



Ndung'u, P. W., Takahashi, T., du Toit, C. J. L., Robertson-Dean, M., Butterbach-Bahl, K., McAuliffe, G. A., Merbold, L., & Goopy, J. P. (2022). Farm-level emission intensities of smallholder cattle (*Bos indicus*; *B. indicus*–*B. taurus* crosses) production systems in highlands and semi-arid regions. *animal*, 16(1), [100445].
<https://doi.org/10.1016/j.animal.2021.100445>

Publisher's PDF, also known as Version of record

License (if available):
CC BY-NC-ND

Link to published version (if available):
[10.1016/j.animal.2021.100445](https://doi.org/10.1016/j.animal.2021.100445)

[Link to publication record in Explore Bristol Research](#)
PDF-document

This is the final published version of the article (version of record). It first appeared online via Elsevier at <https://doi.org/10.1016/j.animal.2021.100445>. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
<http://www.bristol.ac.uk/red/research-policy/pure/user-guides/ebr-terms/>



Farm-level emission intensities of smallholder cattle (*Bos indicus*; *B. indicus*–*B. taurus* crosses) production systems in highlands and semi-arid regions



P.W. Ndung'u^{a,b}, T. Takahashi^{c,d}, C.J.L. du Toit^a, M. Robertson-Dean^e, K. Butterbach-Bahl^f, G.A. McAuliffe^c, L. Merbold^{b,g}, J.P. Goopy^{a,b,*}

^a Department of Animal and Wildlife Sciences, University of Pretoria, 0002, South Africa

^b Mazingira Centre, International Livestock Research Institute, P.O. Box 30709, Nairobi, Kenya

^c Rothamsted Research, North Wyke, Okehampton, Devon EX20 2SB, UK

^d University of Bristol, Langford House, Langford, Somerset BS40 5DU, UK

^e School of Mathematics, University of New England, Armidale, NSW, Australia

^f Institute for Meteorology and Climate Research, Atmospheric Environmental Research, Karlsruhe Institute of Technology, D-82467 Garmisch-Partenkirchen, Germany

^g Agroscope, Research Division Agroecology and Environment, Reckenholzstrasse 191, 8046 Zurich, Switzerland

ARTICLE INFO

Article history:

Received 30 June 2021

Revised 9 December 2021

Accepted 10 December 2021

Available online 10 January 2022

Keywords:

African livestock system

Carbon footprint

CP

Life cycle assessment

Primary data

ABSTRACT

Ruminants are central to the economic and nutritional life of much of sub-Saharan Africa, but cattle are now blamed for having a disproportionately large negative environmental impact through emissions of greenhouse gas (GHG). However, the mechanism underlying excessive emissions occurring only on some farms is imperfectly understood. Reliable estimates of emissions themselves are frequently lacking due to a paucity of reliable data. Employing individual animal records obtained at regular farm visits, this study quantified farm-level emission intensities (EIs) of greenhouse gases of smallholder farms in three counties in Western Kenya. CP was chosen as the functional unit to capture the outputs of both milk and meat. The results showed that milk is responsible for 80–85% of total CP output. Farm EI ranged widely from 20 to >1 000 kg CO₂-eq/kg CP. Median EIs were 60 (Nandi), 71 (Bomet), and 90 (Nyando) kg CO₂-eq/kg. Although median EIs referenced to milk alone (2.3 kg CO₂-eq/kg milk) were almost twice that reported for Europe, up to 50% of farms had EIs comparable to the mean Pan-European EIs. Enteric methane (CH₄) contributed >95% of emissions and manure ~4%, with negligible emissions attributed to inputs to the production system. Collecting data from individual animals on smallholder farms enabled the demonstration of extremely heterogeneous EI status among similar geographical spaces and provides clear indicators on how low EI status may be achieved in these environments. Contrary to common belief, our data show that industrial-style intensification is not required to achieve low EI. Enteric CH₄ production overwhelmingly drives farm emissions in these systems and as this is strongly collinear with nutrition and intake, an effort will be required to achieve an “efficient frontier” between feed intake, productivity, and GHG emissions.

© 2021 The Author(s). Published by Elsevier B.V. on behalf of The Animal Consortium. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Implications

Smallholder cattle production is an important activity in East Africa, but emissions of these farms are thought to be high but not well understood due to a lack of on-farm data. Enteric methane drives on-farm greenhouse gas emissions in smallholder cattle farms, which have great mitigation potential, without the need

for industrial-style intensification. Identification and adoption of practices associated with low emission intensities will facilitate better resource use efficiency and lower greenhouse gas emissions at a regional level.

Introduction

Livestock plays a crucial role in the social and economic growth of Africa (Herrero et al., 2013). Driven by population increase, and improving the gross domestic product, and household incomes (Steinfeld et al., 2006), demand for livestock products is rapidly

* Corresponding author at: Department of Animal and Wildlife Sciences, University of Pretoria, Hatfield, Pretoria, 0002, South Africa.

E-mail address: manofcows@yahoo.com (J.P. Goopy).

growing (Thornton, 2010). The consumption of beef and milk is forecast to increase by 261 and 399%, respectively, between 2010 and 2050 (FAO, 2017). Simultaneously, the supply of livestock products in Africa is constrained by competition with other sectors for scarce natural resources, suboptimal husbandry practices, and unreliability in supply and quality of feed (Thornton, 2010; Nkonya et al., 2016). These conditions are putatively responsible for the characteristically high proportion of regional anthropogenic greenhouse gas (GHG) emissions attributed to animal agriculture (25 compared to 14.5% globally) (Gerber et al., 2011; Gerber et al., 2013). The average carbon footprint of fat- and protein-corrected milk (FPCM) in Africa is estimated to be 7.5 kg CO₂-eq/kg FPCM (Poore & Nemecek, 2018), while the corresponding global mean is ~3.2 kg CO₂-eq/l, leading Opio et al. (2013) to conclude that Sub-Saharan Africa has the least efficient dairy production systems in the world when measure assessed by climate impacts. To date, the accuracy of these estimates has not been confirmed. Additionally, the mechanism behind what drives some farms to pollute more than others has not been elucidated. This is principal because most GHG inventories in Africa have been collated using the Intergovernmental Panel on Climate Change (IPCC) default (Tier I) emission factors (EFs), which results in an annual estimate of GHG emissions per capita by animal class, ignoring (among other factors) variability in production efficiency between individual animals and enterprise management. While this approach is generally necessitated by a lack of detailed field data, it results in a large degree of uncertainty in the presence of seasonality and variability in animal phenotype and feed baskets, conditions almost invariably present in the smallholder context (Herrero et al., 2013; Goopy et al., 2018). African countries where livestock are an important source of GHGs are now committed to quantifying their own EFs, at both national and finer spatial scales (Lee et al., 2017; Ndao et al., 2019), with the objective of providing improved reporting to the United Nations Framework Convention on Climate Change (UNFCCC) following the Paris Climate Agreement.

Several recently completed studies have begun to address the challenge for African countries namely, South Africa (du Toit et al., 2013a; 2013b), Benin (Kouazounde et al., 2015), Kenya (Pelster et al., 2016; Goopy et al., 2018; Zhu et al., 2018; Ndung'u et al., 2019; Goopy et al., 2021; Leitner et al., 2021; Ndung'u et al., 2021), and Senegal (Ndao et al., 2019; Ndao et al., 2020). Nonetheless, accurate estimation of EFs alone does not capture the entire variability in emission impacts across smallholder farms (Goopy et al., 2018; Ndung'u et al., 2019), because in situations where productivity also varies, a farm's overall GHG performance is better assessed by employing emission intensity (EI) (Moran & Wall, 2011) under the life cycle assessment (LCA) framework. This view is particularly pertinent to agricultural systems where the presence of unproductive livestock held for a variety of non-economic reasons has been suggested as a major cause of large on-farm emissions (Weiler et al., 2014). Paradoxically, it has been claimed that these systems are also the ones with the greatest potential to mitigate GHG emissions via increased productivity, and thus are among the most important to critically examine (Parry et al., 2007).

In LCA, environmental burdens such as GHG emissions are referenced to a functional unit (FU) that is the quantity of an output representing the purpose of the system. For livestock systems, the FU has commonly been set as FPCM (Opio et al., 2013; O'Brien et al., 2015; Garg et al., 2016; Rice et al., 2017) or energy-corrected milk (Rotz et al., 2010; Knapp et al., 2014; O'Brien et al., 2014; Ross et al., 2017; Rotz, 2018) in dairy enterprises, or as carcass weight (Rotz et al., 2019), live weight (Desjardins et al., 2012; Opio et al., 2013; Legesse et al., 2016) or live weight gain (McAuliffe et al., 2018a) in beef enterprises. However, the

use of different FUs has been shown to have a profound effect on the EI of a given system (McAuliffe et al., 2018b and 2020), often resulting in multiple and mutually contradictory EIs and arguably, confusion (Weiler et al., 2014). Attempting to resolve this issue, Ross et al. (2017) assessed the suitability of different FUs in a dairy enterprise, finding that energy-corrected milk was generally the most robust measure. However, this conclusion was based on studies conducted in developed countries where comprehensive data-bases are available. In contrast, livestock systems in developing economies typically have multiple functions from a single enterprise and single animals (multi-purpose system).

In Kenya, farming enterprises at a small scale are common throughout the highlands areas of Central and Rift Valley (Thorpe et al., 2000; World.Bank & CIAT, 2015). They are characterised by: (i) crop and livestock interdependence, (ii) small and fragmented land holdings (often < 2 ha) with dependence on access to common land, (iii) keeping a wide variety of livestock phenotypes (indigenous > indeterminate cross-bred > exotic), and (iv) having low inputs and low investment (Thorpe et al., 2000). Commonalities notwithstanding, the resulting system displays a good deal of heterogeneity through differences between farms in resources, production focus (subsistence, >commercial), and technical ability. Individual farms have multiple outputs, of which livestock is only one facet and not well understood.

Using animal-level data collected across multiple seasons on 313 smallholder mixed farms in Western Kenya, this study elucidates the distribution of farm-level EIs as well as their determinants. Although dairy farming is the most developed agricultural sub-sector in Kenya, unintuitively, it is predominantly supported by smallholders in rural areas (Muriuki, 2003). In particular, Western Kenya's Central and Rift Valley highland regions produce 60% of the country's milk supply (Muriuki, 2003), and their systems are representative of wider East Africa where livestock are an integral part of mixed agriculture.

Thus, we hypothesised that:

- (i) GHG EIs in smallholder livestock production systems in Western Kenya do not vary between (a) farms, (b) agro-ecological zones (AEZs) or (c) regions.
- (ii) The contribution of meat production is unimportant to overall farm output as measured by CP production, and
- (iii) EIs are similar to model-based estimates reported in the extant literature.

This work has been presented in a conference proceeding as Ndung'u et al. (2021).

Material and methods

Study sites

Data used in this study were collected from 313 smallholding farms located across three counties in Western Kenya: Nyando (56), Nandi (126), and Bomet (131). Collectively, the study region encompasses seven AEZs (refer to Table 1). Farms were selected randomly for each (county) study (see Supplementary Fig. S1), stratified by AEZ (for full detail refer to Goopy et al. (2018)). Data collection comprised five visits to each farm at an interval of 3 months between visits within 12 months at each site (i.e: 2014–2015 for Nyando, 2015–2016 for Nandi, and 2016–2017 for Bomet). This protocol also captured seasonal changes (local seasons: short rains in November to January; hot dry in February to April; long rains in May to July; and cold dry in August to October) in the feed basket and local pasture quality and abundance that was quantified through the use of harvesting from exclusion cages

Table 1
Description of Agro-ecological Zones where cattle in smallholder farms of Nandi, Bomet, and Nyando were sampled.

| Agro-Ecological Zone | Study Region (s) | Description | Mean Annual Temperature (°C) | Elevation range (metres above sea level) |
|------------------------|------------------|--------------------------------|------------------------------|--|
| Lower Highland 1 (LH1) | Nandi and Bomet | Moderately cool and humid | 15–18 | 1 800/1 900–2 200/2 400 |
| Lower Highland 2 (LH2) | Nandi and Bomet | Moderately cool and sub-humid | | |
| Lower Highland 3 (LH3) | Bomet | Moderately cool and semi-humid | | |
| Upper Midlands (UM) | Nandi and Bomet | Temperate | 18–21 | 1 300/1 500–1 800/1 900 |
| Upper Midland 2 (UM2) | Nyando | Temperate and sub-humid | | |
| Upper Midland 5 (UM5) | Nyando | Temperate and semi-arid | | |
| Lower Midland 2 (LM2) | Nyando | Warm and sub-humid | 21–24 | 800–1 500 |

and subsequent proximate analysis. Details for these procedures and their calculations have been previously published (Goopy et al., 2018; Ndung'u et al., 2019; Ndung'u et al., 2021). Live weight of all cattle were recorded at every visit, and daily milk production was measured for each lactating female. Farm management information, comprising material inputs and animal feeding strategies, was collected on a seasonal basis through farmer interviews during each visit. This approach facilitated the regular recording of animals entering and leaving herds as well as the commencement and completion of lactation, capturing irregular herd dynamics commonly observed among smallholders in the study region. Pasture formed the largest part of cattle diet across all counties, AEZs, and seasons, followed by maize stover and sugarcane tops (see Supplementary Table S1), both residues of crops grown for human consumption. A small amount of fodder crops dominantly Napier grass and Rhodes grass were also grown by some households. In all cases, the Napier grass and Rhodes grass were manually established for 1–20 years using cuttings. The nutritional quality of the resultant feed baskets was analysed using bulked representative samples by season and AEZ and is described elsewhere (Goopy et al., 2018; Ndung'u et al., 2019).

System boundary and functional unit

A cradle-to-farm gate approach was adopted to quantify herd-level EIs associated with cattle (Fig. 1). To eliminate the aggregation bias, or systematic underestimation of disproportionately large climate impacts caused by “weakest link” animals (McAuliffe et al., 2018a), these values were initially calculated on an animal-by-animal basis for each season and subsequently combined across

seasons and then animals in that order (see below). Although cattle data were repeatedly recorded for a period of 12 months, which constitute the temporal boundary of this study, the herd structure of each farm was not always at a steady state due to the movement of animals in and out of the farms in the form of sales, purchases, and temporary relocation to other farms during feed shortages. Across the entire sample, however, this effect was assumed to be largely cancelled out due to the sufficient sample size.

The primary FU for the study was set as CP (kg), encompassing both meat and milk production from multi-purpose cattle. We assumed that all animals sold out of study farms were sold for meat (or sold for further rearing before being on-sold for meat). Commensurably, animals purchased onto study farms were accounted for as an offset to the gross output. Thus, the total CP yield from each animal during the study period was defined as the net growth measured by the embedded CP content (details below) plus the CP content of milk produced.

To estimate the CP content of meat, a dressing percentage of 52.1% of live weight (LW) was assumed based on the locally most relevant information (Muchenje et al., 2008). Meat yield was set at 85% of carcass weight (Department of Agriculture and Rural Development, 2016) with a CP content of 21% (Muchenje et al., 2008). Edible by-products (offal) were also included in the total meat CP yield to reflect the local culinary practice (Table 2). These included the heart, kidneys, liver, lungs, spleen, tripe, tongue, and pancreas. The average offal yield (5.3% LW) and its CP content (18.2%) were obtained from the literature (Nollet & Toldra, 2011).

In addition, FPCM (kg) (IDF, 2010) and bone-free carcass weight (kg) were adopted as auxiliary FUs to facilitate the comparison of results with single-commodity EI studies for milk and meat, respectively. The FPCM was standardised to 4% fat and 3.3% true protein. The bone-free carcass weight was estimated using the assumptions described above.

Inventory analysis

Enteric CH₄ emissions

Enteric CH₄ emissions were calculated according to the approach of Goopy et al. (2018). As discussed, a feed basket with various feedstuff contributing varied proportions to the total feed basket (see Supplementary Table S1) was determined at the AEZ level and per season to produce a representative estimate for seasonal digestibility (see Supplementary Table S2). Similarly, metabolisable energy requirement was determined on an individual animal basis as the sum of metabolisable energy requirement for maintenance, growth, locomotion, and lactation following CSIRO (2007) models per season. The two sets of information (i.e. total metabolisable energy requirements and feed digestibility) were then combined to produce estimates of DM intake for each animal; this value was used to estimate enteric CH₄ production using the conversion factor of Charmley et al. (2016).

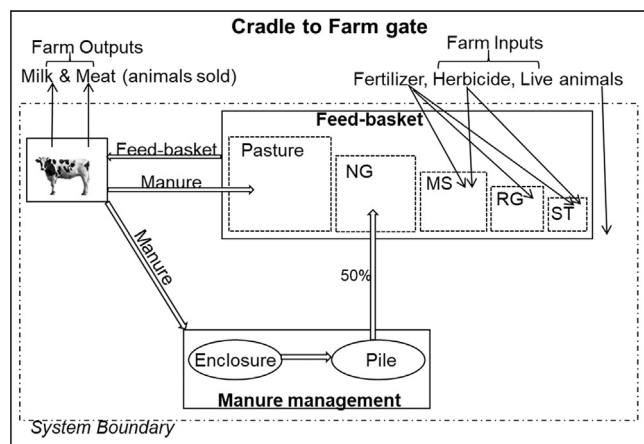


Fig. 1. System boundary for emission intensity assessment of cattle in smallholder farms. Squares show feedstuff in the feed basket where the sizes demonstrate the contribution of each feed to the overall feed basket (NG: Napier grass; MS: maize stover; RG: Rhodes grass; ST: sugarcane tops), ovals show the manure management systems. \longrightarrow shows the flow of raw materials and where the manure is deposited and \rightarrow shows the farm inputs and output.

Table 2
Edible by-products and their percentage yield, proportion, CP and CP yield from slaughtered cattle.

| Offal | Average Yield (%live weight) | Proportion (%) | CP content(g/100 g) | CP (g/Offal Yield) |
|----------|------------------------------|----------------|---------------------|--------------------|
| Heart | 0.4 | 7.58 | 20.00 | 1.52 |
| Liver | 2.75 | 52.13 | 20.00 | 10.43 |
| Kidney | 0.155 | 2.94 | 16.40 | 0.48 |
| Tripe | 0.75 | 14.22 | 13.40 | 1.91 |
| Spleen | 0.185 | 3.51 | 21.17 | 0.74 |
| Lungs | 0.6 | 11.37 | 15.57 | 1.77 |
| Tongue | 0.375 | 7.11 | 16.83 | 1.20 |
| Pancreas | 0.06 | 1.14 | 18.00 | 0.20 |
| Total | 5.275 | 100.00 | | 18.24 |

Manure CH₄ and nitrous oxide (N₂O) emissions

Animals were generally held in yards (bomas) near the farm dwelling overnight for security while grazing away from the home-stead during the day for ~12 h/d. To capture the effect of the practice on manure emissions, we assumed that (i) 50% of the manure was excreted (and left) on pasture that the remainder was excreted in the enclosure and periodically heaped, resulting in equal proportions of (ii) piled (25%) and (iii) unpiled (25%) manure. [Dong et al. \(2006\)](#) states that storage conditions affect both the type and quantum of GHGs from manure and we used the assumed conditions to develop a composite EF for manure deposited in these situations ([Table 3](#)).

Dung excreted was estimated using DM intake and DM digestibility of the relevant feed basket. The carbon content of dung was based on an earlier study carried out under a similar condition ([Zhu et al., 2018; Leitner et al., 2021](#)). The nitrogen excreted was estimated by the difference between the nitrogen intake (derived from DM intake and the nitrogen (N) content of the relevant feed basket) and N embedded in carcasses and milk (see above). The mass ratios between protein and N were assumed to be 6.25 for meat and 6.38 for milk, respectively ([Dong et al., 2006](#)).

Half of the piled manure was assumed to be ultimately applied to Napier grass fields. The proportion of N retained beyond the storage period ([Rufino et al., 2007](#)) and the N₂O emission factor for that retained N at application ([Dong et al., 2006](#)) were obtained from existing studies. Based on interview results, the remaining 50% of piled manure was assumed to be exported for non-economic and non-functional activities outside the system boundary (e.g. home gardens) and therefore not considered at the calculation of post-storage emissions.

Carbon dioxide (CO₂) emissions from farm inputs

Farm management practices, such as agrochemical use and crop/crop by-product yields, were recorded as part of farmer interviews. Land use was quantified through physical surveys. No machinery was used in any of the study farms. Synthetic fertiliser

Table 3
Total yield factors for nitrous oxide (g N-N₂O/100 g N in manure) and methane (g C-CH₄/100 g C in manure) for cattle manure deposited/stored under different management conditions.

| Management conditions | Weighting | Yield Factor (g CH ₄ -C/100 g C in manure (%) or g N ₂ O-N/100 g N in manure) | |
|-----------------------|-----------|---|------------------|
| | | CH ₄ | N ₂ O |
| Pasture | 0.50 | 0.031 | 0.004 |
| Boma | 0.25 | 0.01 | 0.079 |
| Pile | 0.25 | 0.43 | 0.45 |
| Composite Value | 1.00 | 0.126 | 0.134 |

Source: [Leitner et al. \(2021\)](#); N = Nitrogen, C = Carbon.

and herbicides were used on some farms in Nandi ($n = 31$) and Bomet ($n = 17$), (but not Nyando) for the cultivation of RG, maize, and sugarcane. Fertiliser types varied between farms, although the application rates were relatively uniform (and low) due to standardised recommendations from agricultural extension officers ([Mangale et al., 2016](#)). These included: 28.5 kg N/ha as urea (46% N), 32.2 kg N/ha as calcium ammonium nitrate (26% N), 28.5 kg as nitrogen, phosphorus and potassium (NPK, 20% N), and 22.3 kg N/ha as di-ammonium phosphate (18% N and 46% P₂O₅). The background emissions attributable to agrochemical production were obtained from the ecoinvent database V3 ([Wernet et al., 2016](#)). Direct and indirect N₂O emissions associated with fertiliser application were calculated using IPCC emission factors ([Dong et al., 2006](#)). Nitrogen leaching was not considered due to the dry condition in the study region.

Depending on the AEZ and season, the contribution of crop residues to the feed basket ranged between 1 and 42% for maize stover and 9 and 34% for sugarcane tops, respectively ([Goopy et al., 2018; Ndung'u et al., 2019](#)). As outlined, these crops are grown for human consumption and the residues are fed to animals only opportunistically. Thus, GHG emissions from maize and sugarcane fields attributable to livestock production were quantified under the economic allocation method. A harvest index of 0.41 was assumed to estimate maize stover yield ([Remison & Fajemisin, 1982](#)), while sugarcane top yield was estimated as 4.89% of primary crop harvest ([Kapur et al., 2013](#)). Price data used for the final allocation are provided in [Supplementary Table S3](#).

Impact assessment and interpretation

To make the results directly comparable with the largest pool of EIs in the literature ([Poore & Nemecek, 2018](#)), annual emissions attributable to individual animals were converted to global warming potential (GWP) under the 100-year Global Warming Potential (GWP₁₀₀) method, which assumes the characterisation factors of 28 and 265 for CH₄ and N₂O, respectively ([Stocker et al., 2013](#)) thereby reporting emissions using a measure of carbon dioxide equivalent (CO₂-eq). Individual values were aggregated for all animals on a single farm to estimate the farm-level GWP₁₀₀. Finally, the corresponding farm-level EI (CO₂-eq/kg CP) was derived as the ratio between GWP and the total (net) CP output.

Initial analysis of farm EIs ($n = 313$) identified a small number of farms across the three counties ($n = 25$) with nil or negative CP output, resulting in aberrant (infinitely large) EIs. Additionally, a small number of farms ($n = 4$) with positive but very low CP outputs (<3 kg CP per annum) returned extremely high EIs (>3 000 kg CO₂-eq/kg CP). With the upper bound for EIs in livestock systems posited to be ~1 000 kg CO₂-eq/kg CP ([Gerber et al., 2011](#)), the decision was made that EIs above this value would be truncated.

Similarly, the distribution of farm-level EIs was preliminarily studied under a variety of exploratory data analysis methods. As

this revealed that the data were extremely right-skewed without a uniform variance, further investigations to explore the factors contributing to differences in EI were undertaken using quantile regression (Koenker & Hallock, 2001). The motivation for choosing quantile regression was threefold. Firstly, ordinary least square (OLS) regression makes the assumptions of normality and a constant variance (of residuals), neither of which was met in this instance. Secondly, as quantile regression is robust to outliers, the effect of individual farms with truncated EIs on estimators can be minimised. Thirdly, quantile regression provides an opportunity to estimate an individual model for each quantile so that the impact of explanatory variables on EIs can be separated elucidated for low-, intermediate-, and high-performing farms. The following quantiles were used for the present analysis: 0.85, 0.75, 0.5, 0.25, and 0.1 with a model created for each of these quantiles. A quantile of 0.85, for example, corresponds to farms with EIs larger than 85% of sample farms (for a given set of explanatory variables), (with identical characteristics of herd structure, location, AEZ, and protein output), thereby representing relatively low-performing farms. In contrast, $Q_{0.1}$ corresponds to farms with EIs which are lower than 90% of farms, and the model for this quantile represents high-performing farms. Median regression (with a quantile of 0.5), on the other hand, has a similar interpretation to OLS regression. Multicollinearity was investigated for each model, and variables with variance inflation factors >10 were sequentially removed to arrive at a suitable model for each quantile. The explanatory variables considered include herd size, parity, average age (of cattle), milk yield, meat yield, and total GHG emissions. Fixed effects associated with counties and AEZs were also included in the model.

Results

Table 4 describes the herd characteristic and animal performance of the sample population in Nandi, Bomet, and Nyando as well as the weather conditions of these sites. Of the three study sites, Nyando showed the lowest presence of productive females and production levels as compared to Nandi and Bomet. Similarly, the average live weights of the animals in all classes were lower in Nyando and highest in Nandi.

Distribution of farm emission intensities

Median farm EIs were estimated to be 60 (Nandi), 71 (Bomet) and 90 (Nyando) kg CO₂-eq/kg CP. However, the values of individual farms dramatically varied even within each county. There was also substantial variation in the frequency of occurrence of low, intermediate, and high EI farms between counties and AEZs, with Nyando having the greatest proportion of high EI farms (Fig. 2).

Enteric fermentation was by far the largest contributor to total farm emissions in all counties and AEZs, accounting for 96–97% of total GHG emissions. The second greatest contributor was manure emissions, with N₂O and CH₄ responsible for an average of 1.6 and 1.2%, respectively (Fig. 3). Emissions from the production and application of agrochemicals contributed <1% to total GHG emissions. This trend was uniform across all counties and AEZs, except that there was no use of agrochemicals in Nyando as mentioned above.

Milk production was consistently the more important element of CP production, responsible for >80% of the farm-level CP output across all counties and AEZs (Fig. 4). In two AEZs in Nyando (upper midland 5 and lower midland 2), low and negative animal growth rates combined with a slight increase in animal population during the study period (25 purchased vs. 15 sold) resulted in negative CP output.

Table 4
Comparison of climate, and demographic factors, productivity, and ownership of cattle of smallholder farms in Nandi, Bomet, and Nyando.

| Descriptive Factors | Study sites | | |
|---|----------------------|--------------------|--|
| | Nandi ^{a,e} | Bomet ^b | Nyando ^{c,d} |
| Climate Factors | | | |
| Rainfall (mm) | 1 200–2 000 | 1 000–1 400 | 1 200–1 725 |
| Soil Types | | | |
| | Nitisol, Acrisol | Nitisol, Cambisol | Nitisol, Regosol, Leptosol, Vertisol, Arenosol, Adosol, Planosol |
| Average land size (ha) | 1.3 ^a | 1.5 ^b | 2.0 ^c |
| Cattle Productivity | | | |
| Live weight | | | |
| Females > 2 years | 306.9 | 280.6 | 216.3 |
| Males > 2 years | 265.9 | 259.6 | 216.0 |
| Heifers 1–2 years | 186.8 | 167.5 | 154.6 |
| Young Males 1–2 years | 156.9 | 136.5 | 143.5 |
| Calves < 1 year | 73.3 | 69.1 | 73.4 |
| No. of Females/County (% of sample size) | 487 (42.4) | 505 (44.5) | 176 (36.9) |
| No. of Males/county (% of sample size) | 44 (3.83) | 58 (5.12) | 107 (22.43) |
| No. of Heifers/county (% of Sample size) | 159 (13.84) | 121 (10.69) | 15 (3.14) |
| No. of Young males (% of sample size) | 57 (4.96) | 49 (4.33) | 17 (2.94) |
| No. of Calves (% of sample size) | 402 (34.99) | 399 (35.25) | 162 (33.96) |
| Average no. of lactating females/year | 256 | 305 | 39 |
| Percent females that calved down/year | 40.3 | 31.9 | 15.6 |
| Average milk yield (Litres/day) | 4.1 | 3.9 | 2.2 |
| Livestock ownership per household (numbers) | 9.1 | 8.7 | 8.5 |
| No. of cattle sold out | 198 | 243 | 78 |
| No. of cattle bought in | 96 | 197 | 31 |

Source:

- ^a GOK (2013).
- ^b GOK (2018).
- ^c Kirimi et al. (2010).
- ^d Jaetzold and Schmidt (1983).
- ^e Ndung'u et al. (2019).

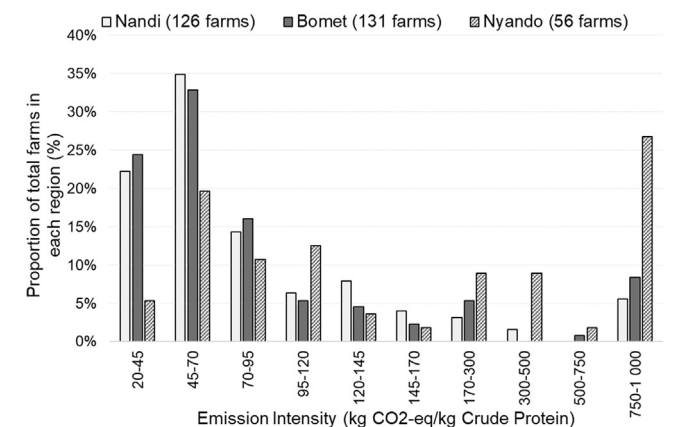


Fig. 2. Distributions of farm-level emission intensities for cattle in smallholder farms in Nandi, Bomet and Nyando.

Factors influencing farm-level emission intensities

Quantile regression revealed several management features that are highly influential to EI at the farm level, irrespective of the

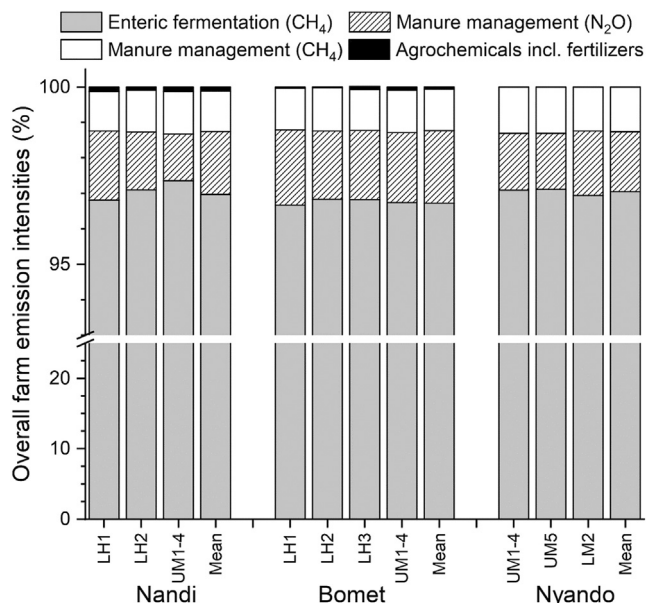


Fig. 3. Relative contribution of enteric CH₄, manure CH₄, and N₂O, and emissions from synthetic fertiliser production and application and agrochemical production to total greenhouse gas emissions related to cattle production by agro-ecological zones (Lower highland 1 (LH1), Lower highland 2 (LH2), Lower highland 3 (LH3), Upper midland 1 to 4 (UM1-4), Upper midland 5 (UM5), Lower midland 2 (LM2)) in Nandi, Bomet, and Nyando.

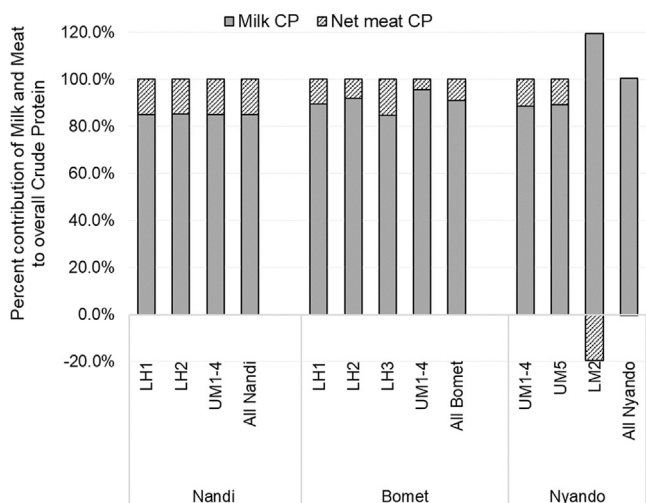


Fig. 4. Relative contribution of milk and meat to the total CP output by agro-ecological zones (Lower highland 1 (LH1), Lower highland 2 (LH2), Lower highland 3 (LH3), Upper midland 1-4 (UM1-4), Upper midland 5 (UM5), Lower midland 2 (LM2)) from cattle in Nando, Bomet, and Nyando.

county or AEZ (Table 5). Some factors were universally important, while others only at some EI quantiles. Despite the uneven contribution to total CP outputs, both meat and milk yields were significant drivers of EI across all quantiles investigated. Mean milk yield per cow, rather than milk production per farm, was found to be the most important driver of EI, with an increase in yield associated with a decrease in EI. An increase in herd size was found to increase EIs for low and medium EI (high and moderate performing) farms (Q_{10} : $\hat{\beta}_{HS} = 1.35, p < 0.005$, Q_{50} : $\hat{\beta}_{HS} = 1.86, p < 0.01$), whereas this tendency was not observed among high EI (low performing) farms. Although the average age of cattle was not impor-

tant to EI, the proportion of females in a herd was negatively related to EI for most quantiles. The effect of calving percentage was only significant—and positive—for high EI farms ($P < 0.005$). Finally, there were no clear differences in EI between AEZs, likely because the intrinsic differences were captured by other variables in the models.

The coefficients for the average milk yield per cow and total farm meat yield across the five quantiles considered are illustrated in Figs. 5 and 6. A negative relationship was observed for both milk and meat, with a 1 kg increase in yield associated with a reduction in EI across all quantiles. The degree to which this occurs increased across quantiles, from $\hat{\beta} = -1.18$ (SE = 0.05) for low EI farms (Q_{10}) to $\hat{\beta} = -6.14$ (SE = 1.33) to high EI farms (Q_{85}) for milk, showing that EIs for low-performing, high EI farms are highly sensitive to changes in average milk (per cow). A similar pattern was also observed for the coefficients for meat yield, such that these models suggest that average milk yield and meat yield become increasingly important contributors to EIs across the quantiles. Thus, attempts to increase protein yield as a means of lowering EIs will be most effective at the upper quantiles, that is, for low-performing farms in terms of EIs. A similar pattern was observed for the coefficients for meat. Additional quantile results and plots are shown in the Supplementary Material S1.

Discussion

Median EIs of milk production for this study (2.3 kg CO₂-eq/kg milk) were less than half of the pan-African estimates of Opio et al. (2013) while perhaps unsurprisingly almost double that for European and North American systems. In some ways, however, nominal comparison of mean/median EIs between different dairy production systems obscures important findings from the present study. While EIs have been reported using total CP output as the primary FU, both milk CP and meat CP alone have been included to facilitate comparison with other studies (Table 6). Our data demonstrate that meat CP makes up 15–25% of farm CP output across systems, and thus to ignore this would result in a substantial overestimation of EI in smallholder farms (unless emissions attributable to the ‘by-product’ (meat) are appropriately allocated out of the system boundary). Next, although several other studies have applied an LCA framework to estimate EIs in African livestock systems (Opio et al., 2013; Weiler et al., 2014; MacLeod et al., 2018; Kiggundu et al., 2019), input data have been derived from a variety of secondary sources in every case, including posthoc farmer estimates, national census statistics, FAOSTAT databases, and modelling based on these secondary data. In contrast, the results reported here are based on measurements of individual animals’ on-farm and actual feed baskets (see Supplementary Table S3). As such, this study provides a far clearer picture of, in particular, the variation of farm-level EI across, but also within, counties and AEZs. This approach, in turn, has led to the revelation that many farms in each of the counties (Nandi – 57%, Bomet – 58%, and Nyando – 20%) of the sampled farms had EIs comparable to European/North American intensified operations, doing so without employing high levels of inputs and mechanisation which are a hallmark of such operations. Exploration of the spectrum of EIs across farms provides insights into factors responsible for low EIs in smallholder farms, (something unachievable in studies relying on secondary data).

Prima facie, the differences between farms at the extremes of EI distribution were attributable to differences in CP output—very low EI enterprises had substantial outputs, whereas very high EI enterprises had little or in some cases no output at all in the course of the year. The absence of lactation females and growing animals resulted in a small number of farms exhibiting exceptionally large

Table 5

Estimated coefficients for each quantile regression model for estimating emission intensities for cattle, with associated *P*-values and pseudo *R*². NA is shown for variables that have been removed from the model based on high GVIF (multicollinearity).

| Variable | <i>q</i> ₁₀ | <i>p</i> | <i>q</i> ₂₅ | <i>p</i> | <i>q</i> ₅₀ | <i>p</i> | <i>q</i> ₇₅ | <i>p</i> | <i>q</i> ₈₅ | <i>p</i> |
|------------------------------|------------------------|----------|------------------------|----------|------------------------|----------|------------------------|----------|------------------------|----------|
| Intercept | 112.63 | 0.00* | 150.58 | 0.00* | 178.85 | 0.00* | 366.23 | 0.04* | 1197.50 | 0.00* |
| County = Nandi ^a | NA | NA | 6.90 | 0.02* | 9.13 | 0.15 | 9.14 | 0.75 | NA | NA |
| County = Nyando ^a | NA | NA | -10.06 | 0.31 | 15.14 | 0.73 | 419.52 | 0.00* | NA | NA |
| AEZ = LH2 ^b | 1.82 | 0.57 | 1.20 | 0.75 | NA | NA | NA | NA | NA | NA |
| AEZ = LH3 ^b | 4.93 | 0.83 | 7.69 | 0.32 | NA | NA | NA | NA | NA | NA |
| AEZ = LM2 ^b | -9.35 | 0.38 | 25.99 | 0.08 | NA | NA | NA | NA | NA | NA |
| AEZ = UM ^b | -5.19 | 0.07 | -2.82 | 0.39 | NA | NA | NA | NA | NA | NA |
| Herd Size | 1.35 | 0.00* | -0.52 | 0.49 | 1.86 | 0.01* | NA | NA | NA | NA |
| Average Parity ^c | -2.59 | 0.09 | -3.33 | 0.08 | NA | NA | -1.61 | 0.95 | -17.33 | 0.61 |
| Age females ^d | 0.01 | 0.95 | -0.19 | 0.20 | -0.33 | 0.05 | -1.68 | 0.43 | -6.00 | 0.01* |
| Age all ^e | NA | NA | 0.29 | 0.23 | NA | NA | 2.18 | 0.61 | 8.25 | 0.01* |
| Average milk yield | -1.18 | 0.00* | -1.33 | 0.00* | -1.52 | 0.00* | -2.28 | 0.03* | -6.14 | 0.00* |
| Meat yield | -0.53 | 0.00* | -0.62 | 0.00* | -0.88 | 0.00* | -1.80 | 0.01* | -4.62 | 0.00* |
| Calving ^f | 5.01 | 0.21 | -0.02 | 1.00 | -6.73 | 0.52 | -62.01 | 0.33 | -275.17 | 0.00* |
| Females ^{g,f} | -47.13 | 0.00* | -79.91 | 0.00* | -79.22 | 0.00* | -252.18 | 0.31 | -813.72 | 0.00 |
| Total Emissions | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Pseudo <i>R</i> ² | 0.82 | | 0.80 | | 0.79 | | 0.71 | | 0.58 | |

GVIF = Generalised Variance Inflation Factor, *q*₁₀ = 10th quantile, *q*₂₅ = 25th quantile, *q*₅₀ = 50th quantile, *q*₇₅ = 75th quantile, *q*₈₅ = 85th quantile, *p* = *P*-value at <0.05, AEZ = agro-ecological zone, LH1 = Lower highland 1, LH2 = Lower highland 2, LH3 = Lower highland 3, UM = Upper midland, LM2 = Lower midland 2, NA = data not available.

* Significant coefficients (*P* < 0.05).

^a Baseline = Bomet

^b Baseline = LH1

^c Average parity was calculated for the adult females in the herd.

^d Average age of adult females in the herd.

^e Average of all animals in the herd.

^f The number of females that calved during the one-year study period.

^g The number of adult females in the herd.

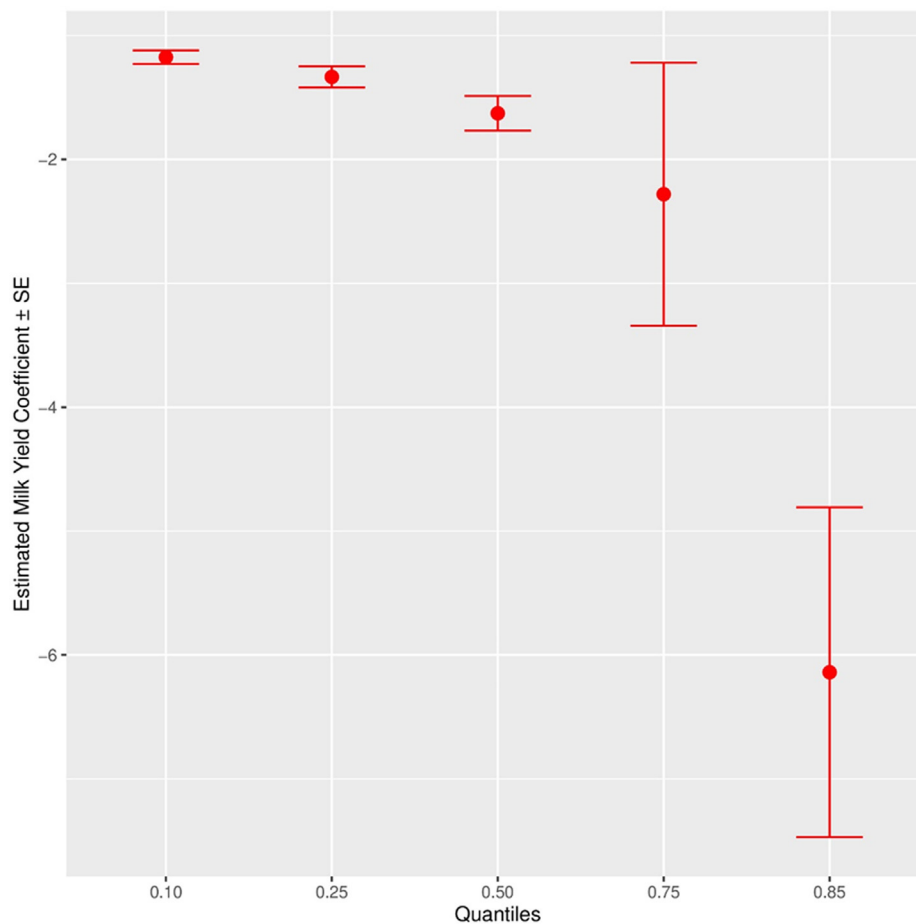


Fig. 5. Estimated coefficients for milk yield (±SE) from cattle across the 5-quantile regression models.

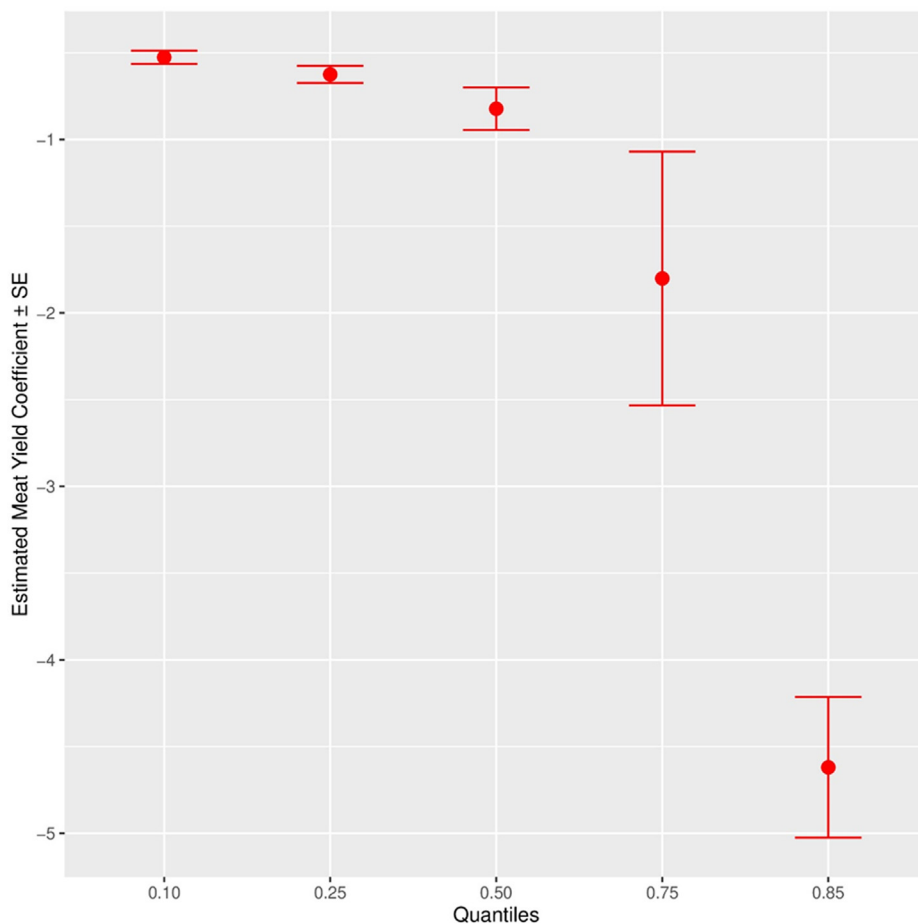


Fig. 6. Estimated coefficients for meat yield (±SE) from cattle across the 5-quantile regression models.

Table 6

Comparison of cattle emissions (kg CO₂-eq.) referenced to different functional units: a kilogram of fat- and protein-corrected milk (FPCM), milk, carcass weight, CP (milk and meat), protein (milk) and protein (meat) across multiple studies.

| Region | FPCM | Milk | Carcass Weight | CP (milk & meat) | Protein (milk) | Protein (meat) | Study |
|------------------|---------|---------|----------------|------------------|----------------|----------------|--------------------------|
| Nandi | 2.1 | 2.1 | 43 | 60 | 76 | 210 | This study |
| Bomet | 2.2 | 2.2 | 52 | 71 | 95 | 241 | This study |
| Nyando | 5.0 | 4.9 | 46 | 90 | 147 | 279 | This study |
| Western Kenya | 2.3 | 2.3 | 47 | 68 | 90 | 232 | This study |
| Kaptumo, Kenya | - | 0.9–4.3 | - | - | - | - | Weiler et al. (2014) |
| Uganda | - | - | - | - | 74.9 | 639.0 | Kiggundu et al. (2019) |
| Africa | 7.5 | - | 71.0 | - | - | - | Opio et al. (2013) |
| India | 1.9–2.3 | - | - | - | - | - | Garg et al. (2016) |
| United States | - | - | 21.3 | - | - | - | Rotz et al. (2019) |
| Ireland | 2.13 | - | - | - | - | - | O'Brien et al. (2014) |
| Western Canada | - | - | 22.0 | - | - | - | Beauchemin et al. (2010) |
| Europe | - | 1.3 | 22.6 | - | - | - | Lesschen et al. (2011) |
| Global Estimates | 2.8 | - | 46.2 | - | - | - | Opio et al. (2013) |

EIs. Although quantile regression mitigates statistical issues arising from the skewness, this observation brings to light a possible limitation of this study, in that we cannot be certain whether farms with very high EIs would continue to keep livestock for no return because their focus is not monetary (see below), or if this is a temporal anomaly caused by the structure of the study. However, to address this question would require longer-term data collection with commensurately greater resource needs and was thus well beyond the capacity of this study.

However, it is incorrect to conclude that EI is simply inverse correlated to livestock output across the EI farm spectrum – bigger is not necessarily better.

Between extremes of EI, the factors affecting farm-level EI seem more nuanced. A curious finding of this study was the presence of farms with very high and very low EI in close proximity to one another, even to the point of adjacent properties, militating against differences in EI being simply agro-ecological or even spatial in nature.

The production of methane from enteric fermentation overwhelmingly drive emissions on all farms in all regions. The importance of enteric CH₄ in the context of SHF is proportionally greater than other reports, especially those from Europe (O'Brien et al., 2014; 2015; Rotz et al., 2019), but also Uganda in the East African region (Kiggundu et al., 2019). There are two readily identifiable reasons for this. Firstly, the livestock systems in this study were

low input in terms of fertilisers, purchased feeds, and mechanisation, which in intensive European farming systems account for 7–20% of total emissions (Opio et al., 2013; O'Brien et al., 2014; 2015). Secondly, emissions from manure management were low (as a result of a drier climate and lower N excretion), compared to those found in Europe under which manure may comprise 5–9% of total emissions (Opio et al., 2013; O'Brien et al., 2014; 2015). Thus, although it has been suggested elsewhere that improved manure management in smallholder farms could be a promising approach for reducing EIs (Weiske et al., 2006; Lesschen et al., 2011; Petersen et al., 2013), it seems unlikely that such a strategy would have a salutary effect on overall emissions (Rotz et al., 2019).

On the other hand, EI is strongly influenced by output or production (Gerber et al., 2011; Lesschen et al., 2011; MacLeod et al., 2018). While we determined that output as such was not inversely proportional to on-farm EI, we identified several production-related factors that were; the proportion of females in a herd, percentage of lactating females, calving percentage, and milk produced per lactating female, were all highly influential, suggesting that herd management is more important than scale in influencing EI at the farm level (Garnsworthy, 2004; Bell et al., 2011; Knapp et al., 2014; MacLeod et al., 2018).

Unfortunately, identifying the factors driving differential EIs at the farm level does not directly explain why these differences exist. Although strictly beyond the scope of this study, we posit that there are three broad elements that are likely causative factors, or important contributors to the observed differences in EI and that further, they are frequently interrelated: Knowledge, Opportunity, and Motivation. Many factors relating to herd management, including growth rate, female fecundity, and milk production, are strongly related to feeding in general and diet quality in particular (Huzzey et al., 2007; Robinson, 2007). Ayantunde et al. (2016) has shown the positive influence of extending grazing time on intake and in turn, animal performance, while Rodney et al. (2018) demonstrated the negative effects on lactation curves (Manzi et al., 2020) and fertility management due to poor nutrition. The positive impact on farm productivity by providing targeted, hands-on training to farmers has been demonstrated (Goopy & Gakige, 2016), while access to the technology and materials to produce good-quality feedstuff from on-farm by-product (Gakige et al., 2020) has the potential to lift production in an affordable manner. However, without access to reliable and trusted markets, many farmers may choose not to invest the required resources to make such improvements. Finally, many SHFs have interests in livestock that go beyond their monetised value and will not be motivated by the financial benefits that improving production can bring.

On-farm changes that increase livestock output also tend to increase total farm emissions because animals require increased energy intake to achieve the new production level. Thus, low EI operations will necessarily require a move towards an "efficient frontier", where increased output is achieved at a minimal increase to emissions per se. As such, milk yield per cow was found to be an important driver of EI in this study, probably because the increased intake of a lactating cow is channelled directly into increased production. Quantile regression has also demonstrated, however, that contrary to European/North American systems, an increase in operational scale does not necessarily lead to improved efficiency in East African smallholding farms. While this situation may not hold true across all of SSA, our findings suggest that climate-driven policy interventions should consider the creation of efficient herd structures (ie: a high proportion of productive females), and optimisation of the feeding of those individuals, rather than the expansion of the farm to ratchet up farm outputs.

Expressing the environmental impact of livestock production systems using an EI approach is important when comparing ostensibly similar farms in the same region (Browne et al., 2011) and is better able to demonstrate the potential of mitigation measures (Rotz et al., 2010) than comparisons based on GHG emissions per area or per animal alone. This approach is a considerably more data-demanding exercise, and thus resource requirements may limit an extensive use of EIs to inform GHG mitigation in developing countries in the immediate future.

Collecting data from smallholder farms facilitated the calculation of emissions attributable to individual animals and of enterprise-level EIs. Based on these data, we demonstrated a high level of variation in farm EI within and across regions, even within ostensibly similar operations. Contrary to existing evidence, certain low-input farms were found to generate notably lower emissions, with EIs comparable to those observed for enterprises in developed economies. Examining the characteristics of these low EI farms provides insight into effective strategies to move smallholder farms towards a low carbon future.

Although this study was limited in its geographic scope, this type of smallholder farming is ubiquitous throughout much of in Eastern Africa, thus the findings of this study have regional significance. Our results indicate that smallholder animal enterprises are not, as has been claimed, inefficient, uniformly high emitters and that exemplars for (relatively) low carbon farming are present in extant operations. Significant mitigation potential exists in improving productivity on a per animal basis and restructuring the herd in favour of productive females with high(er) milk outputs. This, in turn, relies on improved access to and quality of, animal feed. Increasing animal productivity, while retaining a low-input farm model, will not only contribute to reduced carbon footprint but will also likely have social and economic advantages such as increasing household incomes.

Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.animal.2021.100445>.

Ethics approval

All animal data used in this study were collected as part of standard farming practices. As such, no part of this research was subject to the approval of an ethics committee.

Data and model availability statement

Data used for this study are publicly available (Goopy et al. 2018; Ndung'u et al. 2019).

Author ORCIDs

Phyllis Ndung'u: <https://orcid.org/0000-0002-7125-5313>
Taro Takahashi: <https://orcid.org/0000-0002-5434-2407>
Cornelius Jacobus Lindeque Du Toit: <https://orcid.org/0000-0003-1404-228X>
M. Robertson-Dean: <https://orcid.org/0000-0001-8964-773X>
Graham McAuliffe: <https://orcid.org/0000-0001-6031-1394>
Lutz Merbold: <https://orcid.org/0000-0003-4974-170X>
Klaus Butterbach-Bahl: <https://orcid.org/0000-0001-9499-6598>
John Goopy: <https://orcid.org/0000-0001-7177-1310>

Author contributions

The authors confirm the contribution of the paper as follows:

Phyllis Ndung'u: Data collection, data analysis, and interpretation, drafting the paper, critical revision of the paper.

Taro Takahashi: Conceptualising and designing of the study, data interpretation, critical review of the paper, final approval of the version to be published.

Melanie Robertson-Dean: Performed data analysis and interpretation – Statistical analysis

Cornelius Jacobus Lindeque Du Toit: Critical review of the paper.

Graham McAuliffe: Provided data analysis tools.

Lutz Merbold: Funding acquisition.

Klaus Butterbach-Bahl: Data analysis and interpretation, critical review of the paper and final approval of the paper to be published.

John Goopy: Conceptualising and designing of the study, supervision of the study, data interpretation, drafting of the paper, critical review of the paper, final approval of the version to be published.

Declaration of interest

The authors declare no potential conflict of interest associated with this research.

Acknowledgments

The authors acknowledge the CGIAR Fund Council, Australia (ACIAR), Irish Aid, European Union, IFAD, the Netherlands, New Zealand, UK, USAID, and Thailand for funding the CGIAR Research Program on Livestock.

Financial support statement

This study was funded by the International Fund for Agricultural Development (IFAD) through the research projects “Greening Livestock: Incentive-Based Interventions for Reducing the Climate Impact of Livestock in East Africa” (Grant No. 2000000994) and further supported by the German Federal Ministry for Economic Cooperation and Development (BMZ issued through GIZ) Programme of Climate Smart Livestock (PCSL). The authors appreciate funding from the Biotechnology and Biological Sciences Research Council (BBS/E/C/00010320) and the South African National Research Foundation, NRF Thuthuka Grant No. TTK180419322838.

References

- Ayantunde, A.A., Fernández-Rivera, S., Hiernaux, P.H.Y., van Keulen, H., Udo, H.M.J., Chanono, M., 2016. Effect of timing and duration of grazing of growing cattle in the West African Sahel on diet selection, faecal output, eating time, forage intake and live-weight changes. *Animal Science* 72, 117–128. <https://doi.org/10.1017/S1357729800055612>.
- Beauchemin, K.A., Henry Janzen, H., Little, S.M., McAllister, T.A., McGinn, S.M., 2010. Life cycle assessment of greenhouse gas emissions from beef production in western Canada: A case study. *Agricultural Systems* 103, 371–379. <https://doi.org/10.1016/j.agsy.2010.03.008>.
- Bell, M.J., Wall, E., Russell, G., Simm, G., Stott, A.W., 2011. The effect of improving cow productivity, fertility, and longevity on the global warming potential of dairy systems. *Journal of Dairy Science* 94, 3662–3678. <https://doi.org/10.3168/jds.2010-4023>.
- Browne, N.A., Eckard, R.J., Behrendt, R., Kingwell, R.S., 2011. A comparative analysis of on-farm greenhouse gas emissions from agricultural enterprises in south eastern Australia. *Animal Feed Science and Technology* 166–167, 641–652. <https://doi.org/10.1016/j.anifeedsci.2011.04.045>.
- Charmley, E., Williams, S.R.O., Moate, P.J., Hegarty, R.S., Herd, R.M., Oddy, V.H., Reyenga, P., Stauton, K.M., Anderson, A., Hannah, M.C., 2016. A universal

- equation to predict methane production of forage-fed cattle in Australia. *Animal Production Science* 56, 169–180. <https://doi.org/10.1071/AN15365>.
- CSIRO, 2007. *Nutrient Requirements for Domesticated Ruminants*. CSIRO Publishing, Collingwood, VIC, Australia.
- Department of Agriculture and Rural Development, Kwa Zulu Natal (2016). *Useful Statistics: Beef Production*. Pietermaritzburg, South Africa. Retrieved on 19/9/2019 from https://www.kzndard.gov.za/images/Documents/RESOURCE_CENTRE/GUIDELINE_DOCUMENTS/PRODUCTION_GUIDELINES/Beef_Production/Useful%20Statistics.pdf.
- Desjardins, R.L., Worth, D.E., Vergé, X.P.C., Maxime, D., Dyer, J., Cerkowniak, D., 2012. Carbon Footprint of Beef Cattle. *Sustainability* 4, 3279–3301. <https://doi.org/10.3390/su4123279>.
- Dong, H., Mangino, J., McAllister, T.A., Hatfield, J.L., Johnson, D.E., Lassey, K.R., Aparecida de Lima, M., Romanovskaya, A., 2006. Emissions from Livestock and Manure Management. In: Eggelston, S., Buendia, L., Miwa, K., Ngara, T., Tanabe, K. (Eds.), *Agriculture, Forestry and Other Land Use, IPCC Guidelines for National Greenhouse Gas Inventories*. Institute for Global Environmental Strategies (IGES), Hayama, Japan, pp. 87.
- du Toit, C.J.L., Meissner, H.H., van Niekerk, W.A., 2013a. Direct methane and nitrous oxide emissions of South African dairy and beef cattle. *South African Journal of Animal Science* 43, 320–339. <https://doi.org/10.4314/sajas.v43i3.7>.
- du Toit, C.J.L., van Niekerk, W.A., Meissner, H.H., 2013b. Direct greenhouse gas emissions of the South African small stock sectors. *South African Journal of Animal Science* 43, 340–361. <https://doi.org/10.4314/sajas.v43i3.8>.
- FAO, 2017. *Africa Sustainable Livestock 2050 – Technical Meeting and Regional Launch*. Addis Ababa, Ethiopia, 21–23 February 2017. FAO Animal Production and Health Report. No. 12, Rome, Italy. Retrieved on 03/08/2020 from at <http://www.fao.org/3/a-i7222e.pdf>.
- Gakige, J.K., Gachuri, C., Butterbach-bahl, K., Goopy, J.P., 2020. Sweet potato (*Ipomoea batatas*) vine silage: a cost-effective supplement for milk production in smallholder dairy-farming systems of East Africa? *Animal Production Science* 60, 1087–1094. <https://doi.org/10.1071/AN18743>.
- Garg, M.R., Phondba, B.T., Sherasia, P.L., Makkar, H.P.S., 2016. Carbon footprint of milk production under smallholder dairying in Anand district of Western India: a cradle-to-farm gate life cycle assessment. *Animal Production Science* 56, 423–436. <https://doi.org/10.1071/AN15464>.
- Garnsworthy, P.C., 2004. The environmental impact of fertility in dairy cows: a modelling approach to predict methane and ammonia emissions. *Animal Feed Science and Technology* 112, 211–223. <https://doi.org/10.1016/j.anifeedsci.2003.10.011>.
- Gerber, P., Vellinga, T., Opio, C., Steinfeld, H., 2011. Productivity gains and greenhouse gas emissions intensity in dairy systems. *Livestock Science* 139, 100–108. <https://doi.org/10.1016/j.livsci.2011.03.012>.
- Gerber, P.J., Steinfeld, H., Henderson, B., Mottet, A., Opio, C., Dijkman, J., Falcucci, A., Tempio, G., 2013. *Tackling Climate Change Through Livestock: A Global Assessment of Emissions and Mitigation Opportunities*. Food and Agriculture Organization of the United Nations (FAO), Rome, Italy.
- GOK, 2013. Nandi County Profile. Retrieved on 03/11/2016 from nandi.go.ke/wp-content/uploads/2015/04/Nandi-County-Profile.doc.
- GOK, 2018. County Government of Bomet: Bomet County Integrated Development Plan, 2018–2022. Retrieved on 23/5/2019 from <https://www.bometassembly.go.ke/upload/County%20Integrated%20Development%20Plan.pdf>.
- Goopy, J.P., Gakige, J.K., 2016. Improving productivity in Kenyan Smallholder Dairy systems through selective, intensive education and supported adoption. In: *Book of Abstract of the Tropentag 2016 Conference on Solidarity in a Competing World – Fair Use of Resources*, Vienna, Austria, 19–21 September 2016, p. 157.
- Goopy, J.P., Ndung'u, P.W., Onyango, A., Kirui, P., Butterbach-Bahl, K., 2021. Calculation of new enteric methane emission factors for small ruminants in western Kenya highlights the heterogeneity of smallholder production systems. *Animal Production Science* 61, 602–612. <https://doi.org/10.1071/AN19631>.
- Goopy, J.P., Onyango, A., Dickhoeffer, U., Butterbach-Bahl, K., 2018. A new approach for improving emission factors for enteric methane emissions for cattle in smallholder systems of East Africa-results for Nyando, Western Kenya. *Agricultural Systems* 161, 72–80. <https://doi.org/10.1016/j.agsy.2017.12.004>.
- Herrero, M., Grace, D., Njuki, J., Johnson, N., Enahoro, D., Silvestri, S., Rufino, M.C., 2013. The roles of livestock in developing countries. *Animal* 7 (Suppl 1), 3–18. <https://doi.org/10.1017/s1751731112001954>.
- Huzzey, J.M., Veira, D.M., Weary, D.M., von Keyserlingk, M.A., 2007. Parturition behavior and dry matter intake identify dairy cows at risk for metritis. *Journal of Dairy Science* 90, 3220–3233. <https://doi.org/10.3168/jds.2006-807>.
- IDF, 2010. *A Common Carbon Footprint for Dairy*. The IDF Guide to Standard Lifecycle Assessment Methodology for the Dairy Industry Bulletin of the International Dairy Federation 445. International Dairy Federation, Brussels, Belgium.
- Jaezold, R., Schmidt, H., 1983. *Natural Conditions and Farm Management Information. Part A: West Kenya and Part B: Central Kenya*. In: Sine nomine (Ed.), *Farm Management Handbook of Kenya*. Ministry of Agriculture, in Cooperation with German Agriculture Team (GAT) of the German Agency for Technical Cooperation (GTZ), Nairobi, Kenya. Typo-druck, Rossdorf, West Germany, pp. 728.
- Kapur, R., Duttamajumder, S.K., Srivastava, B.L., Madhok, H.L., Kumar, R., 2013. Harvest index and the component of biological yield in sugarcane. *Indian Journal of Genetics and Plant Breeding* 73, 386–391. <https://doi.org/10.5958/j.0975-6906.73.4.058>.
- Kiggundu, N., Ddungu, S.P., Wanyama, J., Cherotich, S., Mpairwe, D., Zziwa, E., Mutebi, F., Falcucci, A., 2019. Greenhouse gas emissions from Uganda's cattle

- corridor farming systems. *Agricultural Systems* 176. <https://doi.org/10.1016/j.agsy.2019.102649> 102649.
- Kirimi, L., Mathenge, M., Olwande, J., Gamba, P., Onyuma, S., Lagat, J., 2010. Globalization and Structural Change in Rural Economies RuralStruc Program – Phase II. Tegemeo Institute, Egerton University Retrieved on 22/2/2021 from <http://documents1.worldbank.org/curated/en/102611468276355485/pdf/779820WPOKENYA00Box377319B00PUBLIC0.pdf>.
- Knapp, J.R., Laur, G.L., Vadas, P.A., Weiss, W.P., Tricarico, J.M., 2014. Invited Review: Enteric methane in dairy cattle production: Quantifying the opportunities and impact of reducing emissions. *Journal of Dairy Science* 97, 3231–3261. <https://doi.org/10.3168/jds.2013-7234>.
- Koenker, R., Hallock, K.F., 2001. Quantile Regression. *Journal of Economic Perspectives* 15, 143–156. <https://doi.org/10.1257/jep.15.4.143>.
- Kouazounde, J.B., Gbenou, J.D., Babatoude, S., Srivastava, N., Eggleston, S.H., Antwi, C., Baah, J., McAllister, T.A., 2015. Development of methane emission factors for enteric fermentation in cattle from Benin using IPCC Tier 2 methodology. *Animal* 9, 526–533. <https://doi.org/10.1017/S1751731114002626>.
- Lee, M.A., Davis, A.P., Chagunda, M.G.G., Manning, P., 2017. Forage quality declines with rising temperatures, with implications for livestock production and methane emissions. *Biogeosciences* 14, 1403–1417. <https://doi.org/10.5194/bg-14-1403-2017>.
- Legesse, G., Beauchemin, K.A., Ominski, K.H., McGeough, E.J., Kroebel, R., MacDonald, D., Little, S.M., McAllister, T.A., 2016. Greenhouse gas emissions of Canadian beef production in 1981 as compared with 2011. *Animal Production Science* 56, 153–168. <https://doi.org/10.1071/AN15386>.
- Leitner, S., Ring, D., Wanyama, G.N., Korir, D., Pelster, D.E., Goopy, J.P., Butterbach-Bahl, K., Merbold, L., 2021. Effect of feeding practices and manure quality on CH₄ and N₂O emissions from uncovered cattle manure heaps in Kenya. *Waste Management* 126, 209–220. <https://doi.org/10.1016/j.wasman.2021.03.014>.
- Lesschen, J.P., van den Berg, M., Westhoek, H.J., Witzke, H.P., Oenema, O., 2011. Greenhouse gas emission profiles of European livestock sectors. *Animal Feed Science and Technology* 166–167, 16–28. <https://doi.org/10.1016/j.anifeedsci.2011.04.058>.
- MacLeod, M., Eory, V., Wint, W., Shaw, A., Gerber, P.J., Cecchi, G., Mattioli, R., Sykes, A., Robinson, T., 2018. Assessing the Greenhouse Gas Mitigation Effect of Removing Bovine Trypanosomiasis in Eastern Africa. *Sustainability* 10, 1633. <https://doi.org/10.3390/su10051633>.
- Mangale, N., Muriuki, A., Kathuku-Gitonga, A.N., Kibunja, C.N., Mutegi, J.K., Esilaba, A.O., Ayuke, F.O., Nguluu, S.N., Gikonyo, E.W., 2016. Field and Laboratory Research Manual for Integrated Soil Fertility Management in Kenya. Kenya Soil Health Consortium, Nairobi, Kenya. Retrieved on 21/10/2020 from <https://www.kalro.org/sites/default/files/KALRO-KSHC-Manual%20final%202016-080118.pdf>.
- Manzi, M., Rydhmer, L., Ntawubizi, M., D'Andre Hirwa, C., Karege, C., Strandberg, E., 2020. Milk production and lactation length in Ankole cattle and Ankole crossbreds in Rwanda. *Tropical Animal Health and Production* 52, 2937–2943. <https://doi.org/10.1007/s11250-020-02311-9>.
- McAuliffe, G.A., Takahashi, T., Harris, P., Orr, R.J., Lee, M.R.F., 2018a. Distributions of emissions intensity for individual beef cattle reared on pasture-based production systems. *Journal of Cleaner Production* 171, 1672–1680. <https://doi.org/10.1016/j.jclepro.2017.10.113>.
- McAuliffe, G.A., Takahashi, T., Lee, M.R.F., 2018b. Framework for life cycle assessment of livestock production systems to account for the nutritional quality of final products. *Food and Energy Security* 7, <https://doi.org/10.1002/fes3.143> e00143.
- McAuliffe, G.A., Takahashi, T., Lee, M.R.F., 2020. Applications of nutritional functional units in commodity-level life cycle assessment (LCA) of agri-food systems. *The International Journal of Life Cycle Assessment* 25, 208–221. <https://doi.org/10.1007/s11367-019-01679-7>.
- Moran, D., Wall, E., 2011. Livestock production and greenhouse gas emissions: Defining the problem and specifying solutions. *Animal Frontiers* 1, 19–25. <https://doi.org/10.2527/af.2011-0012>.
- Muchenje, V., Dzama, K., Chimonyo, M., Raats, J.G., Strydom, P.E., 2008. Meat quality of Nguni, Bonsmara and Aberdeen Angus steers raised on natural pasture in the Eastern Cape, South Africa. *Meat Science* 79, 20–28. <https://doi.org/10.1016/j.meatsci.2007.07.026>.
- Muriuki, H.G., 2003. *Milk and Dairy Products, Post-harvest Losses and Food Safety in Sub-Saharan Africa and the Near East. A Review of the Small Scale Dairy Sector – Kenya. FAO Prevention of Food Losses Programme, Rome, Italy.*
- Ndao, S., Traoré, E.H., Ickowicz, A., Moulin, C.H., 2020. Estimation of enteric methane emission factors for Ndama cattle in the Sudanian zone of Senegal. *Tropical Animal Health and Production* 52, 2883–2895. <https://doi.org/10.1007/s11250-020-02280-z>.
- Ndao, S., Moulin, C.-H., Traoré, E.H., Diop, M., Bocquier, F., 2019. Contextualized re-calculation of enteric methane emission factors for small ruminants in sub-humid Western Africa is far lower than previous estimates. *Tropical Animal Health and Production* 51, 919–928. <https://doi.org/10.1007/s11250-018-1775-4>.
- Ndung'u, P.W., Kirui, P., Takahashi, T., du Toit, C.J.L., Merbold, L., Goopy, J.P., 2021. Data describing cattle performance and feed characteristics to calculate enteric methane emissions in smallholder livestock systems in Bomet County, Kenya. *Data in Brief* 39. <https://doi.org/10.1016/j.dib.2021.107673> 107673.
- Ndung'u, P.W., Bebe, B.O., Ondiek, J.O., Butterbach-Bahl, K., Merbold, L., Goopy, J.P., 2019. Improved region-specific emission factors for enteric methane emissions from cattle in smallholder mixed crop: livestock systems of Nandi County, Kenya. *Animal Production Science* 59, 1136–1146. <https://doi.org/10.1071/AN17809>.
- Ndung'u, P.W., Takahashi, T., Du Toit, C.J.L., Robertson-Dean, M.R., Butterbach-Bahl, K.B., McAuliffe, G.A., Merbold, L., Goopy, J.P., 2021. A partial life cycle assessment of smallholder livestock systems in Western Kenya. In: *Book of abstracts of the 72nd Annual Meeting of the European Federation of Animal Science*, 30 August–3 September, 2021, Davos, Switzerland, p. 232.
- Nkonya, E., Johnson, T., Kwon, H.Y., Kato, E., 2016. *Economics of Land Degradation in Sub-Saharan Africa*. In: Nkonya, E., Mirzababev, A., von Braun, J. (Eds.), *Economics of Land Degradation and Improvement – A Global Assessment for Sustainable Development*. 1st edition. Springer, Cham, Denmark, pp. 215–259.
- Nollet, L.M.L., Toldra, F., 2011. *Handbook of Analysis of Edible Animal By-Products*. CRC Press Taylor & Francis Group, Boca Raton, FL, USA.
- O'Brien, D., Capper, J.L., Garnsworthy, P.C., Grainger, C., Shalloo, L., 2014. A case study of the carbon footprint of milk from high-performing confinement and grass-based dairy farms. *Journal of Dairy Science* 97, 1835–1851. <https://doi.org/10.3168/jds.2013-7174>.
- O'Brien, D., Hennessy, T., Moran, B., Shalloo, L., 2015. Relating the carbon footprint of milk from Irish dairy farms to economic performance. *Journal of Dairy Science* 98, 7394–7407. <https://doi.org/10.3168/jds.2014-9222>.
- Opio, C., Gerber, P., Mottet, A., Falcucci, A., Tempio, G., MacLeod, M., Vellinga, T., Henderson, B., Steinfeld, H., 2013. *Greenhouse gas emissions from ruminant supply chains – A global life cycle assessment*. Food and Agriculture Organization of the United Nations, Rome, Italy.
- Parry, M.L., Canziani, O.F., Palutikof, J.P., van der Linden, P.J., Hanson, C.E., 2007. *Climate Change 2007: Impacts, Adaptation and Vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, UK.
- Pelster, D.E., Gisore, B., Koske, J.K., Goopy, P., Korir, D., Rufino, M.C., Butterbach-Bahl, K., 2016. Methane and Nitrous Oxide Emissions from Cattle Excreta on an East African Grassland. *Journal of Environmental Quality* 45, 1531–1539. <https://doi.org/10.2134/jeq2016.02.0050>.
- Petersen, S.O., Blanchard, M., Chadwick, D., Del Prado, A., Edouard, N., Mosquera, J., Sommer, S.G., 2013. Manure management for greenhouse gas mitigation. *Animal* 7, 266–282. <https://doi.org/10.1017/S1751731113000736>.
- Poore, J., Nemecek, T., 2018. Reducing food's environmental impacts through producers and consumers. *Science* 360, 987–992. <https://doi.org/10.1126/science.aag0216>.
- Remison, S.U., Fajemisin, J.M., 1982. Comparative growth of maize cultivars with different leaf orientation. *Journal of Agricultural Science, Cambridge* 99, 61–66. <https://doi.org/10.1017/S0021859600055039>.
- Rice, P., O'Brien, D., Shalloo, L., Holden, N.M., 2017. Evaluation of allocation methods for calculation of carbon footprint of grass-based dairy production. *Journal of Environmental Management* 202, 311–319. <https://doi.org/10.1016/j.jenvman.2017.06.071>.
- Robinson, J.J., 2007. Nutrition in the Reproduction of Farm Animals. *Nutrition Research Reviews* 3, 253–276. <https://doi.org/10.1079/NRR19900015>.
- Rodney, R.M., Celi, P., Scott, W., Breinhild, K., Santos, J.E.P., Lean, I.J., 2018. Effects of nutrition on the fertility of lactating dairy cattle. *Journal of Dairy Science* 101, 5115–5133. <https://doi.org/10.3168/jds.2017-14064>.
- Ross, S.A., Topp, C., Ennos, R.A., Chagunda, M., 2017. Relative emissions intensity of dairy production systems: employing different functional units in life-cycle assessment. *Animal* 11, 1381–1388. <https://doi.org/10.1017/S1751731117000052>.
- Rotz, C.A., 2018. Modeling greenhouse gas emissions from dairy farms. *Journal of Dairy Science* 101, 6675–6690. <https://doi.org/10.3168/jds.2017-13272>.
- Rotz, C.A., Montes, F., Chianese, D.S., 2010. The carbon footprint of dairy production systems through partial life cycle assessment. *Journal of Dairy Science* 93, 1266–1282. <https://doi.org/10.3168/jds.2009-2162>.
- Rotz, C.A., Asem-Hiablie, S., Place, S., Thoma, G., 2019. Environmental footprints of beef cattle production in the United States. *Agricultural Systems* 169, 1–13. <https://doi.org/10.1016/j.agsy.2018.11.005>.
- Rufino, M.C., Tittone, P., Van Wijk, M.T., Castellanos-Navarrete, A., Delve, R.J., De Ridder, N., Giller, K.E., 2007. Manure as a key resource within smallholder farming systems: Analysing farm-scale nutrient cycling efficiencies with the NUNANCES framework. *Livestock Science* 112, 273–287. <https://doi.org/10.1016/j.livsci.2007.09.011>.
- Steinfeld, H., Wassenaar, T., Jutzi, S., 2006. Livestock production systems in developing countries: status, drivers, trends. *Revue scientifique et technique (International Office of Epizootics)* 25, 505–516.
- Stocker, T.F., Qin, D., Plattner, G.-K., Tignor, M., Allen, S.K., Boschung, J., Nauels, A., Xia, Y., Bex, V., Midgley, P.M., 2013. *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA. pp. 1535. Retrieved on 24/3/2019 from <https://doi.org/10.1017/CBO9781107415324>.
- Thornton, P.K., 2010. Livestock production: recent trends, future prospects. *Philosophical Transactions of Royal Society B* 365, 2853–2867. <https://doi.org/10.1098/rstb.2010.0134>.
- Thorpe, W., Muriuki, H.G., Omoro, A., Owango, M.O., Staal, S., 2000. *Dairy development in Kenya: the past, the present and the future*. Paper presented at the Annual Symposium of the Animal Production Society of Kenya (APSK). Theme: Challenges to Animal Production in this Millennium, KARI Headquarters, 22–23 March 2000. ILRI, Nairobi, Kenya.
- Weiler, V., Udo, H.M.J., Viet, T., Crane, T.A., De Boer, I.J.M., 2014. Handling multi-functionality of livestock in a life cycle assessment: the case of smallholder

- dairying in Kenya. *Current Opinion in Environmental Sustainability* 8, 29–38. <https://doi.org/10.1016/j.cosust.2014.07.009>.
- Weiske, A., Vabitsch, A., Olesen, J.E., Schelde, K., Michel, J., Friedrich, R., Kaltschmitt, M., 2006. Mitigation of greenhouse gas emissions in European conventional and organic dairy farming. *Agriculture, Ecosystems & Environment* 112, 221–232. <https://doi.org/10.1016/j.agee.2005.08.023>.
- Wernet, G., Bauer, C., Steubing, B., Reinhard, J., Moreno-Ruiz, E., Weidema, B., 2016. The ecoinvent database version 3 (part 1): overview and methodology. *International Journal of Life Cycle Assessment* 21, 1218. <https://doi.org/10.1007/s11367-016-1087-8>.
- World Bank, CIAT, 2015. *Climate-Smart Agriculture in Kenya*. CSA Country Profiles for Africa, Asia, and Latin America and the Caribbean Series. The World Bank Group Washington, DC, USA. Retrieved on 7/11/2017 from <https://hdl.handle.net/10568/69545>.
- Zhu, Y., Merbold, L., Pelster, D., Diaz-Pines, E., Wanyama, G.N., Butterbach-Bahl, K., 2018. Effect of Dung Quantity and Quality on Greenhouse Gas Fluxes From Tropical Pastures in Kenya. *Global Biogeochemical Cycles* 32, 1589–1604. <https://doi.org/10.1029/2018gb005949>.