year survival rates have increased from 5-30% in early seventies to more than 80% in the present time.^{1,2} Deployed therapies, such as chemotherapy, radiotherapy, and stem cell transplantation, better stratification, and enhanced supportive care regimens, are responsible for increase in survival rates. However, intensification of treatment is also associated with long-term excess mortality and morbidities in survivors.³ Survivors have a high level of frailty, suggesting their biological age progresses faster than their actual age. Consequently, survivors with an actual mean age of 33 have a biological age of 65 if they are compared with the general population.⁴⁻⁹ At the age of 45-50 years, the prevalence of any chronic health condition is very high, from 95% up to 99%.^{3,10} One of these severe conditions is represented by cardiovascular disease (CVD), which is an important cause of premature death beyond 5 years cancer survival; the standardized mortality risk for CVD ranges from 1.9 to 12.7.11-25

This high risk of cardiovascular death is not only due to treatment effects, such as anthracycline exposure and cardiac irradiation²⁶; survivors are also at high risk of type II diabetes mellitus (T2DM) and the metabolic syndrome (MetS).¹¹ These diseases are independent predictors of CVD and associated with factors such as adiposity, dyslipidemia, insulin resistance, and hypertension. These factors cluster together and form the "deadly quartet," a MetS concept developed by Reaven in 1988.²⁷ The MetS had many definitions ever since.^{11,27–37} Patients with MetS carry a doubled risk of dying from cardio- and cerebrovascular disease.^{11,38} In addition, patients with the MetS are five times more likely to develop T2DM, which subsequently triples the risk of CVD.^{11,39–41}

As survivors develop cardiovascular complications at a relatively young age, there is a need for early diagnosis of MetS, to possibly prevent T2DM and CVD, and to improve long-term survival.¹¹ The occurrence of MetS may be underestimated especially in abdominally irradiated childhood cancer survivors (CCS), who have an unreliable waist circumference, while their MetS risk is even higher.^{11,42-44} Body mass index (BMI) and bioimpedance are alternative methods for body composition measurement but do not specifically measure abdominal fat, rely on hydration status, and often underestimate body fat.42,45-47 Obviously, another alternative option to evaluate adiposity is measuring fat percentage by Dual-energy X-ray Absorptiometry (DXA) scan, which is the gold standard in case of suspected discordance of anthropomorphic measurements and adiposity.^{42,48,49} However, performing DXA scans in all survivors on a routine basis is time-consuming and costly.¹¹ Additionally, there is currently no consensus for the threshold of fat percentage for diagnosing obesity.⁵⁰ Newer serum biomarkers may serve as another alternative for accurate early diagnosis or prediction of (disguised) MetS in CCS. Adult cardiologists currently apply multiple biomarkers that have been shown to improve risk estimation for CVD.51

Therefore, our primary objectives were to evaluate the value of the use of these newer serum biomarkers as (1) diagnostic marker and as (2) additional independent predictor for the occurrence of MetS later in life, in survivors of childhood cancer specifically, and in a relatively young general, non-cancer population (studies with >75% of participants below 65 years). By including this selection of general population studies as well, we aimed to cover all available literature applicable and generalizable to young-adult survivors. To accomplish this, we performed a systematic literature search on adipokines adiponectin and leptin, uric acid, the inflammatory markers high sensitivity C-reactive protein (hsCRP), Tumor Necrosis Factor alpha (TNF-alpha), interleukin 1 (IL-1) and interleukin 6 (IL-6), and the lipid markers apolipoprotein B (apoB) and lipoprotein(a) [Lp(a)] and performed a meta-analysis of these outcomes for relevant recurrently published biomarkers. As secondary purpose, we screened for other new biomarkers that are not enlisted above, in order to reveal additional, potentially useful biomarkers.

2 | METHODS

2.1 | The systematic search

A systematic literature review was performed in PubMed and Embase, to gather all published literature published between the first of October 2009 and September 3, 2020. Details of the search terms are available in Table S1; in general, the search terms were related to adults/general population, as well as to (childhood) cancer survivors, and combined with all enlisted nine separate biomarkers (adiponectin, leptin, uric acid, hsCRP, TNF-alpha, IL-1, IL-6, apoB, and Ip(a)) and the MetS. The AMSTAR checklist for systematic reviews was followed.⁵² All titles and abstracts were screened by two independent reviewers (VP and SSvS), who were blinded to each other's judgment. Studies were included if they had the MetS as outcome, and one or more newer biomarker(s) as independent variable included in the model in predictive studies, or as discriminative variable in diagnostic studies. For studies performed in CCS, no limits were set for sample size or age. General population studies were eligible if the sample size was roughly 250 or larger and if 75% or more of this population was below 65 years of age, as they have comparable levels of frailty to a young adult survivor population.^{5,7,8} We excluded studies with older adults since they are expected to have higher levels of frailty, comorbidities, and aging factors, which may be confounders in the correlation between the newer biomarker and the metabolic syndrome. Multivariable analysis was mandatory for article inclusion of studies that investigated the prediction of MetS.

Studies were excluded if all included patients had an elevated biomarker; if all or none of the subjects had the MetS; if it was a selected cohort with pre-existing comorbidities (i.e., familial hypercholesterolemia, psoriasis, schizophrenia, polycystic ovary syndrome, obesity, and hypertension); if all patients suffered from MetS endpoint(s) such as T2DM, cardiovascular or cerebrovascular disease, or non-alcoholic fatty liver disease; if the article was a review, case study, expert opinion, or conference abstract; if the article was written in a language other than English or Dutch; or if the full text was unavailable (see Appendix S1 for an overview of selection criteria). Studies were only included if the outcome was presence or absence of MetS; those with separate MetS components or MetS risk score as outcome were out of the scope of this review. After all articles were screened based on title and abstract, the judgments were unblinded. Discrepancies were discussed and resolved by the two reviewers (VP and SSvS), and where necessary, two senior experts were consulted (MMvdHE and SJCMMN). A cross-reference check was performed with Scopus, to screen all forward and backward citations of included studies. The articles found by the cross-reference check were screened likewise. A flow diagram with the number of included and excluded articles and reasons for exclusion illustrates this process (Figure 1).

2.2 | Risk of bias assessment

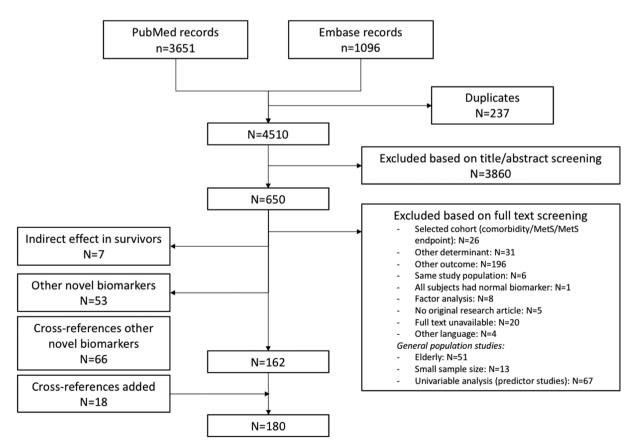
The QUIPS tool was applied for critical appraisal of predictor studies^{53,54} (Table S2) and QUADAS-2 tool for diagnostic studies (Table S3). Definitions for low risk of bias judgment are shown in Appendix S1. In case of doubt, the study was discussed with both reviewers and senior experts (VP, SSvS, MMvdHE, and SJCMMN).

2.3 | Data extraction enlisted novel biomarkers

Data of all included articles were extracted and summarized; the summaries of the enlisted newer biomarkers (adiponectin, leptin, uric acid, hsCRP, TNF-alpha, IL-1, IL-6, apoB, and Ip(a)) are depicted in Table S4A-V. Data of interest are details regarding the size of the population and its type (survivors and their previous diagnosis or general population), the study design (cross-sectional or longitudinal and retrospective or prospective), the biomarker (which and how it was measured), the exact outcome (MetS definition), and statistical analysis of choice. For studies investigating the diagnostic value of the biomarker for MetS, outcomes of interest were area under the curve (AUC) of receiver operating characteristic (ROC) curves, sensitivity, and specificity. For the studies evaluating the predictive value of the biomarker of later development of the MetS, odds ratios (ORs) or beta-coefficients of multivariable logistic regression models, or hazard ratios (HRs) from multivariable Cox Proportional Hazards analysis were extracted from the publications.

2.4 | Summary of evidence

The Grading of Recommendations Assessment Development and Evaluation (GRADE) tool was applied to summarize the quality of the evidence for each biomarker, per clinical research question (diagnosing or predicting MetS) and per population (general population and CCS).⁵⁵ The level of evidence was classified as insufficient, very low, low, moderate, and high (Table S4).⁵⁵ The applied thresholds for biomarkers are shown in Table S5. An overview was made for studies assessing the same independent variables and outcome (Table S6).



2.5 | Data extraction non-enlisted biomarkers

As secondary objective, we screened all articles for other biomarkers than the above enlisted nine biomarkers of our main interest (non-enlisted biomarkers). Details are discussed in Part 2 of Appendix S1. These non-enlisted biomarkers were evaluated for presence of an effect if there were four or more publications with this biomarker in our search. As we did not search for these biomarkers systematically, evidence quality was not assessed with GRADE.

2.6 | Meta-analysis

A meta-analysis was performed of relevant enlisted biomarkers with at least three publications on the same outcome measures and, if applicable, adjusted for the same covariates. Dichotomous outcomes were considered as comparable if the applied threshold differed less than the intra- and inter-assay variability for the biomarker as reported in literature. A random effects model with inverse variance weighting was used to estimate a pooled overall outcome measure. Overall heterogeneity (I-squared) and between-study variance (tau-squared) were calculated.⁵⁶ Meta-analysis was performed with the package *meta* in R.⁵⁷

3 | RESULTS

3.1 | Study selection

As shown in the flow chart (Figure 1), the literature search in PubMed and Embase yielded a total of 4,510 unique records. After title and abstract screening, 650 full-text articles were reviewed, after which 162 relevant studies remained. Backward and forward citation searching identified 18 additional studies. Hence, a total of 180 studies were identified that reported on the diagnostic and/or predictive value of one or more of the enlisted nine biomarkers of interest. Only five studies among the 180 were performed among a population of CCS.^{58–62} All other studies were performed in the general population.

Among 180 studies which included data regarding the 9 enlisted biomarkers, 60 also reported the value of other, non-enlisted newer biomarkers. Furthermore, we identified 119 other studies that only investigated non-enlisted newer biomarkers (other than the nine of our main interest), yielding a total of 179 studies for our secondary objective.

A detailed description of the critical appraisal of each of the 180 included studies for the nine predefined biomarkers is provided in the supporting information (Tables S2 and S3).

3.2 | Used metabolic syndrome definitions

In the included studies, a variety of MetS definitions was used of which the most common are described in Table 1, and the applied definition per study is depicted in Table S4. The applied biomarker thresholds are summarized in Table S5.

3.3 | Evidence for newer, enlisted biomarkers as (additional) diagnostic criterion for metabolic syndrome

Twenty-nine studies reported on the diagnostic value of one or more of the nine enlisted newer biomarkers. These were all performed in the general population without a history of cancer. Six studies had a Caucasian study population.^{63–68} The number of studies per biomarker ranged between 0 [IL-1 and lp(a)] and 12 (adiponectin). The biomarker studied in the largest total number of participants was uric acid (73,190 participants). The relevant data extracted from each study, as well as the summary of evidence scored with the GRADE tool for each biomarker, are provided in the supporting information (Table S4). For each biomarker, a description of the number of studies and participants and a summary of the several diagnostic outcomes are provided in Table 2.

Whereas, ideally, the additional diagnostic value of a biomarker would be tested by comparing the AUC, sensitivity and specificity for a model containing only relevant covariates, versus a model containing covariates and the newer biomarker, this method was used in only two of the 29 studies.^{65,81} One study compared the AUC of the biomarker with the AUC of waist circumference.⁸¹ Most studies, however, only reported the AUC of the biomarker, either unadjusted or adjusted for age, sex, and sometimes BMI or waist circumference. Therefore, interpretation of the additional value is limited by detection and confounding bias for most of the biomarkers.

The overall summary of our findings, with a conclusion about the diagnostic value of each biomarker in the general population and in survivors based on the GRADE assessment, is shown in Figure 2. Of the nine investigated biomarkers, four were identified as valuable diagnostic biomarkers for MetS: leptin (high quality of evidence), uric acid (moderate quality), adiponectin, and hsCRP (both low quality). In addition, apoB may be valuable, although based on only one study with moderate quality of evidence. TNF-alpha and IL-6 appeared to be unusable, based on one low-quality study testing both biomarkers. For IL-1 and Ip(a), no studies were found.

3.4 | Evidence for newer, enlisted biomarkers as independent predictor of metabolic syndrome

In total, 162 general population studies, and 5 survivor studies (two in acute lymphoblastic leukemia [ALL] survivors, two in survivors of hematological malignancies, and one in survivors of heterogeneous tumors),^{58–62} investigated the role of one or more of the nine enlisted, newer biomarkers as independent predictors of MetS. Twenty-six of the general population studies had a Western/Caucasian study population.^{65,67,68,87,93–100,166–171,197–199,218–220,231,233} The number of general population studies per biomarker ranged between

	NCEP ATP III	IDF 2006	Joint interim statement/harmonized definition	Japanese Obesity Society	Chinese diabetes society	Children and adolescents	Modified with BMI instead of waist circumference
Required for MetS diagnosis	3 or more criteria	Obesity plus 2 or more criteria	3 or more criteria	Obesity plus 2 or more criteria	3 or more criteria	3 or more criteria	
Obesity	Waist circumference >102 cm (men) or >88 cm (women)	Waist circumference >90 cm (men) or >80 cm (women)	Waist circumference with ethnic-specific thresholds	Waist circumference >85 cm (men) or >90 cm (women)	Body mass index ≥25 kg/m²	Waist circumference ≥90th percentile	Body mass index ≥30 (Caucasians) or ≥25 (Asians) kg/m ²
Insulin resistance	Fasting plasma glucose ≥5.6 mmol/L or treatment	Fasting plasma glucose ≥5.6 mmol/L or treatment	Fasting plasma glucose ≥5.6 mmol/L or treatment	Fasting plasma glucose ≥6.1 mmol/L or treatment	Fasting plasma glucose ≥6.1 mmol/L or treatment	Fasting plasma glucose ≥5.6 mmol/L or treatment	
Dyslipidemia	Triglycerides ≥1.7 mmol/L or treatment	Triglycerides ≥1.7 mmol/L or treatment	Triglycerides ≥1.7 mmol/L or treatment	Triglycerides ≥1.7 mmol/L, or HDL cholesterol <1 mmol/L (men) or <1.3 mmol/L (women), or treatment	Triglycerides ≥1.7 mmol/L, or HDL cholesterol <0.9 mmol/L (men) or <1.0 mmol/L (women), or treatment	Triglycerides ≥1.7 mmol/L or treatment	
	HDL cholesterol <1 mmol/L (men) or <1.3 mmol/L (women) or treatment	HDL cholesterol <1 mmol/L (men) or <1.3 mmol/L (women) or treatment	HDL cholesterol <1 mmol/L (men) or <1.3 mmol/L (women) or treatment			HDL cholesterol <1 mmol/L or treatment	
Hypertension	≥130/85 mmHg or treatment	≥130/85 mmHg or treatment	≥130/85 mmHg or treatment	≥130/85 mmHg or treatment	≥140/90 mmHg or treatment	≥130/85 mmHg or treatment	

TABLE 1 Commonly used metabolic syndrome definitions in selected studies

	C 3			
Biomarker	Total number of studies and participants	Outcome	Number of studies	Range
Summary of outcomes in diagnostic studies	studies			
Leptin, in general population	6 studies, 8,209 participants ^{68–73}	AUC	568-70,72,73	0.68-0.93
		Sensitivity	3 ^{69,71,73}	48.0-92.6%
		Specificity	3 ^{69,71,73}	56.3-72.0%
Uric acid, in general population	9 studies, 73,190 participants ^{66,73-80}	AUC	766,73-75,77,78,80	0.56-0.85
		Sensitivity	3 ^{73,76,77}	38.0-76.0%
		Specificity	3 ^{73,76,77}	56.0-85.0%
Adiponectin, in general population	12 studies, 21,888 participants ^{63,65,67-}	AUC	1263,65,67-70,81-86	0.55-0.92
	00-10/0/	Sensitivity	2 ^{69,84}	64.7-69.3%
		Specificity	2 ^{69,84}	56.0-66.0%
hsCRP, in general population	7 studies, 18,211 participants ^{64,74,87-91}	AUC	664,74,87-89,91	0.55-0.74
		Sensitivity	3^{89-91}	51.0-69.0%
		Specificity	3 ^{89–91}	56.6-72.0%
ApoB, in general population	1 study, 8,120 participants ⁹²	AUC	1 ⁹²	0.68
TNF-alpha, in general population	1 study, 976 participants ⁶⁴	AUC	164	0.54
IL-6, in general population	1 study, 976 participants ⁶⁴	AUC	164	0.56
IL-1 and lp(a), in general population	No studies	n.a.	n.a.	n.a.
All biomarkers, in survivors	No studies	n.a.	n.a.	n.a.
Summary of outcomes in prognostic studies	studies			
Uric acid, in general population	78 studies, 447,559 participants ^{74–77,79,80,93–164}	OR dichotomous	21 ^{75,96,97,101–105,109,115,118,119,125,} 127,128,135,141,148,157,158,165	1.00-5.17
		OR per unit	1974,76,99,100,105,106,110,111,116,121, 123,124,126,144,146,149,150,159	1.001-2.14
		OR per unit log-transformed	2 ^{77,163}	1.16, 2.08
		OR highest quantile	2494,103,112,114,117-120,122,129,132, 133,136,138-140,142,143,151- 153,157,162,164	1.00-8.04
		HR dichotomous	5 ^{77,80,107,131,161}	1.06-2.99
		HR per unit	479,146,147,155	1.10-2.35
		HR per SD	3 ^{107,108,156}	0.86-1.36
		HR highest quantile	879,107,113,130,147,154–156	0.74-3.47
		HR per unit longitudinal increase	2 ^{107,146}	1.05, 1.31
		RR per unit log-transformed	1^{137}	7.25 for men, 13.26 for women
		RR per SD	1 ⁹³	1.10
		RR per 1.4 mg/dl	1 ⁹⁸	1.54 for men, 1.82 for women
		RR highest quantile	2 ^{93,160}	1.69, 1.76
		PR	2 ^{95,145}	1.47, 2.10
		IRR	1 ⁹⁵	1.73

Summary of outcomes

TABLE 2

PLUIMAKERS ET AL.

Biology (c) Containance of table and porticional of the contact in the cale of colds of c	Total number of studies and participants Momber of studies 2 studies, 5% 6% participants%***** Mess preventers in untra add Q4 vs. Q1-3 Member of studies 3 studies, 5% 6% participants%***** Mess preventers in untra add Q4 vs. Q1-3 2% 3 studies, 5% 6% participants%***** Mess preventers in untra add Q4 vs. Q1-3 2% 3 studies, 5% 6% participants%***** Mess preventers in untra add Q4 vs. Q1-3 2% 3 studies, 5% 6% participants%****** Mess preventers in untra add Q4 vs. Q1-3 2% 3 studies, 5% 6% participants%****** Mess preventers in untra add Q4 vs. Q1-3 2% 3 studies, 139 studies Mess preventers in monos (messe during tollow-up vs. Q1-3 2% 3 studies, 139 studies, 139 studies 1% 1% 3 studies, 139 studies					
2 studies, 500 autrivore ^{44,50} Mills providence in ure acid Qi ko, Qi-3 1% 3 studies, 50,60 participants ^{66,70,10} Mills componention 2 ^{10,21,22,22,72,715,101,114,00,11 3 studies, 50,60 participants^{66,70,10} C per unit 2^{10,21,22,22,72,715,101,114,00,11 2 studies, 50,60 participants^{66,70,10} C per unit 2^{10,21,22,22,72,715,102,114,146,011 2 studies, 50,60 participants^{66,70,10} C per unit 2^{10,21,24,22,72,773,175,114,114,014,114 2 studies, 50,60 participants^{66,70,10} C per unit log-transformed 2^{10,21,24,22,72,773,175,114,114,114 2 studies, 13,03 C per unit log-transformed 2^{10,21,24,24,23,773,173,114,114,114 2 studies, 13,03 C studies 2^{10,21,24,24,24,24 3 studies, 13,03 C studies 2^{10,21,24,24,24,24 3 studies, 13,03 C studies componention 1^{10,21} 3 studies, 46,042 C studies componention 1^{10,21} 3 studies, 13,03 C studies componention 1^{10,11} 3 studies, 13,03 C studies componention 1^{10,11} 3 studies, 13,03 C studies componentin 1^{10,11} 2 st}}}}}}}}	2 Judie, 300 Jurivors ^{46,10} BeS province info and operation in tead of sea Q1-3 1 ¹⁰ 2355,45256 pp. 000 ReS province info and province info an	Biomarker	Total number of studies and participants	Outcome	Number of studies	Range
Referencies Medicionenerision 1 ¹⁰ Referencies Referencies 2 ¹⁷¹¹² Referencies 2 ¹⁷¹¹² 2 ¹⁷¹¹² Referencies 2 ¹⁷¹¹¹¹¹¹² 2 ¹⁷¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹¹	Bit with the set of the properties of the set of the properties of the properis of the properties of the properet of the propertis of	Uric acid, in survivors	2 studies, 390 survivors ^{58,59}	MetS prevalence in uric acid Q4 vs. Q1-3	1 ⁵⁹	28.5% vs. 12.5% ($p=0.0044$)
38,0006 CR dichomos (low adjonnetin) 27013 0.8 per unit 0.8 per unit 27013 0.8 per unit 0.8 per unit 27013 0.8 per unit 0.8 per unit 27013 0.8 per unit 27013 27013 0.8 per unit 27013 27013 0.8 per unit (log transformed 27013 27014 0.8 per unit (log transformed 27013 27014 0.8 per unit (log transformed 27014 27014 0.8 per unit (log transformed 27014 27014 0.8 ber unit (log transformed 27014	Bandless, 56, 656 participants, 56, 756 participants, 576 partionocounding participants, 576 participants, 576 partic			MetS component(s) prevalence high vs. low uric acid	158	60% vs. 24% ($p = 0.04$)
memory Operating Operating <thoperating< th=""> <thoperating< th=""> <thoper< td=""><td>Material Operating <th< td=""><td>Adiponectin, in general population</td><td>38 studies, 56,656 participants^{65,67,69,81-}</td><td>OR dichotomous (low adiponectin)</td><td>2^{170,172}</td><td>0.90, 2.68</td></th<></td></thoper<></thoperating<></thoperating<>	Material Operating Operating <th< td=""><td>Adiponectin, in general population</td><td>38 studies, 56,656 participants^{65,67,69,81-}</td><td>OR dichotomous (low adiponectin)</td><td>2^{170,172}</td><td>0.90, 2.68</td></th<>	Adiponectin, in general population	38 studies, 56,656 participants ^{65,67,69,81-}	OR dichotomous (low adiponectin)	2 ^{170,172}	0.90, 2.68
OR per S units CR per suits 11 OR per unit log transformed 242,134 OR per unit log transformed Z score 11 OR per unit log transformed Z score 12	Circle of soling Circle of soling Circle of solution Circle of		83,86,166-176	OR per unit	967,167,169,174,176,180,181,186,191	0.66-1.08
OR per unit log-transformed dx.dx.12.18 OR per SD OR work of the transformed Z-score 1% OR lower punit log-transformed C-score 1% OR lowe	Of the per unit log-transformed of Rege on the grant formed of the grant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed of the per stant formed			OR per 5 units	1 ⁸¹	0.82 for men, 0.90 for women
OR per SU OR per SU 272.144 OR per unit log transformed Z-score 1 OR lowest quantle 2414 OR lowest quantle 2412 OR lowest quantle 2414 OR lowest quantle 2412 OR lowest quantle 2422 OR lowest qu	0 Rper SD $2^{0.2344}$ 0.60001 0 Rights training 0.60001 0.50001 0 Rights training 0.60001 0.50001 0 Rights training 0.60001 0.50001 0 Rights training 0.60001 0.60001 0 Rights training $0.600000000000000000000000000000000000$			OR per unit log-transformed	486,168,178,183	0.10-0.67
CR per unit log: transformed Z-score 10 CR highest quantile 20 CR highest quantile 21 File developing MES QL vs. Q4 197 The ratio of developing MES QL vs. Q4 197 Autoria 11 Autoria 21	CR per unit log-transformed 2-score 1^{00} C75 for boys, 0.05 for given 3 CR per unit log-transformed 2-score 1^{01} 1^{01} 1^{01} CR holes caratile 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at follow-up 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at hole coontaked 1^{01} 1^{01} 1^{01} 1^{01} R hole creased at hole coontaked 1^{01} 1^{01}			OR per SD	2 ^{192,194}	0.50-0.91
CR highest quantile 10 CR highest quantile 11 CR highest quantile 11 CR howest quantile 64577.1373.1737.1737.103.183.163.102.104-104 CR howest quantile 11 R decreased and forcease during follow-up vs. 11 R decreased at follow-up 11 Time ratio of developing MeIS CQ1 vs. Cd4 11 Baseline and forcease 11 N highest quantile 11 Participants ^{10,00,00,00} 11 Participants ^{10,00,00,00,00,00,00,00,00,00,00,00,00,0}	OR highest quantile 101-0.67 OR highest quantile 647.77.14.17.14.2.14.8.12.0.2.4.4.4.4 101-0.67 PR glowest quantile 7 24.2.4.6.4 23.2.4.6.4 PR glowest quantile 7 24.2.4.6.4.6.4.4.4.4.4.4.4.4.4.4.4.4.4.4			OR per unit log-transformed Z-score	1 ⁶⁹	0.76 for boys, 0.69 for girls
OR lowest quantile GR lowest quantile gs.57.1184484.00.133 HR light baseline and forcessed and forces and forcessed and forces and and forces and for covariants 171 If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of developing MetS QL vs. Q4 1.0° 1.0° If the ratio of the ratio of developing QL vs. Q4 1.0° 1.0° If the ratio of the ratio of the ratio of the ratio of the ra	R web quantifie Genorest q			OR highest quantile	13 ^{81-83,173,175,177,179,182,185,192,194-196}	0.10-0.67
Righ baseline and decrease ouv baseline and decrease in unselline and decrease in non-MetS abjects divided by 1 ⁻¹⁰ Recrease of rollow-up value in non-MetS abjects divided by 1 ⁻¹⁰ Baseline ratio of developing MetS Q1 vs. Qct 1 ⁻¹⁰ Baseline ratio of developing MetS Q1 vs. Qct 1 ⁻¹⁰ Baseline ratio of developing MetS abjects divided by 1 ⁻¹⁰ Baseline ratio of developing MetS abjects divided by 1 ⁻¹⁰ Baseline ratio (value in non-MetS, adjusted for covariates) 1 ⁻¹⁰ Data baseline and forcese 1 ⁻¹⁰ Data baseline and forcese 1 ⁻¹⁰ Data baseline and forcese 1 ⁻¹⁰ Data baseline and forceroninin 1 ⁻¹⁰ Data baseline and forcese 1 ⁻¹⁰ Data baseline and forcese 2 ⁻⁰⁰²⁰³ Data baseline and for covariates 2 ⁻⁰⁰²⁰³ Data baseline and forcese 1 ⁻¹⁰ Data baseline and for covariates 2 ⁻⁰⁰²⁰³ Data baseline	Image: Section of the section and increase during follow-up: 1 ¹⁰ 0.33 Image: Section and increase during follow-up: 1 ¹⁰ 4.37 Image: Section and increase during follow-up: 1 ¹⁰ 0.33 Image: Section and increase during follow-up: 1 ¹⁰ 0.33 Image: Section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and increase during follow-up: 1 ¹⁰ 0.35 Image: Section and increase during d			OR lowest quantile	665,171,184,189,190,193	1.82-18.6
HR decreased at follow-up 171 The ratio of developing MetS QJ vs. Q4 137 The ratio of developing MetS QJ vs. Q4 136 3 studies, 139 survivors ^{98,00,02} Reperson on on-MetS, adjusted for covariates) 146 3 studies, 139 survivors ^{98,00,02} OR highest quantile at baseline and follow-up 12 3 studies, 139 survivors ^{98,00,02} OR highest quantile at baseline and follow-up 12 2 studies, 139 survivors ^{98,00,02} OR highest quantile at baseline and follow-up 12 2 studies, 19,38 0,1,2 of Krustal-Wall components 200,003 2 studies, 19,38 0,1,2 of Krustal-Wall est dual test in the standometer in the stransformed 110,000,000 2 studies, 19,38 0,1,2 of Krustal-Wall est dual test in the stransformed 200,000 2 studies, 19,38 0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,0,	He decreased at follow-up $1^{(1)}$ $43^{(2)}$ The ratio of developing MetS dives $1^{(2)}$ $03^{(2)}$ The ratio of developing MetS dives $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value in works: adjusted tive) $1^{(2)}$ $03^{(2)}$ Baseline anto (value anto (value anto)) $1^{(2)}$ $03^{(2)}$ Baseline anto $1^{(2)}$ $1^{(2)}$ $10^{(2)}$			HR high baseline and increase during follow-up vs. low baseline and decrease	1 ¹⁸⁸	0.33
Time ratio of developing MetS Q1 vs. Q4 1 ⁴⁷ Baseline ratio (value in non-MetS, adjusted for covariates) Baseline ratio (value in non-MetS, adjusted for covariates) 3 studies, 139 survivors ^{36,60,63} QR highest quantile at baseline and follow-up 1 ⁴⁶ 3 studies, 139 survivors ^{36,60,63} QR highest quantile at baseline and follow-up 1 ⁴⁶ 9 studies, 139 survivors ^{36,60,64} QR highest quantile at baseline and follow-up 1 ⁴⁶ 9 studies, 139 survivors ^{36,60,64} QR highest quantile at baseline and follow-up 1 ⁴⁶ 9 studies, 119,138 QR highest quantile at baseline and follow-up 1 ⁴⁹ 9 studies, 119,138 QR highest of three groups (<10, 10-30 and	The ratio of developing MetS CU vs. Q4 1 ¹⁰ 015 (-65% shorter time whet subjects divided by whet mone whet subjects divided by whet whet mone whet subjects divided by whet whet mone whet subjects divided by whet whet mone whet divide whet whet mone whet divide whet whet whet mone whet divide whet whet mone whet divide whet whet whet whet whet whet whet whe			HR decreased at follow-up	1^{171}	4.37
Backine ratio (value in MetS. adjusted for covariates) 1 ¹⁰ 3 studies, 139 survivors ^{36,00,13} CR highest quantile at baseline and folow-up 1 ¹⁰ 3 studies, 139 survivors ^{36,00,13} CR highest quantile at baseline and folow-up 1 ¹⁰ 3 studies, 139 survivors ^{36,00,13} CR highest quantile at baseline and folow-up 1 ¹⁰ 3 studies, 139 survivors ^{36,00,13} CR highest quantile st baseline and folow-up 1 ¹⁰ 3 studies, 139 survivors ^{36,00,13} CR highest quantile st baseline and folow-up 1 ¹⁰ 3 studies, 139,130 CR highest quantile 2 ^{200,203} 3 studies, 139,139,130 CR per unit log-transformed 1 ¹⁰ 3 studies, 139,139,130 CR per unit log-transformed 2 ^{90,213,130,130,130,130,130,130,130,130,130,1}	Baseline ratio (value in Met5 subjects divided by a studies, 139 survhors ^{36,0,0,1} 1,2 1,2 3 studies, 139 survhors ^{36,0,0,1} 0 studies (ris constances) 1,9 0,1 1 studies, 139 survhors ^{36,0,0,1} 0 studies (ris constances) 1,9 0,1 1 studies, 139 survhors ^{36,0,0,1} 0 studies (ris constances) 1,9 0,1 1 studies, 139 survhors ^{36,0,0,1} 0 studies (ris constances) 1,9 0,1 1 studies, 1,9,136 0,1 1,1 1,9 0,1 1 studies, 1,9,136 0 studies, 1,9,136 1,0 0,1 1,1 1 studies, 1,9,136 0 studies, 1,9,136 0,1 0,1 0,1 1 studies, 1,9,136 0 studies, 1,9,136 0,0 0,1 0,1 1 studies, 1,9,136 0 studies, 1,9,136 0,0 0,0 0,0 1 studies, 1,9,136 0 studies, 1,9,136 0,0 0,0 0,0 1 studies, 1,9,136 0 studies (studies, 1,0,0 0,0 0,0 0,0 1 studies, 1,9,136 0 studies (studies, 1,0,0 0,0 0,0 0,0 1 studies, 1,9,136 0 studies (studies, 1,0,0 0,0 0,0 0,0 1 studies, 1,9,136 0 studies (studies, 1,0,0 0,0 0,0 0,0 1 study, studies (studies,			Time ratio of developing MetS Q1 vs. Q4	1 ¹⁸⁷	0.15 (=85% shorter time to develop MetS)
3 studies, 139 survivors ^{9,60,02} OR highest quantile at baseline and follow-up 1 ² R dictonous (low adjonectin) H dictonous (low adjonectin) 1 ⁹ Paule of Kruski-Wallis test median adjonnectin in 1 ⁹ 2 studies, 119,138 O, 1, 2 - 4 Mets components 2 ^{00,000} 2 studies, 119,138 OR dichotmous 2 ^{00,000} 2 studies, 119,138 OR et unit 2 ^{00,000} 2 studies, 119,138 OR et unit log-transformed 2 ^{00,000} 2 studies, 119,138 OR et unit log-transformed 1 ¹¹⁰ 2 studies, 119,138 OR et unit log-transformed 2 ^{00,000} 2 studies, 119,138 OR et unit log-transformed 1 ¹¹⁰ 2 studies, 119,138 OR et unit log-transformed 1 ¹¹⁰ 2 studies, 119 OR lighest of three groups (<10, 10-30 and	3 studies. 139 survivors ^{34,00,02} 0 Rhighest quantile at baseline and follow-up 1^{c2} $0.5(n_{c3})$ for baseline. 0.9R dichotomous (low alponectin)1% $0.1, 2-4$ MeS components $0.5(n_{c3})$ for baseline. 0.9P value of Kruskal-Wallis test median adiponectin1% $0.1, 2-4$ MeS components $0.1, 2-4$ MeS component $0.1, 2-4$ MeS components $0.0, 2-37$ $0.0, 2-37$ $0.1, 2-4$ MeS component $1.2, 2-4$ MeS components $1.2, 2-4$ MeS components $1.2, 2-4$ MeS component $0.1, 2-4$ MeS component $1.2, 2-4$ MeS component $1.2, 2-4$ MeS component $1.2, 2-4$ MeS component $0.1, 0.1, 0.1, 0.1, 0.1, 0.2, 0.1, 0.1, 0.1, 0.2, 0.1, 0.1, 0.1, 0.1, 0.1, 0.1, 0.1, 0.1$			Baseline ratio (value in MetS subjects divided by value in non-MetS, adjusted for covariates)	1166	1.27
He dichotomous (low adiponectin) 1^{50} 0.1.2-4 Mets components 1^{50} 0.1.2-4 Mets components 2^{3} studies.19,1380.1.2-4 Mets components 1^{90} 0.1.2-4 Mets components 2^{3} studies.19,1380.1.2-4 Mets components 1^{90} 2^{3} studies.19,1380.1.2-4 Mets components 2^{20208} 2^{3} studies.19,1380.1.2-4 Mets components 1^{90} 2^{3} studies.19,1380.1.2-4 Mets components 2^{20208} 2^{3} studies.19,1380.1.2-4 Mets components 1^{19} 2^{3} studies.19,1380.1.2-4 Mets components 1^{19} 2^{3} studies.19,1380.1.2-4 Mets components 1^{19} 2^{3} studies.19,1380.1.2-10-3.0 and 1^{19} 2^{3} studies0.1.1-2.0 and 1^{10} 2^{2} studies0.1.1-2.0 and 1^{10}	He dichotomous (now adjouncetin) 1^{0} 6.7 Praise of Kichotomous (now adjouncetin) 1^{0} 1.0 1.0 Praise of Kichotomous $0.1.2$ + Mesic componentis 1^{0} 1.0 Praise of Kichotomous $0.1.2$ + Mesic componentis 1^{0} $1.0.07$ Praise of Kichotomous $0.1.2$ + Mesic componentis 1^{0} $1.0.07$ Praise of Kichotomous $0.1.2$ + Mesic componentis 1^{0} 1.007 Praise of Kichotomous 0.000 2^{0000} 1.007 R per unit log-transformed 2^{0000} 1.007 1.007 R per unit log-transformed 2^{0000} 1.007 1.007 R per unit log-transformed 1^{0000} 1.007 1.07 R per unit log-transformed 1^{0000} 1.07 1.07 R per unit log-transformed 1^{00000} 1.07 1.07 R per unit log-transformed 1^{000000} 1.07 1.07 R per unit log-transformed $1^{000000000000000000000000000000000000$	Adiponectin, in survivors	3 studies, 139 survivors ^{58,60,62}	OR highest quantile at baseline and follow-up	1 ⁶²	0.5 (n.s.) for baseline, 0.9 (n.s.) for follow-up
Pradue of Kruskal-Wallis test median adiponectin in 0, 1, 2-4 MetS components 10 0, 1, 2-4 MetS components 0, 1, 2-4 MetS components 0, 1, 2-4 MetS components 200203 participants ^{37,35,90,80,9110,104} 200203 23 studies, 1,9,138 0 R per unit 0, 1, 2-4 MetS components 200203 0, 1, 2-4 MetS components 200203 140, 2010,104,104 200203 0 R per SD log-transformed 11 0 R per SD log-transformed 11 0 R highest of three groups (<10, 1,0-3,0 and	Product (Kuski-Wallis test median adjoonectin in adjoonectin base) $1^{\circ0}$			HR dichotomous (low adiponectin)	1 ⁵⁸	6.7
32 studies, 119,138 OR dichotomous $2^{200,203}$ participants ^{17,177,121,121,121,121,121,121,121,121,1}	32 studies.119,380 0 R dichotomous 200,203 200,203 1.20,274 Participants************************************			P value of Kruskal-Wallis test median adiponectin in 0, 1, 2–4 MetS components	1 ⁶⁰	n.s.
participants OR per unit OR per unit log-transformed 7979110205206.215.217 1001000000000000000000000000000000000	Participants OR per unit Constants 1007-297 1007-297 Participants Rer unit log-transformed 40156.162.11 1007-297 1007-297 Rer unit log-transformed Rer unit log-transformed 297.11 115-3.2 115-3.2 Rer SD log-transformed 297.21 297.11 117-9.202.214.214 107-7.11 Rer Unit log-transformed 297.21 297.11 107-7.11 107-7.11 Rer Unit log-transformed 117-9.202.214.214 107-7.11 107-7.11 Rer Unit log-transformed 117-9.202.02.214.214 115-6186 107-7.11 Rer Unit log-transformed 117-9.202.02.214.214 115-7.11 116-7.11 Rer Unit log-transformed 117-9.202.02.02.14.214 116-7.11 116-7.11 Rer Unit log-transformed 116-6 116-7 116-7.11 116-7.11	hsCRP, in general population	32 studies, 119,138	OR dichotomous	2 ^{200,203}	1.20, 2.74
OR per unit log-transformed $4^{0126168.211}$ OR per SD OR per SD OR per SD OR per SD OR per SD OR per SD log-transformed OR highest quantile 11^{99} OR highest quantile $2^{99,212}$ OR highest quantile $2^{90,212}$ OR highest quantile $2^{90,212}$ OR highest quantile $1^{199,199,201,204,209-2102,124,1246}$ OR highest of three groups (<1.0, 1.0-3.0 and y	Reper unit log-transformed $4^{0.154.162.11}$ $11^{-5}-32$ Reper SD Reper SD 12^{9} 12^{-1} CR Per SD log-transformed 12^{9} 12^{-1} $036,107$ CR Per SD log-transformed $2^{9,212}$ $036,107$ $107-711$ CR Per SD log-transformed $11^{77,193,01,200,206,202-210,210,210,210}$ $107-711$ CR Per Unit De transformed $11^{77,193,01,200,206,202,202-210,210,210,210,210,210,210,210,210,210,$		participants ^{87,89,90,98,99,110,126,} 130,166,168,179,197-217	OR per unit	787,99,110,205,206,215,217	1.007-2.97
OR per SD OR per SD log-transformed 1^{99} OR per SD log-transformed $2^{90,212}$ OR highest of three groups (<10,10-30 and 31 $3^{92,207,201,204,208-2102,214,216}$ OR highest of three groups (<10,10-30 and 31 $3^{202,207,201,204,208-2102,214,216}$ OR highest of three groups (<10,10-30 and 31 $3^{202,207,201,204,208-2102,214,216}$ OR highest of three groups (<10,10-30 and 31 $1^{11}^{177,198,201,204,208-2102,214,216}$ Dulation 1 1 1^{200} Dulation 1 200 1^{200} Dulation 10 studies, 66,924 participants $^{7/148,218-}$ OR dichotomous Dulation 10 studies, 66,924 participants $^{7/148,218-}$ OR dichotomous Dulation 10^{200} 1^{200} Dulation 10^{200} 1^{200} OR per unit 1^{200} 1^{200} OR per unit 1^{200} 1^{200} OR per sol 1^{200} 1^{200} OR per sol 0^{200} 1^{200} OR per sol 1^{200} 1^{200}	$\begin{tabular}{l l l l l l l l l l l l l l l l l l l $			OR per unit log-transformed	490,126,168,211	1.15-3.2
OR per SD log-transformed $2^{95,212}$ OR highest quantile $1^{179,198,201,204,208-210,212,214,216}$ OR highest of three groups (<10, 10–30 and 3 202,207,211	R per SD log-tansformed $2^{9,212}$ 09,107 R highest quantile CR highest quantile 1177.138.201.201.231.214.216 107-7.11 R highest quantile R highest quantile 1177.138.201.201.231.214.216 107-7.11 N highest quantile R highest quantile 1177.138.201.201.231.214.216 107-7.11 N highest of three groups (<10, 10-3.0 and			OR per SD	1^{199}	1.21
OR highest quantile 11^{173,198,201,204,208-2102,112-214,216} OR highest of three groups (<1,0,1,0-3,0 and -3 µg/m)	$\begin{tabular}{l l l l l l l l l l l l l l l l l l l $			OR per SD log-transformed	2 ^{89,212}	0.96, 1.07
$\label{eq:constraint} Partial and Partiana and Partial and Partial and Parti$	$\label{eq:constant} \end{tabular} tab$			OR highest quantile	$11^{179,198,201,204,208-210,212-214,216}$	1.07-7.11
HR per unit log-transformed 1^{30} R per threefold increase 1^{90} R per threefold increase 1^{90} Baseline ratio 1^{106} P value of likelihood test in multivariable model 1^{106} 1 study. 87 survivors and 87 controls ⁶¹ OR dichotomous 1^{106} pulation 10^{10} studies, 66, 924 participants $97.148.218^{-1}$ OR dichotomous 1^{61} pulation 10^{10} studies, 66, 924 participants $97.148.218^{-1}$ OR dichotomous 1^{61} pulation 10^{10} studies, 66, 924 participants $97.148.218^{-1}$ OR dichotomous 1^{61} pulation 10^{10} studies, 66, 924 participants $97.148.218^{-1}$ OR dichotomous 1^{61} pulation 10^{10} studies, 66, 924 participants $97.148.218^{-1}$ OR per unit 1^{220} pulation 0^{10} studies 0^{10} studies 1^{220} pulation 0^{10} studies 1^{220} 1^{220}	$ \begin{array}{llllllllllllllllllllllllllllllllllll$			OR highest of three groups (<1.0, 1.0–3.0 and >3 µg/ml)	3 202,207,211	1.65-18.86
$\begin{tabular}{lllllllllllllllllllllllllllllllllll$	$\begin{tabular}{l l l l l l l l l l l l l l l l l l l $			HR per unit log-transformed	1 ¹³⁰	1.15
pulation 1^{225} Baseline ratio 1^{166} P value of likelihood test in multivariable model 1^{197} 1 study, 87 survivors and 87 controls ^{6,1} OR dichotomous 1^{97} 1^{106} OR dichotomous 1^{97} OR dichotomous 1^{97} 1^{97} OR per unit 1^{225} OR per 30 mg/dl 1^{220} OR per 30 mg/dl 1^{220} OR per SD OR per SD OR per SD 1^{220}	pulation 1^{166} 1^{167} 1^{166} 1^{197} 1^{167} 1^{166} 1^{197} 1^{198} 1^{19			RR per threefold increase	1 ⁹⁸	1.13
$\begin{tabular}{lllllllllllllllllllllllllllllllllll$	$\begin{tabular}{lllllllllllllllllllllllllllllllllll$			Baseline ratio	1 ¹⁶⁶	0.80 (n.s.)
1 study, 87 survivors and 87 controls ⁶¹ OR dichotomous 1 ⁶¹ pulation 10 studies, 66,924 participants ^{97,168,218-} OR dichotomous 1 ⁹⁷ OR per unit 1 ²²⁵ 1 ⁴⁸ 1 ⁴⁸ OR per unit 1 ²²⁰ 1 ²²⁰ OR per SD 0R per 30 mg/dl 1 ²²⁰ OR per SD OR per 30 mg/dl 1 ²²³ OR per SD OR per SD 1 ²²³	$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$			P value of likelihood test in multivariable model	1 ¹⁹⁷	n.s.
$\begin{array}{ccc} 10 \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ $	$ \begin{array}{ccccc} 10 \ {\rm studies}, 66,924 \ {\rm participants}^{77,168,218-} & {\rm OR \ dichotomous} & 1^{97} & 2.55 \\ {\rm OR \ per \ unit} & 1^{168} & 2.99 \\ {\rm OR \ per \ 30 \ mg/dl} & 1^{220} & 1.76 \ {\rm for \ men}, 2.10 \ {\rm for \ wc} \\ {\rm OR \ per \ SD} & {\rm OR \ per \ SD} & 1^{223} & 0.96-6.03 \\ \end{array} $	hsCRP, in survivors	1 study, 87 survivors and 87 controls 61	OR dichotomous	1 ⁶¹	7.26
OR per unit 1^{168} OR per 30 mg/dl 1^{220} OR per SD 1^{223} OR highest quantile $\delta^{218,221-225}$	OR per unit 1^{168} 2.99 OR per 30 mg/dl 1^{220} 1.76 for men, 2.10 for wc OR per SD 1^{223} 1.56 OR highest quantile $6^{218,221-225}$ 0.96-6.03	ApoB, in general population	10 studies, 66,924 participants $^{97,168,218-}$	OR dichotomous	1 ⁹⁷	2.55
1 ²²⁰ 1 ²²³ 6 ^{218,221-225}	1^{220} 1.76 for men, 2.10 for wo 1^{223} 1.56 $6^{218,221-225}$ 0.96-6.03			OR per unit	1 ¹⁶⁸	2.99
1 ²²³ 6 ^{218,221-225}	1 ²²³ 1.56 6 ^{218,221-225} 0.96-6.03			OR per 30 mg/dl	1 ²²⁰	1.76 for men, 2.10 for women
627-1720129	6 ^{210,221,220} 0.96-6.03			OR per SD	1 ²²³	1.56
	C			OR highest quantile	622-122'0129	0.96-6.03

TABLE 2 (Continued)

IABLE 2 (Continuea)				
Biomarker	Total number of studies and participants	Outcome	Number of studies	Range
		OR highest of three groups (<90, 90−119 and ≥120 mg/dl)	1 ²¹⁹	2.69 for men, 1.69 for women
		RR per SD	1 ²²³	1.17 (n.s.)
		RR highest quantile	1 ²²³	1.79
ApoB, in survivors	No studies	n.a.	n.a.	n.a.
Leptin, in general population	17 studies, 28,797	OR dichotomous	1 ¹⁷²	2.39
	participants ^{00,07,/} 4,100,176,177,100, 183,191,192,196,226-230	OR per unit	4167,180,191,229	0.96-1.91
		OR per 10 ng/ml	1 ²²⁶	1.06 (adjusted for WC), 1.22 (adjusted for BMI)
		OR per unit log-transformed	2 ^{168,183}	1.47, 2.76
		OR per SD	368,192,228	1.01-1.31
		OR per unit log-transformed Z-score	1 ⁶⁹	1.81 for boys, 1.32 for girls
		OR highest quantile	672,179,192,196,227,230	1.16-3.02
Leptin, in survivors	3 studies, 139 survivors ^{58,60,62}	OR highest quantile at baseline and follow-up	162	4.8 for baseline, 5.7 for follow-up
		MetS component(s) prevalence high vs. low leptin	1 ⁵⁸	54% vs. 17% (p $=$ 0.03)
		P value of Kruskal-Wallis test median adiponectin in 0, 1, 2-4 MetS components	1 ⁶⁰	n.s.
IL-6, in general population	5 studies, 3,370	OR per unit	2 ^{67,206}	0.98-1.47
	participants ^{o 7,83,200,231,232}	OR highest quantile	2 ^{83,232}	0.98 (n.s.), 4.10
		P value in multivariable model	1 ²³¹	n.s.
IL-6, in survivors	1 study, 87 survivors and 87 controls 61	OR dichotomous	161	1.53 (n.s.)
Lp(a), in general population	5 studies, 15,162 participants ^{207,233-236}	OR dichotomous	1 ²⁰⁷	8.27
		OR highest of three groups (<18.40, 18.40−33.84 and ≥33.85 µg/ml)	1 ²³⁴	0.82 (n.s.)
		OR per unit	1 ²³⁵	1.0 (n.s.)
		OR highest quantile	1 ²³⁶	0.45
		HR highest quantile	1 ²³³	1.01 (n.s.)
Lp(a), in survivors	No studies	n.a.	n.a.	n.a.
IL-1, in general population	4 studies, 1,594 participants ^{166,206,232,237}	OR per unit	2 ^{206,237}	2.28 (IL-1alpha), 1.009, 2.01 (IL- 1beta)
		OR highest quartile	1 ²³²	0.98 (n.s.)
		Baseline ratio	1 ¹⁶⁶	1.17 (suggests effect in other direction)
TNF-alpha, in general population	3 studies, 1,458 participants ^{206,231,232}	OR per unit	1 ²⁰⁶	1.45
		OR highest quartile	1 ²³²	0.78 (n.s.)
		P value in multivariable model	1 ²³¹	n.s.
TNF-alpha, in survivors	1 study, 87 survivors and 87 controls 61	OR dichotomous	161	0.52 (n.s.)

TABLE 2 (Continued)

Metabolic syndrome biomarkers

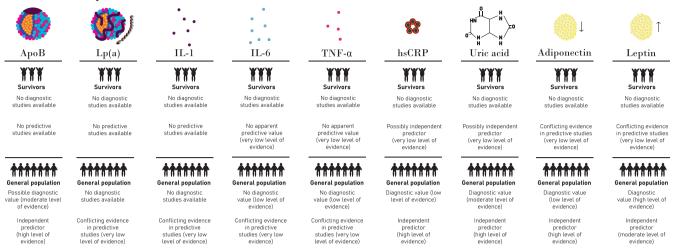


FIGURE 2 Summary of conclusions: predictive and diagnostic value of novel biomarkers for the MetS

3 (TNF-alpha, 1,458 participants in total) and 78 (uric acid, 447,559 participants in total). Two of the survivors studies had a Western/ Caucasian study population^{58,59}; the others were performed in Japan,⁶⁰ Malaysia,⁶¹ and Mexico.⁶² The number of survivors studies per biomarker ranged between zero [IL-1, apoB, and lp(a)] and three (adiponectin and leptin). The biomarker studied in the largest total number of survivors was uric acid (390 survivors). The relevant data extracted from each study, as well as the summary of evidence scored with the GRADE tool for each biomarker, are provided in the supporting information (Table S4). For each biomarker, a description of the number of studies and participants and a summary of the several prognostic outcomes are provided in Table 2.

A common analysis strategy in these studies was to divide the biomarker value in quantiles, with thresholds that may differ per study. Not all participants in the highest or lowest quantile always had a biomarker value that would be classified as abnormal according to reference values. This may attenuate its value in predicting MetS. On the other hand, this bias towards the null hypothesis increases the effect of true positive findings. Also, several studies tested a doseresponse effect by comparing the effect on MetS across the quantiles. Studies can be compared on whether a dose-response effect was observed or not.

Figure 2 shows the overall summary of our findings, consisting of a conclusion about the role of each biomarker as independent predictor of MetS in the general population and in survivors, after GRADE assessment. Five biomarkers were identified as independent predictors of MetS in the general population: uric acid, adiponectin, hsCRP, apoB (all high quality of evidence), and leptin (moderate quality). There is conflicting evidence for the value of TNF-alpha, IL-1, IL-6, and lp(a) (very low quality of evidence). Among survivors, uric acid and hsCRP may be valuable as prognostic biomarkers, based on two and one studies, respectively, with very low quality of evidence. There is conflicting evidence for the prognostic value of adiponectin and leptin (very low quality). TNF-alpha and IL-6 appear not to be independent predictors, based on one very low quality study testing both biomarkers. For IL-1, apoB, and lp(a), no studies were found.

OBESITY

3.5 | Meta-analysis of most relevant findings of enlisted biomarkers

We aimed to perform a meta-analysis of the most promising biomarkers: uric acid, adiponectin, leptin, hsCRP, and apoB. For diagnostic studies, only the AUC is suitable for meta-analysis, due to different thresholds used for sensitivity and specificity (Table S6). For predictor studies, only dichotomous and continuous (per unit or per unit logtransformed) studies are useful. Many studies use quantiles, but these are unsuited for meta-analysis: cut-offs between the quantiles depend on the range and distribution in each study population and are therefore insufficiently comparable between studies to perform a meta-analysis.

A wide variety of outcome measures was used in the studies, and many studies performed an analysis that was unsuited for meta-analysis. Also, there was variance in thresholds used for dichotomous outcomes, as well as in covariates in multivariable models. Therefore, we were unable to retain at least three sufficiently comparable studies for most biomarkers, and for most outcomes, in order to perform a meta-analysis. For a few biomarkers, enough studies were eligible for meta-analysis, because the authors also published crude outcomes, and outcomes that were only age and sex adjusted (Table S6).

We were able to perform a meta-analysis for the prognostic value of uric acid (hyperuricemia and continuous uric acid levels) and for the diagnostic value of hsCRP. We estimated the pooled OR for the association between hyperuricemia and MetS, adjusted for age and sex (four studies,¹⁰¹⁻¹⁰⁴ with threshold variability accepted of

-WILEY 9 of 22

10%,²³⁸ OR 2.94, 95%Cl 2.08–4.15); the pooled OR per unit increase in uric acid, unadjusted (three studies,^{99,105,106} OR 1.086, 95%Cl 1.066–1.106); and the pooled AUC for hsCRP, also unadjusted (three studies,^{74,87,88} AUC 0.71, 95%Cl 0.67–0.74).^{99,106} Forest plots are shown in Figure 3. Unfortunately, many studies could not be included, and the reported estimators are not adjusted for relevant covariates, in particular age and sex for some, and overweight, insulin resistance, and smoking for all.

3.6 | Other, non-enlisted biomarkers

In Table S7, 179 articles for all other biomarkers for diagnosis or prognosis of MetS are enlisted, and the main data are summarized. These included ratios of our studied biomarkers. All studies investigating leptin/adiponectin ratio as prognostic^{62,68,69,167,172,173,239} or diagnostic study^{68–70,173,239,240} showed a possible relevance. Apolipoprotein A1 (apoA1) and apoB/apoA1 ratio seem valuable in predicting the MetS (six studies with a protective effect of apoA1^{97,219–222} and eight studies with an effect of increasing risk of increasing apoB/apoA1 ratio^{92,97,221,222,241–244}). There are two studies reporting a diagnostic value of apoB/apoA1 ratio.^{92,221} Other recurrently reported, potentially useful biomarkers were Gamma GT, (non-high sensitivity) CRP, ferritin, leukocyte count, hemoglobin and urine pH, and sodium excretion.

4 | DISCUSSION

This is the first systematic literature review investigating newer biomarkers for metabolic syndrome (MetS) in CCS, with the aim to obtain the highest level of evidence by including validated tools for risk of

(A) Pooled results for odds ratio of hyperuricemia

Study	Beta	SE		Odd	s R	atio		OR	95%-CI	Weight
Cheserek 2018 Moulin 2017 Porchia 2018 Wei 2015	1.45 0.51 1.28 1.12	0.206 0.224 0.414 0.045			_			1.66 - 3.60	[2.84; 6.38] [1.07; 2.57] [1.60; 8.10] [2.82; 3.37]	25.4% 24.0% 12.3% 38.3%
Random effects model Heterogeneity: $I^2 = 71\%$, $\tau^2 = 0$	0.0784, p	= 0.02	0.2	0.5	1	2	5	2.94	[2.08; 4.15]	100.0%

(B) Pooled results for odds ratio per unit increase in uric acid

Study	Beta	SE	Odds	Ratio	OR	95%-CI	Weight
Liu 2019 Petrikova 2018 Zhang 2017	0.489 0.005 0.007	0.0188 0.0008 0.0003		÷	1.005	[1.571; 1.691] [1.004; 1.007] [1.006; 1.008]	15.8% 42.0% 42.1%
Random effects model Heterogeneity: $I^2 = 100\%$, $\tau^2 =$	= 0.0002, p	< 0.01	1	1.5	1.086 	[1.066; 1.106]	100.0%

(C) Pooled results for AUC of hsCRP

Study	AUC	SE	AUC	AUC	95%-CI Weight
Chen 2019 Kawada 2012 Stefanska 2011	0.68 0.73 0.70	0.018 0.008 0.031	*	0.73	[0.65; 0.71] 33.8% [0.72; 0.74] 45.0% [0.64; 0.76] 21.2%
Random effects model Heterogeneity: $l^2 = 72\%$, $\tau^2 = 0$.0007, p		0.6 0.7 0.8 0.9	0.71	[0.67; 0.74] 100.0%

FIGURE 3 Forest plots for different study-specific outcomes. (A) Odds ratio (OR) for hyperuricemia. (B) OR for per unit increase in uric acid. (C) Area under the curve (AUC) of hsCRP. The sizes of the square boxes on the forest plots are proportional to the total number of patients in the selected trials

bias assessment and summary of evidence, and by performing a meta-analysis.

For five biomarkers, numerous studies with moderate to high quality of evidence were found for diagnosing and predicting MetS: uric acid, adiponectin, leptin, hsCRP, and apoB. The evidence was not sufficient to confirm the value of candidate biomarkers lp(a), IL-1, IL-6, and TNF-alpha.

Meta-analysis of eligible studies showed a predictive value of uric acid for MetS, with a positive association, and a diagnostic value for hsCRP.

These findings suggest that uric acid, adiponectin, leptin, hsCRP, and apoB may be used in a screening setting for CCS, in addition to standard MetS criteria, in order to provide better diagnosis and prediction of MetS (risk). Systematic reviews in other populations have identified not only elevated leptin,²⁴⁵ uric acid,²⁴⁵⁻²⁴⁸ and low (HWM) adiponectin,^{245,249,250} but also II-6²⁴⁵ and TNF-alpha²⁴⁵ as potential MetS biomarkers.

As anticipated, the number of publications for survivors on this topic was rather limited: we identified only five studies in CCS specifically, which found a possible predictive value for hsCRP and uric acid, and conflicting or no evidence for the value of adiponectin, leptin, and TNF-alpha. Disadvantages of these survivor studies were low patient numbers and moderate to high (detection and confounding) bias risk. No studies investigated the diagnostic value of newer biomarkers. Survivor studies with information on altered biomarker values but no direct comparison between biomarker and MetS occurrence were excluded.^{60,251-260} We expected to miss many relevant studies when designing the study, if we based our conclusions only on survivor studies. Therefore, evidence in the younger general adult population without childhood cancer history was included in our search as well, leading to 175 general population studies with relevant data which were generalizable to young adult survivors.

CCS can have an increased risk to develop MetS, in particular after treatment with cranial and/or abdominal radiotherapy, intensive chemotherapy, nephrectomy, adrenalectomy, or stem cell transplantation.^{43,261–269} These therapies can lead to several underlying conditions that can increase the risk for (components of) MetS, such as hypothalamic damage, growth hormone deficiency, pancreatic beta cell dysfunction, hypogonadism, hypothyroidism, and altered body composition with increased abdominal fat.^{43,261–269}

Furthermore, it is well acknowledged that in CCS, the biological age progresses faster than their true age, as can be derived from their high level of frailty.^{4–9} Previous studies have shown that the physiologic reserve of CCS with a median age of 33 is similar to that of adults in the general population who are aged 65 years.⁶ For this reason, we included studies investigating biomarkers for MetS in the general population, with >75% of participants aged below 65 years, as may be very well applicable to CCS. We excluded studies investigating MetS biomarkers among elderly people on purpose, since they have an even higher level of frailty than CCS, comorbidities, and aging factors, which may be confounders in the association between the newer biomarker and metabolic syndrome. We considered that extrapolating conclusions from a general elderly population to CCS

could draw invalid conclusions. Based on this approach, all available literature applicable to survivors is now discussed in this review, as it includes both survivor studies as well as all generalizable data from a reasoned selection of the general population studies.

DBESITY

On the other hand, several studies excluded people with certain chronic illnesses.^{74,75,82,83,167,174–180,226} This may limit applicability of results to the population of CCS, in which the prevalence of comorbidities is high.^{3,25,256,270} This was taken into account when scoring the risk of bias. Additionally, childhood cancer (treatment)-related long-term side effects, such as altered fat distribution, sarcopenic obesity, and hormonal disbalances, may play a survivor specific role in the pathogenesis of MetS¹¹; development of future studies that apply the use of biomarkers in large cohorts of CCS is therefore important.

Due to differences in study designs and statistical analyses, a wide variety of outcome measures was used. There was also substantial diversity in follow-up time in longitudinal studies. By employing the GRADE tool for summarizing evidence, we were able to draw conclusions for each biomarker from this heterogeneity of results. The meta-analysis was based on few studies, as many studies could not be included. Also, heterogeneity was high in the meta-analysis on uric acid per unit increase, as the study of Liu et al.¹⁰⁵ had a remarkably higher OR than the other two studies.

Furthermore, although the ability of different MetS definitions to predict diabetes and CVD appears to be similar,^{271,272} the use of different definitions (Table 1) can lead to differences in occurrence of MetS. There are subtle differences between the definitions that were mostly used in the included studies (Table 1). The potential consequence of choice of definition is illustrated by studies that tested the biomarker use in diagnosing or predicting MetS according to multiple definition used.^{67,71,83,200} Therefore, comparing different studies and interpreting results of the meta-analysis requires some caution, as a full comparison of the studies is often not possible.

Adiposity, and hence the MetS, can be underdiagnosed in survivors, due to altered body composition after radiotherapy, stem cell transplantation, or amputations. For clinical applicability to survivors, it is important that newer biomarkers play an independent role in MetS, and measurement of newer biomarkers is only useful when their effect is not yet captured by established MetS components. Therefore, we did not investigate routine dyslipidemia and insulin resistance markers in our search (e.g., LDL and HOMA-IR). Although apoB and lp(a) are also lipid markers, they are of interest because they are better predictors of atherogenicity than triglycerides, HDL and LDL—particularly apoB, because it gives an estimate of the total number of circulating atherogenic particles.^{273–275}

In this light, it is also favorable that studies adjust for MetS components, such as adiposity and insulin resistance, in order to adjust for potentially major correlations and interactions^{181,276–278} and to yield the independent/additional diagnostic and predictive value of the biomarker. Furthermore, it remains important to evaluate other traditional risk factors, including smoking, physical activity, socio-economic status, and family history^{170,279} In addition, genetic profile may still be relevant for MetS risk, although so far this is not included in standard 12 of 22 WILEY OBESITY

screening.^{280–282} Risk of detection and confounding bias remains high, especially in the diagnostic studies, as many studies did not adjust for MetS components and traditional risk factors. In particular for the diagnostic studies, a risk of (detection and) confounding bias remained.

The MetS is defined as a cluster of symptoms such as obesity, hypertension, impaired glucose tolerance, and dyslipidemia.¹¹ These clustered symptoms are related to each other: an imbalance in energy intake and consumption causes a cascade of increased (visceral) adiposity, increased circulating free fatty acids and decreased adiponectin (which causes also an increase in insulin resistance), and high levels of pro-inflammatory and pro-thrombotic mediators, such as TNF-alpha, IL-1, and IL-6.^{11,34} Insulin resistance is associated with a lowered excretion of uric acid by the kidneys and higher uric acid production.^{283,284} The adipokines leptin and adiponectin are produced by adipocytes.²⁸⁵ Low leptin values trigger metabolic, behavioral, and endocrine responses that aim at a preservation of the fuel reserves of the body.²⁸⁶ Adiponectin enhances insulin sensitization and suppresses inflammation and cell death.²⁸⁶⁻²⁸⁹ Another important molecule is apoB: all atherogenic lipoproteins carry one single apoB molecule as their structural protein, and therefore, apoB represents the atherogenic burden.²⁹⁰ Serum apoB is a strong predictor of cardiovascular risks^{273,291,292} and comes in as an important player for the MetS in this review as well. One of the low density lipoproteins carrying an apoB molecule is Lp(a).²⁹³ The interpretation of Lp(a) values in an individual can be difficult due to a high heterogeneity and wide distribution of Lp(a) concentrations.²⁹⁴ Although evidence for relevance of Lp(a) for MetS evaluation in survivors was unavailable, it remains a marker of interest, since elevated Lp(a) levels were an independent predictor for cardiovascular and cerebrovascular outcomes²⁹⁵⁻³⁰³ and were inversely associated with T2DM.³⁰⁴

An important inflammatory marker is (hs)CRP, which is synthesized by hepatocytes^{305,306} in response to infection, inflammation, tissue damage, and malignant neoplasia.^{305,306} CRP binds to LDL^{305,307} and may have a causal role in atherogenesis,³⁰⁵ as it is present in atherosclerotic plaques.^{305,308} Inflammatory markers may reflect a transient state instead of chronic state of inflammation.²⁰¹ Still, in the study of Oda and Kawai,³⁰⁹ the diagnostic value of hsCRP was reproducible when the measurement was repeated after 1 year. Many studies had a high CRP^{74,166,202,203} or infection^{88,204-206,232} as exclusion criterion. Regarding inflammation, smooth muscle cells, endothelial cells, and macrophages produce cytokines such as IL-1 and IL-6³¹⁰⁻³¹² in reaction to metabolic stress,^{312,313} by other inflammatory mediators such as interferon-gamma and TNF, and cholesterol itself.³¹² Still, the evidence for the usefulness as marker for the MetS is rather limited.

Due to the systemic nature of MetS, our secondary objective to reveal other interesting biomarkers yielded many markers. Interesting markers for further research include Gamma GT, ferritin, leukocytes, and hemoglobin. In several studies, biomarkers were related to each other, as MetS components are related as well.³¹⁴ In one study, leptin was inversely associated with uric acid excretion³¹⁴; in another, a synergistic effect between hsCRP and high molecular weight adiponectin

was found.³¹⁵ Also, ratios of biomarkers (e.g., leptin/adiponectin, apoB/apoA1) include extra information and may be better diagnostic or prognostic agents than single biomarkers. Future studies may investigate the value of combining biomarkers.

Some limitations are present in this systematic literature.

Many of the included studies had a cross-sectional design, which is suboptimal to investigate causality; this was taken into account for the GRADE and level of evidence. Some authors conducted prospective longitudinal studies^{89,107,171,201} and associated MetS risk at end of follow-up with baseline and/or change in biomarker level. Study designs even more suitable for determining prediction and causality include prediction models and Mendelian randomization.^{108,316-318} These study designs require more time and financial resources and large cohorts. These types of studies were either not performed or unsuitable for our research question.

Many studies were performed among Asian cohorts. Asian people are more susceptible to insulin resistance,^{319,320} which is accounted for in lower waist circumference thresholds. Additionally, there may be an ethnicity specific component in the relationship between biomarker and MetS.³²¹⁻³²⁸ This may limit the applicability to a Caucasian population.

For this literature study, we focused on diagnosis and prediction of the full MetS; other outcomes such as resolution of the MetS,³²⁹ components of the MetS, CVD, or T2DM were out of scope.^{302,330-341} Therefore, our findings do not provide a complete overview of the use of the newer biomarkers in diagnosing and predicting cardiovascular risk factors in CCS.

We have two suggestions for future research that are relevant for the implementations of our findings in the follow-up of CCS. The newer biomarkers could be added as a sixth criterion for MetS. This application can be especially of value in cases of doubt of MetS diagnosis for individuals who had abdominal irradiation: it may be valuable to replace waist circumference with the adipokines leptin or adiponectin. This may identify MetS in more survivors and can potentially improve the predictive ability for T2DM and CVD.³³⁷

An important requirement for the applicability of these newer biomarkers in such a screening setting for MetS (risk) in CCS is the determination of a threshold. For uric acid, this is relatively well established (Table S5); for other biomarkers, this is less clear, as is illustrated by the range of applied thresholds (Table S5). This is partly because of the use of different assays and testing of subfractions of a biomarker, such as high molecular weight adiponectin. Also, a tradeoff between sensitivity and specificity may influence the determination of an optimal threshold.

In conclusion, based on this systematic literature search, we suggest to consider the additional use of uric acid, adiponectin, hsCRP, leptin, and apoB in the screening setting for metabolic syndrome in CCS. As our conclusions are largely based on general population studies, studies in CCS are needed. Furthermore, future studies may specifically test the use of newer biomarkers as additional MetS components and define optimal thresholds. The addition of one or more of these newer biomarkers as a criterion for MetS may lead to a newer and better classification and enhanced identification of risk of developing T2DM and CVD, especially in CCS in whom components are difficult to evaluate in the currently applied definitions. Early intervention can delay or prevent complications and hence improve very long-term survival outcomes and quality of life.

FUNDING INFORMATION

This work was supported by the Erasmus Medisch Centrum, Rotterdam, the Netherlands, and Princess Máxima Center for Pediatric Oncology, Utrecht, the Netherlands.

CONFLICT OF INTEREST

The authors have nothing to disclose.

ORCID

Vincent G. Pluimakers b https://orcid.org/0000-0002-3066-3951 Selveta S. van Santen b https://orcid.org/0000-0001-8818-6759 Marta Fiocco b https://orcid.org/0000-0001-5588-0277

Aart J. van der Lelij D https://orcid.org/0000-0002-1059-0126

Marry M. van den Heuvel-Eibrink D https://orcid.org/0000-0002-7760-879X

Sebastian J. C. M. M. Neggers D https://orcid.org/0000-0002-7698-0282

REFERENCES

- Howlader N, Noone AM, Krapcho M, et al. SEER Cancer Statistics Review (CSR) 1975–2014, National Cancer Institute: Bethesda, MD 2017.
- Gibson TM, Mostoufi-Moab S, Stratton KL, et al. Temporal patterns in the risk of chronic health conditions in survivors of childhood cancer diagnosed 1970-99: a report from the Childhood Cancer Survivor Study cohort. *Lancet Oncol.* 2018;19(12):1590-1601.
- Hudson MM, Ness KK, Gurney JG, et al. Clinical ascertainment of health outcomes among adults treated for childhood cancer. JAMA. 2013;309(22):2371-2381.
- Ness KK, Howell CR, Bjornard KL. Frailty and quality of life in adult survivors of childhood cancer. Expert Rev Qual Life Cancer Care. 2017;2(2):79-85.
- Ness KK, Armstrong GT, Kundu M, Wilson CL, Tchkonia T, Kirkland JL. Frailty in childhood cancer survivors. *Cancer*. 2015; 121(10):1540-1547.
- Ness KK, Kirkland JL, Gramatges MM, et al. Premature physiologic aging as a paradigm for understanding increased risk of adverse health across the lifespan of survivors of childhood cancer. J Clin Oncol off J am Soc Clin Oncol. 2018;36(21):2206-2215.
- 7. Ness KK, Wogksch MD. Frailty and aging in cancer survivors. *Transl Res: J Lab Clin Med.* 2020;221:65-82.
- Armenian SH, Gibson CJ, Rockne RC, Ness KK. Premature aging in young cancer survivors. J Natl Cancer Inst. 2019;111(3):226-232.
- Vatanen A, Hou M, Huang T, et al. Clinical and biological markers of premature aging after autologous SCT in childhood cancer. *Bone Marrow Transplant*. 2017;52(4):600-605.
- Bhakta N, Liu Q, Ness KK, et al. The cumulative burden of surviving childhood cancer: an initial report from the St Jude Lifetime Cohort Study (SJLIFE). *Lancet (London, England)*. 2017;390(10112): 2569-2582.
- Pluimakers VG, van Waas M, Neggers S, van den Heuvel-Eibrink MM. Metabolic syndrome as cardiovascular risk factor in childhood cancer survivors. *Crit Rev Oncol Hematol.* 2019; 133:129-141.

 Reulen RC, Winter DL, Frobisher C, et al. Long-term cause-specific mortality among survivors of childhood cancer. JAMA. 2010;304(2): 172-179.

DBESITY

- Tukenova M, Guibout C, Oberlin O, et al. Role of cancer treatment in long-term overall and cardiovascular mortality after childhood cancer. J Clin Oncol off J am Soc Clin Oncol. 2010;28(8): 1308-1315.
- Castellino SM, Geiger AM, Mertens AC, et al. Morbidity and mortality in long-term survivors of Hodgkin lymphoma: a report from the Childhood Cancer Survivor Study. *Blood*. 2011;117(6): 1806-1816.
- Prasad PK, Signorello LB, Friedman DL, Boice JD Jr, Pukkala E. Long-term non-cancer mortality in pediatric and young adult cancer survivors in Finland. *Pediatr Blood Cancer*. 2012;58(3): 421-427.
- Armstrong GT, Oeffinger KC, Chen Y, et al. Modifiable risk factors and major cardiac events among adult survivors of childhood cancer. J Clin Oncol off J am Soc Clin Oncol. 2013;31(29):3673-3680.
- Perkins SM, Fei W, Mitra N, Shinohara ET. Late causes of death in children treated for CNS malignancies. J Neurooncol. 2013;115(1): 79-85.
- Kero AE, Järvelä LS, Arola M, et al. Cardiovascular morbidity in longterm survivors of early-onset cancer: a population-based study. *Int J Cancer*. 2014;134(3):664-673.
- van Laar M, Feltbower RG, Gale CP, Bowen DT, Oliver SE, Glaser A. Cardiovascular sequelae in long-term survivors of young peoples' cancer: a linked cohort study. Br J Cancer. 2014;110(5): 1338-1341.
- Olsen M, Schmidt M, Lash TL, Sorensen K, Pedersen L, Sorensen HT. Cardiovascular disease risk in childhood cancer survivors. Am J Epidemiol. 2014;180(1):120-123.
- Kero AE, Järvelä LS, Arola M, et al. Late mortality among 5-year survivors of early onset cancer: a population-based register study. *Int J Cancer.* 2015;136(7):1655-1664.
- Gudmundsdottir T, Winther JF, de Fine LS, et al. Cardiovascular disease in Adult Life after Childhood Cancer in Scandinavia: A population-based cohort study of 32,308 one-year survivors. *Int J Cancer*. 2015;137(5):1176-1186.
- Bhakta N, Liu Q, Yeo F, et al. Cumulative burden of cardiovascular morbidity in paediatric, adolescent, and young adult survivors of Hodgkin's lymphoma: an analysis from the St Jude Lifetime Cohort Study. *Lancet Oncol.* 2016;17(9):1325-1334.
- Schindler M, Spycher BD, Ammann RA, Ansari M, Michel G, Kuehni CE. Cause-specific long-term mortality in survivors of childhood cancer in Switzerland: A population-based study. *Int J Cancer*. 2016;139(2):322-333.
- Kero AE, Madanat-Harjuoja LM, Järvelä LS, Malila N, Matomäki J, Lähteenmäki PM. Cardiovascular medication after cancer at a young age in Finland: A nationwide registry linkage study. *Int J Cancer*. 2016;139(3):683-690.
- Armstrong GT, Liu Q, Yasui Y, et al. Late mortality among 5-year survivors of childhood cancer: a summary from the Childhood Cancer Survivor Study. J Clin Oncol off J am Soc Clin Oncol. 2009;27(14): 2328-2338.
- 27. Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. *Diabetes*. 1988;37(12):1595-1607.
- 28. Wittcopp C, Conroy R. Metabolic syndrome in children and adolescents. *Pediatr Rev.* 2016;37(5):193-202.
- Kaplan NM. The deadly quartet. Upper-body obesity, glucose intolerance, hypertriglyceridemia, and hypertension. *Arch Intern Med.* 1989;149(7):1514-1520.
- Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. *Eur J Endocrinol*. 2017;178:11-22.

14 of 22 WILEY-Reviews

- Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabet Med: A Journal of the British Diabetic Association. 1998; 15(7):539-553.
- Balkau B, Charles MA. Comment on the provisional report from the WHO consultation. European Group for the Study of Insulin Resistance (EGIR). *Diabet Med: A Journal of the British Diabetic Association*. 1999;16(5):442-443.
- Expert Panel on Detection, Evaluation, And Treatment of High Blood Cholesterol In Adults. Executive summary of the third report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, And Treatment of High Blood Cholesterol In Adults (Adult Treatment Panel III). JAMA. 2001;285(19): 2486-2497.
- Grundy SM, Cleeman JI, Daniels SR, et al. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. *Circulation*. 2005;112(17):2735-2752.
- Alberti KG, Zimmet P, Shaw J. Metabolic syndrome—a new world-wide definition. A Consensus Statement from the International Diabetes Federation. Diabet Med: A Journal of the British Diabetic Association. 2006;23(5):469-480.
- 36. Alberti KG, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation*. 2009; 120(16):1640-1645.
- Oda E. Metabolic syndrome: its history, mechanisms, and limitations. Acta Diabetol. 2012;49(2):89-95.
- Ju SY, Lee JY, Kim DH, et al. Association of metabolic syndrome and its components with all-cause and cardiovascular mortality in the elderly. *Medicine*. 2017;96(45):e8491.
- Alberti G, Zimmet P, Shaw JGS. The IDF consensus worldwide definition of the metabolic syndrome. International Diabetes Federation: 2006.
- Wilson PW, D'Agostino RB, Parise H, Sullivan L, Meigs JB. Metabolic syndrome as a precursor of cardiovascular disease and type 2 diabetes mellitus. *Circulation*. 2005;112(20):3066-3072.
- Ford ES. The metabolic syndrome and mortality from cardiovascular disease and all-causes: findings from the National Health and Nutrition Examination Survey II Mortality Study. *Atherosclerosis*. 2004;173(2):309-314.
- Blijdorp K, van den Heuvel-Eibrink MM, Pieters R, et al. Obesity is underestimated using body mass index and waist-hip ratio in longterm adult survivors of childhood cancer. *PLoS ONE*. 2012;7(8): e43269.
- 43. van Waas M, Neggers SJ, Raat H, van Rij CM, Pieters R, van den Heuvel-Eibrink MM. Abdominal radiotherapy: a major determinant of metabolic syndrome in nephroblastoma and neuroblastoma survivors. PLoS ONE. 2012;7(12):e52237.
- 44. van Waas M, Neggers SJ, Uitterlinden AG, et al. Treatment factors rather than genetic variation determine metabolic syndrome in childhood cancer survivors. *Eur J Cancer (Oxford, England: 1990).* 2013; 49(3):668-675.
- Ward LC. Bioelectrical impedance analysis for body composition assessment: reflections on accuracy, clinical utility, and standardisation. Eur J Clin Nutr. 2019;73(2):194-199.
- Horber FF, Thomi F, Casez JP, Fonteille J, Jaeger P. Impact of hydration status on body composition as measured by dual energy X-ray absorptiometry in normal volunteers and patients on haemodialysis. *Br J Radiol.* 1992;65(778):895-900.

- O'Brien C, Young AJ, Sawka MN. Bioelectrical impedance to estimate changes in hydration status. *Int J Sports Med.* 2002;23(5): 361-366.
- van Santen SS, Olsson DS, Hammarstrand C, et al. Diagnosing metabolic syndrome in craniopharyngioma patients: body composition versus BMI. *Eur J Endocrinol.* 2019;181(2):173-183.
- Perreault L. Determining body composition in adults. In: UpToDate, Pi-Sunyer XF, UpToDate, Waltham, MA, 2020.
- Sommer I, Teufer B, Szelag M, et al. The performance of anthropometric tools to determine obesity: a systematic review and metaanalysis. *Sci Rep.* 2020;10(1):12699.
- Coffman E, Richmond-Bryant J. Multiple biomarker models for improved risk estimation of specific cardiovascular diseases related to metabolic syndrome: a cross-sectional study. *Popul Health Metr.* 2015;13(1):7.
- 52. Shea BJ, Reeves BC, Wells G, et al. AMSTAR 2: a critical appraisal tool for systematic reviews that include randomised or nonrandomised studies of healthcare interventions, or both. BMJ (Clinical Research Ed). 2017;358:j4008.
- Whiting PF, Rutjes AW, Westwood ME, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med.* 2011;155(8):529-536.
- Hayden JA, van der Windt DA, Cartwright JL, Côté P, Bombardier C. Assessing bias in studies of prognostic factors. Ann Intern Med. 2013;158(4):280-286.
- Guyatt G, Oxman AD, Akl EA, et al. GRADE guidelines: 1. Introduction-GRADE evidence profiles and summary of findings tables. *J Clin Epidemiol.* 2011;64(4):383-394.
- DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials. 1986;7(3):177-188.
- 57. R Core Team. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria; 2020. https://www.R-project.org/
- Bielorai B, Weintraub Y, Hutt D, et al. The metabolic syndrome and its components in pediatric survivors of allogeneic hematopoietic stem cell transplantation. *Clin Transplant*. 2017;31(3).
- Cheung YT, Edelmann MN, Mulrooney DA, et al. Uric Acid and Neurocognitive Function in Survivors of Childhood Acute Lymphoblastic Leukemia Treated with Chemotherapy Only. *Cancer Epidemiol Biomarkers Prev.* 2016;25(8):1259-1267.
- Kojima C, Kubota M, Nagai A, Adachi S, Watanabe K, Nakahata T. Adipocytokines in childhood cancer survivors and correlation with metabolic syndrome components. *Pediatr Int.* 2013;55(4): 438-442.
- Ariffin H, Azanan MS, Abd Ghafar SS, et al. Young adult survivors of childhood acute lymphoblastic leukemia show evidence of chronic inflammation and cellular aging. *Cancer*. 2017;123(21):4207-4214.
- Barbosa-Cortes L, Lopez-Alarcon M, Mejia-Arangure JM, et al. Adipokines, insulin resistance, and adiposity as a predictors of metabolic syndrome in child survivors of lymphoma and acute lymphoblastic leukemia of a developing country. *BMC Cancer*. 2017; 17(1):125.
- Eglit T, Lember M, Ringmets I, Rajasalu T. Gender differences in serum high-molecular-weight adiponectin levels in metabolic syndrome. *Eur J Endocrinol.* 2013;168(3):385-391.
- González-Jiménez E, Schmidt-Riovalle J, Sinausía L, Carmen Valenza M, Perona JS. Predictive value of ceruloplasmin for metabolic syndrome in adolescents. *BioFactors (Oxford, England)*. 2016; 42:163-170.
- 65. Henneman P, Janssens AC, Zillikens MC, et al. Menopause impacts the relation of plasma adiponectin levels with the metabolic syndrome. *J Intern Med*. 2010;267(4):402-409.
- 66. Krzystek-Korpacka M, Patryn E, Kustrzeba-Wojcicka I, Chrzanowska J, Gamian A, Noczynska A. Gender-specific association

of serum uric acid with metabolic syndrome and its components in juvenile obesity. *Clin Chem Lab Med*. 2011;49(1):129-136.

- 67. Ntzouvani A, Fragopoulou E, Panagiotakos D, Pitsavos C, Antonopoulou S. Reduced circulating adiponectin levels are associated with the metabolic syndrome independently of obesity, lipid indices and serum insulin levels: a cross-sectional study. *Lipids Health Dis.* 2016;15(1):140.
- Stefanska A, Ponikowska I, Cwiklinska-Jurkowska M, Sypniewska G. Association of FSH with metabolic syndrome in postmenopausal women: a comparison with CRP, adiponectin and leptin. *Biomark Med.* 2014;8(7):921-930.
- Li G, Xu L, Zhao Y, et al. Leptin-adiponectin imbalance as a marker of metabolic syndrome among Chinese children and adolescents: The BCAMS study. *PLoS ONE*. 2017;12(10):e0186222.
- Adejumo EN, Adejumo OA, Azenabor A, et al. Leptin: adiponectin ratio discriminated the risk of metabolic syndrome better than adiponectin and leptin in Southwest Nigeria. *Diabetes Metab Syndr*. 2019;13(3):1845-1849.
- Esteghamati A, Zandieh A, Zandieh B, et al. Leptin cut-off values for determination of metabolic syndrome: third national surveillance of risk factors of non-communicable diseases in Iran (SuRFNCD-2007). *Endocrine*. 2011;40(1):117-123.
- Li WC, Hsiao KY, Chen IC, Chang YC, Wang SH, Wu KH. Serum leptin is associated with cardiometabolic risk and predicts metabolic syndrome in Taiwanese adults. *Cardiovasc Diabetol*. 2011;10(1):36.
- Obeidat AA, Ahmad MN, Haddad FH, Azzeh FS. Leptin and uric acid as predictors of metabolic syndrome in jordanian adults. *Nutr Res Pract*. 2016;10(4):411-417.
- Kawada T. Relationship between several markers and presence of metabolic syndrome or components of the metabolic syndrome in Japanese workers. J Occup Environ Med. 2012;54(8):984-988.
- 75. Jeong J, Suh YJ. Association between serum uric acid and metabolic syndrome in Koreans. *J Korean Med Sci.* 2019;34(48):e307.
- Chiou WK, Wang MH, Huang DH, Chiu HT, Lee YJ, Lin JD. The relationship between serum uric acid level and metabolic syndrome: differences by sex and age in Taiwanese. J Epidemiol. 2010;20(3): 219-224.
- 77. Chen YY, Kao TW, Yang HF, et al. The association of uric acid with the risk of metabolic syndrome, arterial hypertension or diabetes in young subjects—an observational study. *Clin Chim Acta; Int J Clin Chem.* 2018;478:68-73.
- Lee JG, Park SW, Cho BM, et al. Serum amylase and risk of the metabolic syndrome in Korean adults. *Clin Chim Acta; Int J Clin Chem.* 2011;412(19-20):1848-1853.
- Lee YJ, Cho S, Kim SR. A possible role of serum uric acid as a marker of metabolic syndrome. *Intern Med J.* 2014;44(12a): 1210-1216.
- Sun HL, Pei D, Lue KH, Chen YL. Uric acid levels can predict metabolic syndrome and hypertension in adolescents: a 10-year longitudinal study. *PLoS ONE*. 2015;10(11):e0143786.
- Kim JY, Ahn SV, Yoon JH, et al. Prospective study of serum adiponectin and incident metabolic syndrome: the ARIRANG study. *Diabetes Care*. 2013;36(6):1547-1553.
- de Abreu VG, Martins CJM, de Oliveira PAC, Francischetti EA. Highmolecular weight adiponectin/HOMA-IR ratio as a biomarker of metabolic syndrome in urban multiethnic Brazilian subjects. *PLoS ONE*. 2017;12(7):e0180947.
- Ding Y, Li S, Ma RL, et al. Association of homeostasis model assessment of insulin resistance, adiponectin, and low-grade inflammation with the course of the metabolic syndrome. *Clin Biochem.* 2015; 48(7-8):503-507.
- Fujikawa R, Ito C, Tsuboi A. Is the screening of metabolic syndrome using adiponectin possible? *Diabetol Int*. 2015;6(4):313-320.
- 85. Oh J, Kim JY, Park S, et al. The relationship between insulin-like growth factor-1 and metabolic syndrome, independent of

adiponectin. Clin Chim Acta; Int J Clin Chem. 2012;413(3-4): 506-510.

 Saisho Y, Hirose H, Roberts R, Abe T, Kawabe H, Itoh H. C-reactive protein, high-molecular-weight adiponectin and development of metabolic syndrome in the Japanese general population: a longitudinal cohort study. *PLoS ONE*. 2013;8(9):e73430.

DBESITY

- Stefanska A, Sypniewska G, Blaszkiewicz B, Ponikowska I, Cwiklinska-Jurkowska M. Comparison between C-reactive protein and adipocyte fatty acid-binding protein as a component of metabolic syndrome in middle-aged women. *Clin Biochem*. 2011;44(4):304-306.
- Chen H, Xiong C, Shao X, et al. Lymphocyte to high-density lipoprotein ratio as a new indicator of inflammation and metabolic syndrome. *Diabetes Metab Syndr Obes: Targets Ther.* 2019;12: 2117-2123.
- Oda E. High-sensitivity C-reactive protein and white blood cell count equally predict development of the metabolic syndrome in a Japanese health screening population. *Acta Diabetol.* 2013;50(4): 633-638.
- Jeong H, Baek SY, Kim SW, et al. C reactive protein level as a marker for dyslipidaemia, diabetes and metabolic syndrome: results from the Korea National Health and Nutrition Examination Survey. *BMJ Open.* 2019;9(8):e029861.
- Oda E, Kawai R. Comparison between high-sensitivity C-reactive protein (hs-CRP) and white blood cell count (WBC) as an inflammatory component of metabolic syndrome in Japanese. *Intern Med* (*Tokyo, Japan*). 2010;49(2):117-124.
- 92. Jing F, Mao Y, Guo J, et al. The value of Apolipoprotein B/Apolipoprotein A1 ratio for metabolic syndrome diagnosis in a Chinese population: a cross-sectional study. *Lipids Health Dis.* 2014; 13(1):81.
- Bombelli M, Quarti-Trevano F, Tadic M, et al. Uric acid and risk of new-onset metabolic syndrome, impaired fasting glucose and diabetes mellitus in a general Italian population: data from the Pressioni Arteriose Monitorate E Loro Associazioni study. J Hypertens. 2018; 36(7):1492-1498.
- Cohen E, Krause I, Fraser A, Goldberg E, Garty M. Hyperuricemia and metabolic syndrome: lessons from a large cohort from Israel. *Isr Med Assoc J.* 2012;14(11):676-680.
- Goncalves JP, Oliveira A, Severo M, Santos AC, Lopes C. Crosssectional and longitudinal associations between serum uric acid and metabolic syndrome. *Endocrine*. 2012;41(3):450-457.
- Kanagasabai T, Alkhalaqi K, Churilla JR, Ardern CI. The association between metabolic syndrome and serum concentrations of micronutrients, inflammation, and oxidative stress outside of the clinical reference ranges: a cross-sectional study. *Metab Syndr Relat Disord*. 2019;17(1):29-36.
- Luptáková L, Siváková D, Cvíčelová M, et al. Power of biomarkers and their relative contributions to metabolic syndrome in Slovak adult women. Ann Hum Biol. 2013;40(2):132-138.
- Onat A, Can G, Cakr H, et al. Sex-specific predictors of metabolic syndrome independent of its components. J Investig Med: The Official Publication of the American Federation for Clinical Research. 2015; 63(6):796-801.
- Petrikova J, Janicko M, Fedacko J, et al. Serum Uric Acid in Roma and Non-Roma-Its Correlation with Metabolic Syndrome and Other Variables. *Int J Environ Res Public Health*. 2018;15(7).
- Sun D, Li S, Zhang X, et al. Uric acid is associated with metabolic syndrome in children and adults in a community: the Bogalusa Heart Study. *PLoS ONE*. 2014;9(10):e89696.
- Cheserek MJ, Shi Y, Le G. Association of hyperuricemia with metabolic syndrome among university workers: sex and occupational differences. *Afr Health Sci.* 2018;18(4):842-851.
- 102. Moulin SR, Baldo MP, Souza JB, et al. Distribution of Serum Uric Acid in Black Africans and Its Association With Cardiovascular Risk Factors. J Clin Hypertens (Greenwich). 2017;19(1):45-50.

16 of 22 WILEY Reviews

- 103. Wei CY, Sun CC, Wei JC, et al. Association between hyperuricemia and metabolic syndrome: an epidemiological study of a labor force population in Taiwan. *Biomed Res Int.* 2015;2015: 369179.
- Porchia LM, Gonzalez-Mejia ME, Torres-Rasgado E, Ruiz-Vivanco G, Perez-Fuentes R. Low serum uric acid concentration augments insulin effects on the prevalence of metabolic syndrome. *Diabetes Metab Syndr.* 2018;12(3):325-331.
- 105. Liu CW, Chen KH, Tseng CK, Chang WC, Wu YW, Hwang JJ. The dose-response effects of uric acid on the prevalence of metabolic syndrome and electrocardiographic left ventricular hypertrophy in healthy individuals. *Nutr Metab Cardiovasc Dis: NMCD.* 2019;29(1): 30-38.
- Zhang Y, Fu J, Yang S, et al. Prevalence of metabolically obese but normal weight (MONW) and metabolically healthy but obese (MHO) in Chinese Beijing urban subjects. *Biosci Trends.* 2017;11(4): 418-426.
- Oda E. Serum uric acid is an independent predictor of metabolic syndrome in a Japanese health screening population. *Heart Vessels*. 2014;29(4):496-503.
- Wang L, Zhang T, Liu Y, Tang F, Xue F. Association of serum uric acid with metabolic syndrome and its components: a mendelian randomization analysis. *Biomed Res Int.* 2020;2020: 6238693.
- Abbasian M, Ebrahimi H, Delvarianzadeh M, Norouzi P, Fazli M. Association between serum uric acid (SUA) levels and metabolic syndrome (MetS) components in personnel of Shahroud University of Medical Sciences. *Diabetes Metab Syndr*. 2016;10(3): 132-136.
- 110. Ahmadnezhad M, Arefhosseini SR, Parizadeh MR, et al. Association between serum uric acid, high sensitive C-reactive protein and prooxidant-antioxidant balance in patients with metabolic syndrome. *BioFactors (Oxford, England).* 2018;44(3):263-271.
- 111. Ali N, Miah R, Hasan M, et al. Association between serum uric acid and metabolic syndrome: a cross-sectional study in Bangladeshi adults. *Sci Rep.* 2020;10(1):7841.
- 112. Baygi F, Herttua K, Sheidaei A, Ahmadvand A, Jensen OC. Association of serum uric acid with cardio-metabolic risk factors and metabolic syndrome in seafarers working on tankers. *BMC Public Health*. 2020;20(1):442.
- 113. Chen D, Zhang H, Gao Y, et al. Cross-sectional and longitudinal associations between serum uric acid and metabolic syndrome: results from Fangchenggang Area Male Health and Examination Survey in China. *Clin Chim Acta; Int J Clin Chem.* 2015;446:226-230.
- Cho MH, Kim YM, Yoon JH, Kim DH, Lim JS. Serum uric acid in Korean children and adolescents: reference percentiles and association with metabolic syndrome. *Ann Pediatr Endocrinol Metab.* 2020; 25(2):104-111.
- 115. de Oliveira A, Hermsdorff HHM, Guedes Cocate P, et al. The impact of serum uric acid on the diagnostic of metabolic syndrome in apparently healthy brazilian middle-aged men. *Nutr Hosp.* 2014;30(3): 562-569.
- 116. Ferrara LA, Wang H, Umans JG, et al. Serum uric acid does not predict incident metabolic syndrome in a population with high prevalence of obesity. *Nutr Metab Cardiovasc Dis: NMCD.* 2014; 24(12):1360-1364.
- 117. Fu YQ, Yang H, Zheng JS, et al. Positive association between metabolic syndrome and serum uric acid in Wuhan. Asia Pac J Clin Nutr. 2017;26(2):343-350.
- 118. Hara S, Tsuji H, Ohmoto Y, et al. High serum uric acid level and low urine pH as predictors of metabolic syndrome: a retrospective cohort study in a Japanese urban population. *Metabolism: Clinical and Experimental.* 2012;61(2):281-288.
- 119. He SJ, Chan C, Xie ZD, et al. The relationship between serum uric acid and metabolic syndrome in premenopausal and

postmenopausal women in the Jinchang Cohort. Gynecol Endocrinol: The Official Journal of the International Society of Gynecological Endocrinology. 2017;33(2):141-144.

- Huang S, Liu X, Li H, Xu W, Jia H. Sex difference in the association of serum uric acid with metabolic syndrome and its components: a cross-sectional study in a Chinese Yi population. *Postgrad Med.* 2017;129(8):828-833.
- 121. Huang LL, Dou DM, Liu N, et al. Association of erythrocyte parameters with metabolic syndrome in the Pearl River Delta region of China: a cross sectional study. *BMJ Open*. 2018;8(1):e019792.
- 122. Jaipakdee J, Jiamjarasrangsri W, Lohsoonthorn V, Lertmaharit S. Prevalence of metabolic syndrome and its association with serum uric acid levels in Bangkok Thailand. Southeast Asian J Trop Med Public Health. 2013;44(3):512-522.
- 123. Kawada T, Otsuka T, Inagaki H, et al. Association of smoking status, insulin resistance, body mass index, and metabolic syndrome in workers: A 1-year follow-up study. *Obes Res Clin Pract*. 2010;4(3): e163-e246.
- 124. Kawada T, Otsuka T. Factor structure of indices of the second derivative of the finger photoplethysmogram with metabolic components and other cardiovascular risk indicators. *Diabetes Metab J.* 2013; 37(1):40-45.
- 125. Kawada T. Insulin-related biomarkers to predict the risk of metabolic syndrome. *Int J Diabetes Endocrinol Metab.* 2013;11:e10418.
- 126. Kawada T, Otsuka T, Inagaki H, Wakayama Y, Katsumata M. Biological markers, lifestyles and metabolic syndrome in workers. *Diabetes Metab Syndr.* 2015;9(2):71-73.
- 127. Lan Y, Mai Z, Zhou S, et al. Prevalence of metabolic syndrome in China: An up-dated cross-sectional study. *PLoS ONE*. 2018;13(4): e0196012.
- 128. Lee HJ, Park HT, Cho GJ, et al. Relationship between uric acid and metabolic syndrome according to menopausal status. *Gynecol Endocrinol: The Official Journal of the International Society of Gynecological Endocrinology*. 2011;27(6):406-411.
- 129. Lee JM, Kim HC, Cho HM, Oh SM, Choi DP, Suh I. Association between serum uric acid level and metabolic syndrome. *J Prev Med Public Health*. 2012;45(3):181-187.
- Lee JK, Ryoo JH, Choi JM, Park SK. Serum uric acid level and the incidence of metabolic syndrome in middle-aged Korean men: a 5-year follow-up study. J Prev Med Public Health. 2014;47(6): 317-326.
- 131. Lee YB, Jun JE, Lee SE, et al. Utility of serum albumin for predicting incident metabolic syndrome according to hyperuricemia. *Diabetes Metab J.* 2018;42(6):529-537.
- 132. Li Y, Chen S, Shao X, et al. Association of uric acid with metabolic syndrome in men, premenopausal women and postmenopausal women. *Int J Environ Res Public Health*. 2014;11(3): 2899-2910.
- 133. Lim JH, Kim YK, Kim YS, Na SH, Rhee MY, Lee MM. Relationship between serum uric Acid levels, metabolic syndrome, and arterial stiffness in korean. *Korean Circ J.* 2010;40(7):314-320.
- 134. Lin CC, Liu CS, Li CI, et al. The relation of metabolic syndrome according to five definitions to cardiovascular risk factors—a population-based study. *BMC Public Health*. 2009;9(1):484.
- 135. Lin YC, Chen JD, Lo SH, Chen PC. Worksite health screening programs for predicting the development of metabolic syndrome in middle-aged employees: a five-year follow-up study. *BMC Public Health*. 2010;10(1):747.
- Liu PW, Chang TY, Chen JD. Serum uric acid and metabolic syndrome in Taiwanese adults. *Metab Clin Exp.* 2010;59(6): 802-807.
- 137. Liu Y, Fan Y, Liu Q, et al. Sex-specific association of serum uric acid dynamics with the incidence of metabolic syndrome in a health check-up Chinese population: a prospective cohort study. *BMJ Open.* 2020;10(7):e035289.

- 138. Meshkani R, Zargari M, Larijani B. The relationship between uric acid and metabolic syndrome in normal glucose tolerance and normal fasting glucose subjects. *Acta Diabetol.* 2011;48(1): 79-88.
- 139. Nagahama K, Inoue T, Kohagura K, Ishihara A, Kinjo K, Ohya Y. Hyperuricemia predicts future metabolic syndrome: a 4-year followup study of a large screened cohort in Okinawa. *Japan Hypertens Res.* 2014;37(3):232-238.
- 140. Ni W, Wang R, Liu Z, et al. Association of serum uric acid with metabolic syndrome and its components: a cross-sectional study in Chinese coastal population. *Metab Syndr Relat Disord*. 2020;18(2): 103-109.
- 141. Obokata M, Negishi K, Ohyama Y, Okada H, Imai K, Kurabayashi M. A risk score with additional four independent factors to predict the incidence and recovery from metabolic syndrome: development and validation in large Japanese cohorts. *PLoS ONE*. 2015;10(7): e0133884.
- Rhee C, Kim J, Kim JY, et al. Clinical markers associated with metabolic syndrome among military aviators. *Aerosp Med Hum Perform*. 2015;86(11):970-975.
- 143. Safiri S, Qorbani M, Heshmat R, et al. Association of serum uric acid with cardiometabolic risk factors and metabolic syndrome in iranian adolescents: the CASPIAN-III study. *Iran J Kidney Dis.* 2016;10(3): 126-134.
- 144. Salehidoost R, Aminorroaya A, Zare M, Amini M. Is uric acid an indicator of metabolic syndrome in the first-degree relatives of patients with type 2 diabetes? *J Res Med Sci.* 2012;17(11): 1005-1010.
- 145. Schmitt AC, Cardoso MR, Lopes H, et al. Prevalence of metabolic syndrome and associated factors in women aged 35 to 65 years who were enrolled in a family health program in Brazil. *Menopause* (*New York, NY*). 2013;20(4):470-476.
- 146. Song YM, Lee K. Genetic and environmental influences on the associations between uric acid levels and metabolic syndrome over time. *Metab Syndr Relat Disord*. 2018;16(6):299-304.
- 147. Sumiyoshi H, Ohyama Y, Imai K, Kurabayashi M, Saito Y, Nakamura T. Association of uric acid with incident metabolic syndrome in a Japanese general population. *Int Heart J.* 2019;60(4): 830-835.
- 148. Tani S, Matsuo R, Imatake K, et al. The serum uric acid level in females may be a better indicator of metabolic syndrome and its components than in males in a Japanese population. *J Cardiol*. 2020; 76(1):100-108.
- Tao LX, Li X, Zhu HP, et al. Association of hematological parameters with metabolic syndrome in Beijing adult population: a longitudinal study. *Endocrine*. 2014;46(3):485-495.
- Tsai TY, Cheng JF, Lai YM. Prevalence of metabolic syndrome and related factors in Taiwanese high-tech industry workers. *Clinics* (*Sao Paulo*). 2011;66(9):1531-1535.
- 151. Wakabayashi I. Inverse associations between serum urate and glycemic status in a general population and in persons with diabetes mellitus. *Diabetol Metab Syndr.* 2020;12(1):21.
- 152. Wang JY, Chen YL, Hsu CH, Tang SH, Wu CZ, Pei D. Predictive value of serum uric acid levels for the diagnosis of metabolic syndrome in adolescents. *J Pediatr.* 2012;161(4): 753-6.e2.
- 153. Yadav D, Lee ES, Kim HM, et al. Prospective study of serum uric acid levels and incident metabolic syndrome in a Korean rural cohort. *Atherosclerosis*. 2015;241(1):271-277.
- Yang T, Chu CH, Bai CH, et al. Uric acid level as a risk marker for metabolic syndrome: a Chinese cohort study. *Atherosclerosis*. 2012; 220(2):525-531.
- 155. Yu TY, Jee JH, Bae JC, et al. Serum uric acid: a strong and independent predictor of metabolic syndrome after adjusting for body composition. *Metab Clin Exp.* 2016;65(4):432-440.

- 156. Yu TY, Jin SM, Jee JH, Bae JC, Lee MK, Kim JH. The protective effects of increasing serum uric acid level on development of metabolic syndrome. *Diabetes Metab J.* 2019;43(4): 504-520.
- 157. You L, Liu A, Wuyun G, Wu H, Wang P. Prevalence of hyperuricemia and the relationship between serum uric acid and metabolic syndrome in the Asian Mongolian area. J Atheroscler Thromb. 2014; 21(4):355-365.
- Zhang Q, Lou S, Meng Z, Ren X. Gender and age impacts on the correlations between hyperuricemia and metabolic syndrome in Chinese. *Clin Rheumatol.* 2011;30(6):777-787.
- 159. Zhang Z, Bian L, Choi Y. Serum uric acid: a marker of metabolic syndrome and subclinical atherosclerosis in Korean men. *Angiology*. 2012;63(6):420-428.
- 160. Zhang Q, Zhang C, Song X, et al. A longitudinal cohort based association study between uric acid level and metabolic syndrome in Chinese Han urban male population. *BMC Public Health.* 2012; 12(1):419.
- 161. Zhang ML, Gao YX, Wang X, Chang H, Huang GW. Serum uric acid and appropriate cutoff value for prediction of metabolic syndrome among Chinese adults. *J Clin Biochem Nutr.* 2013;52(1):38-42.
- 162. Zhang H, Li Y, Mao Z, et al. Sex-specific associations of serum uric acid with metabolic syndrome in Chinese rural population: the RuralDiab study. *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 2018;480:119-125.
- Zhao Y, Yu Y, Li H, et al. The association between metabolic syndrome and biochemical markers in Beijing adolescents. *Int J Environ Res Public Health*. 2019;16(22).
- 164. Ziaee A, Esmailzadehha N, Ghorbani A, Asefzadeh S. Association between uric acid and metabolic syndrome in Qazvin Metabolic Diseases Study (QMDS), Iran. *Global J Health Sci.* 2012;5(1):155-165.
- Lin KP. The relationship between serum uric acid concentration and metabolic syndrome in university freshmen. J Nurs Res. 2009;17(4): 286-292.
- 166. Ahonen TM, Saltevo JT, Kautiainen HJ, Kumpusalo EA, Vanhala MJ. The association of adiponectin and low-grade inflammation with the course of metabolic syndrome. *Nutr Metab Cardiovasc Dis: NMCD*. 2012;22(3):285-291.
- 167. Delitala AP, Scuteri A, Fiorillo E, Lakatta EG, Schlessinger D, Cucca F. Role of adipokines in the association between thyroid hormone and components of the metabolic syndrome. J Clin Med. 2019;8(6).
- 168. Fernandez-Berges D, Consuegra-Sanchez L, Penafiel J, et al. Metabolic and inflammatory profiles of biomarkers in obesity, metabolic syndrome, and diabetes in a Mediterranean population. DARIOS Inflammatory study. *Rev Esp Cardiol (Engl Ed)*. 2014;67(8): 624-631.
- 169. Juonala M, Saarikoski LA, Viikari JS, et al. A longitudinal analysis on associations of adiponectin levels with metabolic syndrome and carotid artery intima-media thickness. The Cardiovascular Risk in Young Finns Study. Atherosclerosis. 2011;217(1):234-239.
- 170. Khan RJ, Gebreab SY, Sims M, Riestra P, Xu R, Davis SK. Prevalence, associated factors and heritabilities of metabolic syndrome and its individual components in African Americans: the Jackson Heart Study. *BMJ Open*. 2015;5(10):e008675.
- 171. Lindberg S, Jensen JS, Bjerre M, et al. Low adiponectin levels at baseline and decreasing adiponectin levels over 10 years of followup predict risk of the metabolic syndrome. *Diabetes Metab.* 2017; 43(2):134-139.
- 172. Chearskul S, Homsanit M, Chearskul S, et al. Certain hormonal markers in urban Thai adults with metabolic syndrome. *J Med Assoc Thai* = Chotmaihet Thangphaet. 2014;97(1):77-84.
- 173. Yun JE, Won S, Mok Y, Cui W, Kimm H, Jee SH. Association of the leptin to high-molecular-weight adiponectin ratio with metabolic syndrome. *Endocr J.* 2011;58(9):807-815.

18 of 22 WILEY-Reviews

- 174. Baudrand R, Campino C, Carvajal CA, et al. High sodium intake is associated with increased glucocorticoid production, insulin resistance and metabolic syndrome. *Clin Endocrinol (Oxf)*. 2014;80(5): 677-684.
- 175. de Oliveira A, Hermsdorff HH, Cocate PG, Santos EC, Bressan J, Natali AJ. Accuracy of plasma interleukin-18 and adiponectin concentrations in predicting metabolic syndrome and cardiometabolic disease risk in middle-age Brazilian men. *Appl Physiol Nutr Metab = Physiologie Appliquee, Nutrition et Metabolisme.* 2015; 40(10):1048-1055.
- 176. Hirose H, Takayama T, Hozawa S, Hibi T, Saito I. Prediction of metabolic syndrome using artificial neural network system based on clinical data including insulin resistance index and serum adiponectin. *Comput Biol Med.* 2011;41(11):1051-1056.
- Ju H, Zhou Z, Sun M, Chen H. Association of fetuin-A to adiponectin ratio with metabolic syndrome: a cross-sectional study. *Endocrine*. 2017;58(1):190-193.
- 178. Kawada T, Hasegawa M. Predictive ability of serum high-molecularweight adiponectin in combination with serum insulin and serum C-reactive protein for the presence of metabolic syndrome. Ann Hum Biol. 2012;39(2):108-112.
- 179. Abu-Farha M, Behbehani K, Elkum N. Comprehensive analysis of circulating adipokines and hsCRP association with cardiovascular disease risk factors and metabolic syndrome in Arabs. *Cardiovasc Diabetol.* 2014;13(1):76.
- Cheng KH, Huang SP, Huang CN, et al. The impact of estradiol and 1,25(OH)2D3 on metabolic syndrome in middle-aged Taiwanese males. *PLoS ONE*. 2013;8(3):e60295.
- Huffman FG, Knight-Sepulveda K, McLean M, Vaccaro JA, Zarini GG. Serum adiponectin and ghrelin, metabolic syndrome and diabetes status in Cuban Americans. *Int J Health Res.* 2010;3: 93-103.
- 182. Ahn SV, Jung DH, Yadav D, Kim JY, Koh SB. Relative contribution of obesity and menopause to the association between serum adiponectin and incident metabolic syndrome. *Menopause* (*New York*, NY). 2018;25(2):154-159.
- Ayina CNA, Endomba FTA, Mandengue SH, et al. Association of the leptin-to-adiponectin ratio with metabolic syndrome in a sub-Saharan African population. *Diabetol Metab Syndr.* 2017;9(1):66.
- Ben Ali S, Jemaa R, Ftouhi B, et al. Adiponectin and metabolic syndrome in a Tunisian population. *Inflammation*. 2012;35(3): 828-833.
- 185. Cho SA, Joo HJ, Cho JY, et al. Visceral Fat Area and Serum Adiponectin Level Predict the Development of Metabolic Syndrome in a Community-Based Asymptomatic Population. *PLoS ONE*. 2017; 12(1):e0169289.
- 186. Jean-Luc Gradidge P, Norris SA, Jaff NG, Crowther NJ. Metabolic and Body Composition Risk Factors Associated with Metabolic Syndrome in a Cohort of Women with a High Prevalence of Cardiometabolic Disease. *PLoS ONE*. 2016;11(9):e0162247.
- 187. Hata A, Yonemoto K, Shikama Y, et al. Cut-off value of total adiponectin for managing risk of developing metabolic syndrome in male Japanese workers. *PLoS ONE*. 2015;10(2): e0118373.
- Huh JH, Yoon TW, Kang DR, Kim JY. Prospective Study of Sex-Specific Adiponectin Changes and Incident Metabolic Syndrome: The ARIRANG Study. J Clin Med. 2019;8(5).
- 189. Klünder-Klünder M, Flores-Huerta S, García-Macedo R, Peralta-Romero J, Cruz M. Adiponectin in eutrophic and obese children as a biomarker to predict metabolic syndrome and each of its components. BMC Public Health. 2013;13(1):88.
- 190. Koh SB, Park JK, Yoon JH, et al. Preliminary report: a serious link between adiponectin levels and metabolic syndrome in a Korean nondiabetic population. *Metab Clin Exp.* 2010;59(3):333-337.

- 191. Lee SK, Yoon DW, Kim J, et al. Association of adiponectin, ghrelin, and leptin with metabolic syndrome and its metabolic components in Sasang constitutional type. *Eur J Integr Med.* 2018;22:16-21.
- 192. Lee KW, Shin D. Prospective associations of serum adiponectin, leptin, and leptin-adiponectin ratio with incidence of metabolic syndrome: the Korean Genome and Epidemiology Study. *Int J Environ Res Public Health*. 2020;17(9):3287.
- 193. Li P, Jiang R, Li L, Liu C, Yang F, Qiu Y. Correlation of serum adiponectin and adiponectin gene polymorphism with metabolic syndrome in Chinese adolescents. *Eur J Clin Nutr.* 2015;69(1):62-67.
- Song YM, Lee K, Sung J. Adiponectin levels and longitudinal changes in metabolic syndrome: the healthy twin study. *Metab Syndr Relat Disord*. 2015;13(7):312-318.
- 195. Tanihara S, Imatoh T, Momose Y, Miyazaki M, Une H. Inverse correlation between adiponectin and the risk of metabolic syndrome in middle-aged Japanese male workers. Acta Med Okayama. 2009; 63(6):325-330.
- 196. Yu D, Yu Z, Sun Q, et al. Effects of body fat on the associations of high-molecular-weight adiponectin, leptin and soluble leptin receptor with metabolic syndrome in Chinese. *PLoS ONE*. 2011;6(2):e16818.
- 197. Mani P, Ren HY, Neeland IJ, et al. The association between HDL particle concentration and incident metabolic syndrome in the multiethnic Dallas Heart Study. *Diabetes Metab Syndr*. 2017;11(Suppl 1): S175-s79.
- 198. Mazidi M, Toth PP, Banach M. C-reactive protein is associated with prevalence of the metabolic syndrome, hypertension, and diabetes mellitus in US adults. *Angiology*. 2018;69(5):438-442.
- 199. Musani SK, Vasan RS, Bidulescu A, et al. Aldosterone, C-reactive protein, and plasma B-type natriuretic peptide are associated with the development of metabolic syndrome and longitudinal changes in metabolic syndrome components: findings from the Jackson Heart Study. *Diabetes Care*. 2013;36(10):3084-3092.
- 200. Bermúdez V, Rojas J, Salazar J, et al. The Maracaibo city metabolic syndrome prevalence study: Primary results and agreement level of 3 diagnostic criteria. *Rev Latinoam Hipertens*. 2014;9:20-32.
- Yoon K, Ryu S, Lee J, Park JD. Higher and increased concentration of hs-CRP within normal range can predict the incidence of metabolic syndrome in healthy men. *Diabetes Metab Syndr*. 2018;12(6): 977-983.
- 202. Hillman AJ, Lohsoonthorn V, Hanvivatvong O, Jiamjarasrangsi W, Lertmaharit S, Williams MA. Association of high sensitivity C-reactive protein concentrations and metabolic syndrome among Thai adults. Asian Biomed (Res Rev News). 2010;4(3):385-393.
- 203. Kim J, Pyo S, Yoon DW, et al. The co-existence of elevated high sensitivity C-reactive protein and homocysteine levels is associated with increased risk of metabolic syndrome: A 6-year follow-up study. PLoS ONE. 2018;13(10):e0206157.
- Abu-Farha M, Abubaker J, Al-Khairi I, et al. Circulating angiopoietinlike protein 8 (betatrophin) association with HsCRP and metabolic syndrome. *Cardiovasc Diabetol*. 2016;15(1):25.
- 205. Chen T, Chen H, Xiao H, et al. Comparison of the value of neutrophil to high-density lipoprotein cholesterol ratio and lymphocyte to high-density lipoprotein cholesterol ratio for predicting metabolic syndrome among a population in the Southern Coast of China. *Diabetes Metab Syndr Obes*. 2020;13:597-605.
- 206. Mirhafez SR, Pasdar A, Avan A, et al. Cytokine and growth factor profiling in patients with the metabolic syndrome. *Br J Nutr.* 2015; 113(12):1911-1919.
- 207. Chatterjee B, Mehta M, Shah T, Mahant H, Katwa V, Gosai K. Association of lipoprotein(a) and hsCRP levels with metabolic syndrome and its components. *Indian J Clin Biochem*. 2015;30(4): 394-402.
- 208. Darroudi S, Fereydouni N, Tayefi M, et al. Oxidative stress and inflammation, two features associated with a high percentage body

fat, and that may lead to diabetes mellitus and metabolic syndrome. *BioFactors (Oxford, England).* 2019;45(1):35-42.

- 209. Hong GB, Gao PC, Chen YY, et al. High-sensitivity C-reactive protein leads to increased incident metabolic syndrome in women but not in men: a five-year follow-up study in a Chinese population. *Diabetes Metab Syndr Obes: Targets and Therapy.* 2020; 13:581-590.
- Lai MM, Li CI, Kardia SL, et al. Sex difference in the association of metabolic syndrome with high sensitivity C-reactive protein in a Taiwanese population. *BMC Public Health*. 2010;10(1):429.
- 211. Mahajan A, Jaiswal A, Tabassum R, et al. Elevated levels of C-reactive protein as a risk factor for metabolic syndrome in Indians. *Atherosclerosis.* 2012;220(1):275-281.
- 212. Meng G, Zhu Q, Shao J, et al. Comparing the diagnostic ability of inflammatory markers in metabolic syndrome. *Clin Chim Acta; Int J Clin Chem.* 2017;475:1-6.
- Perez CM, Ortiz AP, Guzman M, Suarez E. Distribution and correlates of the metabolic syndrome in adults living in the San Juan Metropolitan Area of Puerto Rico. P R Health Sci J. 2012;31(3): 114-122.
- 214. Song Y, Yang SK, Kim J, Lee DC. Association between C-reactive protein and metabolic syndrome in Korean adults. *Korean J Fam Med*. 2019;40(2):116-123.
- Thompson AM, Zhang Y, Tong W, et al. Association of inflammation and endothelial dysfunction with metabolic syndrome, prediabetes and diabetes in adults from Inner Mongolia, China. BMC Endocr Disord. 2011;11(1):16.
- Yang T, Chu CH, Hsieh PC, et al. C-reactive protein concentration as a significant correlate for metabolic syndrome: a Chinese population-based study. *Endocrine*. 2013;43(2):351-359.
- 217. Ying X, Jiang Y, Qin G, et al. Association of body mass index, waist circumference, and metabolic syndrome with serum cystatin C in a Chinese population. *Medicine*. 2017;96(10):e6289.
- 218. Koskinen J, Magnussen CG, Wurtz P, et al. Apolipoprotein B, oxidized low-density lipoprotein, and LDL particle size in predicting the incidence of metabolic syndrome: the Cardiovascular Risk in Young Finns study. *Eur J Prev Cardiol*. 2012; 19(6):1296-1303.
- Onat A, Komurcu-Bayrak E, Can G, Kucukdurmaz Z, Hergenc G, Erginel-Unaltuna N. Apolipoprotein A-I positively associated with diabetes in women independently of apolipoprotein E genotype and apolipoprotein B levels. *Nutrition (Burbank, Los Angeles County, Calif)*. 2010;26(10):975-980.
- Onat A, Can G, Ornek E, Ayhan E, Erginel-Ünaltuna N, Murat SN. High serum apolipoprotein E determines hypertriglyceridemic dyslipidemias, coronary disease and apoA-I dysfunctionality. *Lipids*. 2013;48(1):51-61.
- Chou YC, Kuan JC, Bai CH, et al. Predictive value of serum apolipoprotein B/apolipoprotein A-I ratio in metabolic syndrome risk: a Chinese cohort study. *Endocrine*. 2015;49(2): 404-414.
- 222. Yang MH, Sung J, Gwak GY. The associations between apolipoprotein B, A1, and the B/A1 ratio and nonalcoholic fatty liver disease in both normal-weight and overweight Korean population. *J Clin Lipidol*. 2016;10(2):289-298.
- 223. Du R, Wu X, Peng K, et al. Serum apolipoprotein B is associated with increased risk of metabolic syndrome among middle-aged and elderly Chinese: A cross-sectional and prospective cohort study. *J Diabetes*. 2019;11(9):752-760.
- Ryoo JH, Park SK. Association of apolipoprotein B and incidence of metabolic syndrome in Korean men: a 5-years' follow-up study. *Atherosclerosis*. 2013;226(2):496-501.
- 225. Yi DW, Jeong DW, Lee SY, Son SM, Kang YH. The association between apolipoprotein A-II and metabolic syndrome in Korean

adults: a comparison study of apolipoprotein A-I and apolipoprotein B. *Diabetes Metab J.* 2012;36(1):56-63.

226. Esteghamati A, Khalilzadeh O, Anvari M, Rashidi A, Mokhtari M, Nakhjavani M. Association of serum leptin levels with homeostasis model assessment-estimated insulin resistance and metabolic syndrome: the key role of central obesity. *Metab Syndr Relat Disord*. 2009;7(5):447-452.

DBESITY

- 227. Choi JR, Kim JY, Huh JH, Kim SH, Koh SB. Contribution of obesity as an effect regulator to an association between serum leptin and incident metabolic syndrome. *Clin Chim Acta; Int J Clin Chem.* 2018; 487:275-280.
- 228. Esteghamati A, Noshad S, Khalilzadeh O, et al. Contribution of serum leptin to metabolic syndrome in obese and nonobese subjects. Arch Med Res. 2011;42(3):244-251.
- Suriyaprom K, Tungtrongchitr R, Thawnasom K. Measurement of the levels of leptin, BDNF associated with polymorphisms LEP G2548A, LEPR Gln223Arg and BDNF Val66Met in Thai with metabolic syndrome. *Diabetol Metab Syndr*. 2014;6(1):6.
- 230. Yun JE, Kimm H, Jo J, Jee SH. Serum leptin is associated with metabolic syndrome in obese and nonobese Korean populations. *Metab Clin Exp.* 2010;59(3):424-429.
- 231. Kosola J, Vaara JP, Ahotupa M, et al. Elevated concentration of oxidized LDL together with poor cardiorespiratory and abdominal muscle fitness predicts metabolic syndrome in young men. *Metab Clin Exp.* 2013;62(7):992-999.
- 232. Orsatti CL, Petri Nahas EA, Nahas-Neto J, Orsatti FL, Giorgi VI, Witkin SS. Evaluation of Toll-Like receptor 2 and 4 RNA expression and the cytokine profile in postmenopausal women with metabolic syndrome. *PLoS ONE*. 2014;9(10):e109259.
- 233. Onat A, Can G, Çoban N, et al. Lipoprotein(a) level and MIF gene variant predict incident metabolic syndrome and mortality. *J Investig Med: The Official Publication of the American Federation for Clinical Research.* 2016;64(2):392-399.
- 234. Bermudez V, Rojas J, Salazar J, et al. Variations of lipoprotein(a) levels in the metabolic syndrome: a report from the Maracaibo City Metabolic Syndrome Prevalence Study. J Diabetes Res. 2013;2013:416451.
- 235. Jain SR, Shah KH, Acharya HN, Barot K, Sharma KH. Prevalence and Predictors of Metabolic Syndrome in Young Asymptomatic Gujarati Population. Int J Chronic Dis. 2015;2015:365217.
- 236. Wu XY, Lin L, Qi HY, et al. Association between lipoprotein (a) levels and metabolic syndrome in a middle-aged and elderly Chinese cohort. *Biomed Environ Sci.* 2019;32(7):477-485.
- 237. Abdel-Moneim A, Mahmoud B, Sultan E, Mahmoud R. Association of erythrocytes indices and interleukin-1 beta with metabolic syndrome components. Univ Toronto Med J. 2020;97:6-13.
- Yu KH, Luo SF, Tsai WP, Huang YY. Intermittent elevation of serum urate and 24-hour urinary uric acid excretion. *Rheumatology* (Oxford). 2004;43(12):1541-1545.
- 239. Yoon JH, Park JK, Oh SS, et al. The ratio of serum leptin to adiponectin provides adjunctive information to the risk of metabolic syndrome beyond the homeostasis model assessment insulin resistance: the Korean Genomic Rural Cohort Study. *Clin Chim Acta; International Journal of Clinical Chemistry*. 2011;412(23-24): 2199-2205.
- Mirza S, Qu HQ, Li Q, et al. Adiponectin/leptin ratio and metabolic syndrome in a Mexican American population. *Clin Invest Med.* 2011; 34:E290.
- 241. Belfki H, Ben Ali S, Bougatef S, et al. The Apolipoprotein B/-Apolipoprotein A 1 ratio in relation to metabolic syndrome and its components in a sample of the Tunisian population. *Exp Mol Pathol*. 2011;91(2):622-625.
- 242. Jung CH, Hwang JY, Yu JH, et al. The value of apolipoprotein B/A1 ratio in the diagnosis of metabolic syndrome in a Korean population. *Clin Endocrinol (Oxf)*. 2012;77(5):699-706.

20 of 22 WILEY Reviews

- Kim SW, Jee JH, Kim HJ, et al. Non-HDL-cholesterol/HDLcholesterol is a better predictor of metabolic syndrome and insulin resistance than apolipoprotein B/apolipoprotein A1. Int J Cardiol. 2013;168(3):2678-2683.
- 244. Zhong L, Li Q, Jiang Y, et al. The ApoB/ApoA1 ratio is associated with metabolic syndrome and its components in a Chinese population. *Inflammation*. 2010;33(6):353-358.
- 245. Srikanthan K, Feyh A, Visweshwar H, Shapiro JI, Sodhi K. Systematic review of metabolic syndrome biomarkers: a panel for early detection, management, and risk stratification in the West Virginian population. *Int J Med Sci.* 2016;13(1):25-38.
- 246. Goli P, Riahi R, Daniali SS, Pourmirzaei M, Kelishadi R. Association of serum uric acid concentration with components of pediatric metabolic syndrome: a systematic review and meta-analysis. *J Res Med Sci.* 2020;25:43.
- 247. Yuan H, Yu C, Li X, et al. Serum uric acid levels and risk of metabolic syndrome: a dose-response meta-analysis of prospective studies. *J Clin Endocrinol Metab.* 2015;100(11):4198-4207.
- Liu Z, Que S, Zhou L, Zheng S. Dose-response relationship of serum uric acid with metabolic syndrome and non-alcoholic fatty liver disease incidence: a meta-analysis of prospective studies. *Sci Rep.* 2015;5(1):14325.
- 249. Liu Z, Liang S, Que S, Zhou L, Zheng S, Mardinoglu A. Meta-analysis of adiponectin as a biomarker for the detection of metabolic syndrome. *Front Physiol.* 2018;9:1238.
- Falahi E, Khalkhali Rad AH, Roosta S. What is the best biomarker for metabolic syndrome diagnosis? *Diabetes Metab Syndr.* 2015;9(4): 366-372.
- 251. Ketterl TG, Chow EJ, Leisenring WM, et al. Adipokines, inflammation, and adiposity in hematopoietic cell transplantation survivors. Blood Marrow Transplant: Journal of the American Society for Blood and Marrow Transplantation. 2018;24(3):622-626.
- 252. Tonorezos ES, Vega GL, Sklar CA, et al. Adipokines, body fatness, and insulin resistance among survivors of childhood leukemia. *Pediatr Blood Cancer.* 2012;58(1):31-36.
- 253. Argyrou C, Hatziagapiou K, Theodorakidou M, Nikola OA, Vlahopoulos S, Lambrou GI. The role of adiponectin, LEPTIN, and ghrelin in the progress and prognosis of childhood acute lymphoblastic leukemia. *Leuk Lymphoma*. 2019;60(9):2158-2169.
- Annaloro C, Airaghi L, Giannarelli D, et al. Prospective evaluation of metabolic syndrome and its features in a single-center series of hematopoietic stem cell transplantation recipients. *Ann Hematol.* 2018;97(12):2471-2478.
- Moschovi M, Trimis G, Vounatsou M, et al. Serial plasma concentrations of adiponectin, leptin, and resistin during therapy in children with acute lymphoblastic leukemia. J Pediatr Hematol Oncol. 2010; 32(1):e8-e13.
- Cepelova M, Kruseova J, Luks A, et al. Accelerated atherosclerosis, hyperlipoproteinemia and insulin resistance in long-term survivors of Hodgkin lymphoma during childhood and adolescence. *Neoplasma*. 2019;66(06):978-987.
- 257. Chow EJ, Simmons JH, Roth CL, et al. Increased cardiometabolic traits in pediatric survivors of acute lymphoblastic leukemia treated with total body irradiation. *Blood Marrow Transplant: Journal of the American Society for Blood and Marrow Transplantation.* 2010;16(12): 1674-1681.
- 258. Cooksey R, Wu SY, Klesse L, et al. Metabolic syndrome is a sequela of radiation exposure in hypothalamic obesity among survivors of childhood brain tumors. J Investig Med: The Official Publication of the American Federation for Clinical Research. 2019;67(2): 295-302.
- Frisk P, Arvidson J, Larsson M, Naessén T. Risk factors for cardiovascular disease are increased in young adults treated with stem cell transplantation during childhood. *Pediatr Transplant*. 2012;16(4): 385-391.

- Grote S, Almstedt HC, Tarleton HP. Cardiometabolic health among cancer survivors: a 13-week pilot study of a combined aerobic and resistance training program. Oncol Nurs Forum. 2016;43(3): 306-315.
- 261. Wei C, Crowne E. The impact of childhood cancer and its treatment on puberty and subsequent hypothalamic pituitary and gonadal function, in both boys and girls. *Best Pract Res Clin Endocrinol Metab.* 2019;33(3):101291.
- Pluimakers VG, van Waas M, Looman CWN, et al. Metabolic syndrome detection with biomarkers in childhood cancer survivors. *Endocr Connect*. 2020;9(7):676-686.
- Marouli E, Graff M, Medina-Gomez C, et al. Rare and low-frequency coding variants alter human adult height. *Nature*. 2017;542(7640): 186-190.
- Chemaitilly W, Cohen LE. Diagnosis of endocrine disease: endocrine late-effects of childhood cancer and its treatments. *Eur J Endocrinol*. 2017;176(4):R183-r203.
- 265. de Vathaire F, El-Fayech C, Ben Ayed FF, et al. Radiation dose to the pancreas and risk of diabetes mellitus in childhood cancer survivors: a retrospective cohort study. *Lancet Oncol.* 2012;13(10): 1002-1010.
- 266. Darzy KH, Shalet SM. Hypopituitarism following radiotherapy revisited. *Endocr Dev.* 2009;15:1-24.
- Oudin C, Simeoni MC, Sirvent N, et al. Prevalence and risk factors of the metabolic syndrome in adult survivors of childhood leukemia. *Blood*. 2011;117(17):4442-4448.
- 268. Rose SR, Horne VE, Howell J, et al. Late endocrine effects of childhood cancer. *Nat Rev Endocrinol*. 2016;12(6):319-336.
- de Haas EC, Oosting SF, Lefrandt JD, Wolffenbuttel BH, Sleijfer DT, Gietema JA. The metabolic syndrome in cancer survivors. *Lancet Oncol.* 2010;11(2):193-203.
- 270. Lipshultz SE, Landy DC, Lopez-Mitnik G, et al. Cardiovascular status of childhood cancer survivors exposed and unexposed to cardiotoxic therapy. *J Clin Oncol off J am Soc Clin Oncol.* 2012; 30(10):1050-1057.
- 271. Meigs JB, Rutter MK, Sullivan LM, Fox CS, D'Agostino RB Sr, Wilson PW. Impact of insulin resistance on risk of type 2 diabetes and cardiovascular disease in people with metabolic syndrome. *Diabetes Care.* 2007;30(5):1219-1225.
- 272. Mottillo S, Filion KB, Genest J, et al. The metabolic syndrome and cardiovascular risk a systematic review and meta-analysis. *J am Coll Cardiol.* 2010;56(14):1113-1132.
- 273. Lamarche B, Moorjani S, Lupien PJ, et al. Apolipoprotein A-I and B levels and the risk of ischemic heart disease during a five-year follow-up of men in the Québec cardiovascular study. *Circulation*. 1996;94(3):273-278.
- 274. Wilson DP, Jacobson TA, Jones PH, et al. Use of lipoprotein(a) in clinical practice: a biomarker whose time has come. A scientific statement from the National Lipid Association. *J Clin Lipidol*. 2019; 13(3):374-392.
- 275. Barter PJ, Ballantyne CM, Carmena R, et al. Apo B versus cholesterol in estimating cardiovascular risk and in guiding therapy: report of the thirty-person/ten-country panel. J Intern Med. 2006;259(3): 247-258.
- 276. Norvik JV, Storhaug HM, Ytrehus K, et al. Overweight modifies the longitudinal association between uric acid and some components of the metabolic syndrome: the Tromsø Study. *BMC Cardiovasc Disord*. 2016;16(1):85.
- 277. Dallmeier D, Larson MG, Vasan RS, et al. Metabolic syndrome and inflammatory biomarkers: a community-based cross-sectional study at the Framingham Heart Study. *Diabetol Metab Syndr*. 2012;4(1):28.
- 278. Côté M, Cartier A, Reuwer AQ, et al. Adiponectin and risk of coronary heart disease in apparently healthy men and women (from the EPIC-Norfolk Prospective Population Study). Am J Cardiol. 2011; 108(3):367-373.

- 279. Goldberg JF, Ness KK, Chi X, et al. Cardiovascular family history increases risk for late-onset adverse cardiovascular outcomes in childhood cancer survivors: a St. Jude Lifetime Cohort Report. *Cancer Epidemiol Biomarkers Prev.* 2020;30(1):123-132.
- Povel CM, Boer JM, Reiling E, Feskens EJ. Genetic variants and the metabolic syndrome: a systematic review. Obes Rev: An Official Journal of the International Association for the Study of Obesity. 2011; 12(11):952-967.
- 281. Monda KL, North KE, Hunt SC, Rao DC, Province MA, Kraja AT. The genetics of obesity and the metabolic syndrome. *Endocr Metab Immune Disord Drug Targets*. 2010;10(2):86-108.
- Fenwick PH, Jeejeebhoy K, Dhaliwal R, et al. Lifestyle genomics and the metabolic syndrome: a review of genetic variants that influence response to diet and exercise interventions. *Crit Rev Food Sci Nutr.* 2019;59(13):2028-2039.
- Al-Meshaweh AF, Jafar Y, Asem M, Akanji AO. Determinants of Blood Uric Acid Levels in a Dyslipidemic Arab Population. *Med Princ Pract.* 2012;21(3):209-216.
- Tsouli SG, Liberopoulos EN, Mikhailidis DP, Athyros VG, Elisaf MS. Elevated serum uric acid levels in metabolic syndrome: an active component or an innocent bystander? *Metabolism: Clinical and Experimental*. 2006;55(10):1293-1301.
- Zhang F, Basinski MB, Beals JM, et al. Crystal structure of the obese protein leptin-E100. *Nature*. 1997;387(6629):206-209.
- Funcke JB, Scherer PE. Beyond adiponectin and leptin: adipose tissue-derived mediators of inter-organ communication. J Lipid Res. 2019;60(10):1648-1684.
- Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem.* 1995;270(45):26746-26749.
- 288. van Andel M, Heijboer AC, Drent ML. Adiponectin and Its Isoforms in Pathophysiology. *Adv Clin Chem*. 2018;85:115-147.
- Ronsley R, Rassekh SR, Fleming A, et al. High molecular weight adiponectin levels are inversely associated with adiposity in pediatric brain tumor survivors. *Sci Rep.* 2020;10(1):18606.
- 290. Young SG. Recent progress in understanding apolipoprotein B. *Circulation*. 1990;82(5):1574-1594.
- Kastelein JJ, van der Steeg WA, Holme I, et al. Lipids, apolipoproteins, and their ratios in relation to cardiovascular events with statin treatment. *Circulation*. 2008;117(23):3002-3009.
- 292. Sniderman AD, Williams K, Contois JH, et al. A meta-analysis of low-density lipoprotein cholesterol, non-high-density lipoprotein cholesterol, and apolipoprotein B as markers of cardiovascular risk. *Circ Cardiovasc Qual Outcomes*. 2011;4(3):337-345.
- Devaraj S, Semaan JR, Jialal I. Biochemistry, Apolipoprotein B. *StatPearls*. StatPearls Publishing Copyright © 2020, StatPearls Publishing LLC.: Treasure Island (FL) 2020.
- 294. Clouet-Foraison N, Marcovina SM, Guerra E, et al. Analytical performance specifications for lipoprotein(a), apolipoprotein B-100, and apolipoprotein A-I Using the biological variation model in the EuBIVAS population. *Clin Chem.* 2020;66(5):727-736.
- 295. Danesh J, Collins R, Peto R. Lipoprotein(a) and coronary heart disease. Meta-analysis of prospective studies. *Circulation*. 2000;102 (10):1082-1085.
- 296. Kar S. Elevated lipoprotein A in South Asians and the associated risk of cardiovascular disease: a systematic review. *Curr Probl Cardiol.* 2020;46(3):100581.
- 297. Craig WY, Neveux LM, Palomaki GE, Cleveland MM, Haddow JE. Lipoprotein(a) as a risk factor for ischemic heart disease: metaanalysis of prospective studies. *Clin Chem.* 1998;44(11): 2301-2306.
- 298. Forbes CA, Quek RG, Deshpande S, et al. The relationship between Lp(a) and CVD outcomes: a systematic review. *Lipids Health Dis.* 2016;15(1):95.

 Smolders B, Lemmens R, Thijs V. Lipoprotein (a) and stroke: a meta-analysis of observational studies. *Stroke.* 2007;38(6): 1959-1966.

DBESITY

- Nave AH, Lange KS, Leonards CO, et al. Lipoprotein (a) as a risk factor for ischemic stroke: a meta-analysis. *Atherosclerosis*. 2015; 242(2):496-503.
- Wang Z, Zhai X, Xue M, Cheng W, Hu H. Prognostic value of lipoprotein (a) level in patients with coronary artery disease: a metaanalysis. *Lipids Health Dis.* 2019;18(1):150.
- Clarke R, Peden JF, Hopewell JC, et al. Genetic variants associated with Lp(a) lipoprotein level and coronary disease. N Engl J Med. 2009;361(26):2518-2528.
- 303. van Iperen EP, Sivapalaratnam S, Holmes MV, Hovingh GK, Zwinderman AH, Asselbergs FW. Genetic analysis of emerging risk factors in coronary artery disease. *Atherosclerosis*. 2016;254:35-41.
- Paige E, Masconi KL, Tsimikas S, et al. Lipoprotein(a) and incident type-2 diabetes: results from the prospective Bruneck study and a meta-analysis of published literature. *Cardiovasc Diabetol.* 2017; 16(1):38.
- 305. Kaptoge S, Di Angelantonio E, Lowe G, et al. C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: an individual participant meta-analysis. *Lancet* (London, England). 2010;375:132-140.
- 306. Pepys MB, Hirschfield GM. C-reactive protein: a critical update. *J Clin Invest*. 2003;111(12):1805-1812.
- Pepys MB, Rowe IF, Baltz ML. C-reactive protein: binding to lipids and lipoproteins. Int Rev Exp Pathol. 1985;27:83-111.
- Zhang YX, Cliff WJ, Schoefl GI, Higgins G. Coronary C-reactive protein distribution: its relation to development of atherosclerosis. *Atherosclerosis*. 1999;145(2):375-379.
- 309. Oda E, Kawai R. Reproducibility of high-sensitivity C-reactive protein as an inflammatory component of metabolic syndrome in Japanese. *Circ J: Official Journal of the Japanese Circulation Society*. 2010;74(7):1488-1493.
- Fearon WF, Fearon DT. Inflammation and cardiovascular disease: role of the interleukin-1 receptor antagonist. *Circulation*. 2008; 117(20):2577-2579.
- Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. Annu Rev Immunol. 2009;27(1):519-550.
- 312. Ridker PM, Howard CP, Walter V, et al. Effects of interleukin-1β inhibition with canakinumab on hemoglobin A1c, lipids, C-reactive protein, interleukin-6, and fibrinogen: a phase IIb randomized, placebo-controlled trial. *Circulation*. 2012;126(23):2739-2748.
- Popovic M, Ebrahimi F, Urwyler SA, Donath MY, Christ-Crain M. The role of IL-1 in the regulation of copeptin in patients with metabolic syndrome. *Endocr Connect*. 2020;9(7):715-723.
- D'Elia L, Giaquinto A, Cappuccio FP, et al. Circulating leptin is associated with serum uric acid level and its tubular reabsorption in a sample of adult middle-aged men. J Endocrinol Invest. 2020;43(5): 587-593.
- 315. Tabara Y, Osawa H, Kawamoto R, et al. Reduced high-molecularweight adiponectin and elevated high-sensitivity C-reactive protein are synergistic risk factors for metabolic syndrome in a large-scale middle-aged to elderly population: the Shimanami Health Promoting Program Study. J Clin Endocrinol Metab. 2008;93(3): 715-722.
- 316. Dai X, Yuan J, Yao P, et al. Association between serum uric acid and the metabolic syndrome among a middle- and old-age Chinese population. *Eur J Epidemiol*. 2013;28(8):669-676.
- 317. White J, Sofat R, Hemani G, et al. Plasma urate concentration and risk of coronary heart disease: a Mendelian randomisation analysis. *Lancet Diabetes Endocrinol.* 2016;4(4):327-336.
- 318. Mente A, Meyre D, Lanktree MB, et al. Causal relationship between adiponectin and metabolic traits: a Mendelian

randomization study in a multiethnic population. *PLoS ONE*. 2013;8 (6):e66808.

- Kodama K, Tojjar D, Yamada S, Toda K, Patel CJ, Butte AJ. Ethnic Differences in the Relationship Between Insulin Sensitivity and Insulin Response. Syst Rev Meta-Anal. 2013;36:1789-1796.
- WHO. Appropriate body-mass index for Asian populations and its implications for policy and intervention strategies. *Lancet (London, England)*. 2004;363:157-163.
- 321. DeBoer MD, Dong L, Gurka MJ. Racial/ethnic and sex differences in the relationship between uric acid and metabolic syndrome in adolescents: an analysis of National Health and Nutrition Survey 1999-2006. *Metabolism: Clinical and Experimental.* 2012; 61(4):554-561.
- 322. Tsimikas S, Clopton P, Brilakis ES, et al. Relationship of oxidized phospholipids on apolipoprotein B-100 particles to race/ethnicity, apolipoprotein(a) isoform size, and cardiovascular risk factors: results from the Dallas Heart Study. *Circulation*. 2009; 119(13):1711-1719.
- 323. Chen W, Srinivasan SR, Bao W, Wattigney WA, Berenson GS. Sibling aggregation of low- and high-density lipoprotein cholesterol and apolipoproteins B and A-I levels in black and white children: the Bogalusa Heart Study. *Ethn Dis.* 1997;7(3):241-249.
- 324. Iso H, Harada S, Shimamoto T, et al. Polymorphism of the apolipoprotein B gene and blood lipid concentrations in Japanese and Caucasian population samples. *Atherosclerosis*. 1996;126(2): 233-241.
- Vermaak WJ, Ubbink JB, Delport R, Becker PJ, Bissbort SH, Ungerer JP. Ethnic immunity to coronary heart disease? *Atherosclerosis*. 1991;89(2–3):155-162.
- 326. Hackler E 3rd, Lew J, Gore MO, et al. Racial differences in cardiovascular biomarkers in the general population. *J am Heart Assoc.* 2019; 8:e012729.
- 327. Osei K, Gaillard T. Disparities in cardiovascular disease and type 2 diabetes risk factors in Blacks and Whites: dissecting racial paradox of metabolic syndrome. *Front Endocrinol.* 2017;8:204.
- 328. Jacobs S, Kroeger J, Schulze MB, et al. Dietary patterns derived by reduced rank regression are inversely associated with type 2 diabetes risk across 5 ethnic groups in the multiethnic cohort. *Curr Dev Nutr.* 2017;1(5):e000620.
- 329. Kim JY, Yadav D, Ahn SV, Koh SB. A prospective study of serum adiponectin and regression of metabolic syndrome: The ARIRANG study. *Biochem Biophys Res Commun.* 2015;466(2):201-205.
- 330. Kramer CK, von Mühlen D, Jassal SK, Barrett-Connor E. Serum uric acid levels improve prediction of incident type 2 diabetes in individuals with impaired fasting glucose: the Rancho Bernardo Study. *Diabetes Care*. 2009;32(7):1272-1273.
- Feig DI, Kang DH, Johnson RJ. Uric acid and cardiovascular risk. N Engl J Med. 2008;359(17):1811-1821.
- Chuang SY, Chen JH, Yeh WT, Wu CC, Pan WH. Hyperuricemia and increased risk of ischemic heart disease in a large Chinese cohort. *Int J Cardiol.* 2012;154(3):316-321.

- Sattar N, Wannamethee G, Sarwar N, et al. Adiponectin and coronary heart disease: a prospective study and meta-analysis. *Circulation*. 2006;114(7):623-629.
- Li S, Shin HJ, Ding EL, van Dam RM. Adiponectin levels and risk of type 2 diabetes: a systematic review and meta-analysis. JAMA. 2009;302(2):179-188.
- Meier U, Gressner AM. Endocrine regulation of energy metabolism: review of pathobiochemical and clinical chemical aspects of leptin, ghrelin, adiponectin, and resistin. *Clin Chem.* 2004;50(9): 1511-1525.
- 336. Sabatine MS, Morrow DA, Jablonski KA, et al. Prognostic significance of the Centers for Disease Control/American Heart Association high-sensitivity C-reactive protein cut points for cardiovascular and other outcomes in patients with stable coronary artery disease. *Circulation*. 2007;115(12):1528-1536.
- 337. Povel CM, Beulens JW, van der Schouw YT, et al. Metabolic syndrome model definitions predicting type 2 diabetes and cardiovascular disease. *Diabetes Care*. 2013;36(2):362-368.
- 338. Yusuf S, Hawken S, Ounpuu S, et al. Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. *Lancet (London, England)*. 2004;364(9438):937-952.
- 339. Gianfagna F, Veronesi G, Guasti L, et al. Do apolipoproteins improve coronary risk prediction in subjects with metabolic syndrome? Insights from the North Italian Brianza cohort study. *Atherosclerosis*. 2014;236(1):175-181.
- 340. Swerdlow DI, Holmes MV, Kuchenbaecker KB, et al. The interleukin-6 receptor as a target for prevention of coronary heart disease: a mendelian randomisation analysis. *Lancet (London, England).* 2012;379:1214-1224.
- Zinman B, Hanley AJ, Harris SB, Kwan J, Fantus IG. Circulating tumor necrosis factor-alpha concentrations in a native Canadian population with high rates of type 2 diabetes mellitus. J Clin Endocrinol Metab. 1999;84(1):272-278.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of this article.

How to cite this article: Pluimakers VG, van Santen SS, Fiocco M, et al. Can biomarkers be used to improve diagnosis and prediction of metabolic syndrome in childhood cancer survivors? A systematic review. *Obesity Reviews*. 2021;22(11): e13312. https://doi.org/10.1111/obr.13312