

# Diversity of the tussock moths (Lepidoptera, Erebidae, Lymantriinae) of the wider Maputaland-Pondoland-Albany, with special reference to the fauna of Mozambique

MICHAL RINDOŠ<sup>1,2</sup>, ALEXANDER SCHINTLMEISTER<sup>3</sup>, ZDENĚK FALTÝNEK FRIC<sup>1</sup>

1 *The Czech Academy of Sciences, Biology Centre, Institute of Entomology, Branišovská 31, 37005 České Budějovice, Czech Republic; E-mail: rindom00@prf.jcu.cz, fric@entu.cas.cz*

2 *University of South Bohemia, Faculty of Science, Branišovská 1760, 37005 České Budějovice, Czech Republic*

3 *Calberla Strasse 3, 1D- 01326 Dresden, Germany; E-mail: schintlm@aol.com*

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**Abstract.** Maputaland-Pondoland-Albany, one of the African biodiversity hotspots, is defined by a high diversity of its fauna and flora. Despite numerous studies published about the Lepidoptera of this region, our knowledge of diversity and distributional patterns of moths still remains inadequate. Our study evaluates the tussock moth richness and endemism of the wider Maputaland-Pondoland-Albany region. We describe for the first time the variety of the tussock moth fauna of Mozambique, with seven newly recorded genera (*Euproctoides* Bethune-Baker, 1911; *Heteronygmia* Holland, 1893; *Leucoma* Hübner, 1822; *Ogoa* Walker, 1856; *Paraproctis* Bethune-Baker, 1911; *Polymona* Walker, 1855; *Tamsita* Kiriakoff, 1954).

## Introduction

Biodiversity hotspots are defined as highly threatened biogeographic regions with the occurrence of endemic species of animals and plants (Myers 1988). Containing approximately half of the world's plant and a significant percentage of world's vertebrate diversity (Myers et al. 2000), conservation and thorough investigation of these 35 remarkable regions distributed worldwide became essential for many researchers due to ongoing species mass extinction (Mittermeier et al. 2011; Eisenhauer et al. 2019). The majority of the African hotspots are located in the Afrotropics (Myers et al. 2000). Maputaland-Pondoland-Albany is one of the three recently recognized biodiversity hotspots in the region of southern Africa (Fig. 1). Diversity and endemism are focused in the three major units: Maputaland in the north, Pondoland in the south and Albany in the south-west, comprising fauna and flora of South Africa, Eswatini (Swaziland) and Mozambique (Perera et al. 2011). The dominant habitats of this region bearing the most unique diversity are tropical and subtropical moist broadleaf forests and mountain grasslands and scrublands. Despite the uniqueness of this area serving as a meeting point for six of the eight southern African biomes, only 25% of the original extent of the natural habitat remains preserved (Mittermeier et al. 2004).

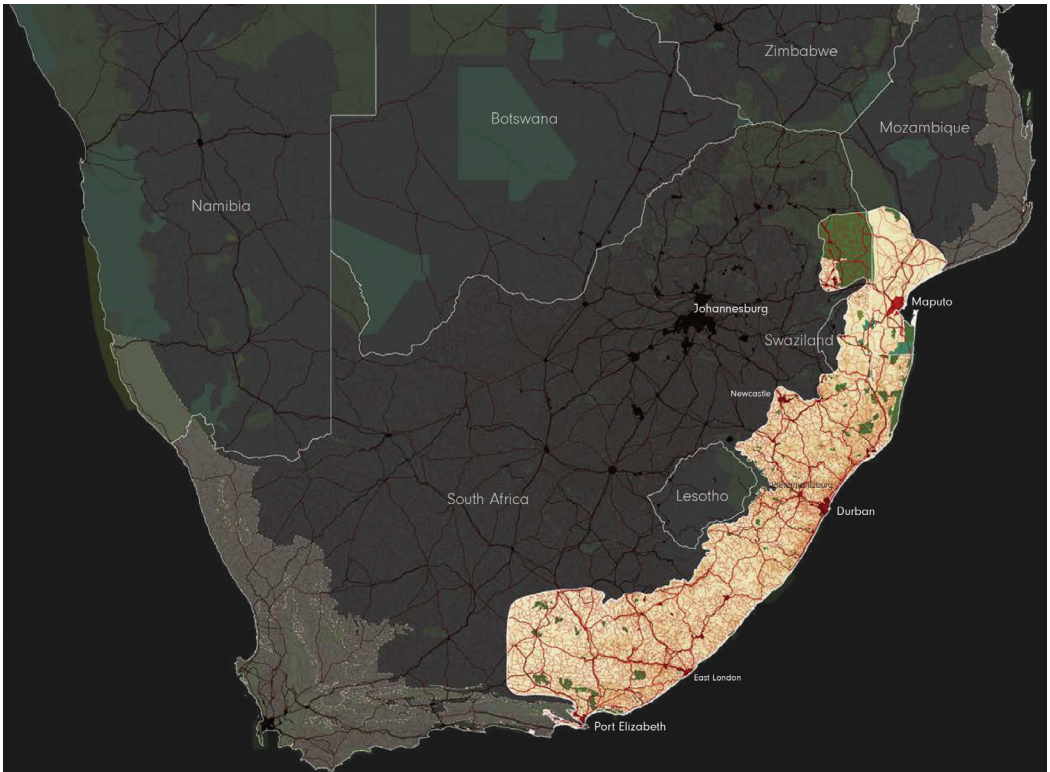
The moth subfamily Lymantriinae (Noctuoidea, Erebidae) occurs worldwide and comprises more than 2600 species in almost 360 genera (Kitching and Rawlins 1998). South Africa can be considered a well-studied part of the Maputaland-Pondoland-Albany, with several studies discussing the tussock moth fauna (Hampson 1905 – 77 spp., Janse 1917 – 109 spp., Pinhey 1975 – 57 spp.,

Vári *et al.* 2002 – 187 spp., Staude *et al.* 2020 – 34 spp.). However, knowledge of the Lymantriinae of the other southern African countries remains fairly insufficient. Presently, approximately 50 species of tussock moths are known to occur in Eswatini (Duke *et al.* 1999) and five species are reported from Lesotho (Kopij 2006). The current knowledge of the fauna of Mozambique is based mainly on past taxonomic works carried out on museum materials (e.g., Collette 1955) counting roughly 50 species. Nonetheless, previously published works also include numerous mistakes in systematics (e.g., published synonyms, identification errors, etc.) and creates “ghost” diversity.

The purpose of this study was to provide an up-to-date overview and fill the gaps in the poor knowledge in the diversity of tussock moths of Afrotropical biodiversity hotspots. Verified distribution data also recovers potential endemic species and thus plays a significant role in the global conservation effort. In addition, we provide a first faunistic summary of the tussock moths of Mozambique, recording seven genera new to the country.

## Material and methods

For the compilation of the list of Lymantriinae of the wider Maputaland-Pondoland-Albany we used presence/absence datasets covering the fauna of South Africa, Lesotho, Eswatini and Mozambique (Table 1). The data were mined from literature (e.g., Vári *et al.* 2002), corrected and supplemented by data obtained by the revision of the following museum collections: African Nat-



**Figure 1.** Map of the studied area, modified from Weller *et al.* (2017). The area of the Maputaland-Pondoland-Albany is highlighted by lighter colour.

**Table 1.** The presence, or absence of tussock moth species of the wider Maputaland-Pondoland-Albany. New genus records are marked with symbol (\*) and endemic species highlighted with bold.

| Species  | Mozambique | Lesotho | South Africa | Eswatini |
|--|------------|---------|--------------|----------|
| <b><i>Aroa anthora</i> (Felder R., 1874)</b>         | 0          | 0       | 1            | 0        |
| <i>Aroa difficilis</i> Walker, 1865                  | 0          | 0       | 1            | 1        |
| <i>Aroa discalis</i> Walker, 1855                    | 1          | 0       | 1            | 1        |
| <i>Aroa melanoleuca</i> Hampson, 1905                | 1          | 0       | 1            | 0        |
| <i>Bracharoa dregei</i> (Herrich-Schäffer, 1854)     | 0          | 1       | 1            | 0        |
| <i>Bracharoa mixta</i> (Snellen, 1872)               | 1          | 0       | 1            | 0        |
| <i>Bracharoa quadripunctata</i> (Wallengren, 1875)   | 1          | 1       | 1            | 1        |
| <i>Bracharoa tricolor</i> (Herrich-Schäffer, 1856)   | 0          | 1       | 1            | 0        |
| <b><i>Cataphractes arenacea</i> (Linnaeus, 1767)</b> | 0          | 0       | 1            | 0        |
| <i>Cataphractes fasciata</i> (Walker, 1855)          | 0          | 0       | 1            | 1        |
| <i>Cimola opalina</i> Walker, 1855                   | 1          | 0       | 1            | 1        |
| <b><i>Conigephyra rikatia</i> Collenette, 1956</b>   | 1          | 0       | 0            | 0        |
| <i>Creagra liturata</i> (Guérin-Ménéville, 1844)     | 1          | 0       | 1            | 1        |
| <i>Cropera phlebitis</i> (Hampson, 1905)             | 1          | 0       | 1            | 0        |
| <i>Cropera sericea</i> (Hampson, 1910)               | 0          | 0       | 1            | 1        |
| <i>Cropera stilpnaroma</i> Hering, 1926              | 0          | 1       | 1            | 0        |
| <i>Cropera testacea</i> Walker, 1855                 | 1          | 0       | 1            | 1        |
| <i>Crorema adspersa</i> (Herrich-Schäffer, 1854)     | 0          | 0       | 1            | 1        |
| <i>Crorema fulvinotata</i> (Butler, 1893)            | 1          | 0       | 1            | 0        |
| <i>Crorema setinoides</i> (Holland, 1893)            | 0          | 0       | 1            | 0        |
| <i>Dasychira albimaculata</i> Hering, 1926           | 0          | 0       | 1            | 1        |
| <i>Dasychira amata</i> Hering, 1926                  | 0          | 0       | 1            | 0        |
| <b><i>Dasychira aurivillii</i> Hering, 1926</b>      | 0          | 0       | 1            | 0        |
| <i>Dasychira batoides</i> (Plötz, 1880)              | 0          | 0       | 1            | 0        |
| <b><i>Dasychira bryophilina</i> Hampson, 1910</b>    | 0          | 0       | 1            | 0        |
| <b><i>Dasychira confinis</i> Distant, 1899</b>       | 0          | 0       | 1            | 0        |
| <i>Dasychira curvivirgata</i> Karsch, 1895           | 0          | 0       | 1            | 1        |
| <b><i>Dasychira didyma</i> Collenette, 1956</b>      | 0          | 0       | 1            | 0        |
| <i>Dasychira diplogramma</i> Hering, 1927            | 0          | 0       | 1            | 0        |
| <b><i>Dasychira eclipses</i> Collenette, 1939</b>    | 0          | 0       | 1            | 0        |
| <i>Dasychira esthlopiis</i> Collenette, 1953         | 0          | 0       | 1            | 0        |
| <i>Dasychira extatura</i> Distant, 1897              | 1          | 0       | 1            | 1        |
| <i>Dasychira extorta</i> Distant, 1897               | 1          | 0       | 1            | 1        |
| <b><i>Dasychira fusca</i> (Walker, 1855)</b>         | 0          | 0       | 1            | 0        |
| <i>Dasychira gephyra</i> Hering, 1926                | 0          | 0       | 1            | 0        |
| <i>Dasychira hamptoni</i> Hering, 1926               | 1          | 0       | 1            | 0        |
| <i>Dasychira herbida</i> (Walker, 1856)              | 0          | 0       | 1            | 0        |
| <b><i>Dasychira hughesi</i> Collenette, 1933</b>     | 0          | 0       | 1            | 0        |
| <i>Dasychira lunensis</i> Hampson, 1905              | 0          | 0       | 1            | 0        |
| <i>Dasychira mediofasciata</i> Hering, 1926          | 0          | 0       | 1            | 0        |
| <i>Dasychira nubifuga</i> (Holland, 1893)            | 0          | 0       | 1            | 1        |
| <i>Dasychira octophora</i> (Hampson, 1905)           | 1          | 0       | 1            | 1        |
| <i>Dasychira phenax</i> Collenette, 1932             | 0          | 0       | 1            | 0        |
| <i>Dasychira pheosia</i> Hampson, 1910               | 0          | 0       | 1            | 1        |
| <i>Dasychira pluto</i> Hering, 1926                  | 0          | 0       | 1            | 0        |
| <i>Dasychira polia</i> Hering, 1926                  | 0          | 0       | 1            | 0        |
| <i>Dasychira postpura</i> Hampson, 1905              | 0          | 0       | 1            | 1        |
| <i>Dasychira pulcherrima</i> Hering, 1926            | 0          | 0       | 1            | 0        |
| <i>Dasychira pyrosoma</i> Hampson, 1910              | 0          | 0       | 1            | 1        |
| <i>Dasychira rocana</i> Swinhoe, 1906                | 1          | 0       | 1            | 1        |
| <i>Dasychira thanatos</i> Hering, 1926               | 0          | 0       | 1            | 0        |
| <i>Dasychira vilis</i> (Felder R., 1874)             | 0          | 0       | 1            | 0        |
| <i>Eudasychira amata</i> (Hering, 1926)              | 0          | 0       | 1            | 0        |

| Species   | Mozambique | Lesotho | South Africa | Eswatini |
|---|------------|---------|--------------|----------|
| <i>Eudasychira georgiana</i> (Fawcett, 1901)              | 1          | 0       | 1            | 0        |
| <i>Eudasychira metathermes</i> (Hampson, 1905)            | 1          | 0       | 1            | 1        |
| <i>Eudasychira poliotis</i> (Hampson, 1910)               | 0          | 0       | 1            | 0        |
| <i>Eudasychira proleprota</i> (Hampson, 1905)             | 0          | 0       | 1            | 0        |
| <i>Euproctis aethiopica</i> Snellen, 1872                 | 0          | 0       | 1            | 1        |
| <b><i>Euproctis aspersum</i> (Felder R., 1874)</b>        | 0          | 0       | 1            | 0        |
| <i>Euproctis beato</i> Bryk, 1934                         | 0          | 0       | 1            | 0        |
| <i>Euproctis bicolor</i> Walker, 1855                     | 1          | 1       | 1            | 1        |
| <b><i>Euproctis chionea</i> Collenette, 1956</b>          | 0          | 0       | 1            | 0        |
| <i>Euproctis crocata</i> (Boisduval, 1847)                | 0          | 0       | 1            | 0        |
| <b><i>Euproctis flavicincta</i> Janse, 1915</b>           | 0          | 0       | 1            | 0        |
| <i>Euproctis haemodetes</i> Hampson, 1905                 | 0          | 0       | 1            | 1        |
| <b><i>Euproctis hardenbergia</i> (Janse, 1915)</b>        | 0          | 0       | 1            | 0        |
| <b><i>Euproctis iridescens</i> Janse, 1915</b>            | 0          | 0       | 1            | 0        |
| <b><i>Euproctis kettlewelli</i> Collenette, 1956</b>      | 0          | 0       | 1            | 0        |
| <i>Euproctis mayottensis</i> Collenette, 1956             | 0          | 0       | 1            | 0        |
| <b><i>Euproctis melanura</i> (Wallengren, 1860)</b>       | 0          | 0       | 1            | 0        |
| <b><i>Euproctis nigripuncta</i> Janse, 1915</b>           | 0          | 0       | 1            | 0        |
| <i>Euproctis pallida</i> (Kirby, 1896)                    | 1          | 0       | 1            | 1        |
| <b><i>Euproctis petavia</i> (Stoll, 1782)</b>             | 0          | 0       | 1            | 0        |
| <i>Euproctis producta</i> (Walker, 1863)                  | 0          | 1       | 0            | 0        |
| <i>Euproctis punctifera</i> (Walker, 1855)                | 1          | 0       | 1            | 0        |
| <i>Euproctis rufopunctata</i> (Walker, 1862)              | 1          | 0       | 1            | 0        |
| <i>Euproctis sanguigutta</i> Hampson, 1905                | 0          | 0       | 1            | 1        |
| <b><i>Euproctis stellata</i> Distant, 1897</b>            | 0          | 0       | 1            | 0        |
| <b><i>Euproctis straminicolor</i> Janse, 1915</b>         | 0          | 0       | 1            | 0        |
| <b><i>Euproctis subalba</i> (Janse, 1915)</b>             | 0          | 0       | 1            | 0        |
| <i>Euproctis terminalis</i> (Walker, 1855)                | 0          | 1       | 1            | 1        |
| <i>Euproctoides ertli</i> (Wichgraf, 1922) *              | 1          | 0       | 1            | 0        |
| <i>Griveaudyria cangia</i> (Druce, 1887)                  | 1          | 0       | 1            | 0        |
| <i>Hemerophanes libyria</i> (Druce, 1896)                 | 1          | 0       | 1            | 1        |
| <i>Heteronygmia dissimilis</i> (Aurivillius, 1910) *      | 1          | 0       | 1            | 0        |
| <b><i>Homochira poecilosticta</i> Collenette, 1938</b>    | 1          | 0       | 0            | 0        |
| <i>Homochira rendalli</i> (Distant, 1897)                 | 1          | 0       | 1            | 1        |
| <b><i>Homoeomeria flavicapilla</i> (Wallengren, 1860)</b> | 0          | 0       | 1            | 0        |
| <i>Homoeomeria nivea</i> Aurivillius, 1909                | 0          | 0       | 1            | 1        |
| <i>Lacipa bizonoides</i> Butler, 1893                     | 1          | 0       | 1            | 0        |
| <b><i>Lacipa exetastes</i> Collenette, 1952</b>           | 1          | 0       | 0            | 0        |
| <i>Lacipa florida</i> (Swinhoe, 1903)                     | 0          | 0       | 1            | 0        |
| <i>Lacipa gracilis</i> Hopffer, 1857                      | 1          | 0       | 1            | 0        |
| <i>Lacipa nobilis</i> (Herrich-Schäffer, 1855)            | 1          | 1       | 1            | 0        |
| <i>Lacipa ostra</i> (Swinhoe, 1903)                       | 1          | 0       | 0            | 0        |
| <b><i>Lacipa picta</i> (Boisduval, 1847)</b>              | 0          | 0       | 1            | 0        |
| <b><i>Lacipa pulverea</i> Distant, 1898</b>               | 0          | 0       | 1            | 0        |
| <i>Lacipa quadripunctata</i> Dewitz, 1881                 | 1          | 1       | 1            | 1        |
| <b><i>Lacipa sarcistis</i> Hampson, 1905</b>              | 0          | 0       | 1            | 0        |
| <b><i>Lacipa sexpunctata</i> Distant, 1897</b>            | 0          | 0       | 1            | 0        |
| <i>Laelia amauro</i> Hering, 1926                         | 0          | 0       | 1            | 0        |
| <b><i>Laelia angustipennis</i> (Walker, 1855)</b>         | 0          | 0       | 1            | 0        |
| <i>Laelia aureus</i> Janse, 1915                          | 0          | 0       | 1            | 1        |
| <i>Laelia bifascia</i> Hampson, 1905                      | 0          | 0       | 1            | 0        |
| <i>Laelia bonaberiensis</i> (Strand, 1915)                | 0          | 0       | 1            | 0        |
| <i>Laelia clarki</i> Janse, 1915                          | 0          | 0       | 1            | 1        |

| Species  | Mozambique | Lesotho | South Africa | Eswatini |
|--|------------|---------|--------------|----------|
| <i>Laelia figlina</i> Distant, 1899                  | 0          | 0       | 1            | 1        |
| <i>Laelia fracta</i> Schaus & Clements, 1893         | 1          | 0       | 1            | 1        |
| <i>Laelia gwelila</i> (Swinhoe, 1903)                | 0          | 0       | 1            | 0        |
| <i>Laelia haemata</i> Hampson, 1905                  | 0          | 0       | 1            | 0        |
| <i>Laelia janenschi</i> Hering, 1926                 | 0          | 0       | 1            | 0        |
| <i>Laelia lavia</i> Swinhoe, 1903                    | 0          | 0       | 1            | 0        |
| <b><i>Laelia melaxantha</i> (Walker, 1865)</b>       | 0          | 0       | 1            | 0        |
| <i>Laelia municipalis</i> Distant, 1897              | 1          | 0       | 1            | 0        |
| <b><i>Laelia nigripulverea</i> Janse, 1915</b>       | 0          | 0       | 1            | 0        |
| <b><i>Laelia punctulata</i> (Butler, 1875)</b>       | 0          | 0       | 1            | 0        |
| <i>Laelia robusta</i> Janse, 1915                    | 0          | 0       | 1            | 1        |
| <i>Laelia rosea</i> Schaus & Clements, 1893          | 0          | 0       | 1            | 0        |
| <i>Laelia subrosea</i> (Walker, 1855)                | 1          | 0       | 1            | 1        |
| <b><i>Laelia subviridis</i> Janse, 1915</b>          | 0          | 0       | 1            | 0        |
| <b><i>Laelia swinnyi</i> Janse, 1915</b>             | 0          | 0       | 1            | 0        |
| <i>Laelioproctis leucosphena</i> Collenette, 1939    | 0          | 0       | 1            | 0        |
| <i>Lepidopalpus hyalina</i> Janse, 1915              | 0          | 0       | 1            | 1        |
| <i>Leptaroa</i> sp.                                  | 1          | 0       | 0            | 0        |
| <i>Leptaroa paupera</i> Hering, 1926                 | 0          | 0       | 1            | 0        |
| <i>Leucoma ogovensis</i> (Holland, 1893) *           | 0          | 0       | 1            | 0        |
| <i>Leucoma parva</i> Plötz, 1880 *                   | 1          | 0       | 1            | 0        |
| <i>Leucoma sevastopuloi</i> Collenette, 1955 *       | 1          | 0       | 0            | 0        |
| <b><i>Lymantria kettlewelli</i> Collenette, 1953</b> | 0          | 0       | 1            | 0        |
| <b><i>Lymantria lutea</i> (Boisduval, 1847)</b>      | 0          | 0       | 1            | 0        |
| <b><i>Lymantria subfusca</i> (Boisduval, 1847)</b>   | 0          | 0       | 1            | 0        |
| <i>Marblepsis flabellaria</i> (Fabricius, 1787)      | 1          | 0       | 1            | 1        |
| <i>Marblepsis melanocraspis</i> (Hampson, 1905)      | 1          | 0       | 1            | 0        |
| <b><i>Micraroa minima</i> Janse, 1915</b>            | 0          | 0       | 1            | 0        |
| <i>Micraroa rufescens</i> Hampson, 1905              | 0          | 0       | 1            | 0        |
| <i>Morasa modesta</i> Walker, 1855                   | 1          | 0       | 1            | 1        |
| <i>Naroma varipes</i> (Walker, 1865)                 | 1          | 0       | 1            | 1        |
| <i>Ogoa simplex</i> Walker, 1856 *                   | 1          | 0       | 1            | 1        |
| <i>Olapa fulviceps</i> Hampson, 1910                 | 1          | 0       | 1            | 0        |
| <i>Olapa nigribasis</i> Janse, 1917                  | 0          | 0       | 1            | 1        |
| <i>Olapa nigricosta</i> Hampson, 1905                | 0          | 0       | 1            | 0        |
| <i>Olapa tavetensis</i> (Holland, 1892)              | 1          | 0       | 1            | 0        |
| <i>Olene basalis</i> (Walker, 1855)                  | 1          | 0       | 1            | 0        |
| <i>Palasea albimacula</i> Wallengren, 1863           | 1          | 0       | 1            | 1        |
| <i>Paraproctis chionozepe</i> Collenette, 1954 *     | 1          | 0       | 1            | 0        |
| <i>Pirga pellucida</i> Wichgraf, 1922                | 1          | 0       | 1            | 0        |
| <b><i>Pirga transvalensis</i> Janse, 1915</b>        | 0          | 0       | 1            | 0        |
| <i>Pirgula atrinotata</i> (Butler, 1897)             | 1          | 0       | 1            | 0        |
| <i>Polymona ruffemur</i> Walker, 1855 *              | 1          | 1       | 1            | 1        |
| <i>Porthesaroa maculata</i> Collenette, 1938         | 1          | 0       | 1            | 0        |
| <i>Psalis securis</i> Hübner, 1823                   | 1          | 0       | 1            | 1        |
| <i>Pseudobazisa perculta</i> (Distant, 1897)         | 0          | 0       | 1            | 1        |
| <i>Pteredoa monosticta</i> (Butler, 1898)            | 1          | 0       | 1            | 1        |
| <i>Pteredoa subapicalis</i> Hering, 1926             | 1          | 0       | 1            | 0        |
| <i>Pteredoa usebia</i> (Swinhoe, 1903)               | 0          | 0       | 1            | 0        |
| <i>Rhyopteryx hemichrysa</i> Collenette, 1960        | 0          | 0       | 1            | 0        |
| <i>Rhyopteryx lugardi</i> (Swinhoe, 1903)            | 0          | 0       | 1            | 0        |
| <i>Rhyopteryx rhodalipha</i> (Felder R., 1874)       | 1          | 0       | 1            | 1        |
| <i>Rhyopteryx rhodea</i> (Hampson, 1905)             | 1          | 0       | 1            | 0        |

| Species   | Mozambique | Lesotho | South Africa | Eswatini |
|---|------------|---------|--------------|----------|
| <i>Rhyopteryx rubripunctata</i> (Weymer, 1892)  | 0          | 0       | 1            | 0        |
| <i>Rhyopteryx tacita</i> (Hering, 1927)         | 0          | 0       | 1            | 1        |
| <i>Rhyopteryx triangulifera</i> (Hampson, 1910) | 0          | 0       | 1            | 0        |
| <i>Ruanda eleuteriopsis</i> Hering, 1926        | 1          | 0       | 0            | 0        |
| <i>Ruanda furva</i> (Hampson, 1905)             | 1          | 0       | 1            | 0        |
| <i>Ruanda nuda</i> (Holland, 1897)              | 0          | 0       | 1            | 0        |
| <i>Stracena bananae</i> (Butler, 1897)          | 1          | 0       | 1            | 0        |
| <i>Tamsita habrotima</i> (Tams, 1930) *         | 1          | 0       | 0            | 0        |

ural History Research Trust (Leominster, United Kingdom), Natural History Museum (London, United Kingdom), Museum Witt (Munich, Germany), Bavarian State Collection of Zoology (Munich, Germany), and Naturalis Biodiversity Centre (Leiden, Netherlands). In total 2352 specimens from South Africa, 2079 specimens from Mozambique, 18 specimens from Lesotho and 61 specimens from Eswatini were revised. The specimens were photographed using a Nikon D90 camera with Nikkor AF Micro 60 mm lens and assembled for our plates using the Corel DRAW X7 computer program.

We compared the faunal diversity of the regions using number of species, and similarity or uniqueness of the species composition expressed as a nestedness of the species and species turnover expressed as binary presence/absence data. We calculated the data in R v. 4.0.3. (R Core Team 2018), by a command “nestedbetajac” (package vegan – Oksanen *et al.* 2019). Species endemism was evaluated based on the data of species occurrence available from online database (De Prins and De Prins 2021) and from literature (e.g., Swinhoe 1903).

## Results

We reported altogether 168 species of tussock moths belonging to 43 genera currently occurring in the Maputaland-Pondoland-Albany biodiversity hotspot. We are listing 158 species from South Africa, 62 species from Mozambique, 48 species from Eswatini and 10 species from Lesotho. The faunal comparison analysis also revealed 44 potentially endemic species from South Africa and six potentially endemic species from Mozambique (Table 1). By the term “potential endemic species” we mean a species reported so far from only one country, but we cannot rule out its distribution in surrounding countries.

The following genera are recorded for the first time from Mozambique: *Euproctoides* Bethune-Baker, 1911; *Heteronygmia* Holland, 1893; *Leucoma* Hübner, 1822; *Ogoa* Walker, 1856; *Paraproctis* Bethune-Baker, 1911; *Polymona* Walker, 1855; *Tamsita* Kiriakoff, 1954 (Fig. 2).

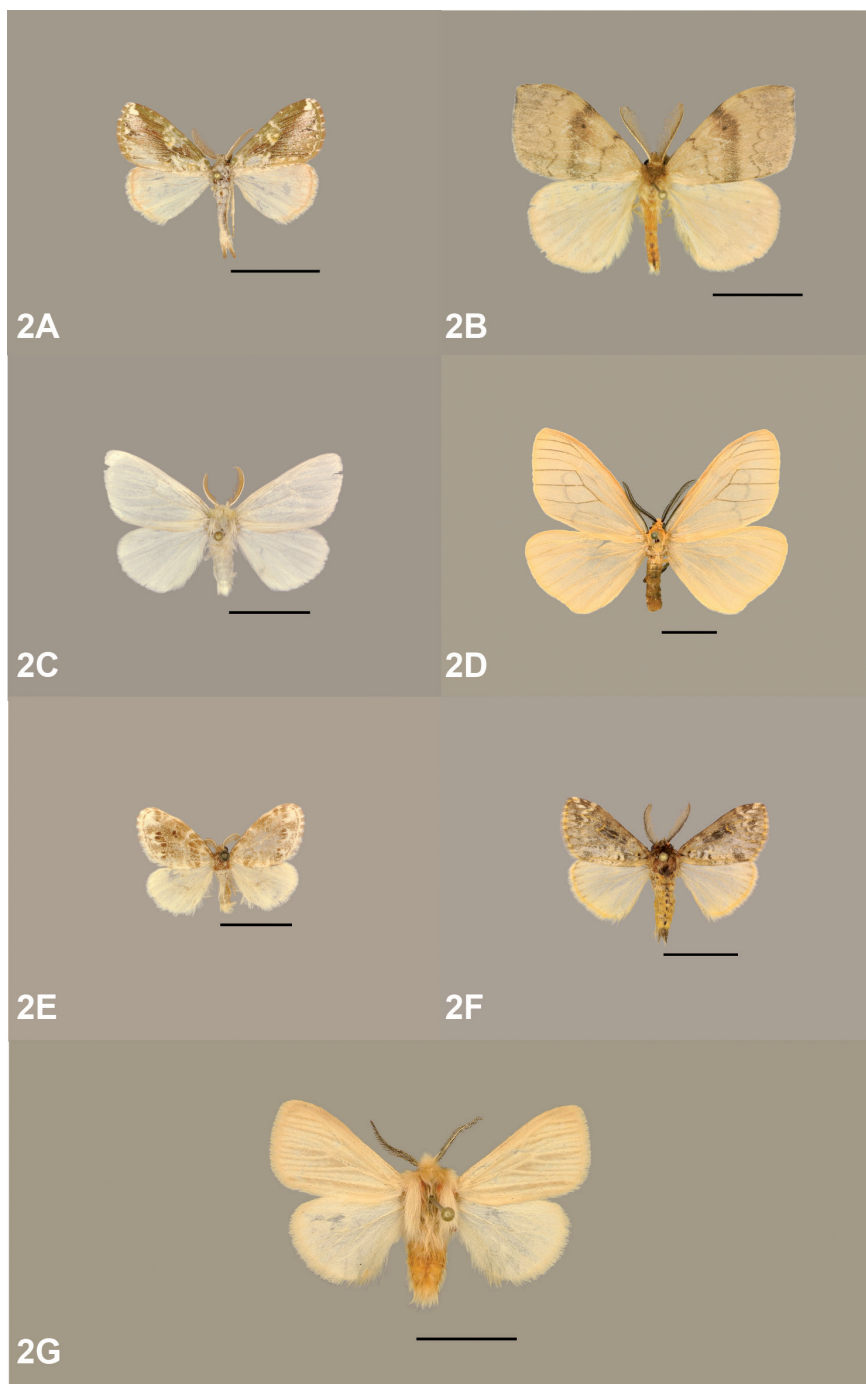
### *Euproctoides* Bethune-Baker, 1911

#### *Euproctoides ertli* (Wichgraf, 1922)

Fig. 2A

**Material.** MOZAMBIQUE • 1♂; Zambezia Province, Mt. Namuli, SW slopes near Mucunha village (forest remnant); 15°22'33"S, 37°05'05.7"E; 15–16 Aug. 2018; 1245 m; MV Light Trap; G. László, W. Miles, A. Vetina leg.; ANHRT: 2018.30, ANHRTUK 00094093.





**Figure 2.** Seven representatives of newly recorded genera of tussock moths in Mozambique. **A.** *Euproctoides ertli* (Wichgraf, 1922); **B.** *Heteronygmia dissimilis* (Aurivillius, 1910); **C.** *Leucoma ogovensis* (Holland, 1893); **D.** *Ogoa simplex* Walker, 1856; **E.** *Paraproctis chionopeza* Collenette, 1954; **F.** *Polymona rufifemur* Walker, 1855; **G.** *Tamsita habrotima* (Tams, 1930). Scale bars: 1 cm.

**Distribution.** Angola (Wichgraf 1922), South Africa (Vári *et al.* 2002).

***Heteronygmia* Holland, 1893**

***Heteronygmia dissimilis* (Aurivillius, 1910)**

Fig. 2B

**Material.** MOZAMBIQUE • 1♂; Manica Province, Chimanimani National Reserve, Moribane Forest, Ndzou Camp (Moist Forest); 19°44'01.4"S, 33°20'15.1"E; 3–5 Aug. 2018; 630 m; Actinic Light Trap; G. László, W. Miles, A. Vetina leg.; ANHRT: 2018.30, ANHRTUK 00046725.

**Distribution.** Kenya (Hering 1926), Malawi (Murphy and Chipeta 2018), Tanzania (Aurivillius 1910), Zimbabwe (Hering 1929).

***Leucoma* Hübner, 1822**

***Leucoma ogovensis* (Holland, 1893)**

Fig. 2C

**Material.** MOZAMBIQUE • 1♂; Maputo Special Reserve, West Gate (Sand Forest); 26°30'14.2"S, 32°42'59.6"E; 21–22 Feb. 2018; 22 m; MV Light Trap; G. László, J. Mulvaney, L. Smith leg.; ANHRT: 2018.2, ANHRT 00027695.

**Distribution.** Cameroon (Aurivillius 1904), Congo (Holland 1920), Gabon (Holland 1893), Ghana (Swinhoe 1903), Kenya (Le Cerf 1922), Nigeria (Swinhoe 1903), South Africa (Hering 1926).

***Ogoa* Walker, 1856**

***Ogoa simplex* Walker, 1856**

Fig. 2D

**Material.** MOZAMBIQUE • 1♂; Maputo Special Reserve, West Gate (Sand Forest); 26°30'14.2"S, 32°42'59.6"E, 13–15 Feb. 2018; 22 m; MV Light Trap; G. László, J. Mulvaney, L. Smith leg.; ANHRT: 2018.2, ANHRTUK 00038737.

**Distribution.** Kenya (Butler 1898), South Africa (Walker, 1856), Tanzania (Grünberg 1907).

***Paraproctis* Bethune-Baker, 1911**

***Paraproctis chionopeza* Collette, 1954**

Fig. 2E

**Material.** MOZAMBIQUE • 1♂; Maputo Special Reserve, Mangrove Camp (Mangrove-Woodland Mosaic); 26°19'35.9"S, 32°42'35.7"E; 7–9 Dec. 2016; 9 m; MV Light Trap; M. Aristophanous, J. Cristóvão, G. László, W. Miles leg.; ANHRT: 2017.22, ANHRTUK 00053987.

**Distribution.** DR Congo (Afromoths, N. Voaden), South Africa (Vári *et al.* 2002), Zambia (Collette 1954), Zimbabwe (Afromoths, R. Butler).



***Polymona* Walker, 1855*****Polymona rufifemur* Walker, 1855**

Fig. 2F

**Material.** MOZAMBIQUE • 1♂; Maputo Special Reserve, Ponta Milibangalala (Dune Grassland – Dune-Forest Ecotone); 26°26'58.6"S, 32°55'29.8"E; 30 Nov.-3 Dec. 2016; 15 m; Light Trap; M. Aristophanous, J. Cristóvão, G. László, W. Miles leg.; ANHRT: 2017.22, ANHRTUK 00055859.

**Distribution.** Botswana (Pinhey 1975), Djibouti (Dall'Asta 1977), Eritrea (Berio 1939), Ethiopia (Collenette 1938), Gambia (Druce 1887), Kenya (Le Cerf 1922), Lesotho (Kopij 2006), Malawi (Swinhoe 1903), Namibia (Dall'Asta 2004), South Africa (Janse 1915), Tanzania (Pinhey 1975), Zambia (Pinhey 1975), Zimbabwe (Swinhoe 1903).

***Tamsita* Kiriakoff, 1954*****Tamsita habrotima* (Tams, 1930)**

Fig. 2G

**Material.** MOZAMBIQUE • 1♂; Zambezia Province, Mt. Namuli, SW slopes near Mucunha village (forest remnant); 15°22'33"S, 37°05'05.7"E; 15–16 Aug. 2018; 1245 m; MV Light Trap; G. László, W. Miles, A. Vetina leg.; ANHRT: 2018.30, ANHRTUK 00050678.

**Distribution.** Kenya (Tams 1930), Uganda (Tams 1930).

**Lymantriinae diversity patterns**

The highest species richness of tussock moths was recognised in the Republic of South Africa (N = 157) representing 93% of all recorded species (Table 2). The highest turnover among studied countries was found in Lesotho (37.8%).

The neighbouring countries Mozambique, Lesotho and Eswatini yielded approximately the same values (60%) of species exchange between each other (Table 3). Surprisingly, neither Leso-

**Table 2.** Evaluation of tussock moth richness patterns in the wider Maputaland-Pondoland-Albany.

|                     | species | nestedness | turnover |
|---------------------|---------|------------|----------|
| <b>Mozambique</b>   | 62      | 0.315      | 0.295    |
| <b>Lesotho</b>      | 10      | 0.265      | 0.379    |
| <b>South Africa</b> | 157     | 0.532      | 0.103    |
| <b>Eswatini</b>     | 12      | 0.380      | 0.238    |

**Table 3.** Faunal exchange (turnover) among studied countries of the Maputaland-Pondoland-Albany.

|                     | Mozambique | Lesotho | South Africa | Eswatini |
|---------------------|------------|---------|--------------|----------|
| <b>Mozambique</b>   | 0.000      | 0.667   | 0.229        | 0.648    |
| <b>Lesotho</b>      | 0.667      | 0.000   | 0.182        | 0.667    |
| <b>South Africa</b> | 0.229      | 0.182   | 0.000        | 0.000    |
| <b>Eswatini</b>     | 0.648      | 0.667   | 0.000        | 0.000    |

**Table 4.** Faunal nestedness among the countries of the Maputaland-Pondoland-Albany.

|              | Mozambique | Lesotho | South Africa | Eswatini |
|--------------|------------|---------|--------------|----------|
| Mozambique   | 0.000      | 0.259   | 0.446        | 0.058    |
| Lesotho      | 0.259      | 0.000   | 0.762        | 0.239    |
| South Africa | 0.446      | 0.762   | 0.000        | 0.696    |
| Eswatini     | 0.058      | 0.239   | 0.696        | 0.000    |

tho nor Eswatini show any faunal exchange with South Africa. All three countries also displayed high species richness differences towards South Africa (Table 4). Due to its geography, the fauna of Lesotho is, not surprisingly, deeply nested in the fauna of South Africa. However, South Africa showed the lowest value in nestedness towards Mozambique (44.6%).

## Discussion

Our study has reviewed the tussock moth fauna hitherto recorded from the wider Maputaland-Pondoland-Albany and analysed the faunal diversity between those four southern African countries abutting the area of the hotspot. We also have presented a first summary of the tussock moths occurring in Mozambique along with seven genera newly recorded in the country. All these were expected to occur in Mozambique because of their presence in neighbouring countries and these records connect their disjunctive distribution ranges with South Africa. The only exception is the genus *Tamsita* Kiriakoff, 1954, the presence of which in Mozambique represents the southernmost occurrence of the genus.

The diversity of the tussock moths in Maputaland-Pondoland-Albany displayed several peculiar distributional trends. In spite of the large amount of faunistic studies published about the South African fauna and flora, only very few of them discussed their distributional or biogeographic patterns with regards to the studied countries. We found that the fauna of Lesotho is deeply nested in the South African fauna, but otherwise it has a high turnover among all other countries. This result outlined the potential origin scenario. Lesotho is a country with the high average altitude and the occurrence of specific alti-montane ecoregions (e.g., Drakensberg grasslands and woodlands) with difficult accessibility and hard conditions. Therefore, it was colonized by tussock moth species from neighbouring countries (Sayre *et al.* 2013). On the other hand, the South African fauna can be considered very similar to the fauna of Mozambique based on the values of nestedness, and simultaneously does not exhibit any faunal exchange towards Eswatini and Lesotho. Thus it forms a gradient of almost the same fauna along the Indian Ocean coast. Our results also highlighted that approximately 27.8% of the South African fauna and 9.6% of the species of Mozambique are potentially endemic to the Maputaland-Pondoland-Albany.

In conclusion, we reviewed and extended the current knowledge of the diversity and distribution patterns of tussock moths in southern Africa. What is more, we also provided a pilot snapshot of species endemism of Lymantriinae in this region, which contributes to future conservation efforts of this remarkable biodiversity hotspot.

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