

1 **Phenotypic plasticity in *Sargassum* forests may not counteract projected biomass losses along a**  
2 **broad latitudinal gradient**

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14  
15 **Highlights**

- 16 • Macroecological environments drive physiological and biomass changes of *S.*  
17 *cymosum*  
18 • Tropical individuals show higher thermal tolerance compared to temperate ones  
19 • Future biomass losses might occur regardless of the higher thermal tolerance

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21  
22 **Abstract**

23 Phenotypic plasticity and local adaptation can adjust individual responses to environmental  
24 changes across species' ranges. Studies addressing the implications of such traits have been  
25 underrepresented in the marine environment. *Sargassum cymosum* represents an ideal model  
26 to test phenotypic plasticity, as populations along the southwestern Atlantic Ocean display a  
27 sharp decrease in abundance towards distributional range limits. We (1) characterized the  
28 macroecological environment of *S. cymosum* across a latitudinal gradient, (2) evaluated  
29 potential differences in ecophysiological adjustments (biomass, photosynthetic pigments,  
30 phenolic compounds, total soluble sugars and proteins, and carbon-nitrogen - CN - content),  
31 and (3) tested for differences in thermal tolerance based on time series analyses produced from  
32 the present to contrasting representative concentration pathways scenarios (RCP) of future

1 climate changes. Our results showed distinct macroecological environments, corresponding  
2 to tropical and warm temperate conditions, driving biomass and ecophysiological adjustments  
3 of *S. cymosum*. Populations from the two environments displayed contrasting thermal  
4 tolerances, with tropical individuals better coping with thermal stress when compared to  
5 more temperate ones (lethal temperatures of 33°C vs. 30°C); yet both populations lose biomass  
6 in response to increasing thermal stress while increasing secondary metabolites (e.g.,  
7 carotenoids and phenolic compounds) and decrease chlorophyll's content, Fv/Fm, total  
8 soluble sugars concentration and CN ratio, owing to oxidative stress. Despite evidence for  
9 phenotypic plasticity, significant future losses might occur in both tropical and warm  
10 temperate populations, particularly under the no mitigation RCP scenario, also known as the  
11 business as usual (i.e., 8.5). In this context, broad compliance with the Paris Agreement might  
12 counteract projected impacts of climate change, safeguarding *Sargassum* forests in the years  
13 to come.

14 Keywords: *Sargassum cymosum*; climate change; centre-edge populations; experimental  
15 essays; global warming; marine heatwave; physiological performance.

## 17 **Introduction**

18 Higher population abundance is predicted in the center of species' geographic distributions  
19 due to the prevalence of more favorable conditions, while towards the edge, abundance is  
20 expected to decrease as conditions become less suitable (Brown 1984; Brussard 1984).  
21 However, this pattern may not be universal, and species may preserve performance along  
22 their ranges owing to phenotypic plasticity, which can be determined by acclimation  
23 (variations in responses of an individual, without genetic changes) and by local adaptation  
24 (selective genetic changes in allele frequencies in a population) (Sagarin and Gaines 2002;  
25 Ntuli and others 2020). Accordingly, populations of the same species may invest differently  
26 in reproduction, growth, and chemical composition under similar conditions due to potential  
27 differences in their niches (e.g., Ferreira and others 2014; King and others 2020). This has  
28 much relevance in the scope of climate change ecology as plasticity might ensure survival in  
29 the face of ongoing and projected environmental changes. Compared to terrestrial systems,  
30 the implications of phenotypic plasticity and adaptation have been underrepresented in the  
31 marine environment (but see King and others 2018).

1 Cellular mechanisms are expected to manifest different outcomes when populations  
2 from contrasting ranges (e.g., center vs. edges) are exposed to extreme conditions (Eggert  
3 2012; Harley and others 2012). For instance, thermal stress can force biochemical and  
4 physiological adjustments in metabolic pathways, affect membrane-associated processes (e.g.,  
5 stability and biochemical composition), and promote greater production of secondary  
6 metabolites (e.g., phenolic compounds and carotenoids) associated with protective strategies  
7 against oxidative stress (Balboa and others 2013; Mannino and others 2016; Hargrave and  
8 others 2017). When exposed to extreme thermal conditions, photosynthesis, resource  
9 utilization, and growth, as well as changes in carbon (C), nitrogen (N), proteins, and storage  
10 of carbohydrate content are straightforward indicators of cell's physiological status (Torres  
11 and others 1991; Wernberg and others 2016; Gouvêa and others 2017; Costa and others 2019).  
12 Ultimately, these indicators can be used to test for phenotypic plasticity between populations  
13 of contrasting ranges under increasing thermal stress.

14 Over the past decades, global climate change has increased ocean temperatures  
15 through marine heatwave events (MHWs). These events have been recorded globally,  
16 severely impacting biodiversity in the Mediterranean Sea, Northwest Atlantic, Northeast  
17 Pacific Ocean, West Australia, Tasman, and Tropical Australian Sea (Mills and others 2013;  
18 Smale and others 2019; Arafeh-Dalmau and other 2020). More frequent and intense MHWs  
19 are anticipated for the future, yet highly dependent on the representative concentration  
20 pathway (RCP) scenarios of greenhouse gas emissions (Oliver and others 2018; Oliver and  
21 others 2019). These can range from high mitigation strategies aligned with the Paris  
22 Agreement initiative, whereby atmospheric temperatures increase up to 2°C above pre-  
23 industrial levels by 2100 (e.g., RCP 2.6), to the no mitigation scenario, also known as the  
24 business as usual, with raising temperatures of up to 4.9 °C (e.g., RCP 8.5; IPCC, 2014).  
25 Accordingly, distinct biodiversity outcomes are expected when considering such contrasting  
26 RCP scenarios. For instance, the projected consequences of future climate changes to marine  
27 forests of large brown algae can vary between moderate losses at lower latitude range margins  
28 to severe range shifts, eroding unique biodiversity hotspots across taxa (Assis and others 2017).

29 *Sargassum* is a canopy-forming macroalga distributed in tropical and subtropical  
30 environments and known for its high biomass and carbon stock (Gouvêa and others 2020). It  
31 forms dominant communities with vital ecological roles in coastal marine ecosystems and  
32 offshore islands. Marine forests of *S. cymosum* represent an ideal model to address potential

1 phenotypic plasticity changes and adaptation across ranges, particularly in the southwestern  
2 Atlantic Ocean, where populations follow the abundant-center hypothesis, with a sharp  
3 decrease in abundance towards the distributional range limit of Brazilian coastlines.

4 The present study evaluated the physiological responses and phenotypic plasticity of  
5 *S. cymosum* forests to increasing thermal stress. In particular, it (1) characterized the  
6 macroecological environment of *S. cymosum* across a broad latitudinal gradient, (2) assessed  
7 for differences in ecophysiological adjustments (biomass, photosynthetic pigments, phenolic  
8 compounds, total soluble sugars and proteins, and CN), and (3) tested for thermal plasticity  
9 based on time series analyses produced from the present to contrasting representative  
10 concentration pathways scenarios (RCP) of future climate changes.

## 11 **Materials and Methods**

### 12 **Ecophysiological responses along a latitudinal gradient**

13  
14 *Sargassum cymosum* (C. Agardh) is widely distributed in both hemispheres, occurring  
15 in the Atlantic and Pacific Oceans, as well as in the Indian islands. Along the coastlines of  
16 Brazil, the species presents a north (warmer region) to south (colder region) decline in  
17 abundance, towards the edge of its distribution (Cardoso beach; SC). To analyze the  
18 macroecological environment of the species along a broad latitudinal gradient (Brazil  
19 coastline; Table S1), i.e., to understand the relationship between its distribution and the  
20 environment, at large spatial scales, samples were collected in nine distinct populations (from  
21 here on referred to as a population). Sampling was performed in the summers (December to  
22 March) of 2016 and 2017 in the upper subtidal zone during low tides. On average, fifteen  
23 0.0625 m<sup>2</sup> quadrats were randomly placed and scraped along a 20 m transect laid across *S.*  
24 *cymosum* forests, in areas distancing 3 meters far apart (120 samples in total). Fresh and dry  
25 weights were recorded using an analytical balance (Marte, AY 220). Epiphytes and epifauna  
26 were removed, and species identification followed dichotomous keys and appropriate  
27 literature (Ouriques 1997; Nunes 1998; Pedrini 2012).

28  
29 Samples were dried at 60°C ( $\pm$  2°C) for 48 hours for biomass measurements. Fresh  
30 weight samples were stored in -80 °C for subsequent chemical analyses. Chemical analyses  
31 for photosynthetic pigments, phenolic compounds, total soluble sugars and proteins, total  
32 organic carbon and nitrogen (CN) were performed according to the protocols of Ritchie

1 (2008), Aman and others (2005), Costa and others (2019), Shannon (1968) and Bradford  
2 (1986). Detailed information on the methods used for each chemical analysis can be found in  
3 S4.

4 A Hierarchical clustering multivariate analysis (HCA) was performed with the  
5 biomass and chemical data (n=4, total of 36 samples) to assess distinct ecophysiological  
6 responses along the latitudinal gradient (nine sites). All variables were standardized and  
7 displayed in a heat map. This analysis used Euclidean distances to examine patterns between  
8 sampled regions and specimens' chemical compounds and biomass values. The analyses were  
9 performed in R (R Development Core Team, 2018).

### 11 **Macroecological environment**

12 To analyze the macroecological environment of the sampled populations (i.e., the nine  
13 sites along the latitudinal gradient), a cluster analysis was performed over a set of biologically  
14 meaningful environmental variables (temperature, nitrate, phosphate, salinity, and light at  
15 bottom) downloaded from Bio-ORACLE v2.0 (Assis and others 2018). The analysis inferred  
16 the environmental dissimilarity between paired samples by using Ward's method over the  
17 Euclidean distance of the normalized values of the five variables (averaged for each site; Hair  
18 and others 2009). Additionally, a distance-based redundancy analysis (dbRDA) was performed  
19 to show the relationship between environmental data (four replicates per site) and the  
20 macroecological clustering of sites. In this process, the environmental variables were  
21 standardized with square root transformation and a site-by-environment dissimilarity matrix  
22 was produced based on Bray-Curtis distance (Legendre and Legendre 2012). To test the  
23 significance of each testable component, an ANOVA-like permutation test for the dbRDA  
24 was performed. Because the correlation between variables was low ( $R < 0.6$ ), no variables  
25 were excluded from both clustering and dbRDA analyses. This analyses were performed in R  
26 (R Development Core Team 2018).

### 28 **Responses to thermal stress between the center and edge populations**

30 Thermal stress responses were tested between the two sampled sites further apart, i.e.,  
31 Coqueiro, a tropical population characterizing the center of the species distribution and

1 Cardoso, a more temperate population characterizing the high latitude edge (Table S1).  
2 Environmental parameters (nutrients, salinity and temperature) were measured in the field  
3 and used as experimental control conditions (data according to Bio-ORACLE v2.0; see Table  
4 S2). Control temperatures (28°C for center and 25°C for the edge population) largely coincide  
5 with the average temperatures of the time series analyses (see Figure 1).

6 Seawater was sterilized by filtering (sand-filters 25 µm, 20 µm, and 5 µm; GF/F  
7 Whatman 0.45 µm filters) and exposure to ultraviolet light (UVC). Dissolved inorganic  
8 nutrients ( $\text{NO}_3^{-2}$ ,  $\text{NH}_4^+$ ,  $\text{PO}_4^{-3}$ ) were determined colorimetrically according to Tréguer and Le  
9 Corre (1975) and Grasshoff (1983), using a spectrophotometer (UV-1100, Pro-Analise ISE  
10 Quimica e Diagnostica Ltda, Brazil). Seawater was changed every 48h, and nutrient levels  
11 were adjusted by adding  $(\text{NH}_4)_2\text{SO}_4$ ,  $\text{Na}_2\text{HPO}_4$ , and  $\text{NaNO}_3$ . *Sargassum cymosum* fronds (~2.0  
12 g, n=5, apical portions of approximately 7 cm) were placed in Erlenmeyer flasks of 250 mL  
13 with sterilized seawater and acclimated for 20 days at the control treatment temperatures of  
14  $28^\circ\text{C} \pm 1^\circ\text{C}$  (tropical samples) and  $25 \pm 1^\circ\text{C}$  (warm temperate samples) at an irradiance of  
15  $110 \pm 10 \mu\text{mol photons.m}^{-2} \cdot \text{s}^{-1}$  (fluorescent lamps, Osram 30 W DayLight), with continuous  
16 aeration, and 12 h photoperiod starting at 6 a.m. The cleaning, identification, and storage of  
17 samples are detailed in the above section of *Ecophysiological responses along a latitudinal*  
18 *gradient* as well as in S4.

19 Experimental treatments included the exposure of both populations to four temperatures (25  
20 °C, 28 °C, 30 °C and 33 °C) for 30 days. Water temperature was manipulated using submersed  
21 heaters (Master, Brazil). Treatments were achieved by increasing temperatures by to 2°C per  
22 hour, and monitored with data loggers (HOBO Pendant Temperature, Onset, Bourne,  
23 USA). These temperatures were chosen based on time-series analyses of covering the period  
24 1993-2100 (Figure 1). This was produced with daily temperature data for the present (period  
25 1993-2019) retrieved from the Global Ocean Physics Reanalysis gridded dataset (Global  
26 Monitoring and Forecasting Center; E.U. Copernicus Marine Service), and daily data for the  
27 future (period 2020-2100) produced for two contrasting RCP scenarios (2.6 and 8.5) by  
28 ensembling distinct models from Coupled Model Intercomparison Project Phase 5  
29 (HadGEM2-ES, MIROC-ESM, IPSL-CM5A-LR) with the change factor approach, as detailed  
30 in Assis and others (2018). These data with daily temporal resolution allows estimating sharp

1 changes in temperature according to the patterns of MHW events (e.g., Oliver and others  
2 2019).

3 Relative growth rate (RGR) changes were determined from the beginning to the end  
4 of the experiment according to the following equation (Lignell and Pedersén 1989):

5 Growth rate (GR) =  $[(Wt/Wi)^{1/t} - 1] \times 100$

6 where  $W_i$  = initial fresh weight,  $W_t$  = final fresh weight, and  $t$  = experimental time

7 results are expressed as a daily percentage [% day<sup>-1</sup>] of biomass production.

8 Photosynthetic performance was evaluated *in vivo* (n=4) via pulse amplitude  
9 modulated (PAM) fluorometry of chlorophyll *a* in photosystem II (PSII) using a Jr-PAM  
10 (Walz, Germany). The parameters set before measuring were gain intensity = 2, measurement  
11 light = 8, light pulse = 0.8 s, n = 3 per replica. The maximum quantum yield (Fv/Fm) was  
12 determined after acclimating algae in the dark for 30 minutes. These are optimal parameters  
13 inferred in preliminary analyses testing a range of values. The final Fv/Fm values were  
14 calculated according to the equations from Schreiber and others (1986). In addition to relative  
15 growth rate and maximum quantum yield, we also tested the effect of increasing temperatures  
16 in the chemical composition of *S. cymosum*, as detailed in S4.

17 All analyses of ecophysiological performance (RGR; Fv/Fm) and chemical  
18 composition (n=4; carotenoids, phenolic compounds, Chl ac, total soluble sugars, and CN) at  
19 different temperatures (25°C, 28°C, 30°C, 33°C) were tested with two-way analysis of variance  
20 (ANOVA). Newman Keuls significant difference post hoc tests were used to identify the  
21 statistically different groups. Homogeneity of variance was tested a priori with the Cochran  
22 test. Variables were not standardized. All analyses were performed in R (R Development Core  
23 Team 2018).

24

## 25 **Results**

26

### 27 **Macroecological characterization and ecophysiological adjustments**

28

29 The cluster analysis using macroecological environmental variables divided the  
30 populations along the latitudinal gradient in two main clusters: tropical, comprising PI, RN,

1 PB BA, ES and warm temperate represented by RJ, SP and SC. The tropical cluster was further  
2 subdivided into two subclusters (Figure 2a).

3 The dbRDA analysis for macroecological clustering (Figure 3;  $R^2 = 0.72$ ) showed  
4 temperature, light and dissolved nutrients strongly segregating data along the first axis (higher  
5 temperature and light levels in tropical regions and higher nutrients in more temperate  
6 regions). Salinity correlated with the second axis (Figure 3) and explained the differentiation  
7 of the two tropical subclusters, although this axis explained much less variability compared  
8 to the first (74.90% vs. 15.45%).

9 The ecophysiological responses along the latitudinal gradient also showed the existence of  
10 two main groups, which coincided with the tropical and warm temperate regions (Figure 2b).  
11 Phenolic compounds, biomass, CN (Table S5), and total soluble sugars presented higher  
12 concentrations in the tropics, while carotenoids, proteins and chlorophylls were higher in the  
13 warm temperate region.

#### 15 **Responses to thermal stress between center and edge populations**

17 The two populations analyzed displayed contrasting thermal tolerances, with  
18 individuals from the tropical (center) region better coping with thermal stress, when  
19 compared to those from the temperate (edge) region (Figure 4; lethal temperatures of 33°C vs.  
20 30°C); yet both lost biomass in response to increasing thermal stress, above control conditions.  
21 The center population lost biomass with temperatures > 30°C and the edge population with  
22 temperatures > 28°C. Future temperature analyses for RCP 2.6 show the center region  
23 frequently exposed to 30°C, especially after 2025, but not to the lethal threshold of 33°C.  
24 Under RCP 8.5 scenario, this region is expected to reach and remain permanently above the  
25 30°C after 2030, and above 33°C by the end of the century (2080). For the edge population,  
26 future projections follow the same trend, with RCP 2.6 temperatures frequently overcoming  
27 the biomass loss threshold of 28°C, especially after 2025, and the RCP 8.5 temperatures  
28 reaching the lethal temperature of 30°C after 2050 (Figure 1).

29 The individuals of both center and edge populations produced higher carotenoids and  
30 phenolic compounds content with rising temperature (Figure 5a, b). The population of the  
31 center region showed 60% more chlorophyll concentration and 10% Fv/Fm higher values  
32 when compared to the edge population at 30°C (Figure 5c, d). On the other hand, total soluble



1 sugars content was reduced with increasing temperatures; the population from the edge lost  
2 more than ~70% when exposed to 28°C and 30°C, when compared to the control treatment  
3 (i.e., 25°C; Figure 5e). This had consequences for the CN ratio, which showed more than ~50%  
4 decrease at these same temperatures (Figure 5f and Table S6). All analyzes showed significant  
5 differences (ANOVA  $p < 0.001$ , Table S3).

## 9 **Discussion**

11 The present study shows marked ecophysiological adjustments of *S. cymosum* forests  
12 driven by distinct macroecological environmental conditions, which coincide with the  
13 previously defined biogeographical provinces (Spalding and others 2007). Tropical  
14 populations exposed to higher levels of irradiation, temperature and lower nutrient conditions  
15 were characterized by higher total soluble sugars content, CN ratio and biomass, while  
16 populations in more temperate environments tend to produce more photosynthetic pigments  
17 and proteins. When exposed to increasing thermal stress, populations lose biomass and change  
18 ecophysiological adjustments, but not to the same extent. Populations from contrasting ranges  
19 (i.e., center vs. edge) showed signs of phenotypic plasticity, with the individuals  
20 characterizing the center of the species' distribution tolerating up to 30°C, a temperature  
21 leading individuals of the edge to perish. These results have much relevance in the scope of  
22 projected environmental changes, as plasticity might ensure long-term survival in the face of  
23 extreme conditions, yet future projections render high relevance to the RCP scenario chosen.  
24 While less demographic changes might occur in the RCP scenario aligned with the Paris  
25 Agreement (i.e., RCP 2.6), potential local extirpations are expected for both center and edge  
26 populations in the no mitigation scenario. Broad compliance with the Paris Agreement might  
27 therefore counteract such expectations, safeguarding *Sargassum* forests in the years to come.

28 Across a broad latitudinal gradient with more than 4000 km, *S. cymosum* forests  
29 inhabit two main macroecological environments, which largely coincided with the Tropical  
30 Atlantic and Warm Temperate Southwestern Atlantic marine provinces defined by Spalding  
31 and others (2007). At a macroecological level, analyses showed the expected general trend of  
32 higher irradiance and temperature levels in the tropics and nutrient enriched conditions

1 (phosphate and nitrate) in the more temperate region. Between these two regions, *S.*  
2 *cymosum* forests exhibited contrasting biomass and ecophysiological adjustments. In  
3 particular, biomass and carbon stocks were higher in the tropics, as the result of physiological  
4 trade-offs between potential growth and structural maintenance (Mauffrey and others 2020).  
5 This pattern is in the light of the abundant-center hypothesis (but see Sagarin and Gaines  
6 2002; Ntuli and others 2020), which predicts higher fitness of individuals and more abundant  
7 populations at the center of the species distribution, owing to more favorable conditions  
8 (Brown 1984; Brussard 1984). There, populations might have increased photosynthetic  
9 capacity and fixed carbon available for growth, resulting in higher biomass, CN and total  
10 soluble sugars content (Manns and others 2017). Also, sugar content in macroalgae has been  
11 shown to be inversely related to nutrient conditions (Marinho-Soriano and others 2006) as a  
12 consequence of the reallocation of carbon skeletons necessary to reduce nitrate and form  
13 amino acids (Huppe and Turpin 1994). On the other hand, the populations from the  
14 temperate region produced more pigments and proteins. Previous findings also support these  
15 results, showing that pigment's response has been attributed to high nutrient concentration  
16 in *Sargassum* forests (Gaol and Nakahara 1990).

17 The tropical populations (i.e., center of distribution) also showed increased  
18 production of phenolic compounds content (Hargrave and others 2017), suggesting that *S.*  
19 *cymosum* may adjust chemical defensive barriers against oxidative stress (Costa and others  
20 2015, 2019), in response to environments characterized by elevated solar radiation, as shown  
21 in the macroecological analysis. Previous studies with brown seaweeds along the same  
22 latitudinal gradient support the observed pattern of higher photoprotective compounds  
23 towards the tropics (Schmitz and others 2018). Higher herbivory pressure could also explain  
24 this (Poore and others 2012; Zarco-Perello and others 2017; Vasconcelos and others 2019).  
25 Despite the lack of data to test this, the general latitudinal herbivory-defense hypothesis  
26 predicts higher herbivore pressure toward the tropics, with tropical populations developing  
27 higher levels of defenses than those at higher latitudes (Gao and others 2019). Increasing  
28 thermal stress led to populations from both center and edge regions to increase carotenoid  
29 production, which acts in dissipating the excess light energy through the xanthophyll cycle  
30 and also as accessory pigments of photosynthesis, which together with chlorophylls, sustains  
31 photosynthetic activity (Takaichi 2011; Balboa and others 2013). Also, carotenoids can be  
32 degraded inversely to the chlorophyll content, i.e., in the treatments at higher temperatures,

1 reduced chlorophyll concentration was observed while carotenoids increased, indicating their  
2 potential role as an energy source for maintaining physiological integrity. The reduction in  
3 chlorophylls content observed in the edge population under thermal stress also impacted  
4 Fv/Fm, as a possible consequence of light-harvesting capacity modification affecting net  
5 carbon fixation and re-allocation of energy derived from photosynthesis (Lalegerie and others  
6 2020). In general, the carbohydrates reserve is a ratio between accumulation from  
7 photosynthesis and consumption for energy production, biosynthesis, and growth. Therefore,  
8 CN imbalance can cause adverse effects to brown algae, impacting photosynthetic pigments,  
9 carbohydrates and proteins (Urrea-Victoria and others 2020).

10 *S. cymosum* populations from contrasting ranges (i.e., center vs. edge) showed  
11 different growth and physiological responses when exposed to increasing thermal stress.

12 At control conditions, both populations exhibited maximum growth rates, as observed for  
13 additional *Sargassum* species (*S. vestitum* in Poore and others 2016; *S. polycystum* in Graba-  
14 Landry and others 2020), as well as increased photosynthesis, chlorophylls, CN ratio and total  
15 soluble sugars. However, thermal tolerances differed by up to 3°C, with the individuals from  
16 the center of the species distribution more adapted to warmer conditions. This supports the  
17 idea of phenotypic plasticity, i.e., the ability of an individual to change the phenotype in  
18 response to environmental changes (king and others 2017). Yet, our data does not allow  
19 concluding whether local adaptation might also be implied, as this process acts through the  
20 selection and changing of allele frequencies towards a local optimum (king and others 2017).  
21 In such conditions, the species may have adapted to local temperature conditions, resulting  
22 in an ecotype with different optimal temperatures for photosynthesis and growth in center,  
23 compared to edge populations (Gerard and Du Bois 1988; Pakker and others 1996; Gouvêa  
24 and others 2017). Thermal tolerance differences between populations from contrasting ranges  
25 are widespread in marine macrophytes (King and others 2018), but the observed pattern of  
26 more warm-tolerant populations in the center, completely contrasts with previous studies,  
27 which were mostly performed with cold-temperate species with range edges typically  
28 exposed to warmer conditions than the center (e.g., Gerard and Du Bois 1988; Bennett and  
29 others 2015; Saada and others 2016).

30 Despite the evidence of phenotypic plasticity, significant future losses are anticipated  
31 for both center and edge populations, but severity depends on the future scenario. Under the  
32 mitigation scenario complying with the Paris agreement (RCP 2.6), global warming is not

1 expected to reach lethal thresholds, but biomass losses are anticipated for both populations.  
2 Such losses are already evident in field observations and are expected to further increase by  
3 2025. *Sargassum* is already declining 2.6% per year in the last decades, and overall losses in  
4 biomass sum up to 52% (ranging from 20% to 89%) since systematic records began in 1969 on  
5 the Brazilian coastline (Gorman and others 2020). Potential loss of the *Sargassum* forests could  
6 trigger cascading ecosystem turnovers, impacting the local biodiversity and turning these rich  
7 and complex ecosystems to, e.g., sea urchin barrens, unable to support the ecological services  
8 of the previous forests (Arafeh-Dalmau and others 2020; Bernal-Ibáñez and others 2021).  
9 Under the no-mitigation scenario (RCP 8.5), global warming impacts on *S. cymosum* are  
10 anticipated to be more severe. Biomass loss thresholds are projected to be permanently crossed  
11 by 2050 and lethal thresholds by the end of the century, leading the *S. cymosum* populations  
12 of both center and edge ranges to perish. This may further contribute to the loss of genetic  
13 diversity due to population bottlenecks, as observed for additional macroalgal species (Gurgel  
14 and others 2020), possibly reducing the evolutionary capacity of the species as a whole  
15 (Hampe and Petit 2005). For instance, *Sargassum fallax* and *Scytothalia dorycarpa* have lost  
16 between 30% to 65% of their average genetic diversity due to population declines while  
17 exposed to a heatwave event (Gurgel and others 2020). Considering that *Sargassum* species  
18 can disperse over long distances (Mattio and others 2013), potential gene flow from more  
19 adapted tropical to warm-temperate populations could improve regional thermal fitness. This  
20 would depend on direction of ocean current transport (Molinos and others 2017), and on  
21 warming not exceeding the inferred critical thermal thresholds.

22         The knowledge about divergent ecophysiological responses between populations  
23 along species' distributional ranges is crucial to estimate the potential impacts of climate  
24 change. Distributional and abundance changes have been reported and projected worldwide,  
25 both in the central and edge populations of marine and terrestrial species, beyond the  
26 *Sargassum* forests here addressed (Galaiduk and others 2013; Vergés and others 2014;  
27 Poloczanska and others 2016; Cavanaugh and others 2019; Osland and others 2021). Because  
28 phenotypic plasticity / local adaptation has been increasingly reported elsewhere (e.g.,  
29 crustacean and echinoid; Paganini and others 2014), additional species could display the  
30 pattern of *S. cymosum*, of drastic climate-induced impacts, regardless of distinct population  
31 ecophysiological responses.

1 Overall, our findings represent a baseline to predict the survival of *S. cymosum*  
2 populations in the future, so coastal management strategies focused on this macroalgae may  
3 help maintaining ecosystem services at higher structural and functional levels. In this context,  
4 broad compliance with the Paris Agreement is critical to reducing the potential impacts of  
5 climate change, as it may be more important than plasticity in safeguarding biodiversity in  
6 the years to come.

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**Figure legends:**

Figure 1: Time series of sea surface temperature for tropical and warm temperate regions for present-day conditions (in grey) and contrasting scenarios of projected climate change (in blue RCP 2.6, and in red RCP 8.5). Temperature data for the present (period 1993-2019) were accessed from the Global Monitoring and Forecasting Center (Copernicus Marine Service), while projected conditions were accessed from the Coupled Model Intercomparison Project Phase 5 (models HadGEM2-ES, MIROC-ESM, IPSL-CM5A-LR).

Figure 2: (a) Sites where *S. cymosum* samples were collected along a latitudinal gradient and cluster analysis of environmental data (maximum temperature, nitrate, phosphate, light at bottom and salinity) separating the sites into two main regions. Abbreviations correspond to

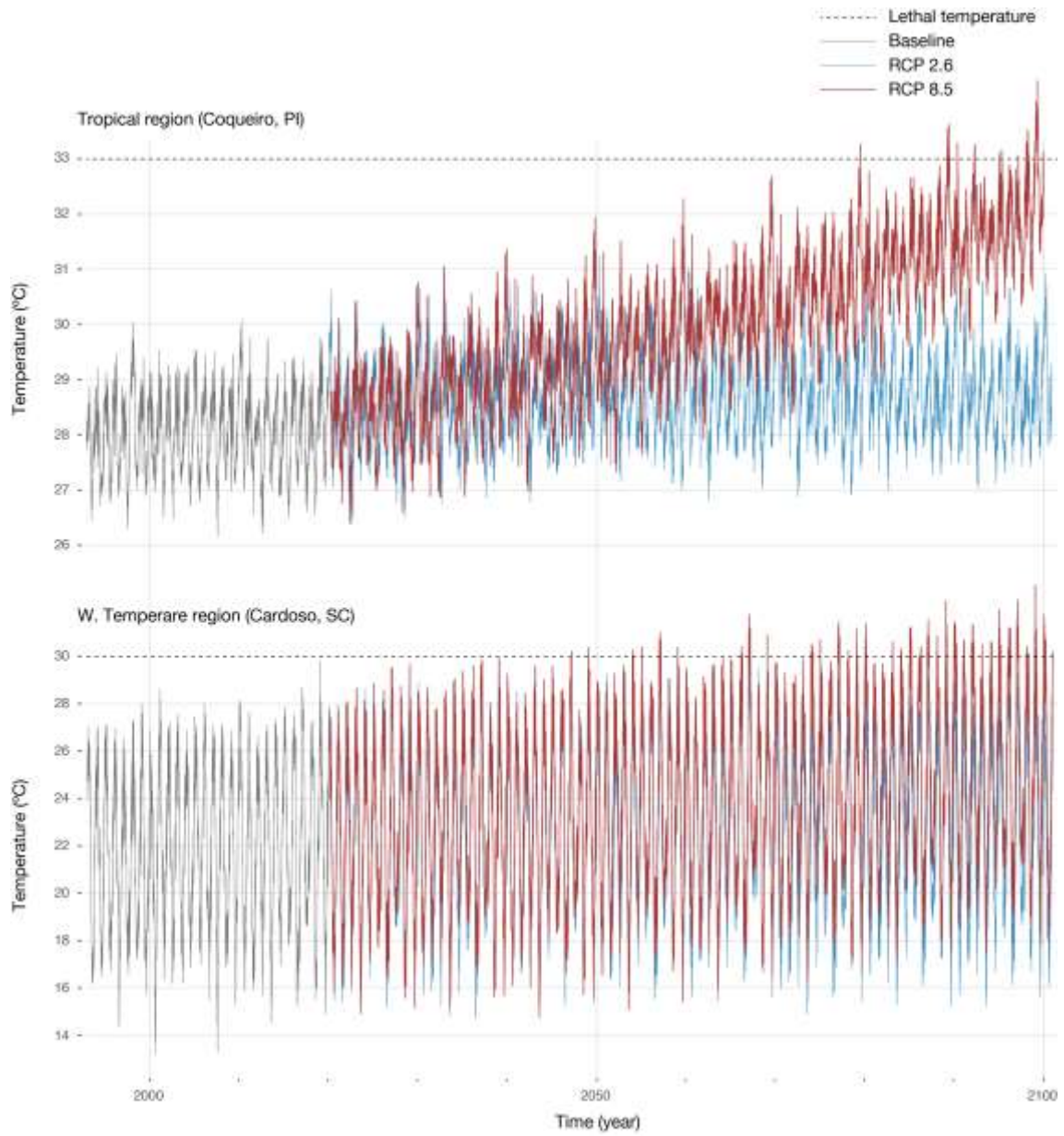
1 Piauí (PI), Rio Grande do Norte (RN), Paraíba (PB), Bahia (BA), Espírito Santo (ES), Rio de  
2 Janeiro (RJ), São Paulo (SP) and Santa Catarina (SC). (b) Relationship between environmental  
3 clustering and chemical composition and biomass inferred for 36 *S. cymosum* specimens  
4 sampled along the latitudinal gradient in Brazilian coastline (9 sites, n=4, two replicates in  
5 SC). Dendrograms depict hierarchical clustering between regions and samples. Gradient from  
6 blue to orange shows values from low to high chemical concentrations and biomass values (-  
7 3 and +3 are standardized values). The tropical province was characterized by the high content  
8 of phenolic compounds, total soluble sugars, CN and biomass while warm temperate by  
9 carotenoids, Chl ac and total soluble proteins.

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12 Figure 3: Distance-based redundancy analysis (dbRDA) biplot of first and second axes  
13 depicting the multivariate dissimilarity of macroecological environment data of *S. cymosum*  
14 populations in association with tropical (i.e., central populations; gray circles) and warm  
15 temperate (i.e., edge populations; black circles) regions.

16  
17 Figure 4: Relative growth rate (RGR, % d<sup>-1</sup>) of warm temperate and tropical *S. cymosum* under  
18 the temperature stress experiment (30 days exposure under 25°C, 28°C, 30°C and 33°C). The  
19 temperatures of 25 °C and 28 °C were used as control treatment to warm temperate and  
20 tropical populations, respectively. Data are shown as mean ±SD (n = 5). Different letters  
21 indicate significant differences, p < 0.05 (Newman–Keuls test, two-way ANOVA).

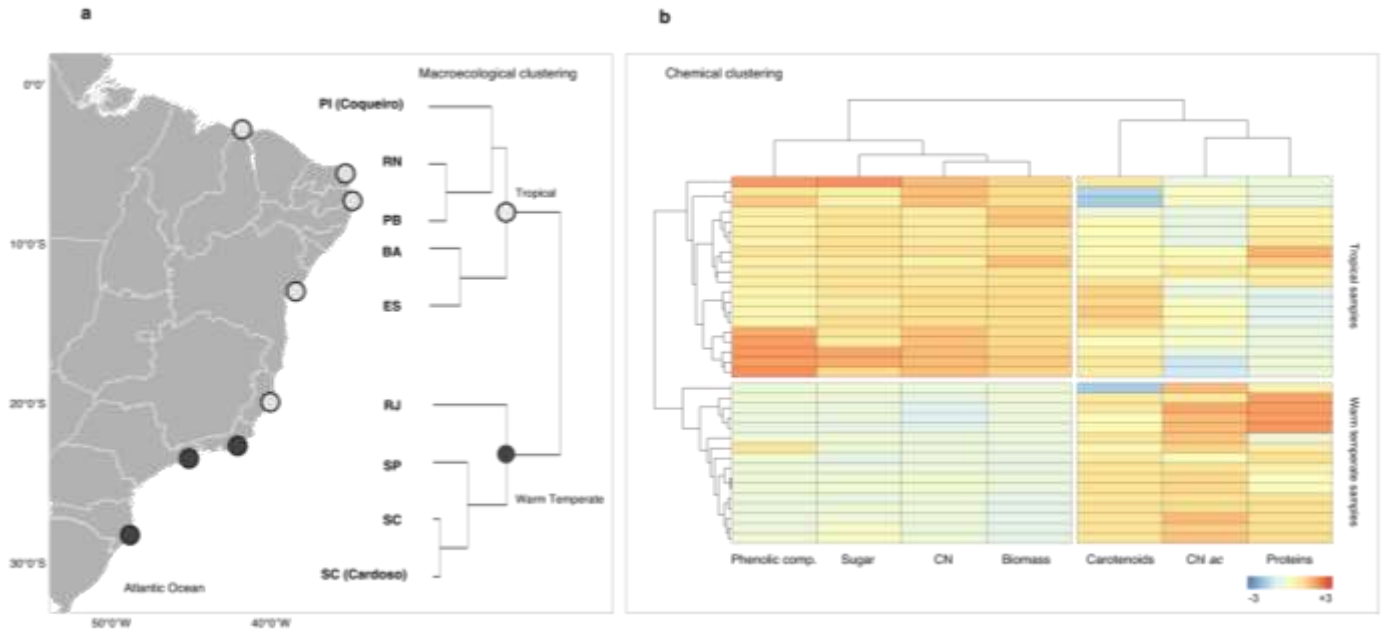
22  
23 Figure 5: Chemical composition of warm temperate and tropical *S. cymosum* samples under  
24 the temperature stress experiment (30 days exposure under 25°C, 28°C, 30°C and 33°C). The  
25 temperatures of 25 °C and 28 °C were used as control treatment to warm temperate and  
26 tropical populations, respectively. Data are shown as mean ±SD (n = 4). Different letters  
27 indicate significant differences, p < 0.05 (Newman–Keuls test, two-way ANOVA). (a)  
28 Carotenoids, (b) Phenolic compounds, (c) Chlorophylls a and c, (d) Fv/Fm, (e) Total soluble  
29 sugars and (f) CN ratio present in *S. cymosum*.

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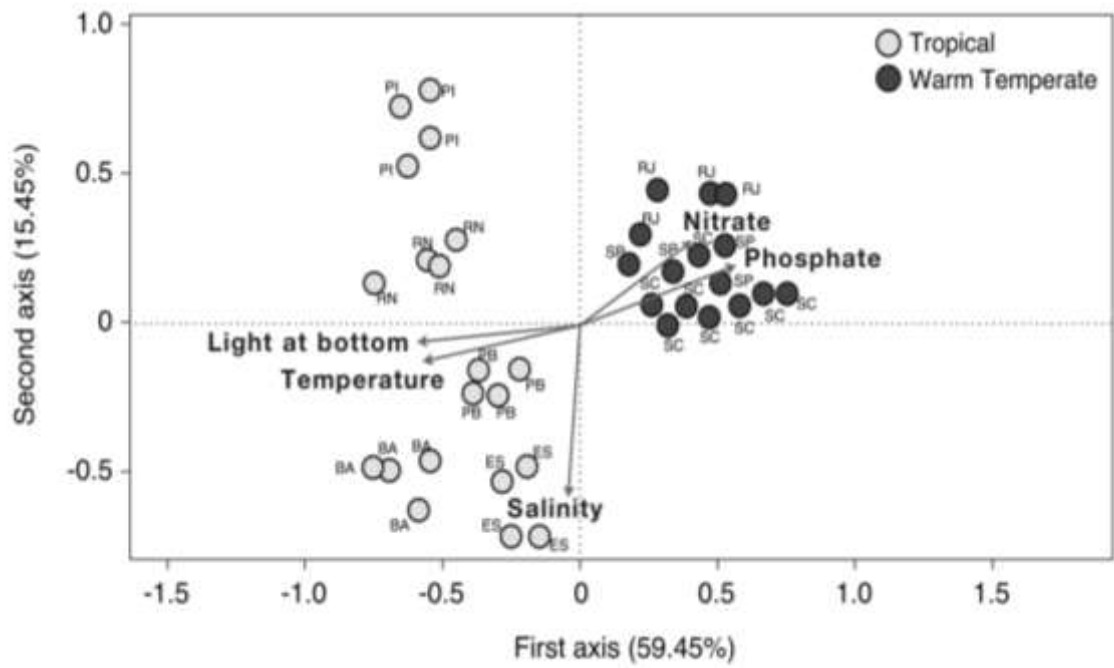




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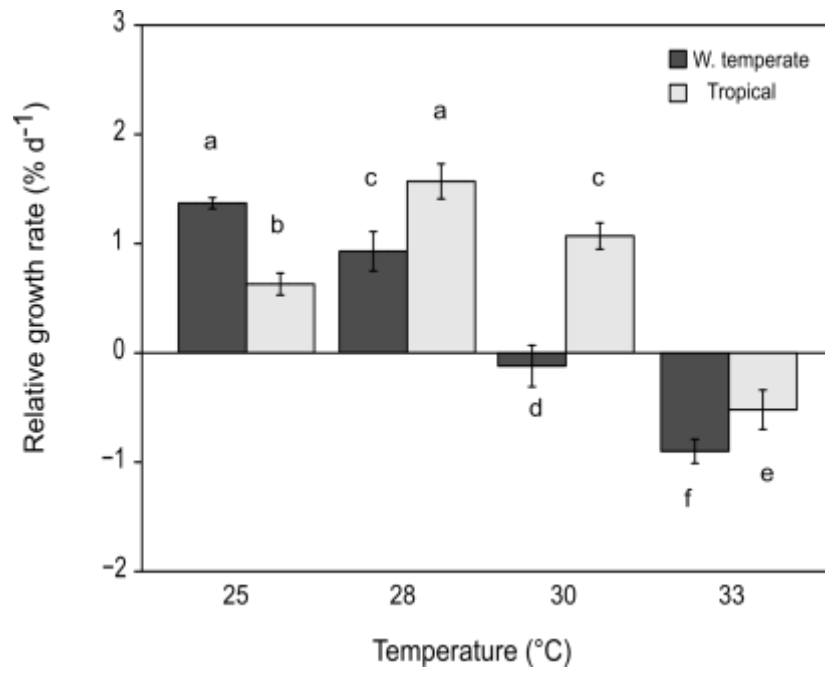
Figure 2

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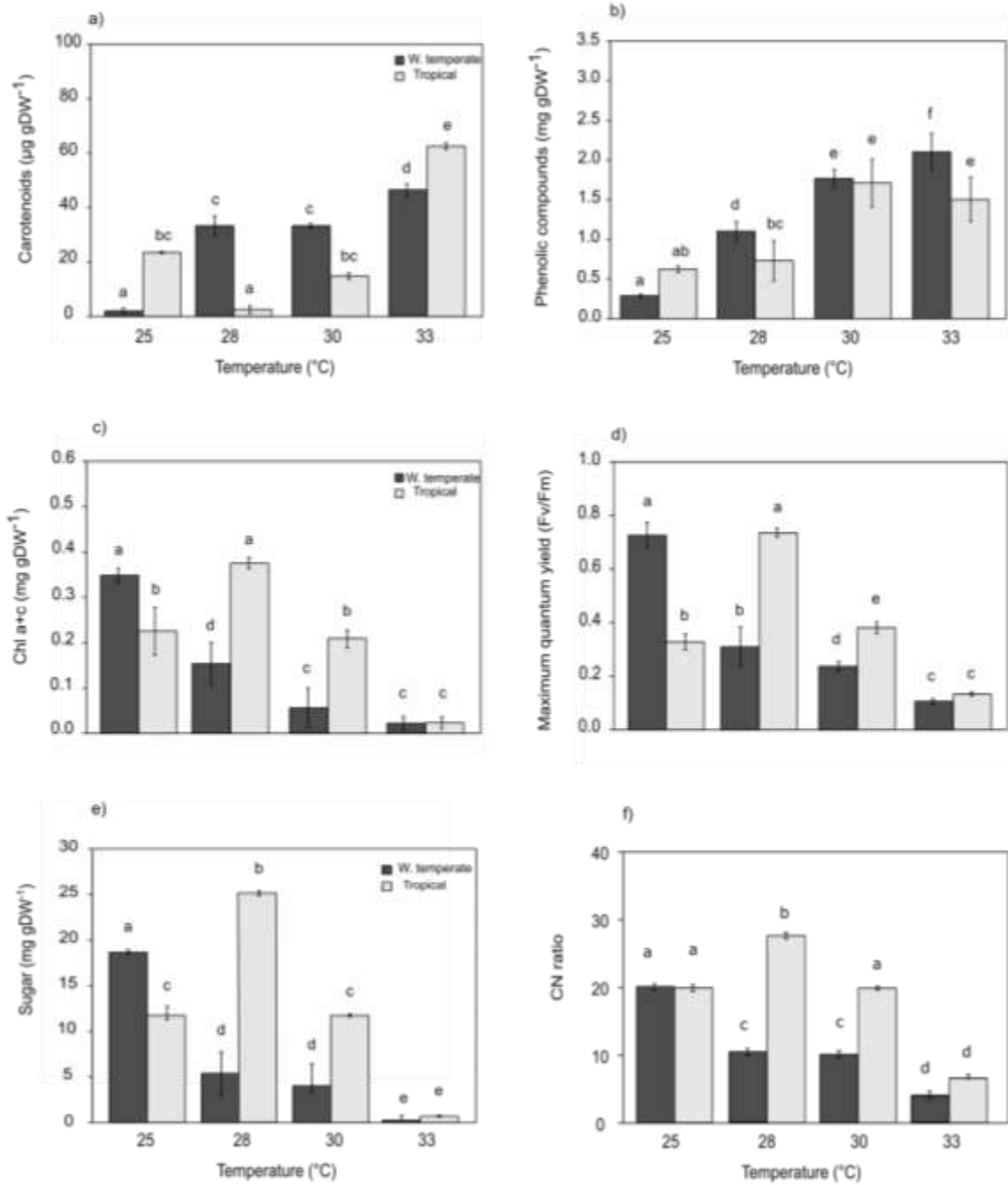


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