# **Review Article**

# Cell and Molecular Mechanisms of Retinal Ganglion Cell Degeneration in Glaucoma

Yalda Yaghooti<sup>1</sup>, MS; Bita Shalbafan<sup>2,\*</sup>, MD; Fatemeh Abdi<sup>3</sup>, MS

1. Faculty of pharmacy, Tehran University of Medical Sciences. Tehran, Iran.

2. Clinical Research Development Center of Labbafinejad Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

3. Master of Internal Surgery Nursing, Hamadan University of Medical Science, Hamadan, Iran.

\*Corresponding Author: Bita Shalbafan

E-mail: shalbafan.b@gmail.com

## Abstract

Glaucoma is an eye disorder in which intraocular pressure is elevated and retinal ganglion cells therefore degenerate. It is a multifaceted ailment with multiple cell types and pathways involved, all working together and giving rise to optic nerve degeneration. Current drugs used in the treatment of glaucoma all work by lowering intraocular pressure and only slowing the progression of the optic nerve damage. No drugs have yet been shown to effectively target retinal ganglion cells and help regain the lost vision. It is of great importance to understand the cellular and molecular processes involved in glaucomatous neurodegeneration to be able to identify potential targets of treatment. The current review attempts to provide insight into these processes. First, an overview of the disease is provided and then, cell types other than retinal ganglion cells (RGCs) that contribute to the neurodegeneration process (including lamina cribrosa cells, astrocytes, oligodendrocytes, and microglia) and cellular and molecular events in the RGCs leading to their degeneration and death (such as mitochondrial dysfunction, axonal transport disruption, calcium dyshomeostasis, oxidative stress, apoptosis, and endothelial reticulum stress) are explained.

**Keywords:** Glaucoma; Retinal Ganglion Cell; Neurodegeneration; Apoptosis; Cellular Components; Signaling Pathways.

Article Notes: Received: Sep. 16, 2019; Received in revised form: Nov. 6, 2019; Accepted: Jan. 2, 2020; Available Online: Apr. 4, 2020.

**How to cite this article:** Yaghooti Y, Shalbafan B, Abdi F. Cell and molecular mechanisms of retinal ganglion cell degeneration in glaucoma. Journal of Ophthalmic and Optometric Sciences . 2020;4(2): 55-71.

Journal of Ophthalmic and Optometric Sciences. Volume 4, Number 2, Spring 2020

#### Introduction

Glaucoma is one of the leading causes of permanent blindness worldwide and it affects about 76 million people around the globe. Increased intraocular pressure (IOP) is a major risk factor for the damage of retinal ganglion cells (RGCs) in the optic nerve, which leads to blindness <sup>2,3</sup>. Other risk factors include Old age, family history, black or Asian race, diseases such as diabetes and hypertension, and use of systemic or topical corticosteroids <sup>4,5</sup>.

There are several types of glaucoma including open-angle, angle-closure, and normal-tension glaucoma. Open-angle glaucoma is the result of the ineffective outflow of the aqueous humor through trabecular and uveoscleral pathways in the anterior chamber of the eye and in contrast to angle-closure, the angle between the iris and the cornea is normal in this type of glaucoma. It is the most common type and patients suffering from it are usually asymptomatic until the damage to the optic nerve is severe and peripheral vision is lost. In angle-closure glaucoma, on the other hand, closure of this angle occurs due usually to agerelated thickening of the lens and as a result, the drainage of the aqueous humor through the trabecular meshwork is blocked. IOP increases rapidly in angle-closure glaucoma and can cause vision loss within a day of the onset of aqueous outflow blockage. The decreased aqueous outflow in angle-closure and openangle glaucoma then results in increased IOP which then damages the optic nerve. Unlike these two types, IOP is not elevated in normaltension glaucoma and it is believed that the optic nerve damage could be the consequence of either pressure-sensitivity of the optic nerve or nerve ischemia due to vascular insufficiency. Accordingly, all types of glaucoma cause neurodegeneration in the retinal nerve fiber layer <sup>4</sup>.

The optic nerve consists of RGC axons, glial cells, the connective tissue of the lamina cribrosa, blood vessels. Although and neurodegeneration glaucomatous occurs throughout the whole visual pathway from the retina to the brain, the optic nerve head (the location where RGC axons exit the eye to form the optic nerve) seems to be the initial site of RGC injury in glaucoma<sup>4</sup>. Figure 1 demonstrates the structure of the optic nerve head, its cellular components, and the effects of increased IOP on them.

The only drug treatments shown to be efficacious in glaucoma, reduce the intraocular pressure and include prostaglandin analogs, beta-blockers, alpha-2 agonists, carbonic anhydrase inhibitors, miotic agents, and more recently Rho-kinase inhibitors and nitric-oxide donating medications <sup>6</sup>. These medications only prevent the progression of the disease and do not reverse vision loss 7. Moreover, the IOPlowering effect of these medications wears off in some patients and despite controlled intraocular pressure, the IOP fluctuation in response to these drugs could lead to the progression of vision loss <sup>8</sup>. Many drugs including antioxidants, neuroprotective and immunomodulatory agents have been tested in clinical trials but none were shown to be efficacious <sup>9</sup> and the efforts made to develop effective medications targeting the optic nerve have been unsuccessful due to a lack of precise understanding of the underlying molecular events in glaucomatous neuropathy. Some pathways and events such as inflammation, ECM remodeling, RGC axonal degeneration, and apoptosis are proven to participate in glaucomatous RGC degeneration. However, the importance of some events such as RGC excitotoxicity and declined activity of insulin receptor signaling pathway is not quite clear and other pathways involved in the



Figure 1: The Optic Nerve Head and Events Contributing to IOP-induced RGC Axonal Degeneration and Apoptosis. A) A normal optic nerve head with inactivae glial cells (green), B) IOP-induced stress (black arrows) on the retinal ganglion cells and activation of glial cells, C) Axonal demyelination and degeneration and apoptosis of some retinal ganglion cells due to glial activation, D) Optic nerve head cupping, backward displacement of lamina cribrosa, glial-induced neuroinflammation and death of most retinal ganglion cells. Adopted from: (1)

pathogenesis of glaucomatous neuropathy are yet to be discovered.

The purpose of the current review is to provide a clear view of the cellular processes known to participate in the optic nerve degeneration following increased intraocular pressure observed in most glaucoma patients. First, we will discuss the cell types and then the cellular and molecular events in RGCs and these cell types which have important roles in glaucomatous neurodegeneration. A summary of the upcoming sections is shown in table 1.

# Cell types involved in glaucomatous neurodegeneration

Multiple cell types other than RGCs are involved in the neurodegeneration process in glaucoma. Normally, they are responsible for providing biomechanical, trophic, metabolic, and immunomodulatory protection to RGCs but due to changes in glaucoma, they tend to increase RGCs vulnerability to injury or even cause the injury themselves. These cell types, which are all present in the optic nerve head, include lamina cribrosa cells, glial cells (astrocytes, microglia, and oligodendrocytes), and vascular endothelial cells <sup>10</sup>. Figure 2 depicts the signaling pathways and cellular events of these cell types involved in glaucomatous neurodegeneration and each one is discussed in the following sections.

## Lamina cribrosa

The optic nerve head is provided with mechanical support by lamina cribrosa which consists of parallel series of fibroelastic connective tissue plates (see figure 3). It also provides a scaffold for different cell types like lamina cribrosa cells, glial cells, and vascular endothelial cells. Elevated IOP causes a structural reconfiguration in the lamina cribrosa which is the reason for the cupping 

 Table 1: Cellular and molecular events involved in glaucomatous neurodegeneration of retinal ganglion cells, cell types involved in each, and their inducers and consequences. Up and down arrows indicate increase and decrease of the item on their right, respectively

Cellular or molecular event	Cell(s) involved	Inducer(s)	Consequence(s)
Activation of stretch- activated K+ channels	Lamina cribrosa cells	Mechanical stress	Intracellular calcium $\uparrow$
Axonal transport dysfunction	RGCs	$ATP\downarrow$	Mitochondrial dysfunction, NMNAT $\downarrow$
Bax activation	RGCs	NF-kB activation, JNK pathway activation, insulin receptor activation, ER stress	Caspase cascade activation
Caspase cascade activation	RGCs	Bax activation, ER stress	Apoptosis
Cytoskeleton degeneration	RGCs	Oxidative stress, proteasome activation	Axonal degeneration
ECM production $\uparrow$	Lamina cribrosa cells, astrocytes	Mechanical stress, TGF-β, oxidative stress ↑	Tissue deformation
ER stress	RGCs	Abnormal protein aggregation	Activation of pro-apoptotic proteins, mitochondrial dysfunction
Extracellular ATP	RGCs	RGC damage	Purinergic receptor activation
Extracellular HSP	RGCs	RGC damage	TLR activation
IL-1 $\beta$ receptor activation	RGCs	IL-1 $\beta$ secretion $\uparrow$	NF-κB activation
IL-1 $\beta$ secretion $\uparrow$	Microglia	Vanilloid receptor activation, NLRP3 activation	IL-1 $\beta$ receptor activation
Insulin receptor activation ↓	RGCs, Microglia, Astrocytes	Unknown	Mitochondrial dysfunction, expression of pro- inflammatory mediators, apoptosis, impaired astroglial metabolic support
Intracellular calcium ↑	Lamina cribrosa cells RGCs	Activation of stretch- activated K+ channels ↓ ATP, NMDA receptor activation, SARM1 activation, NAD+↓	Expression of profibrotic proteins Proteasome activation
JNK pathway activation	RGCs	axonal cytoskeleton distortion, neurotrophin deprivation, energy failure, or neuroinflammation	Activation of pro-apoptotic proteins
Mechanical stress	RGCs, lamina cribrosa cells, astrocytes, microglia, oligodendrocytes, vascular endothelial cells	IOP ↑	Tissue deformation in the optic nerve head, ECM production, ↑ TGF-β↑

Journal of Ophthalmic and Optometric Sciences. Volume 4, Number 2, Spring 2020

Yaghooti et al.

# Cell and molecular mechanisms of retinal ganglion cell degeneration

Cellular or molecular event	Cell(s) involved	Inducer(s)	Consequence(s)
Mitochondrial dysfunction	RGCs	ER stress, Axonal transport dysfunction, vascular dysfunction, activation of pro-apoptotic proteins	ATP, oxidative stress $\downarrow$
MMP ↑	Astrocytes	TGF-β ↑	Tissue deformation
Neurotrophin deprivation	RGCs	Axonal transport dysfunction	Apoptosis
NF-κB activation	RGCs	TNF- $\alpha$ receptor activation, IL-1 $\beta$ receptor activation	Bax activation
NMDA receptor activation	RGCs	Unknown	Intracellular calcium ↑
NMNAT↓	RGCs	Axonal transport dysfunction, JNK signaling	SARM1 activation, NAD+ $\downarrow$
NLRP3 activation	Microglia	Purinergic receptor activation	IL-1β ↑
Oxidative stress	RGCs, lamina cribrosa cells, astrocytes, microglia, oligodendrocytes, vascular endothelial cells	Mitochondrial dysfunction	ECM production, damage to proteins & DNA, cytoskeleton degeneration ↑
Proteasome activation	RGCs	Intracellular calcium ↑	Cytoskeleton degeneration
Purinergic receptor activation	Microglia	Extracellular ATP	NLRP3 activation
SARM1 activation	RGCs	NMNAT $\downarrow$	Intracellular calcium ↑
Tenascin-C production	Astrocytes	IOP	TLR activation
TGF-β ↑	Lamina cribrosa cells	Mechanical stress	ECM production, $\uparrow$ MMP $\uparrow$
Tissue deformation	RGCs, lamina cribrosa cells, astrocytes, microglia, oligodendrocytes, vascular endothelial cells	MMP, ↑ IOP, ↑ ECM ↑ production	Activation of stretch-activated K+ channels, TLR activation, vascular dysfunction
TLR activation	Microglia	Tenascin-C production, tissue deformation	TNF-α ↑
TNF- $\alpha$ receptor activation	RGCs	TNF- $\alpha$ secretion $\uparrow$	NF-κB activation
TNF- $\alpha$ secretion $\uparrow$	Microglia	TLR activation, Vanilliod receptor activation	TNF- $\alpha$ receptor activation
Vanilloid receptor activation	Microglia	$IOP\uparrow$	IL-1 $\beta$ , $\uparrow$ TNF- $\alpha$ $\uparrow$
Vascular dysfunction	RGCs, lamina cribrosa cells, astrocytes, microglia, oligodendrocytes, vascular endothelial cells	↑ IOP, ↓ insulin signaling, ECM production, tissue deformation	Oxidative stress



**Figure 2:** Signaling pathways and other cellular events of retinal ganglion cells, lamina cribrosa cells, microglia, and astrocytes. Proteins and other entities demonstrated in this figure mostly participate in pathways leading to inflammation (with proteins such as IL-1 $\beta$ , TNF- $\alpha$ , TLR, and NLRP3 complex) and extracellular matrix remodeling (with proteins such as MMP and TGF). Abbreviations: ECM: extracellular matrix, HSP: heat-shock protein, IL-1 $\beta$ : Interleukin-1 $\beta$ , IOP: intraocular pressure, LC: lamina cribrosa, RGC: retinal ganglion cell, ROS: reactive oxygen species, TGF: transforming growth factor, TLR: toll-like receptor, TNC: Tenascin-C, TNF- $\alpha$ : tumor necrosis factor. Created with BioRender.com

and enlargement of the optic disc observed in glaucomatous eyes <sup>11</sup>. Elevated IOP generates mechanical stress on the lamina and sclera and results in the backward displacement of lamina cribrosa which then deforms the resident cells <sup>12</sup> and eventually, damages the RGC axons and optic nerve head capillaries <sup>13</sup>. Due to individual differences in extracellular matrix components and therefore differences in the biomechanical properties of the sclera <sup>14</sup>, the stress experienced by RGC axons, and glial and vascular cells' response to stress could vary <sup>11</sup>. This explains the racial differences in susceptibility of RGCs to IOP-induced injury <sup>15</sup>.

Lamina cribrosa cells, which are located between the connective tissue plates of the lamina cribrosa <sup>16</sup>, secrete components of the extracellular matrix (ECM) including fibronectin, collagen, and elastin and along with astrocytes, which are explained later, these cells are responsible for ECM remodeling and fibrosis in the optic nerve head. The production of ECM proteins by lamina



**Figure 3:** Longitudinal section (A) and cross-section (B) of lamina cribrosa in the optic nerve head. A) Surface nerve fiber layer (A), Prelaminar region (B), Lamina cribrosa region containing connective tissue and elastic fibers (C), and Retrolaminar region (D). B) Fenestrated structure of lamina cribrosa with blood vessels passing through the central openings (arrow) and passage of retinal ganglion cell axons through the sorriunding fenestrae. S: superior, T: temporal. Adopted from: Entokey.com

cribrosa cells and astrocytes is increased in response to IOP-induced mechanical stretch, TGF- $\beta$ , and oxidative stress. Secretion of TGF- $\beta$  is induced by increased IOP, while elevated levels of reactive oxygen species and oxidative stress are a result of reduction of the optic nerve head blood flow <sup>12</sup>.

Lamina cribrosa cells are also able to sense the IOP-induced deformation and after their stretch-activated potassium channels open in response to the increased pressure, intracellular calcium levels elevate <sup>17</sup> and promote the profibrotic processes <sup>18</sup>.

In rodents and non-human primates, the mechanical support for the optic nerve is provided by a network of astroglial processes (called glia lamina), and not lamina cribrosa, as it isn't well developed in these species <sup>19</sup>. The topography of RGC damage in rodents matches the localized damage of the axon bundles at the glia lamina, and a similar pattern is seen in ocular hypertension-induced RGC damage, regardless of the structural differences of the optic nerve head. Thus, lamina cribrosa

is not required for glaucomatous neuropathy, and glial pro-fibrotic and cellular processes (mostly related to astrocytes) are responsible for the observed optic nerve head damage <sup>11</sup>.

#### Glial cells

Glial cells are important elements of the nervous system and provide support for all neurons including retinal ganglion cells. There are several types of glial cells such as astrocytes and microglia which possess crucial roles in the central nervous system. Astrocytes are responsible for maintaining the blood-brain barrier and providing support for neurons <sup>20</sup>, while microglia play important roles in inflammation and brain infections<sup>21</sup>. Evidence shows that astrogliosis (activation of astrocytes) and microgliosis (activation of microglia) play a major role in the development of neurodegeneration 11 glaucomatous Astrogliosis and microgliosis also lead to the loss of oligodendrocytes, another type of glial cell responsible for myelination of neurons in the CNS, through secretion of inflammatory

cytokines by microglia <sup>22</sup>. Two phenomena are known to activate glial cells in glaucoma: increased IOP-induced stress and signals from RGCs. Glial mechanosensitive ion channels, such as vanilloid receptors, purinergic receptors, and pannexin channels, sense the increased IOP and lead to the initiation of inflammatory signaling <sup>23</sup>. They can also be activated due to the release of damage-associated molecules like ATP <sup>24</sup> and heat-shock protein (HSP) <sup>25</sup> from injured RGCs as explained later in more detail. We will now discuss the inducers and consequences of astrocytes and microglia activation, respectively.

Increased IOP leads to alterations in the activity of astrocytes. This response by astrocytes is called reactive astrogliosis 26. In short term, astrocytes provide more trophic and metabolic support to the uninjured axons and assist in tissue repair in the injured ones by eliminating the injured dendritic structures with the help of microglia, but the prolonged effects of reactive astrogliosis are destructive rather than protective. Prolonged activation of glial cells results in diminished structural, trophic, and bioenergetic support from astrocytes, and neuroinflammation and formation of a toxic environment for RGCs by microglia <sup>27</sup>. However, Diminished support for RGCs is not the only aspect of astrogliosis. When sensing the IOP-induced tissue deformation, astrocytes produce more extracellular matrix and cause an imbalance between the production of matrix metalloproteinase (proteolytic enzymes that degrade ECM components) and their inhibitors. The resulting extracellular matrix remodeling increases the mechanical stress on the optic nerve axons <sup>28</sup>. This could also explain the existing distress of vascular properties of the optic nerve. Initially, it was thought that the direct mechanical stress of elevated IOP on optic nerve head blood vessels

is the reason behind vascular dysfunction in glaucoma, but it is now evident that it could also be explained by astrocytes activity. The basal lamina produced by astrocytes surrounds RGCs and microcapillaries which besides the tissue stiffening, alters the intracellular signaling between astrocytes and both RGCs and endothelial cells of the microcapillaries <sup>13</sup>. Increased inflammatory signaling is an example thatlipid leads to the cytotoxic and phagocytic activity of glial cells and optic nerve neurodegeneration <sup>11</sup>.

Microglia are responsible for neuroinflammation cellular and death detected in RGCs of glaucomatous eyes, but they aren't the only phagocytes involved in this process. Monocytes in the blood can enter the optic nerve head via the activation of leukocyte transendothelial migration pathway in the early stages of glaucoma and together with microglia<sup>29</sup>, secret proinflammatory cytokines such as TNF- $\alpha$  and IL-1 $\beta$  which activate the NF- $\kappa$ B pathway in retinal ganglion cells and result in apoptosis and axonal degeneration 27. Secretion of IL-1 $\beta$  by microglia is a result of NLRP3 inflammasome activation through stimulation of purinergic receptors by damage-associated molecules like ATP and ROS production by the mitochondria <sup>30</sup>. TNF- $\alpha$  secretion is in response to the stimulation of Toll-like receptors (TLRs) which are a class of pattern recognition receptors (PRRs) that initiate the innate immune response of microglia <sup>31</sup> by sensing stress-related ligands like heat-shock protein and Tenascin-C secreted by RGCs and astrocytes, respectively 29, 32. There are two mechanisms by which TNF- $\alpha$  can cause damage to RGCs: the caspase-dependent apoptosis pathway which is briefly explained later, and the caspase-independent pathway in which mitochondrial dysfunction and oxidative

stress occur <sup>33</sup>. An alternate mechanism by which microglia can contribute to the loss of RGCs dendrites and synapses is through the activation of the complement system <sup>34</sup>.

Microglia, like many other cells, communicate with each other via exosomes which are extracellular vesicles that may contain miRNAs, proteins, or lipids <sup>35</sup>. Active microglia produce these exosomes and result in the activation of other microglia and therefore, exacerbate the inflammation and RGC death <sup>36</sup>. ATP release from RGCs stimulates the exosome production by microglia which could explain why elevated IOP doubles the exosome release <sup>37</sup>.

Besides innate immunity, glial cells can induce adaptive immunity which results in the abnormal activity of T-cells and increased production of autoantibodies against ocular antigens. Thus, the cytotoxicity mediated by the autoantibodies could also lead to toxicity to RGCs <sup>38</sup>.

# Cellular processes in RGCs involved in glaucomatous neurodegeneration

Multiple cellular processes are involved in glaucomatous neurodegeneration including impaired axonal transport and depletion of neurotrophic factors, SARM1-induced axonal degeneration, cytoskeletal disruption, neuroinflammation, vascular dysregulation, mitochondrial dysfunction and energy failure, reticulum oxidative stress, endothelial (ER) dysfunction, calcium dyshomeostasis, excitotoxicity, and insulin resistance <sup>11</sup>. These processes and how they are linked together are discussed in the following section and a summary of them is shown in figure 3.

# Axonal transport and mitochondrial dysfunction

Increased IOP disrupts the axoplasmic flow which is crucial for the retrograde transport of

neurotrophic factors from the brain and this lack of neurotrophic factors leads to apoptosis. Dysfunctional axonal transport also leads to the accumulation of some proteins like amyloid precursor protein (APP) and  $\gamma$ -synuclein <sup>39</sup> in the axons, which are proteins known for their contribution to neurodegeneration. The compromised transport could be a result of mitochondrial dysfunction which itself could either be caused by deficient vascular nutrient supply due to compromised capillaries in the optic nerve head or increased mitochondrial permeability (which occurs due to activation of pro-apoptotic proteins such as Bax and Bak resulting in the activation of the caspase cascade, as later discussed) and therefore mitochondrial swelling and oxidative stress due to activated self-destruction program <sup>40</sup>. The axonal selfdestruction is activated because of the lack of anterograde axonal survival factors supplied from the brain <sup>41</sup>. Also, the compromised transport can lead to the accumulation of dysfunctional mitochondria which are normally eliminated from the cytoplasm in a process called mitophagy <sup>11</sup>. Mitophagy is decreased in aging <sup>42</sup> which could be one of the reasons why aging is a risk factor for glaucoma. Dysfunctional axonal transport also leads to decreased transport of mitochondria to sites of injury in the RGCs and therefore, fails to compensate for the increased energy demand in these sites <sup>43</sup>. Other than mitochondrial and capillary impairment, decreased metabolic supply by astrocytes is also speculated to cause inadequate energy supply <sup>44</sup>.

Besides energy generation, mitochondria are critical for the regulation of intracellular calcium homeostasis, apoptosis signaling, and oxidative stress, all of which contribute to RGC degeneration in glaucoma <sup>11</sup>. Studies also suggest that dysfunctional mitochondria could stimulate neuroinflammation <sup>45</sup> by



Figure 4: Cellular and signaling pathways involved in axons (right) and somas (left) of retinal ganglion cells in glaucoma and their relationships Created with BioRender.com

Abbreviations: ATP: adenosine triphosphate, Ca: calcium, Casp: caspase, IL-1R: interleukin-1 receptor, NAD+: nicotinamide adenine dinucleotide, NMNAT: nicotinamide mononucleotide adenylyltransferase, ROS: reactive oxygen species, SARM1: sterile alpha and TIR motif-containing protein 1, TNF: tumor necrosis factor

contributing to inflammasome activation <sup>46</sup>. Inflammasome activity may provoke mitochondrial injury and create a pathological cycle between mitochondrial dysfunction and neuroinflammation <sup>47</sup>.

# Calcium dyshomeostasis and cytoskeletal disruption

The enzyme nicotinamide mononucleotide adenylyltransferase 2 (NMNAT2) which is responsible for the production of nicotinamide adenine dinucleotide (NAD+), decreases due to injury-induced disrupted axonal transport. NAD+ is crucial in calcium regulation and its decreased amounts cause an elevation in the intracellular concentration of calcium. Dysregulation of calcium could also be a result of ATP depletion and therefore the reversed activity of the ATP-dependent Na+/Ca++ exchanger. The increased intraaxonal calcium then over-activates the ubiquitin-proteasome system and therefore leads to protease-mediated cytoskeletal breakdown and the destruction of axonal structure <sup>41</sup>. Calcium could also activate calpains (cysteine proteases) which also participate in proteolysis in the axonal cytoskeleton <sup>48</sup>.

## SARM1-dependent axonal degeneration

As explained before, axonal transport is disrupted in RGCs of glaucomatous eyes and consequently, turnover of the enzyme NMNAT2 is decreased. Normally, NMNAT2 inhibits the enzyme SARM1 (sterile alpha and TIR motif-containing protein 1)<sup>49</sup>

which hydrolyzes NAD+ to form NAM (nicotinamide) and either ADPR (adenosine diphosphate ribose) or cADPR (cyclic ADPR)<sup>50</sup>. When the NMNAT2 amounts are reduced, SARM1 activates and produces more ADPR and cADPR. These two molecules then bind to calcium channels and increase calcium influx and therefore, as explained in the last section, lead to axonal degeneration <sup>51</sup>. Other mechanisms by which NMNAT2 activity is decreased are the activation of the JNK signaling pathway 52 and mitochondrial dysfunction in RGCs which could also decrease the activity of STMN2 (Stathmin 2), another SARM1 inhibitor 53. TNF-a released from microglia can also activate SARM1 and lead to axonal degeneration 54.

The whole process of SARM1-induced axonal degeneration following lowered activity of NMNAT2 is known as Wallerian degeneration <sup>53</sup> which eventually leads to RGC axonal loss, RGC death, and oligodendrocyte loss. However, oligodendrocyte loss appears to be secondary to SARM1-induced axonal loss <sup>54</sup>.

## **Oxidative** stress

Increased generation of reactive oxygen species is another consequence of mitochondrial dysfunction. The resulting oxidative stress can impact cell survival through oxidative damage to proteins <sup>33</sup> and boost neurodegenerative inflammation <sup>55</sup>. Antioxidant treatment in experimental glaucoma models has been shown to act as an immunomodulator and protect RGCs somas and axons <sup>56</sup>.

# Apoptosis and other signaling pathways

Apoptosis in RGCs occurs following the activation of the caspase cascade. Caspase could either be triggered by Bax, a member of the pro-apoptotic Bcl-2 family of proteins

which is activated by energy depletion due to mitochondrial dysfunction and neurotrophin deficiency, or by binding of microglia-derived TNF-α to its receptor on RGCs 57. TNF-α could also act independently of caspase and cause mitochondrial dysfunction and oxidative stress <sup>33</sup>. When Bax and Bak are activated, they aggregate at the mitochondrial outer membrane (MOM), undergo conformational changes, and insert into the membrane. Then, they form homodimers and cause membrane destabilization and therefore, proteolipid pores are formed in the mitochondrial membrane <sup>58</sup>. As a result, pro-apoptotic signaling molecules such as cytochrome C exit the intermembrane space and enter the cytosol. This results in the activation of the caspase cascade which leads to apoptosis of the RGC 59. Other important pathways include the JNK pathway and Death receptor-6 (DR6) signaling.

The JNK pathway is important in signal transduction after cellular stress. In glaucoma, it can be activated by distortion of axonal cytoskeleton, neurotrophin deprivation, energy failure, or neuroinflammation. It appears that this pathway is involved in both axonal degeneration and RGC soma apoptosis <sup>60</sup> by initiating the apoptotic transcriptional program and interacting with the Bcl-2 family of genes <sup>61</sup>.

In the Death receptor-6 (DR6) signaling pathway, Activation of DR6 receptor by surface ligands like amyloid precursor protein (APP) due to axon injury results in the activation of caspase-6 (which is independent of the caspase cascade activated during apoptosis) and therefore activates axonal destruction program and leads to axonal degeneration <sup>62</sup>. APP is a protein involved in the pathophysiology of Alzheimer's disease. The involvement of APP, the accumulation of tau protein (a key mediator of neurotoxicity in Alzheimer's

65

Journal of Ophthalmic and Optometric Sciences. Volume 4, Number 2, Spring 2020

disease) in glaucoma <sup>63</sup>, and other similarities between glaucoma and Alzheimer's disease demonstrate that these two are more related than previously thought.

#### ER stress

ER is the major intracellular organelle that senses cellular stresses and environmental changes, coordinates signaling pathways, and controls cell survival <sup>64</sup>. Studies indicate the presence of chronic ER stress in glaucoma. Abnormal protein aggregation which could be a result of aging leads to the activation of unfolded protein response (UPR) <sup>65</sup> by which the ER attempts to clear out the misfolded proteins. The UPR consists of the activation of chaperones, inhibition of mRNA translation, and transportation of the unfolded proteins to the cytosol for ubiquitination <sup>66</sup>. Some ER stress-related signaling proteins control cell fate either by activating pro-apoptotic molecules in response to cell burden 67.

ER and mitochondria are in contact at membrane contact sites (MCSs) 68. These MCSs are important in some processes such as calcium transfer from the ER to the mitochondria(essential for many mitochondrial activities such as oxidative phosphorylation) and lipid transfer which are crucial for cell homeostasis <sup>43, 69</sup>. During the UPR, MCSs between the ER and the mitochondria are augmented and therefore, calcium transfer to the mitochondria is increased for the production of more ATP needed in the UPR. Calcium overload in the mitochondria then leads to increased release of cytochrome C to the cytosol and apoptosis <sup>70</sup>. Other than induction of apoptosis, loss of cytochrome C protein also leads to less ATP production and hence, production of ROS and cell damage <sup>43</sup>. The existence of more MCSs could also cause apoptosis due to more aggregation of Bax and

Bak at these sites <sup>70</sup>.

## **Excitotoxicity**

The association of excessive NMDA glutamate receptor activation and therefore excitotoxicity with glaucoma is believed to be related to the OPTN gene which encodes Optineurin. Optineurin is a protein involved in controlling glutamate receptor signaling and is often mutated in patients with primary open-angle glaucoma (POAG)<sup>71</sup>. However, there isn't much evidence demonstrating excitotoxicity as a primary mechanism for RGC death in glaucoma.

#### Insulin receptor (IR) signaling

IR signaling has been shown to be important in the pathogenesis of glaucoma. In the RGC, declined activity of the insulin signaling pathway promotes dendritic retraction, mitochondrial dysfunction, tau hyperphosphorylation, and apoptosis. In microglial cells, insulin resistance induces the expression of pro-inflammatory mediators and therefore neuroinflammation in RGCs. It also contributes to vascular dysfunction by causing nitric oxide/endothelin-1 imbalance and endothelial cell apoptosis. In astrocytes, decreased insulin signaling causes depletion of glycogen stores which impairs astroglial metabolic support for RGCs. However, it is not quite clear whether insulin signaling directly contributes to the pathogenesis of glaucomatous neurodegeneration or is only a consequence of it 72.

#### Conclusion

Multiple cellular components are involved in glaucomatous neurodegeneration consisting of neuroinflammation, vascular dysfunction, mitochondrial and axonal transport dysfunction, cytoskeletal disruption, and many other

processes. In glaucomatous neurodegeneration, different cellular pathways are activated in different cellular compartments, multiple cell types (including glial cells and lamina cribrosa cells) are involved and asynchrony of neurodegeneration is seen among different RGCs, meaning the degeneration does not affect all RGCs at once. Thus, recognizing the earliest molecular events in glaucomatous neurodegeneration is very challenging. Given the complexity of these processes, systems biology and integration of different '-omics' data could be tremendously helpful in understanding the precise sequence of cell type-specific events. The resulting spatiotemporal knowledge of pathological events in glaucomatous neurodegeneration could lead to the identification of new pharmacological targets in retinal ganglion cells that could slow the progress of degeneration more effectively or even reverse the optic nerve damage done before diagnosis.

#### **ORCID IDs**

Yalda Yaghooti:

b https://orcid.org/0000-0003-0835-4279

Bita Shalbafan:

https://orcid.org/0000-0002-7748-0538

#### References

 Kwon YH, Fingert JH, Kuehn MH, Alward WL. Primary open-angle glaucoma. N Engl J Med. 2009;360(11):1113-24. Epub 2009/03/13.
 Allison K, Patel D, Alabi O. Epidemiology of Glaucoma: The Past, Present, and Predictions for the Future. Cureus. 2020;12(11):e11686.

 Geyer O, Levo Y. Glaucoma is an autoimmune disease. Autoimmun Rev. 2020;19(6):102535.
 Weinreb RN, Aung T, Medeiros FA. The

pathophysiology and treatment of glaucoma: a review. JAMA. 2014;311(18):1901-11.

5. McMonnies CW. Glaucoma history and risk factors. J Optom. 2017;10(2):71-8.

6. Dietze J, Blair K, Havens SJ. Glaucoma. StatPearls. Treasure Island (FL): StatPearls PublishingCopyright © 2021, StatPearls Publishing LLC.; 2021.

7. Schuster AK, Erb C, Hoffmann EM, Dietlein T, Pfeiffer N. The Diagnosis and Treatment of Glaucoma. Dtsch Arztebl Int. 2020;117(13):225-34.

8. Shalaby WS, Shankar V, Razeghinejad R, Katz LJ. Current and new pharmacotherapeutic

approaches for glaucoma. Expert Opin Pharmacother. 2020;21(16):2027-40.

9. Tsai JC. Innovative IOP-Independent Neuroprotection and Neuroregeneration Strategies in the Pipeline for Glaucoma. J Ophthalmol. 2020;2020:9329310.

10. Lopez NN, Clark AF, Tovar-Vidales T. Isolation and characterization of human optic nerve head astrocytes and lamina cribrosa cells. Exp Eye Res. 2020;197:108103.

11. Tezel G. A broad perspective on the molecular regulation of retinal ganglion cell degeneration in glaucoma. Prog Brain Res. 2020;256(1):49-77.

12. Wallace DM, O'Brien CJ. The role of lamina cribrosa cells in optic nerve head fibrosis in glaucoma. Exp Eye Res. 2016;142:102-9.

13. Burgoyne CF, Downs JC, Bellezza AJ, Suh JK, Hart RT. The optic nerve head as a biomechanical structure: a new paradigm for understanding the role of IOP-related stress and strain in the pathophysiology of glaucomatous optic nerve head damage. Prog Retin Eye Res. 2005;24(1):39-73.

14. Murienne BJ, Jefferys JL, Quigley HA,

Journal of Ophthalmic and Optometric Sciences. Volume 4, Number 2, Spring 2020

Nguyen TD. The effects of glycosaminoglycan degradation on the mechanical behavior of the posterior porcine sclera. Acta Biomater. 2015;12:195-206.

15. Park HL, Kim JH, Jung Y, Park CK. Racial Differences in the Extracellular Matrix and Histone Acetylation of the Lamina Cribrosa and Peripapillary Sclera. Invest Ophthalmol Vis Sci. 2017;58(10):4143-54.

16. Tovar-Vidales T, Wordinger RJ, Clark AF. Identification and localization of lamina cribrosa cells in the human optic nerve head. Exp Eye Res. 2016;147:94-7.

17. McElnea EM, Quill B, Docherty NG, Irnaten M, Siah WF, Clark AF, et al. Oxidative stress, mitochondrial dysfunction and calcium overload in human lamina cribrosa cells from glaucoma donors. Mol Vis. 2011;17:1182-91.

 Irnaten M, Zhdanov A, Brennan D, Crotty T, Clark A, Papkovsky D, et al. Activation of the NFAT-Calcium Signaling Pathway in Human Lamina Cribrosa Cells in Glaucoma. Invest Ophthalmol Vis Sci. 2018;59(2):831-42.
 Howell GR, Libby RT, Jakobs TC, Smith RS, Phalan FC, Barter JW, et al. Axons of retinal ganglion cells are insulted in the optic nerve early in DBA/2J glaucoma. J Cell Biol. 2007;179(7):1523-37.

20. Siracusa R, Fusco R, Cuzzocrea S. Astrocytes: Role and Functions in Brain Pathologies. Front Pharmacol. 2019;10:1114.

21. Wake H, Moorhouse AJ, Nabekura J. Functions of microglia in the central nervous system--beyond the immune response. Neuron Glia Biol. 2011;7(1):47-53.

22. Domingues HS, Portugal CC, Socodato R, Relvas JB. Oligodendrocyte, Astrocyte, and Microglia Crosstalk in Myelin Development, Damage, and Repair. Front Cell Dev Biol. 2016;4:71.

23. Albalawi F, Lu W, Beckel JM, Lim JC, McCaughey SA, Mitchell CH. The P2X7

Receptor Primes IL-1beta and the NLRP3 Inflammasome in Astrocytes Exposed to Mechanical Strain. Front Cell Neurosci. 2017;11:227.

24. Reigada D, Lu W, Zhang M, Mitchell CH. Elevated pressure triggers a physiological release of ATP from the retina: Possible role for pannexin hemichannels. Neuroscience. 2008;157(2):396-404.

25. Luo C, Yang X, Kain AD, Powell DW, Kuehn MH, Tezel G. Glaucomatous tissue stress and the regulation of immune response through glial Toll-like receptor signaling. Invest Ophthalmol Vis Sci. 2010;51(11):5697-707.

26. Lye-Barthel M, Sun D, Jakobs TC. Morphology of astrocytes in a glaucomatous optic nerve. Invest Ophthalmol Vis Sci. 2013;54(2):909-17.

27. Bariş M, Tezel G. Immunomodulation as a Neuroprotective Strategy for Glaucoma Treatment. Curr Ophthalmol Rep. 2019;7(2):160-9.

28. Roberts MD, Liang Y, Sigal IA, Grimm J, Reynaud J, Bellezza A, et al. Correlation between local stress and strain and lamina cribrosa connective tissue volume fraction in normal monkey eyes. Invest Ophthalmol Vis Sci. 2010;51(1):295-307.

29. Howell GR, Soto I, Zhu X, Ryan M, Macalinao DG, Sousa GL, et al. Radiation treatment inhibits monocyte entry into the optic nerve head and prevents neuronal damage in a mouse model of glaucoma. J Clin Invest. 2012;122(4):1246-61.

30. Coyle S, Khan MN, Chemaly M, Callaghan B, Doyle C, Willoughby CE, et al. Targeting the NLRP3 Inflammasome in Glaucoma. Biomolecules. 2021;11(8).

31. Nie L, Cai SY, Shao JZ, Chen J. Toll-Like Receptors, Associated Biological Roles, and Signaling Networks in Non-Mammals. Front Immunol. 2018;9:1523. 00.

32. Tezel G. A decade of proteomics studies of glaucomatous neurodegeneration. Proteomics Clin Appl. 2014;8(3-4):154-67.

33. Tezel G, Yang X. Caspase-independent component of retinal ganglion cell death, in vitro. Invest Ophthalmol Vis Sci. 2004;45(11):4049-59.

34. Williams PA, Tribble JR, Pepper KW, Cross SD, Morgan BP, Morgan JE, et al. Inhibition of the classical pathway of the complement cascade prevents early dendritic and synaptic degeneration in glaucoma. Mol Neurodegener. 2016;11:26.

35. Aires ID, Ribeiro-Rodrigues T, Boia R, Catarino S, Girao H, Ambrosio AF, et al. Exosomes derived from microglia exposed to elevated pressure amplify the neuroinflammatory response in retinal cells. Glia. 2020;68(12):2705-24.

36. Paolicelli RC, Bergamini G, Rajendran L. Cell-to-cell Communication by Extracellular Vesicles: Focus on Microglia. Neuroscience. 2019;405:148-57.

37. Rodrigues-Neves AC, Aires ID, Vindeirinho J, Boia R, Madeira MH, Goncalves FQ, et al. Elevated Pressure Changes the Purinergic System of Microglial Cells. Front Pharmacol. 2018;9:16.

38. Lorenz K, Beck S, Keilani MM, Wasielica-Poslednik J, Pfeiffer N, Grus FH. Longitudinal Analysis of Serum Autoantibody-Reactivities in Patients with Primary Open Angle Glaucoma and Optic Disc Hemorrhage. PLoS One. 2016;11(12):e0166813..

39. Surgucheva I, McMahan B, Ahmed F, Tomarev S, Wax MB, Surguchov A. Synucleins in glaucoma: implication of gamma-synuclein in glaucomatous alterations in the optic nerve. J Neurosci Res. 2002;68(1):97-106.

40. Barrientos SA, Martinez NW, Yoo S, Jara JS, Zamorano S, Hetz C, et al. Axonal

degeneration is mediated by the mitochondrial permeability transition pore. J Neurosci. 2011;31(3):966-78.

41. Wang JT, Medress ZA, Barres BA. Axon degeneration: molecular mechanisms of a self-destruction pathway. J Cell Biol. 2012;196(1):7-18..

42. Diot A, Morten K, Poulton J. Mitophagy plays a central role in mitochondrial ageing. Mamm Genome. 2016;27(7-8):381-95.

43. Muench NA, Patel S, Maes ME, Donahue RJ, Ikeda A, Nickells RW. The Influence of Mitochondrial Dynamics and Function on Retinal Ganglion Cell Susceptibility in Optic Nerve Disease. Cells. 2021;10(7).

44. SaabAS, Tzvetavona ID, TrevisiolA, Baltan S, Dibaj P, Kusch K, et al. Oligodendroglial NMDA Receptors Regulate Glucose Import and Axonal Energy Metabolism. Neuron. 2016;91(1):119-32.

45. Clarke LE, Liddelow SA, Chakraborty C, Münch AE, Heiman M, Barres BA. Normal aging induces A1-like astrocyte reactivity. Proc Natl Acad Sci U S A. 2018;115(8):E1896-e905. 46. Dela Cruz CS, Kang MJ. Mitochondrial dysfunction and damage associated molecular patterns (DAMPs) in chronic inflammatory diseases. Mitochondrion. 2018;41:37-44.

47. Tezel G. Molecular regulation of neuroinflammation in glaucoma: Current knowledge and the ongoing search for new treatment targets. Prog Retin Eye Res. 2021:100998.

48. Ma M, Ferguson TA, Schoch KM, Li J, Qian Y, Shofer FS, et al. Calpains mediate axonal cytoskeleton disintegration during Wallerian degeneration. Neurobiol Dis. 2013;56:34-46.

49. Mink M, Fogelgren B, Olszewski K, Maroy P, Csiszar K. A novel human gene (SARM) at chromosome 17q11 encodes a protein with a SAM motif and structural similarity to Armadillo/beta-catenin that is conserved

in mouse, Drosophila, and Caenorhabditis elegans. Genomics. 2001;74(2):234-44.

50. Loring HS, Thompson PR. Emergence of SARM1 as a Potential Therapeutic Target for Wallerian-type Diseases. Cell Chem Biol. 2020;27(1):1-13.

51. Adalbert R, Morreale G, Paizs M, Conforti L, Walker SA, Roderick HL, et al. Intraaxonal calcium changes after axotomy in wildtype and slow Wallerian degeneration axons. Neuroscience. 2012;225:44-54.

52. Walker LJ, Summers DW, Sasaki Y, Brace EJ, Milbrandt J, DiAntonio A. MAPK signaling promotes axonal degeneration by speeding the turnover of the axonal maintenance factor NMNAT2. Elife. 2017;6.

53. Loreto A, Hill CS, Hewitt VL, Orsomando G, Angeletti C, Gilley J, et al. Mitochondrial impairment activates the Wallerian pathway through depletion of NMNAT2 leading to SARM1-dependent axon degeneration. Neurobiol Dis. 2020;134:104678.

54. Ko KW, Milbrandt J, DiAntonio A. SARM1 acts downstream of neuroinflammatory and necroptotic signaling to induce axon degeneration. J Cell Biol. 2020;219(8).

55. Tezel G. The immune response in glaucoma: a perspective on the roles of oxidative stress. Exp Eye Res. 2011;93(2):178-86.

56. Yang X, Hondur G, Tezel G. Antioxidant Treatment Limits Neuroinflammation in Experimental Glaucoma. Invest Ophthalmol Vis Sci. 2016;57(4):2344-54.

57. Tezel G, Wax MB. Increased production of tumor necrosis factor-alpha by glial cells exposed to simulated ischemia or elevated hydrostatic pressure induces apoptosis in cocultured retinal ganglion cells. J Neurosci. 2000;20(23):8693-700.

58. Qian S, Wang W, Yang L, Huang HW. Structure of transmembrane pore induced by Bax-derived peptide: evidence for lipidic pores. Proc Natl Acad Sci U S A. 2008;105(45):17379-83.

59. Xu XP, Zhai D, Kim E, Swift M, Reed JC, Volkmann N, et al. Three-dimensional structure of Bax-mediated pores in membrane bilayers. Cell Death Dis. 2013;4:e683.

60. Syc-Mazurek SB, Fernandes KA, Libby RT. JUN is important for ocular hypertension-induced retinal ganglion cell degeneration. Cell Death Dis. 2017;8(7):e2945.

61. Watkins TA, Wang B, Huntwork-Rodriguez S, Yang J, Jiang Z, Eastham-Anderson J, et al. DLK initiates a transcriptional program that couples apoptotic and regenerative responses to axonal injury. Proc Natl Acad Sci U S A. 2013;110(10):4039-44.

62. Ito Y, Shimazawa M, Tsuruma K, Mayama C, Ishii K, Onoe H, et al. Induction of amyloidbeta(1-42) in the retina and optic nerve head of chronic ocular hypertensive monkeys. Mol Vis. 2012;18:2647-57.

63. Chiasseu M, Cueva Vargas JL, Destroismaisons L, Vande Velde C, Leclerc N, Di Polo A. Tau Accumulation, Altered Phosphorylation, and Missorting Promote Neurodegeneration in Glaucoma. J Neurosci. 2016;36(21):5785-98.

64. Schwarz DS, Blower MD. The endoplasmic reticulum: structure, function and response to cellular signaling. Cell Mol Life Sci. 2016;73(1):79-94.

65. Jing G, Wang JJ, Zhang SX. ER stress and apoptosis: a new mechanism for retinal cell death. Exp Diabetes Res. 2012;2012:589589.

66. Bravo R, Parra V, Gatica D, Rodriguez AE, Torrealba N, Paredes F, et al. Endoplasmic reticulum and the unfolded protein response: dynamics and metabolic integration. Int Rev Cell Mol Biol. 2013;301:215-90.

67. Wang X, Olberding KE, White C, Li C. Bcl-2 proteins regulate ER membrane permeability to luminal proteins during ER stress-induced apoptosis. Cell Death Differ. 2011;18(1):38-47.

68. Prinz WA, Toulmay A, Balla T. The functional universe of membrane contact sites. Nat Rev Mol Cell Biol. 2020;21(1):7-24.

69. Ozturk Z, O'Kane CJ, Perez-Moreno JJ. Axonal Endoplasmic Reticulum Dynamics and Its Roles in Neurodegeneration. Front Neurosci. 2020;14:48.

70. Peña-Blanco A, García-Sáez AJ. Bax, Bak and beyond - mitochondrial performance in apoptosis. Febs j. 2018;285(3):416-31.

71. Doucette LP, Rasnitsyn A, Seifi M, Walter MA. The interactions of genes, age, and

environment in glaucoma pathogenesis. Surv Ophthalmol. 2015;60(4):310-26.

72. Al Hussein Al Awamlh S, Wareham LK, Risner ML, Calkins DJ. Insulin Signaling as a Therapeutic Target in Glaucomatous Neurodegeneration. Int J Mol Sci. 2021;22(9).

#### **Footnotes and Financial Disclosures**

#### **Conflict of interest:**

The authors have no conflict of interest with the subject matter of the present manuscript. The authors have no conflict of interest with the subject matter of the present study.