

INHIBITION OF Ca²⁺-INDEPENDENT PHOSPHOLIPASE A₂ BY 2-OXOAMIDES BASED ON DIPEPTIDES AND ETHER PSEUDODIPEPTIDES

Anneta Smyrniotou,^a Efrosini Barbayianni,^b Ishita Shah,^c Violetta Constantinou-Kokotou,^a Edward A. Dennis,^c George Kokotos^b

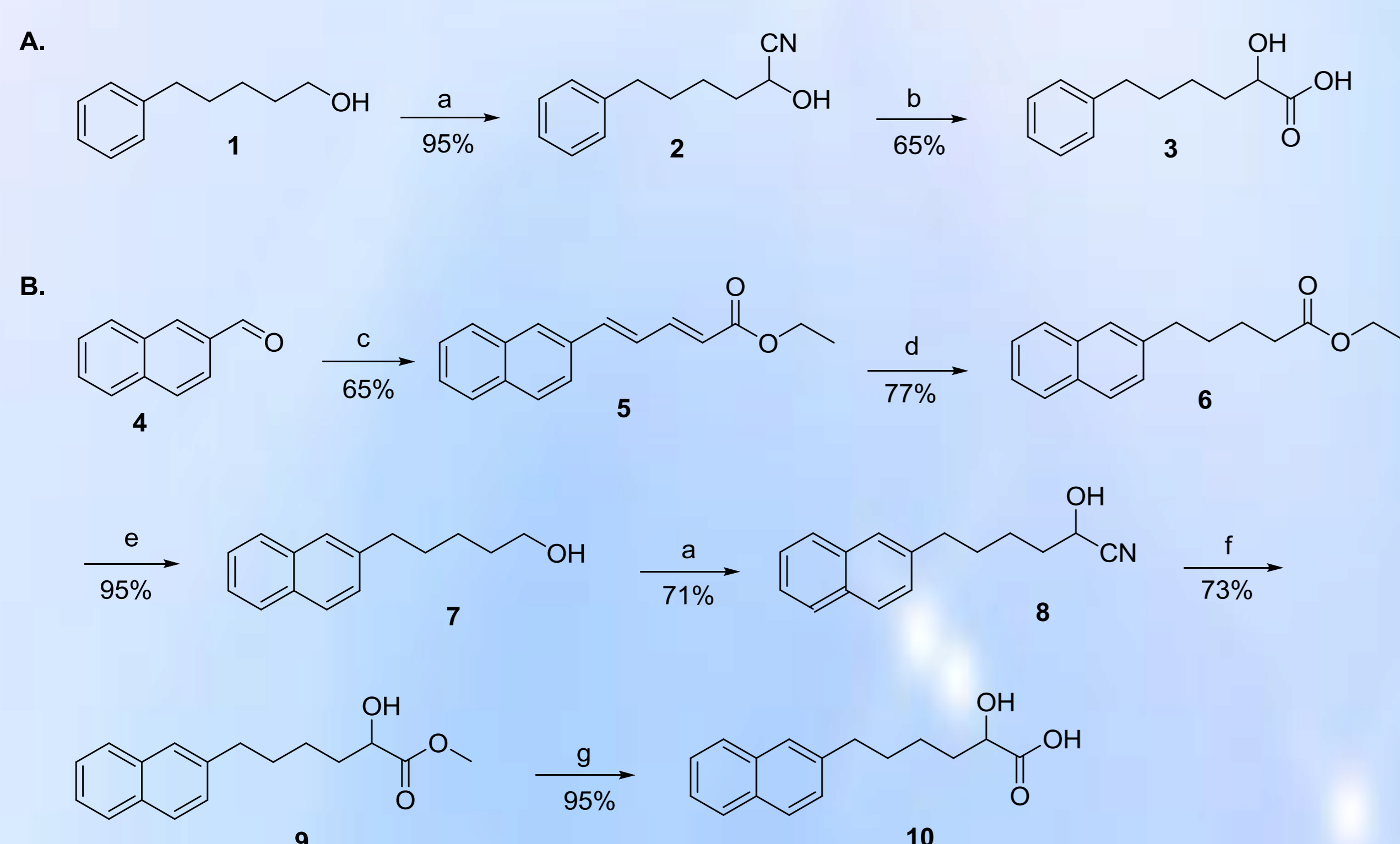
^aChemical Laboratories, Agricultural University of Athens, Athens 11855, Greece; ^bLaboratory of Organic Chemistry, Department of Chemistry, University of Athens, Panepistimiopolis, Athens 15771, Greece; ^cDepartment of Chemistry and Biochemistry and Department of Pharmacology, School of Medicine, University of California, San Diego, La Jolla, California 92093-0601, USA.

Introduction

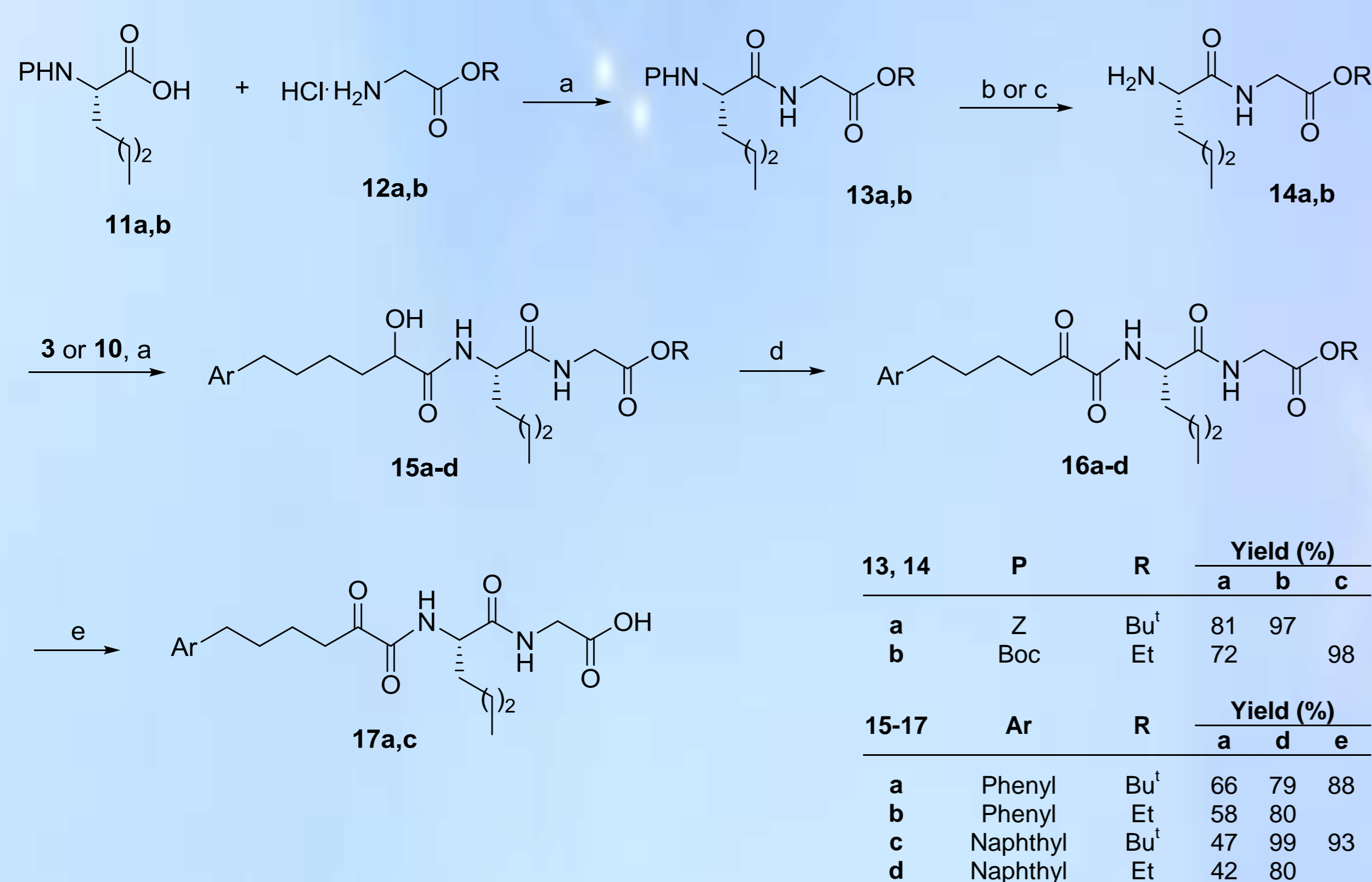
Phospholipases A₂ (PLA₂) catalyze the hydrolysis of the *sn*-2 ester bond of glycerophospholipids producing free fatty acids and lysophospholipids. The free arachidonic acid (AA) that is released may be converted to a variety of proinflammatory eicosanoids. Therefore, inhibiting AA release is of great therapeutic relevance for the development of new anti-inflammatory drugs. Moreover, selective inhibition of the less studied enzyme of the three main categories, GVIA iPLA₂, may offer valuable information about the enzyme's physiological role. A new class of PLA₂ inhibitors has been developed by our group: long chain 2-oxoamides based on amino acids.¹⁻³ Among others, we have shown that two molecules based on a pseudodipeptide ethyl ester and a dipeptide *tert*-butyl ester are the first 2-oxoamide derivatives, which preferentially inhibit GVIA iPLA₂ with X₁(50) values of 0.017 and 0.011 μM, respectively.⁴ To extend our studies, we synthesized a variety of 2-oxoamides based on dipeptides and ether pseudodipeptides, bearing an aryl terminal group on their 2-oxoamide aliphatic chain and we studied their *in vitro* activity on three human PLA₂ classes: GIVA cPLA₂, GVIA iPLA₂ and GV sPLA₂.

Synthesis

As starting materials for the synthesis of 2-hydroxy-acids we used 5-phenyl-1-pentanol (**1**) and 2-naphthaldehyde (**4**). Alcohol **1** was oxidized to aldehyde using the AcNH-TEMPO method, which was treated with NaHSO₃/KCN to provide the corresponding cyanhydrin **2**. The latter was converted to the 2-hydroxy-acid **3** after treatment with condensed hydrochloric acid and subsequently with potassium hydroxide in a solution of ethanol/water. 2-Naphthaldehyde (**4**) underwent a Horner – Wadsworth – Emmons olefination, in order to extend the carbon chain, followed by catalytic hydrogenation. The resulting ester **6** was reduced to the corresponding alcohol **7** using DIBALH. Following the same chemical steps as previously, cyanhydrin **8** was obtained, which was converted to the 2-hydroxy-methylester **9** after treatment with methanolic hydrochloride. The desired 2-hydroxy-acid **10** was produced after saponification (Scheme 1). *N*-Protected L-norleucines **11a,b** and glycine esters **12a,b** were coupled using WSCI.HCl as a condensing agent in the presence of HOBt to provide the desired dipeptides (**13a,b**) (Scheme 2), while the pseudodipeptide **20** was obtained after reaction of L-norleucinol with *tert*-butyl bromoacetate under phase transfer conditions (Scheme 3).

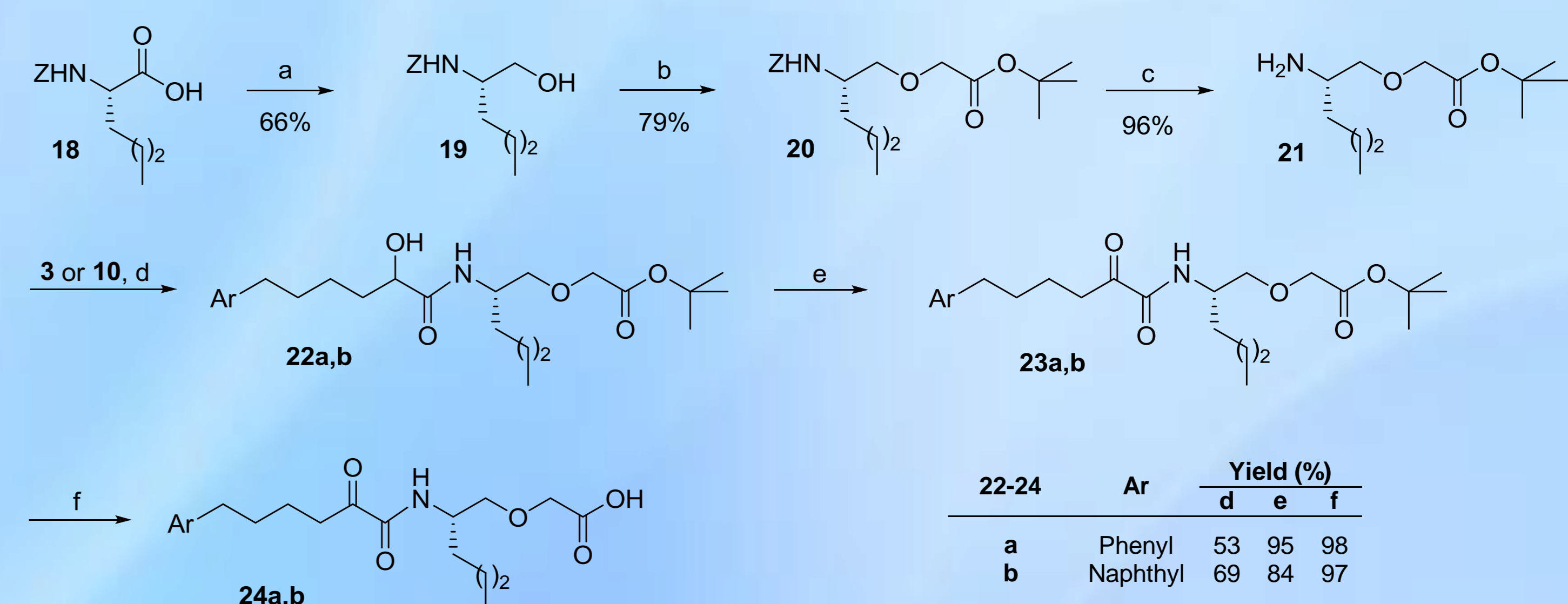


Scheme 1. Reagents and conditions: a) i. NaOCl, NaBr, AcNH-TEMPO, NaHCO₃, AcOEt/toluene/H₂O 3:3:0.5, -5 °C; ii. NaHSO₃, KCN, CH₂Cl₂, H₂O; b) i. conc. HCl, ii. KOH, EtOH/H₂O; c) C₂H₅OOCCH=CHCH₂P(=O)(OC₂H₅)₂, LiOH, THF, reflux; d) H₂, 10% Pd/C, EtOH; e) DIBALH, anhydrous Et₂O; f) 3N HCl/MeOH; g) NaOH 1N, MeOH.



Scheme 2. Reagents and conditions: a) WSCI, HOBt, Et₃N, CH₂Cl₂; b) H₂, 10% Pd/C, THF; c) 4N HCl/Et₂O; d) Dess-Martin periodinane, CH₂Cl₂; e) 50% TFA/CH₂Cl₂.

Coupling reaction between the free amino group of the dipeptides or the ether pseudodipeptide and the 2-hydroxy acid took place in the presence of WSCI/HOBt and the resulting 2-hydroxy amides **15a-d** and **22a,b** were oxidized to 2-oxoamides **16a-d** and **23a,b** using the Dess-Martin method. Oxoamides **17a,c** and **24a,b** containing a free carboxyl group, were obtained by treatment of **16a,c** and **23a,b** with trifluoroacetic acid (Schemes 2 and 3).



Scheme 3. Reagents and conditions: a) i. NMM, ClCOOEt, THF; ii. NaBH₄, CH₃OH; b) BrCH₂COOBu^t, 50% NaOH, Bu₄NHSO₄, benzene; c) H₂, 10% Pd/C; d) WSCI, HOBt, Et₃N, CH₂Cl₂; e) Dess-Martin periodinane, CH₂Cl₂; f) 50% TFA/CH₂Cl₂.

Table 1. *In vitro* results of the human phospholipases A₂ inhibition by 2-oxoamides.

Compound	Structure	% Inhibition		
		cPLA ₂	iPLA ₂	sPLA ₂
16a		46.9	91.1	43.4
16c		57.5	89.1	44.8
16d		70.5	77.1	54.4
17a		11.2	-	-
17c		43.1	-	-
23a		82.3	76.0	57.5
23b		85.5	88.4	63.9
24 ^a		72	X ₁ (50) 0.011	59
25 ^a		52	X ₁ (50) 0.017	81

^a Data taken from Ref. 4

Results and discussion

Preliminary PLA₂ inhibition results are summarized in Table 1. Derivatives **24** and **25** have been also included for comparison reasons. Among the seven compounds tested, **16a** and **16c**, based on Nle-Gly *tert*-butyl ester, bearing phenyl and naphthyl side chain terminal moieties respectively, appear to inhibit preferentially GVIA iPLA₂. This observation is in full agreement with our previous report, that the 2-oxoamide derivative based on Nle-Gly *tert*-butyl ester (**24**) is potent and selective inhibitor of GVIA iPLA₂.⁴ The other two *tert*-butyl esters of this series, namely **23a** and **23b**, based on ether pseudodipeptides, exhibited considerable but lower activity towards GVIA iPLA₂, though they inhibit also GIVA cPLA₂ to a similar level. 2-Oxoamide derivatives containing free carboxylic acids (**17a** and **17c**) do not inhibit GVIA iPLA₂.

References

- Stephens, D.; Barbayianni, E.; Constantinou-Kokotou, V.; Peristeraki, A.; Six, D. A.; Cooper, J.; Harkewicz, R.; Deems, R. A.; Dennis, E. A.; Kokotos, G. *J. Med. Chem.* **2006**, *49*, 2821.
- Six, D. A.; Barbayianni, E.; Loukas, V.; Constantinou-Kokotou, V.; Hadjipavlou-Litina, D.; Stephens, D.; Wong, A. C.; Magrioti, V.; Moutevelis-Minakakis, P.; Baker, S.; Dennis, E. A.; Kokotos, G. *J. Med. Chem.* **2007**, *50*, 4222.
- Antonopoulou, G.; Barbayianni, E.; Magrioti, V.; Cotton, N.; Stephens, D.; Constantinou-Kokotou, V.; Dennis, E. A.; Kokotos, G. *Bioorg. Med. Chem.* **2008**, *16*, 10257.
- Barbayianni, E.; Stephens, D.; Grkovich, A.; Magrioti, V.; Hsu, Y.-H.; Cotton, N.; Dolatzas, P.; Kalogiannidis, D.; Dennis, E. A.; Kokotos, G. *Bioorg. Med. Chem.* **2009**, *17*, 4833.

Acknowledgements

This research has been co-financed by the European Union (European Social Fund – ESF) and Greek national funds through the Operational Program "Education and Lifelong Learning" of the National Strategic Reference Framework (NSRF) - Research Funding Program: Heracleitus II. Investing in knowledge society through the European Social Fund.

