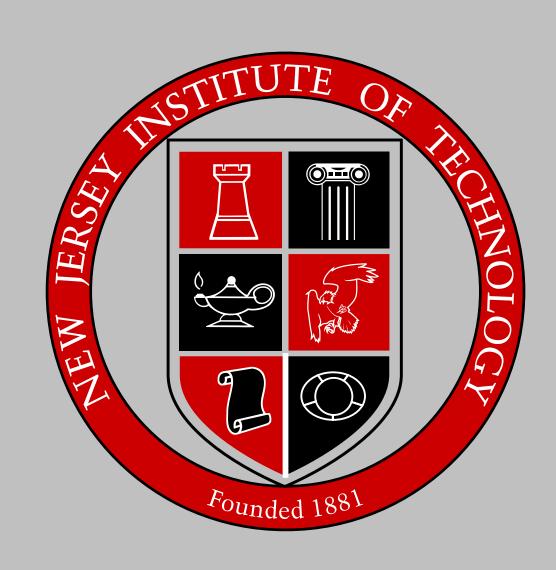


# Novel methodology for the investigation of Dmrt3a interneurons in larval zebrafish

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### Background

Understanding the neural circuits that underlie locomotion is key to understanding the causes of motor behaviors and disorders. This project explores the role of spinal interneurons (Fig. 1), linked to the gene Dmrt3a, in larval zebrafish. Past studies suggest that these interneurons are linked to speed-shifting and locomotor coordination.

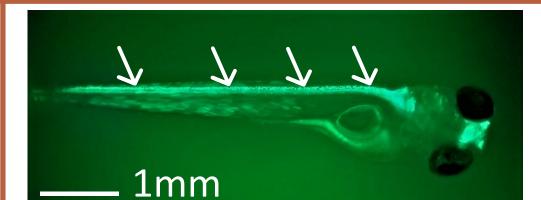


Figure 1. Dmrt3a-HS:Gal4;
Botox-GFP;nacre -/- zebrafish
larvae taken under Olympus
MVX10 disection scope at
6.3x. White arrows indicate
Dmrt3a neurons

## **Preliminary Results**

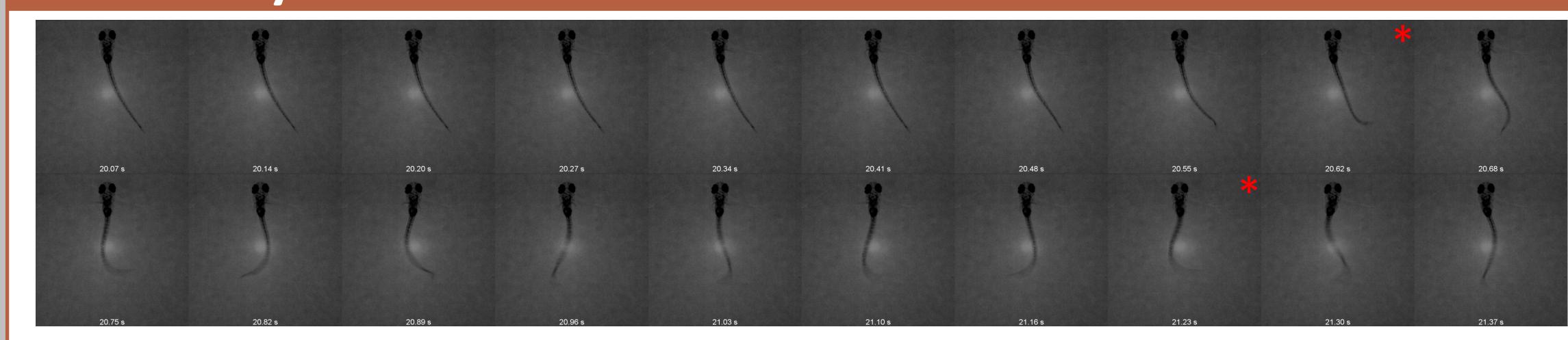


Figure 4. Example of coordinated motion in AB wildtype fish across gaits. This high speed footage shows both independant and coordinated motion of tail and fins. Of note are possible gait transitions at the marked (\*) frames.

#### Methods

#### **Control:**

8 AB wildtype 6 days post fertilization (dpf) zebrafish **Experimental:** 

6 Dmrt3a-HS:Gal4;Botox-GFP;nacre -/- 6 dpf zebrafish

#### Preparation:

Zebrafish larvae were **head-embeded** in 2% low melting point agarose

#### Video Capture:

Fish were stimulated to swim using a projected OMR

grating at speeds 10mm/s, 20mm/s, and 30mm/s

Video captured on a custom experimental rig (Fig. 2)

#### Analysis:

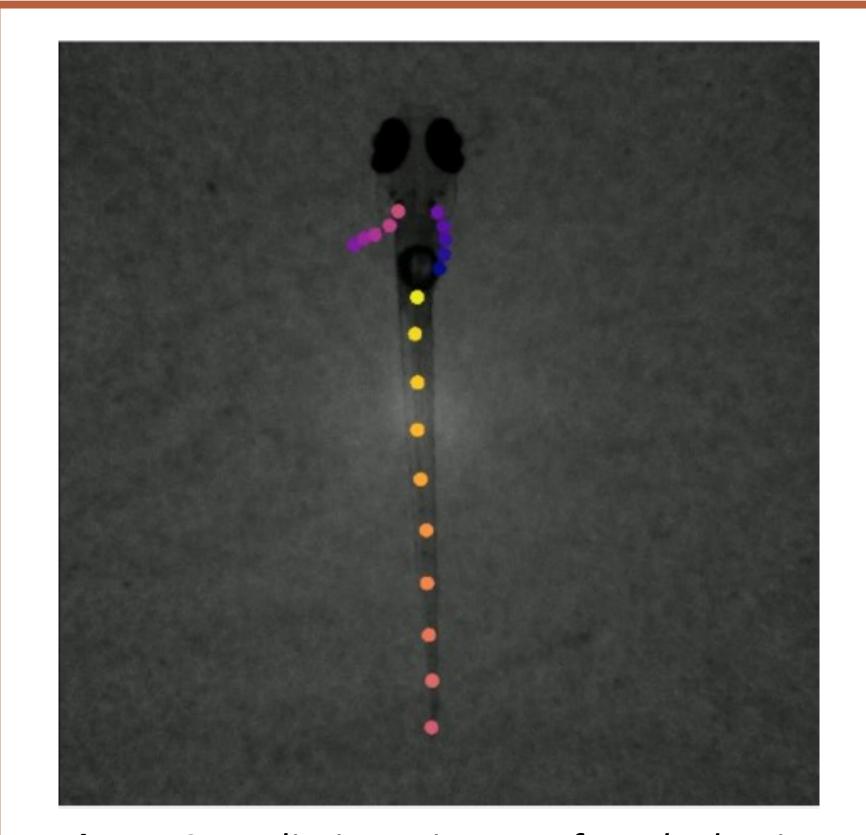
Raw video files were converted from .tiff to uncompressed .AVI

DeepLabCut was used to track points along the bilateral pec fin and tails

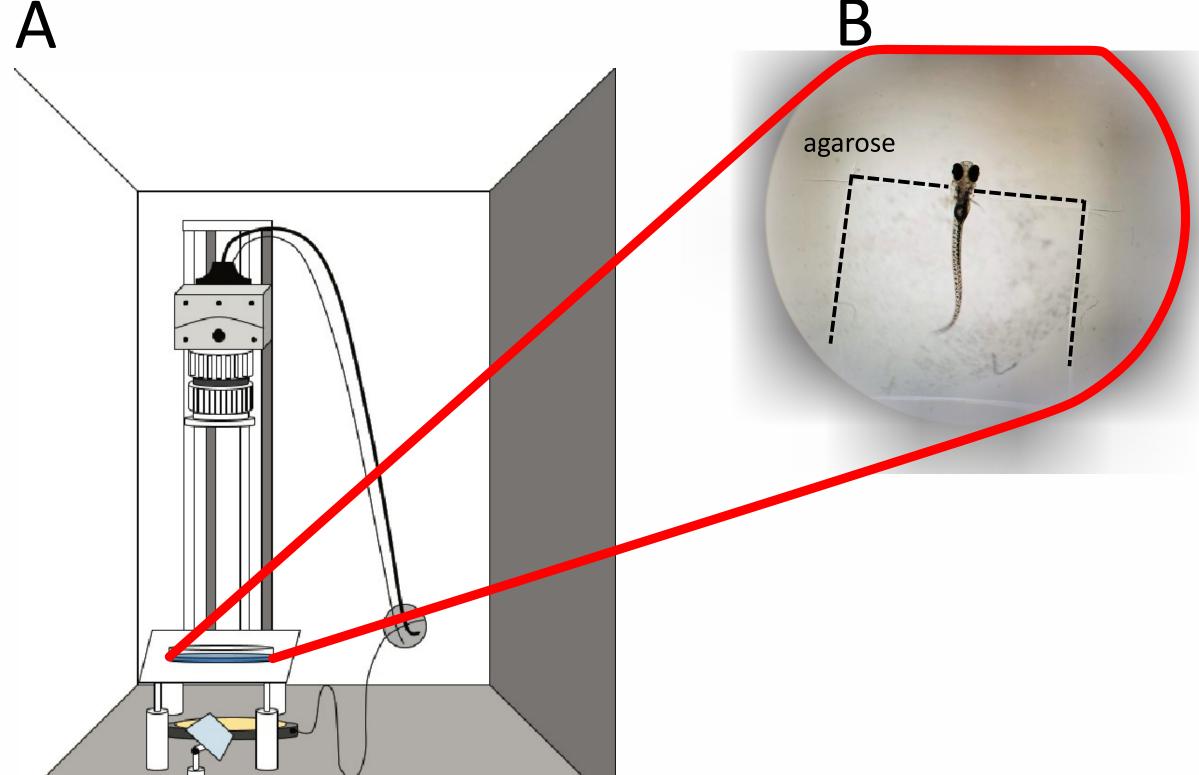
Data was processed and analyized in Python and R

# Tail and Fin Bend Angles Tail and Fin Tracing Body Part Right Fin Tail and Fin Tracing 450 400 400 250 250 1000 2000 3000 4000 5000

**Figure 5.** Comparison of bend angles from one AB wildtype trial at 20 mm/s (A). Comparison of tail and fin tracings from the same fish (B). In this wildtype example, fins and tail are coordinated in "bouts" of motion.



**Figure 6.** Preliminary image of tracked points along tail and pec fins



**Figure 2.** Behavior rig schematic (A) with insert of head embedded zebrafish (B)



Figure 3. Dmrt3a-HS:Gal4;Botox-GFP;nacre
-/- 6 dpf zebrafish larvae pictured with
light (A) and flourescent (B) microscopy.
All control and experimental fish were
screened using Olympus MVX10 disection
scope at 6.3x.

# Discussion & Future Directions

Preliminary data indicates that there may be differences in coordination between AB wildtype fish and Dmrt3a-Botox fish. In order to further explore our hypothesis and confirm the presence of any statistically significant differences several steps are being taken:

- 1. In order to make meaningful comparisions between fish and trials, individual bouts of movement must be extracted. This is currently in progress.
- 2. Additional experiments utilizing channelrhodopsin2 (ChR2) in trangenic fish are planned. These experiments will use a similar head embedded process to allow selective stimulation of Dmrt3a neurons with a fiberoptic light source. This setup will allow for more precise control Dmrt3a activity.
- 3. Data from free swimming behavioral experiments is currently being analyzed

Use of DLC has allowed for more extensive tracking over more frames and trials then would be practical if working by hand. We are currently exploring how to make use of this broad data set and how to take advantage of machine-learning assisted data collection in future experiments.

# Acknowledgements

Dmrt3a line courtesy of Shin-Ichi Higashijima's lab. Botox-GFP line provided by Clair Wyart's lab