

Total phenolic compounds, antioxidant activity and toxicity of selected medicinal and aromatic plants

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The importance of dietary antioxidant components for the prevention of some diseases and health quality improvement has attracted much research attention through the last decade. Vegetables and herbal infusions have been recognized as important antioxidant sources. Food industry shows significant interest in application of plant bioactive compounds for flavoring but also for preservation purposes, but attention should be given in case of high doses. In the current study were investigated the total phenolic content, the antioxidant activity and the toxicity of selected medicinal aromatic plants that are being consumed as decoctions or used as food additives.

Sample preparation

2 g dry mass of each plant species were:
a) steeped in 200 ml (1 cup) a) at 85 °C for 15 min,

b) at room temperature for 15 min and,

c) at room temperature with the assistance of ultrasound (35 MHz) for 15 min. The herbal infusions were then filtered through a Whatman filter No. 1.

All extracts were then further extracted by petroleum ether.

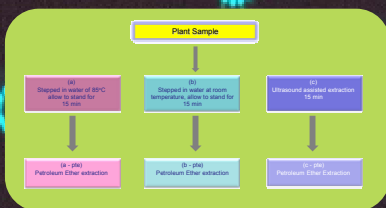


Fig 1. Schematic Plan of extraction procedures

Total phenolic content and antioxidant activity

Total phenolic content (in terms of caffeic acid) was determined using a Folin-Ciocalteu assay. Antioxidant activity determined applying both the ABTS assay - 2,2-azino-bis-(3-ethylbenzothiazoline-6-sulfonic acid), and the DPPH assay 2,2-diphenyl-1-picrylhydrazyl radical.

Toxicity tests

Toxicity assessment of the plant extracts has been performed using MICROTOX® toxicity analyzer. MICROTOX® uses the marine bioluminescence bacteria *Vibrio fischeri* as a reference test species for the measurement of bioluminescence inhibition. The protocol used was the Basic Test 81.9% (AZUR, 1997) with appropriate primary dilution of samples in order the EC₅₀ (Effective Concentration where bioluminescence inhibition is 50%) not to be obtained by extrapolated data.

Table 1. Name and use of plant species examined

Code	Species	Common name	Plant part used	Use in diet
MOF	<i>Melissa officinalis</i>	Lemon balm	leaves	Decoctions
OVU	<i>Origanum vulgare</i>	Oregano	leaves	Herb in salads (fresh or dry), roasted, stewed
ODI	<i>Origanum dictamnus</i>	Dittany	Leaves/flowers	Decoctions
SOF	<i>Salvia officinalis</i>	Sage	Leaves	Decoctions, as a herb in cooking
HOF	<i>Hyssopus officinalis</i>	Hyssop	Leaves/flowers	Decoctions

Table 2. Toxicity of plant species expressed as EC₅₀ (mg/ml) after 15 minutes incubation time

Sample	Treatments					
	(a)	(a-pte) ^{*1}	(b)	(b-pte) ^{*1}	(c)	(c-pte) ^{*1}
MOF	119.75	210.68	200.61	216.30	113.09	126.93
HOF	38.31	139.60	150.67	162.75	96.05	101.47
ODI	10.56	42.87	60.46	110.75	62.61	96.87
OVU	8.61	36.38	40.96	52.32	22.09	77.69
SOF	52.17	107.77	199.45	198.78	137.19	224.87

Table 3. Total Phenolic Content, Antioxidant Activity estimated by DPPH and ABTS method

Sample Code	Treatment	Antioxidant Activity DPPH		Antioxidant Activity ABTS	
		mg Caffeic acid / 200 ml	µmole Trolox / 200 ml	µmole Trolox / 200 ml	µmole Trolox / 200 ml
MOF	<i>Melissa officinalis</i>	197.0	1268.3	1321.7	
HOF	<i>Hyssopus officinalis</i>	34.9	206.4	206.8	
OVU	<i>Origanum vulgare</i>	128.6	631.1	669.3	
ODI	<i>Origanum dictamnus</i>	63.9	299.7	321.0	
SOF	<i>Salvia officinalis</i>	64.5	328.2	326.1	
MOF	<i>Melissa officinalis</i>	185.9	1240.1	1319.5	
HOF	<i>Hyssopus officinalis</i>	31.7	182.5	190.2	
OVU	<i>Origanum vulgare</i>	105.1	565.2	595.2	
ODI	<i>Origanum dictamnus</i>	51.3	227.7	311.9	
SOF	<i>Salvia officinalis</i>	62.0	306.9	323.1	
MOF	<i>Melissa officinalis</i>	135.8	644.5	766.8	
HOF	<i>Hyssopus officinalis</i>	10.7	51.5	69.2	
OVU	<i>Origanum vulgare</i>	63.5	290.1	309.5	
ODI	<i>Origanum dictamnus</i>	28.3	153.5	169.6	
SOF	<i>Salvia officinalis</i>	15.5	77.3	99.2	
MOF	<i>Melissa officinalis</i>	119.1	605.5	684.3	
HOF	<i>Hyssopus officinalis</i>	10.4	39.2	71.0	
OVU	<i>Origanum vulgare</i>	53.9	262.8	267.1	
ODI	<i>Origanum dictamnus</i>	27.9	123.7	139.1	
SOF	<i>Salvia officinalis</i>	15.5	66.3	89.6	
MOF	<i>Melissa officinalis</i>	159.9	752.7	871.8	
HOF	<i>Hyssopus officinalis</i>	13.1	49.7	85.2	
OVU	<i>Origanum vulgare</i>	68.3	323.5	344.5	
ODI	<i>Origanum dictamnus</i>	34.5	177.0	179.6	
SOF	<i>Salvia officinalis</i>	16.9	83.7	123.1	
MOF	<i>Melissa officinalis</i>	114.7	676.0	834.9	
HOF	<i>Hyssopus officinalis</i>	15.0	55.7	86.1	
OVU	<i>Origanum vulgare</i>	57.6	276.3	288.7	
ODI	<i>Origanum dictamnus</i>	32.9	155.5	160.3	
SOF	<i>Salvia officinalis</i>	28.0	68.6	107.9	

Table 4. Concentration that cause 20% luminescence inhibition and maximum dry plant mass per reference volume

Code	Treatment	Concentration that cause 20% luminescence inhibition mg/ml	Plant mass per reference volume that cause 20% luminescence inhibition (g)	Reference volume (ml)	Use in diet
MOF	(b)	92.33	18.47	200	Decoction
	(a)	48.00	9.60	200	
OVU	(b)	12.95	6.48	500	In salads, appetizers and meals
	(a)	3.29	0.66	200	
ODI	(a)	16.5	16.5	5000	During cooking process
	(b)	21.03	4.21	200	
SOF	(a)	4.04	0.81	200	Decoction
	(b)	83.79	16.76	200	
HOF	(a)	18.95	3.79	200	Decoction
	(b)	94.75	13.02	5000	
HOF	(a)	65.10	13.02	200	Decoctions
	(b)	11.90	2.38	200	

Results

Total phenolic content was estimated in both infusions and the aqueous phase of their extraction by petroleum ether, expressed in mg of caffeic acid / 200 ml. Maximum values were given by *Melissa officinalis*. The ranking of antioxidant activity regardless of the method that is being determined DPPH or ABTS is the same with the ranking of the total phenolic content. Results show that there is a scaling increase in total phenolic content among the species tested. *Melissa officinalis* exhibited the higher phenolic content and *Hyssopus officinalis* the lowest regardless the extraction procedure. Ranking remains the same in the aquatic phase of the extraction by petroleum ether for all the infusions (table 2) tested.

Antioxidant activity when determined by ABTS method expressed also µmole Trolox / 200 ml (table 2). Maximum values were given by *Melissa officinalis*. The ranking of antioxidant activity regardless of the method that is being determined DPPH or ABTS is the same with the ranking of the total phenolic content.

Toxicity evaluation of the plant extracts was performed by applying the Basic Test protocol obtained after 15 min exposure expressed as EC₅₀ value, at which 50% loss of luminescence is obtained. EC₅₀ was also estimated, at which a 20% loss of light emission is observed, as according to the ISO guidelines (ISO 11348, 1998) a toxic sample shows an effect percentage greater than 20%. Results show that *Origanum vulgare* gave the lower EC₅₀ values in all treatments. Respectively, higher values were obtained by *Melissa officinalis* in treatments (a), (a-pte), (b), (b-pte), and by *Salvia officinalis* in treatments (c) and (c-pte) (table 3).

Depending on the use of each plant tested in nutrition, was calculated the maximum dry plant mass, in 200 ml for decoctions, in 500ml in case of use in salads, and in 5l when is being used in cooking processes (table 4). Furthermore, was calculated the synergism ratio (SR) as follows:

$$SR_{\text{total phenolics}} = \frac{\text{Total phenolics} - \text{pte}}{\text{Total phenolic mixture}}$$

$$SR_{\text{antioxidant activity}} = \frac{\text{Antioxidant activity} - \text{pte}}{\text{Antioxidant activity mixture}}$$

$$SR_{\text{toxicity}} = \frac{EC_{50, \text{pte}}}{EC_{50, \text{mixture}}}$$

Table 5. SR_{total phenolics}, SR_{antioxidant activity}, SR_{toxicity} indexes for sample tested

Sample Code	Treatment	SR total phenolics	Effect ^{*1}	SR DPPH	Effect ^{*1}	SR ABTS	Effect ^{*1}	SR toxicity ^{*2}	Effect
MOF	(b)	0.9	Synergism	0.9	Synergism	0.9	Synergism	1.1	Synergism
	(c)	0.7	Synergism	0.9	Synergism	1.0	Additive	1.1	Synergism
HOF	(a)	0.9	Synergism	1.0	Additive	1.0	Additive	1.8	Synergism
	(b)	1.0	Additive	0.8	Synergism	1.0	Additive	1.1	Synergism
HOF	(c)	1.1	Antagonism	1.1	Antagonism	1.0	Additive	1.1	Synergism
	(a)	0.9	Synergism	0.9	Synergism	0.9	Synergism	3.6	Synergism
ODI	(b)	1.0	Additive	0.8	Synergism	0.8	Synergism	1.8	Synergism
	(c)	1.0	Additive	0.9	Synergism	0.9	Synergism	1.5	Synergism
OVU	(a)	0.8	Synergism	0.8	Synergism	1.0	Additive	4.1	Synergism
	(b)	0.8	Synergism	0.9	Synergism	0.9	Synergism	1.3	Synergism
OVU	(c)	0.8	Synergism	0.9	Synergism	0.8	Synergism	3.5	Synergism
	(a)	0.8	Synergism	0.9	Synergism	0.9	Synergism	4.2	Synergism
SOF	(b)	1.0	Additive	0.9	Synergism	0.9	Synergism	1.0	Additive
	(c)	1.7	Antagonism	0.8	Synergism	0.9	Synergism	1.6	Synergism
SOF	(a)	1.0	Additive	0.9	Synergism	1.0	Additive	2.1	Synergism

*1: In case of Total Phenolic Content and Antioxidant Activity, SR<1 indicates synergism, SR>1 indicates antagonism and SR = 1 indicates additive action

*2: In case of toxicity, SR<1 indicates antagonism, SR<toxicity> 1 indicates synergism and SR = 1 indicates additive action

Conclusions. Results of this study indicate that between plant species examined, *Melissa officinalis* presented the higher, and *Hyssopus officinalis* the lower phenolic content and antioxidant activity. Extraction procedure seems to influence significantly the extraction of phenolic compounds and antioxidant activity respectively as great differences were observed between extractions in high and room temperature. A small increase was observed in ultrasound assisted extraction. Similar results were obtained when infusions extracted by petroleum ether. Linear positive correlation has been found between total phenolic content and antioxidant activity in the infusions studied.

Toxicity of infusions seems to be also influenced by the temperature and extraction procedure, but no correlation was detected between toxicity and the total phenolic content or antioxidant activity. Interactions of soluble substances and the essential oil of plants were remarkable in case of toxicity in contrast to the case of total phenolic content and antioxidant activity where interaction was almost neutral.

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This research has been co-financed by the European Union (European Social Fund – ESF) and Greek national funds through the Operational Program "Education and Lifelong Learning" of the National Strategic Reference Framework (NSRF) – Research Funding Program: Heracleitus II. Investing in knowledge society through the European Social Fund.

