# **Correlation of Microclimate of West Java on Caffeine and Chlorogenic acid in** *Coffea canephora* var. *robusta*

Suci Awaliyah<sup>\*</sup>, Sri Nanan B. Widiyanto, Rijanti R. Maulani, Asep Hidayat, Ujang Dinar Husyari, Tati S . Syamsudin, Erly Marwani<sup>\*</sup>

School of Life Sciences and Technology, Institut Teknologi Bandung

\*) Corresponding author; e-mail: suciawaliyah16@gmail.com, erly@sith.itb.ac.id Received: 2022-02-24 Accepted for publication: 2022-04-13

#### Abstract

Caffeine and chlorogenic acid (CGA) are two compounds that play a role in determining the quality of coffee. The amount of the two compounds may vary depending on the environment where they are grown. This study aimed to determine the correlation between the local microclimatic condition and the concentration of caffeine and CGA in green and roasted beans of Robusta coffee from six different cultivation areas in West Java, Indonesia (i.e., Ciamis, Tasikmalaya, Sumedang, Kuningan, Cianjur, and Bogor). Samples of green beans and roasted beans were extracted with 70% methanol for caffeine analysis and ethyl acetate for CGA analysis. Caffeine and CGA were analyzed by UV-HPLC using a C18 shimpack gist shimadzu column, with an isocratic elution of methanol:water (1:1) at a 1 mL/min flow rate. Detection was performed at  $\lambda 272 \text{ nm}$  and  $\lambda 324 \text{ nm}$  for caffeine and chlorogenic acid, respectively. Principal component analysis (PCA) was used to evaluate the correlation between microclimate with caffeine and chlorogenic acid. Results indicated that the concentration of caffeine ranged from 7.67 to 16.52% and 10.79 to 15.56% in the green and roasted bean coffee, respectively. Based on PCA analysis, the most influential microclimate on the caffeine concentration were the humidity, temperature, and altitude, with the total variance of PC1 and PC2 of 76.3%. However, there was no positive correlation between the measured microclimate and the CGA concentration. In conclusion, Robusta coffee's caffeine content is positively affected by the microclimatic condition (i.e., humidity, temperature, and altitude).

Keywords: caffeine, chlorogenic acid, Robusta, microclimate, Java coffee

#### 1. Introduction

Indonesia is known as the third largest coffee producer globally after Brazil and Vietnam [1,2,3,4]. The region of Coffee production exists in almost all parts of Indonesia, including West Java. Based on the Regional Rule of the Province of West Java concerning Guidelines for the Implementation of Cultivation, coffee is a commodity that has an essential role in building the economy of the society of West Java [5,6,7].

Coffee in West Java consists of two types, i.e., Arabica and Robusta coffee. Arabica coffee is more suitable for being grown at high altitude, while Robusta coffee can be grown at lower altitudes. The average productivity of Robusta coffee is 784 kg/Ha. The area cultivation of the Robusta coffee in West Java is around 15.750 Ha [35]. West Java Robusta coffee has been marketed to local and international markets. Robusta coffee from Ciamis, Sumedang, Tasikmalaya, and Bogor has been locally marketed to other provinces in Indonesia. In contrast, Robusta coffee from Cianjur and Kuningan has been internationally marketed to other countries such as Saudi Arabia, Morocco, Singapore, Turkey, and France.

In general, Robusta coffee from various regions in West Java has different flavors, such as sweetness and acidity [30]. Many factors influence the differences in Robusta coffee flavors, such as natural and artificial factors [35]. Natural factors include geographical conditions, genetics or varieties/species, microclimates conditions, and fruit maturity. Artificial factors such as post-harvest processing include preparation, grinding, drying, washing, and roasting temperature [8,9,26,35].

The demand for West Java Robusta coffee generally is related to the coffee quality, especially the taste of coffee. The taste of coffee is influenced by the composition of the metabolites in the beans [13,14]. Some coffee beans' metabolites, e.g. sugar, caffeine, theobromine, theophylline, trigonelline, and chlorogenic acid, can significantly affect the taste [27,28,29]. Two of them were caffeine and chlorogenic acid [15,16,17]. Caffeine plays a role in determining the bitter taste of coffee, and chlorogenic acid in coffee's sour taste [18,19,20,21,22,36]. An increase in caffeine concentration in the coffee beans indicates a good quality of coffee. In contrast, the rise of CGA concentration in the coffee beans indicates poor quality [8].

According to previous studies, environmental factors such as altitude and microclimate (i.e., temperature, humidity and rainfall) correlate with caffeine and CGA concentration [10,11,12,17]. However, the concentration of caffeine and CGA negatively correlates with altitude [37]. In addition, lower temperatures and low intensity delay the coffee maturity, thus, inhibiting some metabolites' accumulation in coffee beans [11,23]. Moreover, no significant effect was found between CGA concentration and rainfall, but the temperature positively correlates with the caffeine concentration and CGA [23,24].

Research on the analysis of caffeine and CGA concentrations in coffee has been done previously by Jeszka et al. [34], Awwad et al. [36], Girma et al. [37], and Hagos et al. [38]. None of those researches analyzed caffeine and CGA concentration of Robusta coffee beans from West Java. Therefore, the objective of this study was to analyze the caffeine and CGA of Robusta coffee from West Java and to determine the correlation between microclimates with the concentrations of caffeine and CGA. Cultivation area of Robusta coffee in West Java selected which has area more than 1000 Ha, including the areas of Bogor, Ciamis, Cianjur, Kuningan, Sumedang, and Tasikmalaya.

In this study, High-Performance Liquid Chromatography (HPLC) was used to analyze caffeine and CGA as HPLC is an effective method for analyzing caffeine and chlorogenic acids in the coffee bean and roasted bean [32,33]. The concentration of caffeine and CGA were correlated to the microclimate (temperature, rainfall, and humidity) and altitude using a multivariate statistical. The Principal Component Analysis (PCA) in this study was used to determine the correlation between each variable and the concentration of caffeine and CGA.

## 2. Methodology

The sample of six green and six roasted coffee beans were obtained from six different areas in West Java (Bogor, Ciamis, Cianjur, Kuningan, Sumedang, and Tasikmalaya). Robusta Coffee from those cultivation areas was subjected to caffeine and CGA analysis using HPLC. The concentration of caffeine and CGA was correlated to microclimates (temperature, rainfall, and humidity) and the altitude of where the coffee originated. Multivariate statistical analysis was used to interpret the data through Principal Component Analysis (PCA).

#### 2.1. Samples preparation

Green beans Robusta coffee were obtained from smallholder farmers from six different cultivation areas in West Java (i.e., Ciamis, Tasikmalaya, Sumedang, Kuningan, Cianjur, and Bogor). Roasted samples of Robusta coffee were prepared using a coffee roaster machine. The samples of coffee beans were ground using a coffee grinder, Cyprus grinder GR0063. The ground green bean coffee was put quickly into a 30 ml dark glass bottle to avoid the evaporation of volatile compounds. Bottles containing samples of roasted and green bean coffee were stored at room temperature before further processing.

## 2.2. Extraction

0.25 g of green beans and roasted beans from each location were separately extracted with 30 mL of methanol/water (70:30 v/v) for caffeine analysis and ethyl acetate for chlorogenic acid analysis. The extracts were sonicated at  $65^{\circ}$ C for 10 minutes and centrifuged at 1372 xg for 3 minutes. This procedure was repeated three times, and the supernatant was collected at the end of each centrifugation cycle. The supernatant from each sample was evaporated using a vacuum evaporator to obtain a crude extract. Then, it was filtered using a polytetrafluoroethylene (PTFE) syringe filter with a size of 0.22µm, according to the method of Ceylan et al. [3].

## 2.3. Caffeine and CGA analysis

The analysis of caffeine and chlorogenic acid in the green and roasted bean samples was conducted by Shimadzu Prominence type 20A HPLC. Twenty  $\mu$ L of each sample was applied to the HPLC using Shimpack C18 gist shimadzu column (5  $\mu$ m particles, 4.6 mm internal diameter, and 25 cm length). The analysis was carried out at 27 °C using isocratic elution of methanol:water (50:50) with a flow rate of 1 mL/min. A UV detector was used at  $\lambda$ 272 nm and  $\lambda$ 324 nm to detect the caffeine and chlorogenic acid peaks, respectively.

Standard solutions of caffeine ad CGA were prepared in separate flask by appropriate dilution of 0.1 mg/mL (100ppm) stock solutions with the same solvent to contain 0.5,0.25,0.125, and 0. 0625 mg/ml final concentration. Each standard solution was injected into HPLC in triplicate. Calibration curves were obtained by plotting the peak areas of each standard solution versus the concentration of the injected standard solution.

# 2.4. Microclimate

Data of temperature, humidity, and rainfall were average per year obtained from the data center of the National Statistics Agency on the website (https://www.bps.go.id/), and data on altitude was obtained from Google Earth 2021.

# 2.5. Statistical analysis

The data were processed by Minitab version 17 followed by principal component analysis (PCA) using Minitab version 17 to evaluate the correlation between microclimate with caffeine and chlorogenic acid.

# 3. Results and discussion

# 3.1. Caffeine and CGA concentration

Results showed that the caffeine concentration in green beans and roasted beans from six coffee plantation areas ranged from 7.67-16.52% in green beans and 10.79-15.56% in roasted beans, respectively (Table 1.). The lowest caffeine concentration was found in coffee samples from Ciamis, both for green bean coffee (7.67%) and roasted bean coffee (10.79%). The highest caffeine concentration was found in the green bean coffee sample from Cianjur (16.52%) and the roasted bean coffee sample from Bogor (15.46%). In this study, the Robusta coffee has a high concentration of caffeine. This finding was in line with the study by Cheng et al. [17] and Sunarhanum et al. [8], who mentioned that the concentration of caffeine in Robusta coffee was higher than in Arabica coffee. The study also showed that the concentration of caffeine in coffee beans before and after the roasting process was not much different. This result was correlated with a previous study by Cheng et al.[17] and Jeszka et al. [34] and supported by the theory that caffeine has thermostable characteristics [17].

Chlorogenic acid concentration ranged from 0.74-3.03% in green bean coffee and 0.25-0.77% in roasted bean coffee, respectively (Table 1.). The lowest concentration of chlorogenic acid was found in the green bean coffee sample from Cianjur (0.74%) and the roasted bean coffee sample from Tasikmalaya (0.25%). The highest concentration of chlorogenic acid was found in the green bean coffee sample from Sumedang (3.03%) and in the roasted bean coffee sample from Kuningan (0.77%). The study also showed that the concentration of chlorogenic acid in coffee beans before and after the roasting process is not much different. This result agrees with the study of Cheng et al. [17] and is supported by the theory that chlorogenic acid has thermostable characteristics [17].

Caffeine concentration increased after the roasting process, while CGA concentration decreased (Table 1.). The change

in caffeine and chlorogenic acid concentration after roasting was affected by high temperatures during the roasting process, as reported by Sunarharum et al. [8]. The temperature and duration of the roasting process affect caffeine concentration and a drop of chlorogenic acid [8]. The roasting process plays a role in determining the taste and quality of brew coffee. Several volatile and nonvolatile compounds were formed during the roasting process and contribute significantly to the sensory characteristics of coffee drinks. Certain compounds' intensity varies depending on the temperature of roasting processed, particularly caffeine and chlorogenic acid. During the roasting process, chlorogenic acid was degraded into lactone, which caused the quantity of CGA to decrease [25]. On the other hand, caffeine acid releases caffeine during the roasting process, increasing the total caffeine concentration [31].

# 3.2. Correlation of caffeine and CGA with altitude and microclimates

The PCA analysis reveals that the caffeine and chlorogenic acid concentrations were affected by different microclimatic conditions. Caffeine concentration was positively correlated with humidity, altitude, and temperature. On the other hand, caffeine does not correlate with rainfall. The chlorogenic acid concentration was not correlated with the microclimate. PCA results showed that the most influential microclimatic variable to caffeine concentration was the humidity, altitude, and temperature, with a variance value of 47.8% (PC1) and 28.5% (PC2), or 76.3% in total (Figure 1).

The microclimate variables, altitude, temperature, rainfall, and humidity were not correlated with the concentration of CGA, with a variance of 37.2% (PC1) and 34.1 (PC2) or a total of 71.3% (Figure 2.). Caffeine concentration was with humidity, positively correlated altitude, and temperature, while CGA concentrations were not correlated with microclimatic variables (humidity, temperature, and rainfall) and altitude. Altitude was negatively correlated with CGA concentration [17,37,38]. Indeed, the coffee from Cianjur with the highest altitude (713 masl) showed the lowest concentration of CGA (0.74%). A similar result was also reported by Girma et al. [37] and Hagos et al. [38].

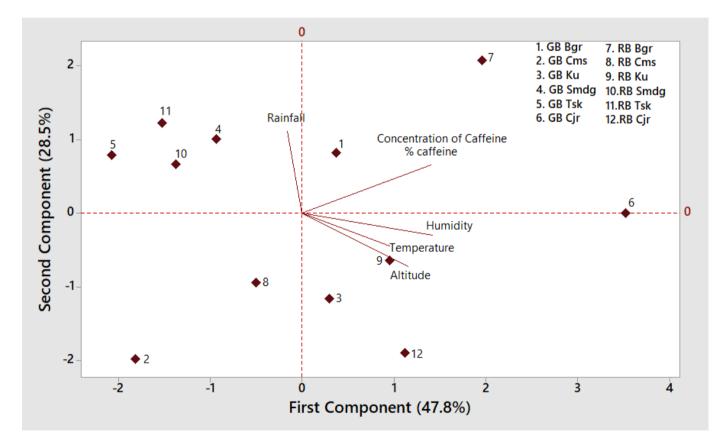
According to Odeny et al. [10], environmental factors influence caffeine accumulation [10]. For example, lower temperatures, high altitude, and high humidity delay coffee maturity for about one month and cause the biochemical composition to variate significantly at the stage of bean development. Delay coffee maturity causes the accumulation of caffeine and CGA [23,11]. However, no relationship was found between caffeine and CGA concentration and rainfall (Figure 2.). A similar result was also reported in another study by Joet et al. [23].

Sample coffee	Region	Concentration (%) (v/v)		water content (%)	Microclimates*			Altitude (masl**)
		CGA (%)	Caffeine (%)	_ ` `	Temp . (°C)	Humidity (%)	Rainfall (mm/year)	_
Green	Bogor	1,86	11,69	8,13	25,5	83	3930	700
Bean	Ciamis	3,0	7,67	4,28	25	74,8	1309	607
	Kuningan	3,03	11,07	4,99	27	74	1682	657,5
	Sumedang	2,28	12,13	4,85	24,7	69	2570	551
	Tasikmalaya	1,77	10,14	3,31	25,7	67	2750	398,5
	Cianjur	0,74	16,52	4,04	26,5	88	1250	713
Roaste	Bogor	0,27	15,46	1,81	25,5	83	3930	700
d Bean	Ciamis	0,32	10,79	0,92	25	74,8	1309	607
	Kuningan	0,77	12,62	3,10	27	74	1682	657,5
	Sumedang	0,32	11,09	1,57	24,7	69	2570	551
	Tasikmalaya	0,25	11,43	1,10	25,7	67	2750	398,5
	Cianjur	0,49	10,82	3,53	26,5	88	1250	713

**Table 1.** The concentration of caffeine and chlorogenic acid in the green bean and roasted bean Robusta coffee samples from six areas of West Java.

\*Data of microclimates were obtained from the data center of the National Statistics Agency on the website (<u>https://www.bps.go.id/</u>), while data on altitude was obtained from google earth 2021.

\*\*Above the sea level



**Figure 1**. PCA biplot of microclimatic (altitude, humidity, rainfall, temperature) on caffeine concentration (GB = green beans, RB = roasted beans, Bgr = Bogor coffee samples, Cms = Ciamwas coffee samples, Ku = Kuningan coffee samples, Smdg = sample of Sumedang coffee, Tsk = sample of Tasik coffee and Cjr = sample of Cianjur coffee).

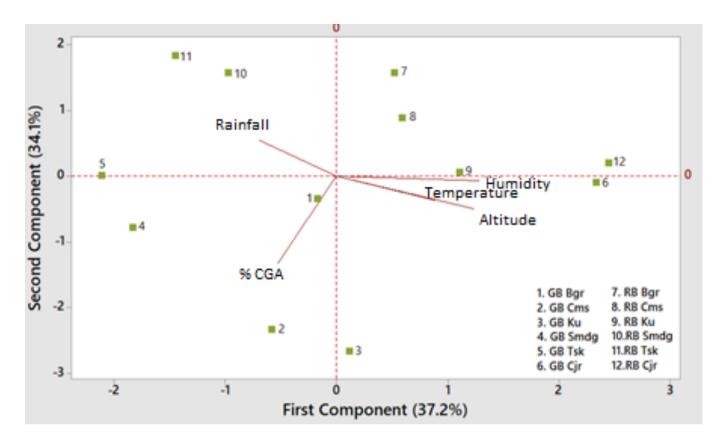


Figure 2. PCA biplot of microclimatic (altitude, humidity, rainfall, temperature) on CGA concentration (GB = green beans, RB = roasted beans, Bgr = Bogor coffee samples, Cms = Ciamwas coffee samples, Ku = Kuningan coffee samples, Smdg = sample of Sumedang coffee, Tsk = sample of Tasik coffee and Cjr = sample of Cianjur coffee).

#### 4. Conclusion

There is a positive correlation between humidity, temperature, altitude, and caffeine concentration. However, no correlation between the microclimate with the concentration of CGA.

## Acknowledgments

The authors acknowledge financial grants provided by P2MI Institut Teknologi Bandung, Indonesia to Erly Marwani. We also acknowledge to Lembaga Pengelola Dana Pendidikan (LPDP) who gave Master degree program scholarship to Suci Awaliyah.

## References

 Jeszka-Skowron M, Zgoła-Grześkowiak A, Grześkowiak T. Analytical methods applied for the characterization and the determination of bioactive compounds in coffee. European Food Research and Technology. 2015 Jan;240(1):19-31. DOI: https://doi.org/10.1007/s00217-014-2356-z

- [2] Dulsat-Serra N, Quintanilla-Casas B, Vichi S. Volatile thiols in coffee: A review on their formation, degradation, assessment and influence on coffee sensory quality. Food Research International. 2016 Nov 1;89:982-8. DOI: https://doi.org/10.1016/j.foodres.2016.02.008
- [3] Esquivel P, Jimenez VM. Functional properties of coffee and coffee by-products. Food research international. 2012 May 1;46(2):488-95. DOI: https://doi.org/10.1016/j.foodres.2011.05.028
- [4] Bhupathiraju SN, Pan A, Malik VS, Manson JE, Willett WC, van Dam RM, Hu FB. Caffeinated and caffeine-free beverages and risk of type 2 diabetes. The American journal of clinical nutrition. 2013 Jan 1;97(1):155-66. DOI: https://doi.org/10.3945/ajcn.112.048603
- [5] Cano-Marquina A, Tarín JJ, Cano A. The impact of coffee on health. Maturitas. 2013 May 1;75(1):7-21. DOI: https://doi.org/10.1016/j.maturitas.2013.02.002
- [6] Jiang L, Ding Y, Jiang F, Li L, Mo F. Electrodeposited nitrogen-doped graphene/carbon nanotubes nanocomposite as enhancer for simultaneous and sensitive voltammetric determination of caffeine and vanillin. Analytica chimica acta. 2014 Jun 23;833:22-8. DOI: https://doi.org/10.1016/j.aca.2014.05.010
- [7] Manalu DS, Harianto H, Suharno S, Hartoyo S. Permintaan Kopi Biji Indonesia di Pasar Internasional. AGRIEKONOMIKA. 2020 Jun 29;9(1):114-26. DOI: https://doi.org/10.21107/agriekonomika.v9i1.7346

- [8] Sunarharum WB, Williams DJ, Smyth HE. Complexity of coffee flavor: A compositional and sensory perspective. Food Research International. 2014 Aug 1;62:315-25. DOI: https://doi.org/10.1016/j.foodres.2014.02.030
- [9] Choi MY, Choi W, Park JH, Lim J, Kwon SW. Determination of coffee origins by integrated metabolomic approach of combining multiple analytical data. Food Chemistry. 2010 Aug 15;121(4):1260-8. DOI: https://doi.org/10.1016/j.foodchem.2010.01.035
- [10] Odeny D, Chemining'wa G, Shibairo S. Beverage quality and biochemical components of shaded coffee. In25th International Conference of Coffee Science 2014.
- [11] Kumar, A., Simmi, P., Naik, G. K., Giridhar, P., RP-HPLC and transcript profile indicate increased leaf caffeine in Coffea canephora plants by light, Journal of Biology and Earth Sciences, 2015, 5(1), pp. 1-9.
- [12] Putri SP, Irifune T, Fukusaki E. GC/MS based metabolite profiling of Indonesian specialty coffee from different species and geographical origin. Metabolomics. 2019 Oct;15(10):1-1. DOI: https://doi.org/10.1007/s11306-019-1591-5
- Farah A, de Paulis T, Trugo LC, Martin PR. Effect of roasting on the formation of chlorogenic acid lactones in coffee. Journal of agricultural and food chemistry. 2005 Mar 9;53(5):1505-13. DOI: https://doi.org/10.1021/jf048701t
- [14] Monteiro MC, Farah A. Chlorogenic acids in Brazilian Coffea arabica cultivars from various consecutive crops. Food Chemistry. 2012 Sep 1;134(1):611-4. DOI: https://doi.org/10.1016/j.foodchem.2012.02.118
- [15] Reuter JA, Spacek DV, Snyder MP. High-throughput sequencing technologies. Molecular cell. 2015 May 21;58(4):586-97. DOI: https://doi.org/10.1016/j.molcel.2015.05.004
- [16] Tran HT, Lee LS, Furtado A, Smyth H, Henry RJ. Advances in genomics for the improvement of quality in coffee. Journal of the Science of Food and Agriculture. 2016 Aug;96(10):3300-12. DOI: https://doi.org/10.1002/jsfa.7692
- [17] Cheng B, Furtado A, Smyth HE, Henry RJ. Influence of genotype and environment on coffee quality. Trends in Food Science & Technology. 2016 Nov 1;57:20-30. DOI: https://doi.org/10.1016/j.tifs.2016.09.003
- [18] Rodrigues CI, Marta L, Maia R, Miranda M, Ribeirinho M, Máguas C. Application of solid-phase extraction to brewed coffee caffeine and organic acid determination by UV/HPLC. Journal of Food Composition and Analysis. 2007 Aug 1;20(5):440-8. DOI: https://doi.org/10.1016/j.jfca.2006.08.005
- [19] Dawidowicz AL, Typek R. Transformation of chlorogenic acids during the coffee beans roasting process. European Food Research and Technology. 2017 Mar;243(3):379-90. DOI: https://doi.org/10.1007/s00217-016-2751-8
- [20] Upadhyay R, Mohan Rao LJ. An outlook on chlorogenic acids—occurrence, chemistry, technology, and biological activities. Critical reviews in food science and nutrition. 2013 Jan 1;53(9):968-84. DOI: https://doi.org/10.1080/10408398.2011.576319
- [21] Lepelley, M., Cheminade, G., Tremillon, N., Simkin, A., Caillet, V. and McCarthy, J., 2007. Chlorogenic acid synthesis in coffee: An analysis of CGA content and realtime RT-PCR expression of HCT, HQT, C3H1, and CCoAOMT1 genes during grain development in C.

canephora. *Plant Science*, *172*(5), pp.978-996. DOI: https://doi.org/10.1016/j.plantsci.2007.02.004

- [22] Lepelley M, Mahesh V, McCarthy J, Rigoreau M, Crouzillat D, Chabrillange N, de Kochko A, Campa C. Characterization, high-resolution mapping and differential expression of three homologous PAL genes in Coffea canephora Pierre (Rubiaceae). Planta. 2012 Jul;236(1):313-26. DOI: https://doi.org/10.1007/s00425-012-1613-2
- [23] Joët T, Salmona J, Laffargue A, Descroix F, Dussert S. Use of the growing environment as a source of variation to identify the quantitative trait transcripts and modules of coexpressed genes that determine chlorogenic acid accumulation. Plant, cell & environment. 2010 Jul;33(7):1220-33. DOI: https://doi.org/10.1111/j.1365-3040.2010.02141.x
- [24] Tovar MJ, Romero MP, Girona J, Motilva MJ. l-Phenylalanine ammonia-lyase activity and concentration of phenolics in developing olive (Olea europaea L cv Arbequina) fruit grown under different irrigation regimes. Journal of the Science of Food and Agriculture. 2002 Jun;82(8):892-8. DOI: https://doi.org/10.1002/jsfa.1122
- [25] Ginz M, Balzer HH, Bradbury AG, Maier HG. Formation of aliphatic acids by carbohydrate degradation during roasting of coffee. European Food Research and Technology. 2000 Nov;211(6):404-10. DOI: https://doi.org/10.1007/s002170000215
- [26] Ruan YL. Sucrose metabolism: gateway to diverse carbon use and sugar signaling. Annual review of plant biology. 2014 Apr 29;65:33-67. DOI: https://doi.org/10.1146/annurev-arplant-050213-040251
- [27] Yashin A, Yashin Y, Xia X, Nemzer B. Chromatographic methods for coffee analysis: a review. Journal of Food Research. 2017;6(4):60-82. DOI: https://doi.org/10.5539/ifr.v6n4p60
- [28] Bicho NC, Leitão AE, Ramalho JC, de Alvarenga NB, Lidon FC. Identification of chemical clusters discriminators of Arabica and Robusta green coffee. International Journal of Food Properties. 2013 May 19;16(4):895-904. DOI: https://doi.org/10.1080/10942912.2011.573114
- [29] Perrois C, Strickler SR, Mathieu G, Lepelley M, Bedon L, Michaux S, Husson J, Mueller L, Privat I. Differential regulation of caffeine metabolism in Coffea arabica (Arabica) and Coffea canephora (Robusta). Planta. 2015 Jan;241(1):179-91. DOI: https://doi.org/10.1007/s00425-014-2170-7
- [30] Farah A, Monteiro MC, Calado V, Franca AS, Trugo LC. Correlation between cup quality and chemical attributes of Brazilian coffee. Food chemistry. 2006 Jan 1;98(2):373-80. DOI: https://doi.org/10.1016/j.foodchem.2005.07.032
- [31] Seninde DR, Chambers E. Coffee flavor: A review. Beverages. 2020 Sep;6(3):44. DOI: https://doi.org/10.3390/beverages6030044
- [32] Cordoba N, Fernandez-Alduenda M, Moreno FL, Ruiz Y. Coffee extraction: A review of parameters and their influence on the physicochemical characteristics and flavour of coffee brews. Trends in Food Science & Technology. 2020 Feb 1;96:45-60. DOI: https://doi.org/10.1016/j.tifs.2019.12.004
- [33] Ceylan R, Zengin G, Guler GO, Aktumsek A. Bioactive constituents of Lathyrus czeczottianus and ethyl acetate and water extracts and their biological activities: An endemic plant to Turkey. South African Journal of Botany. 2021 Dec 1;143:306-11. DOI:

https://doi.org/10.1016/j.sajb.2020.11.023

- [34] Jeszka-Skowron M, Sentkowska A, Pyrzyńska K, De Peña MP. Chlorogenic acids, caffeine content and antioxidant properties of green coffee extracts: influence of green coffee bean preparation. European Food Research and Technology. 2016 Aug;242(8):1403-9. DOI: https://doi.org/10.1007/s00217-016-2643-y
- [35] Dinas Perkebunan Jawa Barat, http://disbun.jabarprov.go.id/post/view/118-idpengembangan-kopi-di-jawa-barat, 2016.
- [36] Awwad S, Issa R, Alnsour L, Albals D, Al-Momani I. Quantification of Caffeine and Chlorogenic Acid in Green and Roasted Coffee Samples Using HPLC-DAD and Evaluation of the Effect of Degree of Roasting on Their Levels. Molecules. 2021 Jan;26(24):7502. DOI: https://doi.org/10.3390/molecules26247502
- [37] Girma B, Gure A, Wedajo F. Influence of Altitude on Caffeine, 5-caffeoylquinic acid, and nicotinic acid contents of arabica coffee varieties. Journal of Chemistry. 2020 Aug 12;2020. DOI: https://doi.org/10.1155/2020/3904761
- [38] Hagos M, Redi-Abshiro M, Chandravanshi BS, Ele E, Mohammed AM, Mamo H. Correlation between caffeine contents of green coffee beans and altitudes of the coffee plants grown in southwest Ethiopia. Bulletin of the Chemical Society of Ethiopia. 2018 Apr 12;32(1):13-25. DOI: https://doi.org/10.4314/bcse.v32i1.2