



## Article

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*Varesi, Angelica, Pierella, Elisa, Romeo, Marcello, Piccini, Gaia Bavestrello, Alfano, Claudia, Bjørklund, Geir, Oppong, Abigail, Ricevuti, Giovanni, Esposito, Ciro et al (2022) The Potential Role of Gut Microbiota in Alzheimer's Disease: from Diagnosis to Treatment. Nutrients, 14 (3). e668.*

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<http://dx.doi.org/10.3390/nu14030668>

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Review

# The Potential Role of Gut Microbiota in Alzheimer's Disease: from Diagnosis to Treatment

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**Citation:** Varesi, A.; Pierella, E.; Romeo, M.; Piccini, G.B.; Alfano, C.; Bjørklund, G.; Oppong, A.; Ricevuti, G.; Esposito, C.; Chirumbolo, S.; Pascale, A. The Potential Role of Gut Microbiota in Alzheimer's Disease: from Diagnosis to Treatment. *Nutrients* **2022**, *14*, 668. <https://doi.org/10.3390/nu14030668>

Academic Editors: Luisa Cigliano, Maria Stefania Spagnuolo and Arianna Mazzoli

Received: 21 January 2022

Accepted: 3 February 2022

Published: 5 February 2022

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**Abstract:** Gut microbiota is emerging as a key regulator of many disease conditions and its dysregulation is implicated in the pathogenesis of several gastrointestinal and extraintestinal disorders. More recently, gut microbiome alterations have been linked to neurodegeneration through the increasingly defined gut microbiota brain axis, opening the possibility for new microbiota-based therapeutic options. Although several studies have been conducted to unravel the possible relationship between Alzheimer's Disease (AD) pathogenesis and progression, the diagnostic and therapeutic potential of approaches aiming at restoring gut microbiota eubiosis remain to be fully addressed. In this narrative review, we briefly summarize the role of gut microbiota homeostasis in brain health and disease, and we present evidence for its dysregulation in AD patients. Based on these observations, we then discuss how dysbiosis might be exploited as a new diagnostic tool in early and advanced disease stages, and we examine the potential of prebiotics, probiotics, fecal microbiota transplantation, and diets as complementary therapeutic interventions on disease pathogenesis and progression, thus offering new insights into the diagnosis and treatment of this devastating and progressive disease.

**Keywords:** Alzheimer's disease; gut microbiota; dysbiosis; gut-brain axis; biomarker; prebiotics; probiotics; diet; fecal microbiota transplantation

## 1. Introduction

Alzheimer's disease (AD), which affects approximately 50,000,000 people worldwide, is the most frequent cause of dementia, constituting a real global health problem [1]. The disease is characterized by the progressive deposition of beta amyloid (A $\beta$ )

plaques and tangles of hyperphosphorylated tau neurofibrils, leading to neuroinflammation and progressive cognitive decline [2]. Synaptic dysfunction and neuronal death are at least in part due to the excessive or non-resolving activation of the immune response and any infections or traumatic events affecting the brain (traumatic brain injury) can interfere with central immune homeostasis and accelerate the progression of the disease [3]. Although several hypotheses have been formulated about the causes of AD pathogenesis and progression, both the onset and the evolution of the disease remain not entirely clear. Therefore, although different therapeutic options have been proposed, many have failed in clinical trials and have not been found to produce significant benefits [4–6]. It is widely thought that an early diagnosis could be essential to act at the earliest disease stages, but effective and reproducible biomarkers are still far from clinical application [7,8].

In recent years, the gut microbiota brain axis (GMBA) has been at the center of biomedical research and it has been suggested as a potential therapeutic target for disorders affecting the central nervous system, including AD [9–11].

The term “gut microbiota” refers to the commensal microbial community that colonizes the gastrointestinal tract and is constituted by bacteria, fungi, archaea, viruses, and protozoans living in symbiotic relationship with our intestine [9,12–14]. Thanks to their active role in regulating host’s homeostasis and disease, they are becoming more and more important in the pathogenetic mechanisms of neurodegenerative disorders, such as AD [15–18]. Indeed, even though for a long time it was believed that the brain was a totally isolated organ, recent evidence shows that the gut microbiota is at the center of a bidirectional communication between intestine and brain, the so-called microbiota gut–brain axis [15,19–21]. This interplay involves the central nervous system (CNS), the autonomic nervous system, the enteric nervous system (ENS), and the hypothalamus-pituitary-adrenal axis (HPA), and it has been reported to be implicated in a number of physiological and pathological processes such as satiety, food intake, glucose and fat metabolism, insulin sensitivity, and stress [22]. Although the mechanisms underlying this interaction are not fully understood, targeting the microbiota might represent a new diagnostic and therapeutic strategy in AD and in other neurodegenerative diseases [23]. However, despite several published papers having reviewed possible microbiome-based therapies, to our knowledge a comprehensive view of gut microbiota-based diagnostic and therapeutic approaches is still lacking. Here, based on the main studies addressing gut microbiota dysregulation in AD, we discuss how the microbiota-derived biomarkers might be exploited for early disease detection, and we review the potentiality of probiotics, prebiotics, diet, and fecal microbiota transplantation as complementary therapeutic options for this devastating and progressive disease.

## 2. Main

### 2.1. The Gut–Brain Axis: An Overview

The gut–brain axis (GBA) consists of a signaling pathway between the gastrointestinal (GI) tract and the CNS, which allows a bidirectional communication between the two systems. Its primary role is to monitor and integrate intestinal functions as well as to link, through immune and neuro-endocrine mediators, the emotional and cognitive centers of the brain with peripheral intestinal mechanisms such as immune activation, intestinal permeability, enteric reflex, and entero-endocrine signaling [20]. In this communicating network, the brain affects gut movement, sensory, and secretion functions, and in turn the signals from the gut affect brain function [24]. This relationship is therefore of outmost importance for the maintenance of gut homeostasis, and it has been reported to be also involved in the etiology of several metabolic and mental (psychiatric and neurological) dysfunctions and disorders [21,25].

Different routes of communication between the gut microbiota and the brain have been suggested:

- Through incoming and outgoing branches of the vagus nerve [26], which represents the major modulatory pathway [27].
- Through the generation of metabolites and bioactive peptides (such as short-chain fatty acids) as well as the modulation of transmitters (e.g., serotonin and acetylcholine) by the microbiota [26–28].
- Through the secretion of cortisol by the HPA in case of stress, which can affect intestinal motility, integrity, and mucus production, leading to changes in gut microbiota composition. This alteration, in turn, may affect the CNS through the modulation of stress hormones [28].
- Through pro-inflammatory cytokines and chemokines [29].
- Immunity is also critically involved. Specifically, toll-like receptors (TLRs) and peptidoglycans (PGNs) mediate the immune response towards microbes by acting as sensors of microbial components [30,31]. A local immune activation can, throughout different pathways, lead to an immune activation in different organs, including the brain [32]. This low-grade immune activation has been implicated in the pathophysiology of some forms of depression and neurodegenerative disorders such as AD and Parkinson's disease (PD) [26].

Given this complex interplay, it is not surprising that the gut–brain axis, and therefore the gut microbiota as main component of this crosstalk, directly or indirectly affects neuropsychiatric illnesses [33].

## 2.2. The GMBA in Alzheimer's Disease: What's New?

The role of gut microbiota and GMBA in AD is of utmost importance [34]. The composition of the gut bacteria affects dramatically any age-related neurological disorder, such as AD, and mood disorders. Extrinsic factors including diet, lifestyle, or also pro-inflammatory insults, along with intrinsic components including genetic polymorphism, immunity, metabolites, and hormones, profoundly affect the composition of the gut microflora, which in turn produces signaling molecules such as short chain fatty acids (SCFAs), tryptophan, choline, and hormones (such as ghrelin, leptin) in the GI tract able to regulate CNS functions [35]. Aging has a strong impact on gut microbiota composition favoring the development of pro-inflammatory bacteria (such as *Bacillus fragilis*, *Faecalibacterium prausnitzii*, *Eubacterium rectale*, *Eubacterium hallii*, and *Bacteroides fragilis*) to the detriment of anti-inflammatory bacteria, a condition that induces local systemic inflammation then leading to enhanced permeability of the gastrointestinal tract, an impairment in the blood–brain barrier (BBB), finally promoting neuroinflammation. Indeed, Cattaneo et al. observed such pro-inflammatory bacteria in amyloid-positive patients when compared to healthy subjects [36]. In transgenic mice (mutant human APP) when infected with *Salmonella enterica* and in transgenic *Caenorhabditis elegans* (human A $\beta$ 42 peptide) infected with *Candida albicans*, the authors reported a susceptibility to further infections although they died later with respect to wild type animals. Probably, this was due to the antimicrobial activity of A $\beta$  peptide, as the heparin-binding motif of A $\beta$  oligomers make easier the binding to the glycosyl group of the carbohydrate moiety in the bacterial cell wall, so preventing its adhesion to the host cell and the induction of microbial agglutination [37]. Also, bacteria-derived amyloids have been reported to be causative factors for A $\beta$  peptide aggregation in AD. For example, amyloids produced by bacteria such as curli (*E. coli*), TasA (*Bacillus subtilis*), CsgA (*S. Typhimurium*), FapC (*Pseudomonas fluorescens*), phenol soluble modulins (*Staphylococcus aureus*), etc., have been shown to contribute to the development of AD pathology particularly by promoting A $\beta$  oligomers and fibrils formation [38].

Besides bacteria-derived amyloids, further components contribute to the onset and pathogenesis of AD. For instance, lipopolysaccharides (LPS) from bacteria inoculated in experimental animals (in the fourth ventricle of the brain) generated a symptomatology very akin to AD [39]. Even the injection of LPS in mice induces elevation of A $\beta$  in the

hippocampal area causing cognition defects, thus supporting the role of LPS in amyloid fibrillogenesis [40,41]. When in circulation, LPS has been found to activate the TLR4 pathway, thus triggering immune cells to secrete pro-inflammatory cytokines and IgM/IgA, exacerbating systemic inflammation [42–45]. In this perspective, gut inflammation may be a cause of AD pathogenesis.

The relationship among gut microbiome composition, inflammation, further neuroinflammation, and AD onset, is a fundamental matter of debate in AD etiopathogenesis. A certain number of investigations reported the presence of pathogens in the post mortem brains of AD patients [46–49]. Among them are herpes simplex virus type 1 and bacteria such as *Chlamydomyxa pneumoniae*, *Borrelia burgdorferi*, or other spirochetes [50–52]. Furthermore, a significant increase in the level of *Helicobacter pylori*-specific IgG antibodies, found in the cerebrospinal fluid and in the serum of AD patients, was reported [53]. In this context, novel therapeutic approaches can be envisaged by investigating the crucial role of some gut microbiota compositions leading to AD with the aim of promoting the prevalence of health-associated species, also adjusting both dietary habits and lifestyle, that could help to prevent disease development/progression [54–56].

### 2.3. Gut Microbiota Alterations in AD

Intestinal dysbiosis is a condition of microbial imbalance caused by an overgrowth of “bad” bacteria inside the gut, associated with potential negative outcomes such as the incorrect production of essential metabolites or even the genesis of harmful metabolites [14,57]. Although the composition of a “healthy microbiota” has not yet been defined, a balanced environment between the host and microorganisms is known to be essential to carry on the necessary immunological and metabolic functions [58]. Over the past years, dysbiosis has been reported to be implicated in the development of several disorders, such as obesity, diabetes, chronic fatigue syndrome, intestinal bowel syndrome, cancer, autoimmune diseases, depression, anxiety, PD, multiple sclerosis, amyotrophic lateral sclerosis, and other neuropsychiatric disorders [59–69]. Recently, many studies have shown that gut microbiota alterations directly influence cognitive decline, actively participating in AD pathogenesis and progression [36,70–73]. Generally, AD patients are often characterized by a decreased gut microbial diversity, with a significant shift in favor of pro-inflammatory taxa at the expense of the more beneficial anti-inflammatory ones, similar to what has been observed in both mouse and human aging [25,36,70–75]. For example, when fecal microbiota 16S rRNA sequencing was performed on 97 individuals [33 AD, 32 MCI (mild cognitive impaired) and 32 controls], a significant decrease in *Firmicutes* was accompanied with a higher *Proteobacteria*, *Gammaproteobacteria*, and *Enterobacteria* abundance in patients with neurodegeneration compared to healthy subjects. Interestingly, a pronounced difference in *Enterobacteriaceae* has been reported also between MCI and AD patients, thus indicating a progressive change in the gut microbiota composition during disease progression [72]. Similarly, Vogt et al. detected a significant dampen in *Firmicutes* and *Bifidobacteria* in the fecal samples of AD patients, and this decrease was counterbalanced by the overgrowth of *Bacteroidetes* species in the same individuals [70]. Alterations in the gut microbiota composition during neurodegeneration has also been reported by Zhuang et al. when comparing 43 AD patients with age- and sex-matched controls: enriched *Bacteroidetes* and decreased *Actinobacteria* at the phylum level were paralleled by enhanced *Ruminococcaceae*, *Enterococcaceae*, and *Lactobacillaceae*, together with less *Lachnospiraceae*, *Bacteroidaceae*, and *Veillonellaceae* at the family level [76]. However, in contrast with this evidence, lower *Bacteroides*, *Lachnospira*, and *Ruminiclostridium* and higher levels of *Prevotella* have been reported in another study [77]. Although reductive, this discrepancy might be at least in part explained by the different geographical origin of the participants, since regional identity may strongly affect gut microbiota composition, as well as other comorbidities [78]. In this respect, larger studies are certainly needed to establish standard and reproducible inclusion criteria, possibly excluding also the possible confounding effect of other comorbidities.

A growing body of evidence indicates that gut microbiota dysfunctions are involved in the early disease stages of AD pathogenesis, enhancing immuno-senescence, oxidative stress, cytokine secretion, and neuroinflammation [79]. In this respect, Cattaneo et al. report that patients with AD show an increase in pro-inflammatory endobacteria species of *Escherichia/Shigella* and a decrease in the anti-inflammatory taxon *E. rectale*, and that this microbiota alteration is associated with amyloidosis and peripheral inflammation [36]. Moreover, when stool samples were collected from 108 nursing home elders and analyzed with metagenomic sequencing, a decline in butyrate-synthesizing bacteria was paralleled by a rise in pro-inflammatory taxa in AD elders, thus possibly exacerbating local and systemic inflammation [71]. Interestingly, these data have been correlated with low levels of expression of the P-glycoprotein, an essential molecule required for intestinal homeostasis, therefore indicating a clear nexus between microbiome dysregulation and intestinal inflammation [71]. These results further support the concept that changes in gut microbiota composition also reflect in alterations in intestinal function. Indeed, differences in gut microbiota population may influence tryptophan and serotonin levels in the body and may affect the synthesis of some key molecules useful for the brain, such as dopamine, norepinephrine, and brain-derived neurotrophic factor (BDNF) [80–82]. As mentioned, another beneficial role exerted by the gut microbiota is the production of SCFAs, including butyrate, propionate, and acetate, essential for energy production, gut epithelia homeostasis, and immune regulation [83]. When their production is altered as a consequence of dysbiosis, A $\beta$  plaques deposition, metabolic dysfunctions, and microglia dysregulation is favored, thus promoting cognitive decline [84–86]. Moreover, a decrease in butyrate-producing bacteria, as reported in AD, has been linked to T cell imbalance, epithelial barrier leakage (so called “leaky gut”), and increased bacterial translocation [80,87,88]. Consequently, circulating Gram negative endobacteria-derived LPS, also known as metabolic endotoxemia, triggers systemic inflammation via TLR4 and promotes BBB disruption, thus fostering neuroinflammation. [89,90]. Intestinal dysbiosis can also contribute to the increase of harmful substances such as amyloid and trimethylamine N-oxide (TMAO). TMAO is a microbial metabolite that has been recently implicated in increased formation of beta amyloid, peripheral immune response activation, enhanced oxidative stress, platelet hyperactivity, intestinal mucosal barrier dysfunction, and BBB permeability, thus promoting the consequent passage of bile acids produced by bacteria and cholesterol in the brain [91–95]. Finally, the ability of some endobacteria to produce gaseotransmitter molecules, such as nitric oxide (NO), hydrogen (H<sub>2</sub>), ammonia (NH<sub>3</sub>), methane (CH<sub>4</sub>), and hydrogen sulfide (H<sub>2</sub>S) seems to be fundamental for the proper neuronal function, and its alteration participates to AD pathogenesis [96,97]. Overall, these data indicate that the dialogue between gut microbiota and brain is much more complicated than previously thought, and only its entire understanding can provide insights into new diagnostic and therapeutic interventions.

#### 2.4. Gut Microbiota-Based AD Biomarkers

One of the major concerns in AD research is to find predictive, sensitive, non-invasive accurate, and accessible biomarkers for early disease diagnosis [98,99]. Although many studies focus on fluid biomarkers for early disease detection, we are still far from having found an effective and consistent assay to be used in the clinical practice [100]. As mentioned above, the gut microbiota has emerged as a key player in regulating both physiological and non-physiological conditions, thus gut microbiota-related biomarkers may represent a promising alternative/complementary tool to assess disease conditions [101]. Indeed, although initially hypothesized for gastrointestinal disorders [102], gut microbiome-derived biomarkers have also been considered for psychological and neurodegenerative diseases (i.e., bipolar disorder, multiple Sclerosis, and PD), reporting powerful predictivity and differential diagnosis ability [103–105]. Regarding AD, promising results have recently been obtained, and Table 1 summarizes the main findings [72,73,106–115] (Table 1). Whilst species of *Prevotella* and *Helicobacter* have been shown to be significantly

different between APP/PS1 transgenic mice and controls, *Actinobacteria* and TM7 phylum seem to be more accurate in diagnosing AD when using the triple transgenic mouse model [106,107,109]. Changes in beta diversity and variations in circulating metabolites involved in inflammatory pathways and metabolism of nucleotides, lipids, and sugars (i.e., glutamate, hypoxanthine, thymine, hexanoyl-CoA, and leukotrienes) have also been considered in the same studies, showing promising results [106,107]. Remarkably, when the gut microbiota of APP/PS1 mice at different ages was compared to matched controls, huge shifts in the abundance of the families *Proteobacteriaceae*, *Verrucomicrobiaceae*, *Bifidobacteriaceae*, *Erysipelotrichaceae*, *Prevotellaceae*, *Bacteroidaceae*, and *Rikenellaceae* could be detected far before any plaque deposition in the brain, suggesting a great potentiality for early diagnosis [110]. Although nowadays, mice clearly represent the most used animal model, some evidence obtained with *Drosophila melanogaster* indicate *Wolbachia* as a potential AD biomarker, while *Stenotrophomonas* appears to exert a beneficial role in preventing neurodegeneration [111].

In humans, when a cohort of individuals with AD and/or MCI were compared to healthy controls, significant differences in microbial diversity and in the fecal and blood abundance of 11 genera were observed [112,113]. Importantly, Li et al. report no major variation in the analyzed gut microbiota biomarkers between MCI and AD groups, suggesting a better ability in early detection rather than in clinical progression monitoring [113]. Similarly, cerebrospinal fluid levels of the gut microbiome-dependent metabolite TMAO were not different in AD compared to MCI patients, although significantly higher than controls [114]. On the contrary, other studies report the capability of some biomarkers (i.e., *Enterobacteriaceae*, SCFAs, and indole-3-pyruvic acid abundance) to clearly differentially diagnose between a mild symptomatic disease (MCI) and a more advanced stage, thus leaving the debate open [72,115]. Other suggestions might come from the evidence of a progressive shift from *Faecalibacterium* to *Bifidobacterium* genera in AD, thus offering the possibility to follow the ratio of butyrate/lactate producing genera as a disease marker of neurodegeneration [73]. Although the data are still limited, it would be interesting to see the results of the currently undergoing Emory Healthy Aging and Emory Healthy Brain Studies, aimed at following 50–75 years old individuals (without AD or any other cognitive impairment), to identify early disease biomarkers, comprised the gut microbiome ones [116]. Biosignatures from the gut microbiota might also be exploited for patients' stratification and therapy in clinical trials aimed at applying precision medicine in AD treatment, but the information remains for now limited [117]. Finally, although our review focuses on the gut microbiota, it is important to mention that oral microbiota has been implicated in AD pathogenesis and might also represent a source of novel salivary biomarkers in AD, possibly in a combinatorial approach with the gut microbiota ones [118–122].

Although these preliminary data may appear promising, several limitations still exist and must be accounted. First, when looking for a new biomarker, large cohorts should always be preferred over smaller ones, and the evidence obtained should be confirmed on a validating group [100]. Secondly, since the gut microbiome composition changes widely according to nationality, lifestyle, and dietary habits, it is not often easy to distinguish between real evidence and confounding factors, thus questioning the relevance of the results [123]. Additionally, the importance of age- and gender-matched control groups should not be underestimated, and the respective cohorts should be designed accordingly [100]. To partially solve these limitations, a combination of different biomarkers could be adopted. For example, Zhang et al. report how gut microbiota composition, serum miRNAs and dietary quality scores can be used together to improve reproducibility and consistency [112]. In this respect, it would be interesting to investigate whether SCFAs, in combination with other fluid biomarkers, might prove effective in disease diagnosis and clinical monitoring, as some evidence already suggested [108].

Overall, although there are still some limitations, these data indicate that gut microbiota-based biomarkers might represent an alternative and/or an integration to the existing ones and should encourage scientists to plan larger investigations in humans.

**Table 1.** Gut microbiota-based biomarkers for AD.

Ref	Journal	Study Cohort and Design	Analysis Performed	Results	Biomarker/s Proposed
Yan et al., 2021 [106]	Front. Aging Neurosci.	APP/PS1 transgenic mice (8 months old, $n = 7$ ) receiving fasudil (ADF group) or saline (ADNS group) were compared to age- and gender- matched WT mice	Fecal metagenomic and metabolites	<p>↑ <i>Firmicutes/Bacteroidetes</i> in ADNS compared to WT</p> <p>↓ <i>Firmicutes/Bacteroidetes</i> in ADF compared to WT</p> <p>↑ Metabolites involved in metabolism of nucleotides, lipids, sugars and inflammation</p>	<ul style="list-style-type: none"> <li>• <i>s_Prevotella_sp_CA G873</i> as ADF biomarker</li> <li>• <i>s_Helicobacter_typhlonius</i> and <i>s_Helicobacter_sp_MIT_03-1616</i> as ADNS biomarkers</li> <li>• Glutamate, hypoxanthine, thymine, hexanoyl-CoA, and leukotrienes in ADF or ADNS</li> </ul>
Bello-Medina et al., 2021 [107]	Front. Neurosci.	Mice 3xTg-AD 3 and 5 month-old ( $n = 10$ females and $n = 10$ males) compared to matched controls	Fecal sample collection, $\alpha$ and $\beta$ diversity, LDA and LEfSe	<p>↓ <i>Actinobacteria</i> and TM7 in 3xTg-AD compared to controls at 3 month-old</p> <p>≠ <math>\beta</math> diversity in female and male 3xTg-AD mice compared to controls</p>	<ul style="list-style-type: none"> <li>• <i>Actinobacteria</i> and TM7 phylum alterations</li> <li>• <math>\beta</math> diversity changes</li> <li>• Increase in the bacteria families and genera: <i>Gemella</i>, <i>Allobaculum</i> and <i>Selenomonas</i></li> </ul>
Gu et al., 2021 [108]	Alzheimers Res. Ther.	APP/PS1 transgenic mice ( $n = 11$ ) were compared to WT	16S rRNA sequencing of the gut microbiome and integrated metabolomics	<p>↓ SCFA-producing bacteria (i.e., <i>Parasutterella</i> and <i>Blautia</i>) in APP/PS1 mice compared to controls</p> <p>↑ Gut dysbiosis in APP/PS1 mice compared to controls</p> <p>↑ <i>Firmicutes/Bacteroidetes</i> in APP/PS1 compared to WT</p>	<ul style="list-style-type: none"> <li>• Inflammatory factors (IL-6 and INF-<math>\gamma</math>), phosphatidylcholines and SCFA-producing bacteria as combinatorial biomarker for AD</li> </ul>
Shen et al., 2017 [109]	J. Alzheimers Dis.	APP/PS1 transgenic mice were compared to WT	16S rRNA sequencing	<p>↓ Gut microbiota diversity in APP/PS1 mice compared to controls</p>	<ul style="list-style-type: none"> <li>• Gut microbiota signature in AD and controls</li> </ul>



				<p>↓ <i>Prevotella</i> in APP/PS1 compared to controls</p> <p>↑ <i>Helicobacteraceae</i> and <i>Desulfovibrionaceae</i> in APP/PS1 compared to controls</p>
Chen et al., 2020 [110]	Biomed. Res. Int.	APP/PS1 transgenic mice were compared to WT controls ( $n = 14-24$ at 1–2–3–9 months and $n = 31-34$ at 6 months)	16S rRNA sequencing from fecal samples	<p>↑ <i>Proteobacteriaceae</i>, <i>Verrucomicrobiaceae</i>, <i>Bifidobacteriaceae</i>, <i>Erysipelotrichaceae</i> and <i>Prevotellaceae</i> in APP/PS1 mice</p> <p>↓ <i>Bacteroidaceae</i> and <i>Rikenellaceae</i> in APP/PS1 mice</p> <ul style="list-style-type: none"> <li>• Changes in gut microbiota composition precede plaque deposition: early biomarker</li> </ul>
Tan et al., 2020	Benef. Microbes	<i>Drosophila melanogaster</i> AD model compared to WT controls	Gut microbiota composition analysis	<p>↑ <i>Wolbachia</i> in AD flies compared to controls</p> <p>↓ Gut microbiota diversity in AD flies compared to controls</p> <ul style="list-style-type: none"> <li>• <i>Wolbachia</i> as a potential biomarker for AD</li> <li>• <i>Stenotrophomonas</i> negatively correlated with neurodegeneration</li> </ul>
Zhang et al., 2021 [111]	Am. J. Clin. Nutr.	Humans: 75 MCI individuals and 52 healthy controls	Changes in gut microbiota and serum miRNA expression	<p>↓ Microbial diversity, <i>Faecalibacterium</i>, <i>Ruminococcaceae</i>, <i>Alipstes</i> in MCI compared to controls</p> <p>↑ <i>Proteobacteria</i> and <i>Gammaproteobacteria</i> in MCI compared to controls</p> <ul style="list-style-type: none"> <li>• Differential gut microbiota composition, diet quality scores and serum miRNA as combinatorial biomarker for MCI patients</li> </ul>
Li et al., 2019 [113]	Alzheimers Dement.	Humans: AD patients ( $n = 30$ ), MCI patients ( $n = 30$ ), healthy controls ( $n = 30$ ).	Analysis of microbiota community in the faeces and blood via 16S rRNA sequencing	<p>↓ Microbial diversity in AD and MCI compared to controls</p> <p>≠ 11 genera in the feces and in the blood between AD/MCI and controls</p> <p>= Genera in the blood and feces between AD and MCI</p> <ul style="list-style-type: none"> <li>• Changes in gut microbiota as early diagnosis in AD</li> </ul>

Liu et al., 2019 [72]	Brain Behav. Immun.	Humans: AD patients ( $n = 33$ ), MCI patients ( $n = 32$ ) and healthy controls ( $n = 32$ )	16S rRNA MiSeq sequencing and phylogenetic investigation of communities by reconstruction of unobserved states	<p>↓ Microbial diversity in AD compared to MCI and controls</p> <p>↓ <i>Firmicutes</i> in AD compared to controls</p> <p>↑ <i>Proteobacteria</i> in AD compared to controls</p> <p>↑ <i>Gammaproteobacteria</i>, <i>Enterobacteriales</i> and <i>Enterobacteriaceae</i> in AD &gt; MCI &gt; controls</p>	<ul style="list-style-type: none"> <li>• The abundance of the <i>Enterobacteriaceae</i> family as a differential diagnostic tool for AD, MCI and healthy individuals.</li> </ul>
Ling et al., 2021 [73]	Front. Cell Dev. Biol.	Humans: 100 AD patients and 71 age- and gender-matched healthy controls	16S rRNA Miseq sequencing of fecal microbiota	<p>↓ Microbial diversity in AD compared to controls</p> <p>↓ Butyrate producing bacteria (<i>Faecalibacterium</i>)</p> <p>↑ Lactate producing bacteria (<i>Bifidobacterium</i>)</p>	<ul style="list-style-type: none"> <li>• Microbiota shift from butyrate producer to lactate producer genera (from <i>Faecalibacterium</i> to <i>Bifidobacterium</i>)</li> </ul>
Vogt et al., 2018 [114]	Alzheimer Res. Ther.	Humans: AD patients ( $n = 40$ ), MCI patients ( $n = 35$ ) and healthy controls ( $n = 335$ )	Cerebrospinal TMAO levels measurement	<p>↑ TMAO in AD and MCI compared to controls</p>	<ul style="list-style-type: none"> <li>• TMAO levels in the cerebrospinal fluid</li> </ul>
Wu et al., 2021 [115]	Nutrients	Humans: AD patients ( $n = 27$ ), MCI patients ( $n = 22$ ) and healthy controls ( $n = 28$ )	LC/GC/MS metabolomics profiling of fecal microbiota	<p>↓ Tryptophan metabolites in MCI and, more pronounced, in AD compared to controls</p> <p>↓ SCFAs in MCI and, more pronounced, in AD compared to controls</p>	<ul style="list-style-type: none"> <li>• Indole-3-pyruvic acid and five SCFAs for pre-onset and progression of AD</li> </ul>

Abbreviations: APP/PS1: APPswe/PSEN1dE9 transgenic; GC: gas chromatography; LDA: linear discriminant analysis; LEfSe: linear discriminant analysis effect size; LC = liquid chromatography; MCI: mild cognitive impaired; MS: mass spectrometry; SCFAs: short chain fatty acids; 3xTg; triple-transgenic mouse model of AD; TMAO: Trimethylamine N-oxide; WT: wild type; ↑: increase; ↓: decrease.

### 2.5. Prebiotics

Prebiotics are non-digestible organic substances (i.e., short-chain carbohydrates) capable of selectively stimulating the growth and/or activity of one or a limited number of beneficial bacteria present in the gut [124]. Being used as food from the gut microbiota, they stimulate the production of SCFAs, thus influencing both gastrointestinal and extra-intestinal functionality [125]. A growing body of evidence suggests their potentiality as

adjuvant therapy in different neurological and psychiatric conditions, such as anxiety, depression, and PD [126]. Recently, some studies are also considering the use of prebiotics for AD prevention/therapy, with promising results [124,126–136]. For example, yeast beta glucans administration to mouse models of AD proved effective in re-establishing the balance between pro-inflammatory and anti-inflammatory gut microbiome species, promoting SCFAs production and limiting neuroinflammation and insulin resistance [127]. Reduced neuroinflammation and improved short-term memory and cognitive ability in mice resembling AD features were also reported upon pre-treatment with lactulose and melibiose, two trehalose analogues, possibly via enhanced autophagy function [129]. Furthermore, 5xFAD mice fed for eight weeks with mannan oligosaccharide were shown capable of favoring the growth of *Lactobacillus* species and decreasing *Helicobacter* abundance, therefore preventing LPS leakage and intestinal epithelial barrier and BBB dysfunctions [130]. Interestingly, this prebiotic-driven reshaping of the gut microbiota was also accompanied by reduced A $\beta$  accumulation in different brain areas (i.e., cortex, hippocampus, and amygdala), re-established redox homeostasis, and increased butyrate levels [130]. Similar results were also obtained in both rats and mice models of AD via oral administration of *Marinanda officinalis*-derived oligosaccharides, reporting improved memory and learning ability, together with a decrease in plaque formation, oxidative stress, and overall inflammation [134,135]. Although the mechanism of action of the above-mentioned prebiotics is not totally clear, the capability of these molecules to sustain gut microbiota diversity and stability might be at the basis of these improvements [127,132,134]. This hypothesis is reinforced by recent evidence showing that a combination of probiotics and prebiotics (so-called synbiotics) seems to be more effective in increasing neurogenesis and reducing local and systemic inflammation compared to prebiotics alone [132].

Regarding humans, data on a large multi-ethnic longitudinal study comprising 1837 elderly people with no evidence of neurodegeneration have shown that daily administration of fructan, a well-known prebiotic, reduces the risk of AD development, confirming the previous evidence in mice [131]. However, despite this study being conducted normalizing for age, gender, recruitment time, ethnicity, daily caloric intake, education, and APOE genotype, other authors point out that the evidence for the use of prebiotics in the clinical practice still lacks robustness [133]. Altogether, these data suggest that prebiotics may be helpful as preventative/adjuvant therapy for AD, but more human clinical trials are needed before drawing any conclusion.

## 2.6. Probiotics

In 1965, Lilly and Stillwell introduced for the first time in the literature, the term “probiotics”, defining them as “living microorganisms with a low or zero pathogenicity that provide beneficial effects on the health of the host” [137]. Studies on human and animal models have shown that probiotics can modulate intestinal ecosystem homeostasis, regulate intestinal epithelial functions by helping to maintain the epithelial barrier, producing SCFAs, supporting cell survival, enhancing protective immune response, and inhibiting the production of pro-inflammatory cytokines [83,138–147]. Many of these responses arise from the regulation of specific intracellular signaling ways by probiotics, such as mitogen-activated protein kinases (MAPK) and nuclear factor (NF)- $\kappa$ B in intestinal epithelial cells [83,138–147]. Probiotic bacteria, through the modulation of the intestinal microbial ecosystem, have shown capable of playing an important role in immune response regulation by Th1, Th2, Th17, Treg cells, and NK and B cells stimulation [148]. Several studies have also confirmed the anti-inflammatory capacity of specific probiotics, by modulating the cytokine network and the macrophage tissue pattern, to reduce the mucosal inflammatory process and modulate the local immune response [149].

Probiotics can also modulate the gut–brain axis. The so-called psychobiotics, a new class of probiotics with potential applications in the treatment of psychiatric diseases, are able to modulate the bidirectional communication between brain and gut through the

modulation of neurotransmitters and proteins, including gamma-aminobutyric acid, serotonin, glutamate, and the brain-derived neurotrophic factor, which play important roles for the functionality of our central nervous system, mood, cognitive functions, learning and memory processes [150–153]. The administration of a probiotic mixture modified the gut microbiota in an animal model of AD by increasing *Actinobacteria* and *Bacteroides* with a significant impact on the enhancement of long-term memory, inflammation, and neural plasticity [154]. Mitochondrial dysfunction, excessive production of reactive oxygen species, and increased apoptosis have been implicated in the pathogenesis of AD. In this respect, several studies have highlighted the role of superoxide anion, hydroxyl radical, hydrogen peroxide, and nitric oxide in neurodegeneration mediated by oxidative stress in AD [155,156]. Recently, a study on transgenic AD mice demonstrates that the administration of a probiotic formulation (SLAB51) significantly reduces oxidative stress by inducing SIRT-1-dependent mechanisms [157]. In addition, the probiotic integration of a multi-species mixture of *Lactobacillus* and *Bifidobacterium* has proven capable of modifying specific brain metabolites such as  $\gamma$ -aminobutyric acid and glutamate [158]. Immune response and neural inflammation were also suppressed after probiotic integration with short A1 strain of *Bifidobacterium* [159]. Furthermore, the integration of *L. acidophilus*, *L. fermentum*, *B. lactis*, and *B. longum* improved learning disability and oxidative stress of rats subjected to intra-hippocampal injection of A $\beta$ 1-42 [160].

Although these studies on animal models show that probiotics may play an important role in two-way communication between gut and brain and support the potential role of probiotics in improving cognitive health, the results of clinical studies in subjects with AD or MCI are controversial.

In a recent randomized, double-blind, clinically controlled trial, 60 AD patients were divided into two groups and administered milk (control group) or probiotics (probiotic group). After 12 weeks of daily administration of 200 mL of a mixture of *Lactobacillus acidophilus*, *Lactobacillus casei*, *Bifidobacterium bifidum*, and *Lactobacillus fermentum*, a significant improvement in the mini-mental state exam (MMSE) score was reported compared to controls ( $P < 0.001$ ). Changes in plasma malondialdehyde, serum C-reactive protein, beta cells function, serum triglycerides, and differences in the quantitative control index of insulin sensitivity were also improved in the individuals receiving the probiotic mixture [161]. Similarly, data from another meta-analysis report a significant amelioration in cognition and a consistent reduction in post-intervention levels of malondialdehyde and high sensitivity C-reactive protein in subjects receiving probiotics compared to controls [162]. Although these results indicate potential benefit of probiotics in the management of patients with AD, other studies show contrary data. For example, in a recently published randomized, double-blind, placebo-controlled clinical trial, AD patients (between 65 and 90 years old) were supplemented with placebo (control group,  $n = 23$ , 13 females and 10 males) or a probiotic mixture (probiotic groups,  $n = 25$ , 18 females and 7 males). Two different probiotic capsules were used in this study: one containing *Lactobacillus fermentum*, *Lactobacillus plantarum*, and *Bifidobacterium lactis*, and one containing *Lactobacillus acidophilus*, *Bifidobacterium bifidum*, and *Bifidobacterium longum*. After 12 weeks of alternate day administration, the levels of proinflammatory (TNF- $\alpha$  and IL-6) and anti-inflammatory (IL-10) cytokines, as well as the levels of oxidizing (MDA and 8-OHdG) and antioxidants factors (TAC, GSH), were not significantly changed between the two groups. Of note, no improvements in cognitive functions were reported in the probiotic group compared to the placebo one, suggesting insensitivity to probiotic supplementation for severe AD patients [163].

In conclusion, even if there are several studies that show the influence of gut microbiota in neurological and psychiatric pathologies, the mechanisms of action and the effects of probiotics rest largely unknown, and several gaps and inconsistencies remain. Therefore, human studies need to be further developed and need to include analysis of the gut microbiota composition in specific populations of patients by identifying probiotic bacteria strains able to significantly affect gut–brain axis and assess their safe use.

### 2.7. Diet

Diet is a rapid and direct way of modifying the gut microbiota composition and function, reducing inflammation, and helping in eubiosis maintenance [123,164,165]. Given the evidence of association between neuropsychiatric conditions and gut microbiota dysregulation, it is worth speculating that dietary interventions could represent effective candidates for preventing and delaying the pathogenesis and progression of AD [166–186] (Table 2). Here, we present some of the most promising dietary therapies proposed in the literature, with a particular focus on Mediterranean and ketogenic diets.

**Table 2.** Evidence of diet as a possible complementary therapy in AD.

References	Type of Studies	Dietary Intervention	Aim	Outcomes
Duplantier et al., <i>Nutrients</i> , 2021 [166]	27 ObS, 5 RCT	Medi or DASH or MIND	Association between diet and cognitive health	Promising results for Medi diet but inconsistent outcomes. Lack of accuracy and standard tools
Bartochowski et al., <i>Curr. Nutr. Rep.</i> , 2020 [167]	4 RCT 24 RCT	Medi or MIND Vitamins and supplements (curcumin, EGb761, EPA, DHA)	Association between diet and AD	Protective and promising therapeutic role of Medi. Not enough evidence for MIND. No statistically significant results; promising evidence for vitamin D supplementation and curcumin use.
Gutierrez et al., <i>Nutrients</i> , 2021 [168]	61 RCT	Different dietary patterns	Effects of nutrition on cognitive function	Healthy food consumption (Medi Diet) improves cognitive function. Polyphenols have protective effects. Low evidence for PUFAs, vitamin D and other supplements.
Limongi et al., <i>J. Am. Med. Dir. Assoc.</i> , 2020 [169]	38 LS and 7 RCT	Medi	Association between diet and late-life cognitive disorders	Protective and promising therapeutic role of Medi diet for cognitive impairment.
Kheirouri et al., <i>Critical Reviews in Food Science and Nutrition</i> , 2021 [170]	9 CS, 3CrS, 1 RCT	MIND	Association between diet and neurodegenerative delay and cognitive functions	Improvement in cognition; limited number of studies and lack of mechanistic aspects in humans.
Lilamand et al., <i>Curr. Opin. Clin. Nutr. Metab. Care</i> , 2021 [171]	8 IS	KD or KS	Association between diet and cognitive and biological/neuropathological outcomes	Evident improvement: decrease in cerebral inflammation, A $\beta$ -amyloid, aggregates of tau protein.
Grammatikopoulou et al., <i>Adv. Nutr.</i> , 2020 [172]	10 RCT	KD or KS	Effects of KD on patients with AD/mild cognitive impairment	Improvement in acute and long-term cognition.
Pavón et al., <i>Nutr. Rev.</i> , 2021 [173]	N/A	KD or KS	Effect of KD on cognitive skills in patients with	Improvements in memory, cognitive performance and learning capabilities

			AD, PD, refractory epilepsy, and type 1 glucose deficiency syndrome	
Jensen et al., Int. J. Mol. Sci., 2020 [174]	N/A	KD or KS	Effects of KD on brain metabolism and function in neurodegenerative diseases	Reduction in AD symptoms.
Christensen et al., Nord. J. Psychiatry, 2021 [175]	24 RCT	KD or KS or modified Atkins diet	Effects of KD on CNS diseases	Modified-Atkins diet significantly improved memory in AD patients.
Moreira et al., Dement. neuropsychol., 2020 [176]	32 RCT	Omega-3, nutritional formula including ginseng, inositol and coconut oil	Association between diet and cognitive performance in AD	Omega-3 fatty acids showed positive effects at different doses. Probiotic, Ginseng, Inositol and specialized nutritional formulas might have a positive effect on cognition.
Zhang et al., Nutrients, 2020 [177]	12 CS, 3 case-control, 13 CrS, 1 IS	Meat	Association between meat (red meat, processed meat and poultry) consumption and cognitive functions	No significant association.
Dimache et al., Nutrients, 2021 [178]	21 (ObS, LS, CrS, IS)		Association between triglycerides with cognitive, vascular cognitive impairment and amyloid accumulation	In longitudinal studies: TG level is associated with cognitive decline. In cross sectional studies no correlation.
Gkatzamanis et al., Psychiatriki, 2020 [179]	4 RCT 6 RCT	Omega-3 polyphenols	Effect of supplementation on dementia	Promising preventative but not therapeutic effect.
El Gaamouch et al., Neurochem. Int., 2021 [180]	N/A	Grape polyphenols	Association between grape polyphenols and AD	No significant results from interventions.
Colizzi et al., Alzheimers Dement. (N Y), 2019 [181]	24 RCT	Polyphenols	Association between polyphenols and AD	12 studies found a positive correlation with reduced cognitive decline; 5 studies did not find any correlation and 7 studies reported mixed results.
Mielech et al., Nutrients, 2020 [182]	8 CS/RCT 3 CS/RCT	Vitamins B Vitamin A	Association between antioxidant vitamins	4 studies: beneficial effect slowing cognitive decline; 4 studies: no differences Protective effect for cognitive functions in 2 studies.

	7 CS/RCT	Vitamins C and E	and AD and cognitive decline	Protective effect for AD in 5 studies.
	7 CS/RCT	Vitamin D		Low level in the serum associated with increased risk of cognitive decline; no positive correlation with supplementation.
Szczechowiak et al., Pharmacology Biochemistry and Behavior, 2019 [183]	N/A	Pro-inflammatory (rich in saturated fats, meat) vs anti-inflammatory (rich in vitamins, antioxidants, probiotics) diet	Association between pro- and anti-inflammatory diets and AD prevention and treatment	Overconsumption of foods rich in d-AGEs (Dietary Advanced Glycosylation End-products), saturated fats and red and processed meat have a pro-inflammatory influence on AD patients' brains.
Kosti et al., Nutr. Rev., 2021 [184]		Fish, EPA/DHA supplementation	Associations between fish intake and AD dementia or AD and the effect of EPA/DHA supplementation on cognitive performance.	Regular consumption of fish up to 2 portions per week seems to be more protective than EPA/DHA supplementation.
Haider et al., International Journal of Geriatric Psychiatry, 2020 [185]	4 RCT	Vitamins B and E, omega-3, polyunsaturated fatty acids.	Effects of nutritional supplementation on neuropsychiatric symptoms among people with dementia	No significant results.
Arbo et al., Front. Aging Neurosci., 2020 [186]	3 RCT, 1 retrospective study	Resveratrol	Effect of resveratrol as potential treatment in AD and PD	No significant results in human trails.

Abbreviations: CrS: cross sectional study; CS: cohort studies; DHA: docosahexaenoic acid; EGb 761: Ginkgo biloba extract 761; EPA: eicosapentaenoic acid; KD: ketogenic diet; KS: ketogenic supplement; IS: interventional study; LS: longitudinal study; Obs: observational studies; RCT: randomized controlled trial.

### 2.7.1. Mediterranean, DASH (Dietary Approaches to Stop Hypertension), and MIND (Mediterranean-DASH Intervention for Neurodegenerative Delay)

The renowned and ancient Mediterranean diet (Medi), rich in vegetables, fruit, whole grains, nuts, olive oil, moderate consumption of fish and poultry and limited consumption of red meat and sweets, have been extensively described for their protective role against non-communicable diseases [187]. DASH diet (Dietary Approaches to Stop Hypertension), designed for hypertension treatment, overlaps the Medi diet in composition, with more attention on salt introduction (less than 2.4 g/day) [188]. Similarly, MIND diet (Mediterranean-DASH Intervention for Neurodegenerative Delay) is a combination of both, DASH and Medi, specifically developed to delay neurodegeneration. Besides being rich

in fruits, vegetables and legumes, the MIND includes the consumption of single dietary components, i.e., green leafy vegetables and berries, which have displayed a superior effect against cognitive impairment and decline compared to other vegetables and fruits [189].

Although Randomized Clinical trials (RCT) and Observational Cohort Studies (ObS) have been conducted to unravel the potential therapeutic effect of Medi, DASH, and MIND in AD, the results are still unclear [166]. Two large RCT conducted in Spain in 2013 and 2015 have demonstrated a positive correlation between ‘Medi diet plus olive oil’ or ‘Medi diet plus nuts’ with cognitive performance [190,191]. More recently, an additional RCT study also associated Medi diet with improved cognition [192]. However, differential results from another RCT did not show any significant association [193]. Further narrative, systematic reviews, and meta-analyses have evidenced the protective and promising therapeutic role of Medi diet in AD disease, confirming its ability to hinder cognitive impairment [167,169]. Generally, any dietary pattern rich in fruits, vegetables, and legumes and poor in saturated fats and sweets seems to provide protective effects [194]. Similarly, results presented by Barbaresko et al. on 20 systematic reviews and meta-analyses, highlighted the benefits of the Medi diet as a protective factor for AD [195].

So far, besides promising results for Medi diet, the role of DASH diet in AD prevention and therapy is still unveiled [168], and more studies should be carried out before driving any conclusion. Moreover, standard tools for assessing food intake and cognitive decline are needed to state which dietary pattern might be the most effective in protecting and delaying the onset of neurodegenerative diseases, and to ensure reproducibility [166].

Concerning MIND diet, Morris et al. were the first to show that a moderate adherence to this dietary habit slows cognitive decline compared to a moderate adherence to Medi and DASH diets; however, they have also confirmed that a high adherence to Medi and DASH diets can reduce AD risk [189]. Potential neuroprotective mechanisms shared by those dietary regimes are the presence of antioxidant and anti-inflammatory compounds, which contribute to a reduction in brain inflammation and oxidative stress, high abundances of fibers, vitamin C, beta-carotene, and folate, which lead to a better brain integrity and increase in brain tissue volume [196,197]. Also, the scarcity in saturated and trans fatty acids can reduce BBB dysfunction and amyloid aggregation [183,198,199].

A significant improvement in cognition was also reported among older adults following the MIND diet, confirming the effectiveness of this approach [170]. Despite those findings, the lack of evidence on the correlation between MIND diet and brain-related mechanisms, and given the similarities with the Medi and DASH diets in terms of nutrients composition, MIND diet cannot be disclosed as more proactive than Medi and DASH diets.

On the whole, as previously mentioned, the protective and potential therapeutic effect of Medi (and similar diets) might be based on the consumption of much food rich in vitamins and polyphenols, i.e., fruits, vegetables, legumes and whole grains, a moderate amount of fish, and less meat and food rich in trans and saturated fats. Regarding meat, so far, the majority of the studies did not report any significant association with cognitive impairment or decline [177]. Differently, fish intake is inversely associated with AD—likely related to omega-3 (EPA/DHA) contents [195]. Interestingly, the regular consumption of fish up to two portions per week seems to be more protective than EPA/DHA supplementation [184]. Even though many studies are supporting the protective effect of unsaturated fatty acids EPA/DHA, their role in the brain is still under debate [200–204]. Medi diet is also connected to an improved lipid profile. Overall, lipid dysregulation might contribute to AD pathogenesis, enhancing synaptic loss, BBB dysfunction, mitochondrial disruption, oxidative stress, and inflammation [198,205]. Indeed, in large longitudinal studies, high levels of triglycerides and cholesterol in the serum are significantly associated with cognitive impairment [178]. Again, a recent cross-sectional study with 689 participants including AD and healthy patients, revealed that reduced levels of triglycerides were related to better cognitive performance and a reduction in brain dysfunction and



atrophy [206]. In conclusion, even if the interplay between dietary lipids and AD pathogenesis is not straightforward, Medi diet with consequent improvement in lipid dysregulation through dietary changes is strongly recommended.

Dietary regimens based on a daily integration of the essential nutrients and vitamins are also of interest, but the data remain limited. Cross-sectional and longitudinal cohort studies on vitamins C and E consumption showed promising effects in reducing cognitive decline, but no difference has been identified in intervention trials [182]. Similarly, low levels of vitamin D in the serum seem to be associated with an increased risk of cognitive decline, but its supplementation did not provide any difference [182]. Vitamin B (folic acid, pyridoxine, and cobalamin) consumptions lead to ambiguous results, with only a few RTC displaying beneficial effects in slowing the cognitive decline [182]. Finally, even though vitamin A supplementation might reduce the risk of cognitive decline, there are not enough consistent data to confirm its protective and therapeutic effect in AD [182]. Overall, it seems that some vitamin supplementation might delay the progression of AD and dementia; nonetheless, due to the lack of statistically significant results and limited scientific evidence analyzing the role of vitamins in older adults [167,185], it is not possible, at least until now, to point out their specific protective and therapeutic effects in AD.

Besides micronutrients and omega-3, further nutritional formula including ginseng, inositol, and coconut oil have been recently studied as potential therapy in AD patients, but the effects are inconclusive [176].

Polyphenols are receiving growing interest in AD research due to their antioxidant, anti-inflammatory, and neurotrophic properties supported by preclinical evidence. Nonetheless, so far, there is no conclusive evidence on the association between polyphenols and AD in humans. On 24 RCT conducted on AD patients exposed to polyphenols (mainly flavonoids), only 12 have shown a reduction in cognitive decline [181]. Again, further trials carried out in people with mild cognitive impairment consuming grape juice or blueberries rich in polyphenols showed minimal benefits in memory or no significant results [179,180]. A polyphenol that might contribute to neuroprotection is resveratrol. This phenolic compound promotes synthesis of glutamate receptors, enhances synaptic transmission, activates SIRT1, exerts antioxidant and anti-inflammatory actions [186,207]. Results from *in vitro* and *in vivo* (mice and rats) studies underscored resveratrol as a potential treatment for AD; however, its effectiveness is only partially understood in humans [208]. Although some research groups have performed trials in humans to test the potential protective effect of resveratrol, results have failed to demonstrate a positive correlation. The lack of a substantial number of clinical trials and issues related to clinical applications, e.g., dosage, bioavailability, side effects, etc., emphasize the need of further investigation [186].

### 2.7.2. Ketogenic Diet

Ketogenic diet (KD) is a nutritional program rich in fats and low in carbohydrates and proteins (ideally, 90% fat, 4% carbohydrates, 6% proteins) developed in the early 1990s as a treatment for epilepsy, with numerous studies consistently supporting its effectiveness [209]. Recently, the application of KD as potential treatment for other neurological diseases, such as PD and AD, has been investigated *in vitro* and *in vivo* [210–212]. The sugar-shortage leads the body to break down and oxidate fats with the production of ketone bodies, used as an alternative energy-substrate to glucose by many organs, including the brain [213]. In mice models, ketone bodies influence neurotransmission, channels modulation, increase BDNF, reduce neuroinflammation and oxidative stress, improve mitochondrial functions, reduce amyloid accumulation, and improve learning and memory abilities [213–216]. In humans, results from RCT reported that KD might be beneficial in people with mild cognitive impairment or AD [171,172]. Similar to KD in terms of mechanisms (i.e., ketone bodies production), medium-chain triglyceride (MCT) diet/supplementation and the modified Atkins diet are effective in counteracting cognitive decline in AD, symptoms such as fatigue and daytime sleepiness in PD, epileptic seizures and mood

swings in depression [173–175,217,218]. Moreover, the modified Atkins diet, which does not restrict protein intake as the KD protocol, allows a much more nutritional flexibility than classic KD. Indeed, overall, dietary patterns that lead to ketone bodies production seem to represent a promising therapy for AD, but more investigation to unveil protective mechanisms in humans and adverse aspects is needed—including the lack of flexibility and variability of the alimentary regimen easily leading to a drop-out, the scarcity of plant-based food rich in vitamins, and other antioxidant compounds [219].

Even if further well-designed human clinical trials are needed to better understand the role of diet for the prevention and treatment of AD, up to now, most of the diet-related beneficial effects in AD patients seem to be in favor of the Medi diet. Vitamins and other phenolic compounds might represent potential boosts for AD patients.

### 2.7.3. The Role of Diet in AD Mediated Through Gut Microbiota

Diet is the most impactful modulator of gut microbiota across lifespan. Considering that, as previously mentioned, AD is associated with changes in microbiota composition, it is reasonable to assume that dietary interventions, and the related gut microbiota composition shifts, might constitute a future complementary tool to prevent or manage dementia. However, the proof of a cause–effect relationship among gut microbiota, diet, and neurodegeneration is very poor, with a low number of clinical studies analyzing the interplay among those elements.

Current evidence shows that the Medi diet beneficially impacts the gut microbiota composition in elderly adults reducing frailty [220,221]. The abundance of specific “protective” taxa (e.g., *Faecalibacterium* or *Roseburia*) was positively associated with improved cognitive function and negatively associated with pro-inflammatory markers, possibly related to an increase in SCFAs [165]. Medi diet is rich in polyphenols, and emerging evidence supports the beneficial role of polyphenols in preventing and/or ameliorating AD progression, reducing plaques formation and protecting blood brain barrier disruption.[222]. Wine polyphenols, for instance, may lead to an increase in *Bacteroides*, *Bifidobacteria*, and *Lactobacilli*, re-storing a ‘healthy’ microflora composition in AD patients [223].

Improvement in the microbiota profile as a result of KD was initially showed in pre-clinical animal models, where mice subjected to KD diet displayed an increase in *Akkermansia* and *Lactobacillus* and a parallel enhancement in vascular brain function [224]. An interventional study, where MCI patients followed a modified Medi-KD, demonstrated that dietary regimen positively affects the gut microbiota composition, with an increase in the abundance of *Enterobacteriaceae*, *Akkermansia*, *Christensenellaceae*, and in SCFAs production, with a consequent improvement in cognitive symptoms [225]. Nonetheless, while a recent study in an animal model of AD revealed that KD might exacerbate gut dysbiosis, a diet rich in carbohydrates seemed to improve the microbiota profile with an increase in *Bacteroidetes* and a reduction in *Proteobacteria* [226,227]. Dietary patterns that allow not-refined carbohydrates consumption, but still lead to ketone bodies production, e.g., intermittent fasting, might be a promising protective dietary strategy for dementia[226].

Decoding the interplay between microbiota and diet in neurogenerative disease patients seem to be promising; however, all the multi-faceted aspects of dietary patterns on human health should be examined in depth, considering the body as a superorganism, made of human and microbial cells.

### 2.8. Fecal Microbiota Transplantation

Fecal microbiota transplantation (FMT) is a procedure where a solution of fecal material from a donor is transferred (through colonoscopy, nasogastric tube, or oral pills) into the intestinal tract of a recipient, aimed at directly changing the gut microbiota composition [227]. Reprogramming the gut microbiota *eubiosis* by FMT has been already used to successfully treat *C. difficile* infections and could be an innovative therapy for various neurological diseases in an imminent future [228]. So far, most of the limited number of

studies have been conducted in mice/rats, with promising but not conclusive results (Table 3) [10,229–239].

**Table 3.** Murine and human studies performing FMT in AD.

Ref	Journal	Study Cohort/Sample Size	Donor	Recipient	Transplantation Technique	Results
Hazan et al., 2020 [229]	J. Int. Med. Res.	Case study ( $n = 1$ )	85-year-old woman (recipient's wife)	82-year-old man with recurrent CDI and AD	Single 300 mL FMT infusion	<ul style="list-style-type: none"> <li>↑ Cognitive function (MMSE test)</li> <li>↑ Memory</li> <li>↑ Mood</li> </ul>
Park et al., 2021 [230]	Curr. Med. Res. Opin.	Case study ( $n = 1$ )	27-year-old healthy man	90-year-old woman with AD and severe CDI	Colonoscopy (60 g of stool suspension for 2 times).	<ul style="list-style-type: none"> <li>↑ Cognitive function tests (MMSE, MCA and CDR tests)</li> <li>↑ Microbiota <math>\alpha</math> diversity</li> <li>= Microbiota <math>\beta</math> diversity</li> <li>↑ SCFAs</li> </ul>
Kim et al., 2021 [231]	Brain. Behav. Immun.	Mouse ( $n = 8$ )	5xFAD mice	C57BL/6 mice	Oral gavage (200 $\mu$ l for 5 consecutive days)	<ul style="list-style-type: none"> <li>↓ Adult hippocampal neurogenesis and BDNF expression</li> <li>↑ p21 expression</li> <li>↑ Microglia activation</li> <li>↑ TNF-<math>\alpha</math> and IL-1<math>\beta</math></li> <li>↑ Colon and plasma pro-inflammatory cytokines</li> </ul>
Sun et al., 2019 [232]	Transl. Psychiatry	Mice ( $n = 8$ )	WT mice	APPswe/PS1dE9 transgenic (Tg) mouse model	Intragastrically (0.2 mL of fresh fecal solution once daily for 4 weeks)	<ul style="list-style-type: none"> <li>↑ Cognitive function (MWM and ORT tests)</li> <li>↓ Amyloid <math>\beta</math> brain deposition (A<math>\beta</math>40 and A<math>\beta</math>42)</li> <li>↓ Tau protein phosphorylation</li> <li>↑ Synaptic plasticity (increased PSD-95 and synapsin I)</li> <li>↓ COX2 and CD11b</li> </ul>

						↑ SCFA and microbiota composition
Wang et al., 2021 [233]	Brain. Behav. Immun.	Mice ( $n = 4$ )	16 months old APP <sup>SWE</sup> /PS1 <sup>ΔE9</sup> mice	3 months old APP <sup>SWE</sup> /PS1 <sup>ΔE9</sup> mice	Antibiotic cocktails for 2 weeks by gavage and then FMT for 7 consecutive days by oral gavage	↑ Aβ plaques ↓ Astrocyte activation around Aβ plaques
Kim et al., 2020 [10]	Gut	Mice ( $n = 16$ )	WT mice	ADLP <sup>APT</sup> transgenic mouse model	Fresh fecal matters for 16 weeks by oral gavage or for 4 weeks in mice pre-treated with antibiotics	↓ Aβ plaques ↓ Neurofibrillary tangles ↓ Glial reactivity ↓ Cognitive impairment ↓ Circulating blood inflammatory monocytes
Harach et al., 2017 [234]	Sci. Rep.	Mice ( $n = 6$ )	12 month-old CONVR-WT or CONVR-APP <sup>PS1</sup> mice	4 month-old GF-APP <sup>PS1</sup> mice	Oral gavage of fecal contents on day 1 and day 4	↑ Cerebral Aβ pathology
Fujii et al., 2019 [235]	Biosc. Biotechnol. Biochem.	Humanized mice ( $n = 7$ )	4-weeks old germ-free C57BL/6N mice	Human healthy volunteers (76-year-old female) or AD patients (82-year-old male)	Oral inoculation	↓ OLT and ORT in mice colonized with AD microbiome ↓ γ-aminobutyrate, taurine and valine in mice colonized with AD microbiome
Zhan et al., 2018 [236]	Aging	Mice ( $n = 8$ )	SAMP8 or SAMR1 mice	pseudo germ-free mice	0.2 mL fecal suspension by gavage for 14 days	↑ Behaviour (only from SAMR1 transplant) ↑ α diversity and β diversity (only from SAMR1 transplant) ↓ Abnormal microbiota
Dodiya et al., 2019 [237]	J. Exp. Med.	Mice ( $n = 9$ )	age-matched APP <sup>PS1-21</sup>	ABX-treated APP <sup>PS1-21</sup> male	0.2 mL fecal slurry by gastric gavage daily starting on P25 until sacrifice	↓ Aβ pathology ↑ Microglial physiology
Cui B. et al., 2018 [238]	Journal of Neuroinflammation	Mice ( $n = 6$ )	Low intensity noise (LN) exposure SAMP8	male 3-month-old SAMP8 mice	0.1 mL fecal preparation via oral gavage	↑ CLDN1 and ZO-1 in intestine

			mice (control group) and high intensity noise (HN) exposure (AD model group)		twice per week for 30 days	and hippocampus of HN microbiota recipient ↑ Aβ in hippocampus of the HN microbiota recipient
Valeri et al., 2021 [239]	Microorganisms	Mice (n = 10)	Either 4 months old or 1 year old wild type mice	5xFAD mice (4-month old)	150 μ fecal preparation via oral gavage one time after antibiotics-treatment	↑ <i>Enterobacteriaceae</i> , <i>Lactobacillaceae</i> , serum LPS binding protein ↓ <i>Firmicutes</i> ↑ Plaques in dentate gyrus and prefrontal cortex

ABX: antibiotic cocktail; *APP<sup>SWE/PS1<sup>L166P</sup></sup>*: APPPS1-21; BDNF: brain derived neurotrophic factor; CDI: *Clostridioides difficile* infection; CDR: Clinical Dementia Rating assessment; CLDN1: claudin 1; CONVR-APPPS1: conventionally-raised transgenic APPPS1 mice; COX2: cyclooxygenase 2; FMT: fecal microbiota transplantation; LPS: lipopolysaccharide; MCA: Montreal Cognitive Assessment; MMSE: Mini-Mental State Examination; MWM: Morris water maze test; OLT: object location test; ORT: object recognition test; PSD-95: postsynaptic density protein 95; SAMP8: senescence-accelerated mouse prone 8; SAMR1: senescence-accelerated mouse resistant 1; SCFAs: short chain fatty acids; ZO-1: Tight junction protein-1; ↑: increase; ↓: decrease.

Impaired neurogenesis, decreased BDNF expression, increased memory impairment, enhanced circulating pro-inflammatory cytokines, and Aβ plaques deposition were detected when feces from AD-model donor mice were transplanted in healthy mice [231,233]. Moreover, FMT from senescence-accelerated mice or from senescence-accelerated-resistant mice into germ-free (GF) mice revealed significant differences on behaviors, cognitive performance, and gut microbiota composition, with a better profile in recipient mice receiving the microbiota from senescence-accelerated-resistant donors compared to senescence-accelerated mice [236]. Similarly, GF mice receiving fecal material from APPPS1 transgenic mice developing cerebral Aβ-deposition showed an increase in plaques formation [234,235]. When FMT was carried out from an AD patient into GF mice, accelerated cognitive decline and a decrease in microbiota-derived metabolites important for the nervous system function were reported [235].

Successfully, researchers confirmed that interventions aimed at manipulating gut microbiota influence brain disorders. Indeed, transplanting healthy fecal microbiota from wild-type mice to mouse models of AD documented a decrease in cognitive impairment, amyloid accumulation, and circulating levels of pro-inflammatory markers [10]. Improved cognition, reduced amyloid accumulation and tau expression, enhanced synaptic plasticity, and increased SCFAs-producing gut endobacteria were also confirmed in another study [232]. Dodiya et al. reported the effectiveness of FMT in restoring microbiota composition in the APP/PS1 transgenic mouse model of AD, improving microglia and Aβ deposition profile [237].

Regarding humans, only two case-studies showing promising results have been conducted so far [229,230]. Hazan et al. demonstrated an improvement in AD symptoms (cognitive function, memory, and mood) in a 82-year-old man after FMT from a 85-year-old woman (recipient's wife) [229]. A second case-study, involving a 90-year-old woman with AD and severe *C. difficile* infection who received FMT from a 27-year-old healthy man,

also showed an improvement in cognitive function, microbiota diversity, and SCFAs production [230].

Despite the potential application of FMT in AD treatment, several limitations still exist. Standardization of the therapeutic protocols, timings and length of administration, short and term risks, and inclusion criteria are all points that should be considered and addressed [68,240–244].

In conclusion, although the promising results obtained in mice certainly prove that the gut microbiota is involved in the pathogenesis and progression of neurological diseases, more human studies are needed before pointing out FMT as an AD complementary therapy.

### 3. Conclusions

AD is a neurodegenerative disorder, often occurring in the elderly, which has a fundamental causative source in the impairments in the GMBA. Recent data, which are to be further deepened and improved in any investigation planning, reported to date a close relationship between gut microbiota composition (then affected by nutritional habits) and AD onset, usually derived from neuroinflammation caused by bacteria products or bacterial brain migration, a circumstance that normally occurs to contribute to the regulation of brain synaptogenesis and development, besides mood and cognition evolution. Given the close cross talk between gut bacteria and brain, here we reviewed that gut microbiota dysregulations, often reported in AD patients, can be exploited to investigate both new diagnostic and therapeutic approaches for this devastating disease. However, despite promising results have been published, more research is needed to limit interstudy inconsistencies and enhances reproducibility before considering a clinical application.

**Author Contributions:** Conceptualization, A.V. and G.R.; Writing—original draft preparation, A.V., G.B.P., E.P., M.R., S.C., G.B., A.O. and C.A.; Writing—review and editing, A.V. and A.P.; Supervision, A.P., C.E., and G.R. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** We kindly thank the Department of Internal Medicine and Therapeutics of the University of Pavia, Pavia, Italy, and MR MEDICA SRL for supporting the publication costs.

**Conflicts of Interest:** The authors declare no conflicts of interest.

### References

1. Hodson, R. Alzheimer's Disease. *Nature* **2018**, *559*, S1. <https://doi.org/10.1038/d41586-018-05717-6>.
2. Mattson, M.P. Pathways towards and away from Alzheimer's Disease. *Nature* **2004**, *430*, 631–639. <https://doi.org/10.1038/nature02621>.
3. Ashraf, G.M.; Tarasov, V.V.; Makhmutova, A.; Chubarev, V.N.; Avila-Rodriguez, M.; Bachurin, S.O.; Aliev, G. The Possibility of an Infectious Etiology of Alzheimer Disease. *Mol. Neurobiol.* **2019**, *56*, 4479–4491. <https://doi.org/10.1007/s12035-018-1388-y>.
4. Oxford, A.E.; Stewart, E.S.; Rohn, T.T. Clinical Trials in Alzheimer's Disease: A Hurdle in the Path of Remedy. *Int. J. Alzheimers Dis.* **2020**, *2020*, 1–13. <https://doi.org/10.1155/2020/5380346>.
5. Yaari, R.; Hake, A. Alzheimer's Disease Clinical Trials: Past Failures and Future Opportunities. *Clin. Investig.* **2015**, *5*, 297–309. <https://doi.org/10.4155/cli.14.127>.
6. Rosenblum, W.I. Why Alzheimer Trials Fail: Removing Soluble Oligomeric Beta Amyloid Is Essential, Inconsistent, and Difficult. *Neurobiol. Aging* **2014**, *35*, 969–974. <https://doi.org/10.1016/j.neurobiolaging.2013.10.085>.
7. Ausó, E.; Gómez-Vicente, V.; Esquiva, G. Biomarkers for Alzheimer's Disease Early Diagnosis. *J. Pers. Med.* **2020**, *10*, 114. <https://doi.org/10.3390/jpm10030114>.
8. Rasmussen, J.; Langerman, H. Alzheimer's Disease—Why We Need Early Diagnosis. *Degener. Neurol. Neuromuscul. Dis.* **2019**, *9*, 123–130. <https://doi.org/10.2147/DNND.S228939>.

9. Cryan, J.F.; O’Riordan, K.J.; Cowan, C.S.M.; Sandhu, K.V.; Bastiaanssen, T.F.S.; Boehme, M.; Codagnone, M.G.; Cusotto, S.; Fulling, C.; Golubeva, A.V.; et al. The Microbiota-Gut-Brain Axis. *Physiol. Rev.* **2019**, *99*, 1877–2013. <https://doi.org/10.1152/physrev.00018.2018>.
10. Kim, M.-S.; Kim, Y.; Choi, H.; Kim, W.; Park, S.; Lee, D.; Kim, D.K.; Kim, H.J.; Choi, H.; Hyun, D.-W.; et al. Transfer of a Healthy Microbiota Reduces Amyloid and Tau Pathology in an Alzheimer’s Disease Animal Model. *Gut* **2020**, *69*, 283–294. <https://doi.org/10.1136/gutjnl-2018-317431>.
11. Long-Smith, C.; O’Riordan, K.J.; Clarke, G.; Stanton, C.; Dinan, T.G.; Cryan, J.F. Microbiota-Gut-Brain Axis: New Therapeutic Opportunities. *Annu. Rev. Pharmacol. Toxicol.* **2020**, *60*, 477–502. <https://doi.org/10.1146/annurev-pharmtox-010919-023628>.
12. Rhee, S.H.; Pothoulakis, C.; Mayer, E.A. Principles and Clinical Implications of the Brain–Gut–Enteric Microbiota Axis. *Nature Nat. Rev. Gastroenterol. Hepatol.* **2009**, *6*, 306–314. <https://doi.org/10.1038/nrgastro.2009.35>.
13. Sonnenburg, J.L.; Sonnenburg, E.D. Vulnerability of the Industrialized Microbiota. *Science* **2019**, *366*, eaaw9255. <https://doi.org/10.1126/science.aaw9255>.
14. Pascale, A.; Marchesi, N.; Marelli, C.; Coppola, A.; Luzi, L.; Govoni, S.; Giustina, A.; Gazzaruso, C. Microbiota and Metabolic Diseases. *Endocrine* **2018**, *61*, 357–371. <https://doi.org/10.1007/s12020-018-1605-5>.
15. Kowalski, K.; Mulak, A. Brain-Gut-Microbiota Axis in Alzheimer’s Disease. *J. Neurogastroenterol. Motil.* **2019**, *25*, 48–60. <https://doi.org/10.5056/jnm18087>.
16. Liu, S.; Gao, J.; Zhu, M.; Liu, K.; Zhang, H.-L. Gut Microbiota and Dysbiosis in Alzheimer’s Disease: Implications for Pathogenesis and Treatment. *Mol. Neurobiol.* **2020**, *57*, 5026–5043. <https://doi.org/10.1007/s12035-020-02073-3>.
17. Burokas, A.; Moloney, R.D.; Dinan, T.G.; Cryan, J.F. Microbiota Regulation of the Mammalian Gut–Brain Axis. *Adv. Appl. Microbiol.* **2015**, *91*, 1–62.
18. de J.R. De-Paula, V.; Forlenza, A.S.; Forlenza, O.V. Relevance of Gutmicrobiota in Cognition, Behaviour and Alzheimer’s Disease. *Pharmacol. Res.* **2018**, *136*, 29–34. <https://doi.org/10.1016/j.phrs.2018.07.007>.
19. Stilling, R.M.; Dinan, T.G.; Cryan, J.F. Microbial Genes, Brain & Behaviour – Epigenetic Regulation of the Gut-Brain Axis. *Genes Brain Behav.* **2014**, *13*, 69–86. <https://doi.org/10.1111/gbb.12109>.
20. Carabotti, M.; Scirocco, A.; Maselli, M.A.; Severi, C. The Gut-Brain Axis: Interactions between Enteric Microbiota, Central and Enteric Nervous Systems. *Ann. Gastroenterol.* **2015**, *28*, 203–209.
21. Kaur, G.; Behl, T.; Bungau, S.; Kumar, A.; Uddin, M.S.; Mehta, V.; Zengin, G.; Mathew, B.; Shah, M.A.; Arora, S. Dysregulation of the Gut-Brain Axis, Dysbiosis and Influence of Numerous Factors on Gut Microbiota Associated Parkinson’s Disease. *Curr. Neuropharmacol.* **2020**, *19*, 233–247. <https://doi.org/10.2174/1570159X18666200606233050>.
22. Heck, A.L.; Handa, R.J. Sex Differences in the Hypothalamic–Pituitary–Adrenal Axis’ Response to Stress: An Important Role for Gonadal Hormones. *Neuroendocrinology* **2019**, *44*, 45–58. <https://doi.org/10.1038/s41386-018-0167-9>.
23. Cerovic, M.; Forloni, G.; Balducci, C. Neuroinflammation and the Gut Microbiota: Possible Alternative Therapeutic Targets to Counteract Alzheimer’s Disease? *Front. Aging Neurosci.* **2019**, *11*, 284. <https://doi.org/10.3389/fnagi.2019.00284>.
24. Wang, H.-X.; Wang, Y.-P. Gut Microbiota-Brain Axis. *Chin. Med. J.* **2016**, *129*, 2373–2380. <https://doi.org/10.4103/0366-6999.190667>.
25. Pascale, A.; Marchesi, N.; Govoni, S.; Barbieri, A. Targeting the Microbiota in Pharmacology of Psychiatric Disorders. *Pharmacol. Res.* **2020**, *157*, 104856. <https://doi.org/10.1016/j.phrs.2020.104856>.
26. Margolis, K.G.; Cryan, J.F.; Mayer, E.A. The Microbiota-Gut-Brain Axis: From Motility to Mood. *Gastroenterology* **2021**, *160*, 1486–1501. <https://doi.org/10.1053/j.gastro.2020.10.066>.
27. Bravo, J.A.; Forsythe, P.; Chew, M.V.; Escaravage, E.; Savignac, H.M.; Dinan, T.G.; Bienenstock, J.; Cryan, J.F. Ingestion of Lactobacillus Strain Regulates Emotional Behavior and Central GABA Receptor Expression in a Mouse via the Vagus Nerve. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 16050–16055. <https://doi.org/10.1073/pnas.1102999108>.
28. Heijtz, R.D.; Wang, S.; Anuar, F.; Qian, Y.; Bjorkholm, B.; Samuelsson, A.; Hibberd, M.L.; Forssberg, H.; Pettersson, S. Normal Gut Microbiota Modulates Brain Development and Behavior. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 3047–3052. <https://doi.org/10.1073/pnas.1010529108>.
29. Rea, K.; Dinan, T.G.; Cryan, J.F. The Microbiome: A Key Regulator of Stress and Neuroinflammation. *Neurobiol. Stress* **2016**, *4*, 23–33. <https://doi.org/10.1016/j.ynstr.2016.03.001>.
30. Dichter, G.S.; Damiano, C.A.; Allen, J.A. Reward Circuitry Dysfunction in Psychiatric and Neurodevelopmental Disorders and Genetic Syndromes: Animal Models and Clinical Findings. *J. Neurodev. Disord.* **2012**, *4*, 19. <https://doi.org/10.1186/1866-1955-4-19>.
31. Banks, W.A.; Erickson, M.A. The Blood–Brain Barrier and Immune Function and Dysfunction. *Neurobiol. Dis.* **2010**, *37*, 26–32. <https://doi.org/10.1016/j.nbd.2009.07.031>.
32. Silver, J.; Schwab, M.E.; Popovich, P.G. Central Nervous System Regenerative Failure: Role of Oligodendrocytes, Astrocytes, and Microglia. *Cold Spring Harb. Perspect. Biol.* **2014**, *7*, a020602. <https://doi.org/10.1101/cshperspect.a020602>.
33. Kim, Y.-K.; Shin, C. The Microbiota-Gut-Brain Axis in Neuropsychiatric Disorders: Pathophysiological Mechanisms and Novel Treatments. *Curr. Neuropharmacol.* **2018**, *16*, 559–573. <https://doi.org/10.2174/1570159X15666170915141036>.
34. Hu, X.; Wang, T.; Jin, F. Alzheimer’s Disease and Gut Microbiota. *Sci. China Life Sci.* **2016**, *59*, 1006–1023. <https://doi.org/10.1007/s11427-016-5083-9>.
35. Mancuso, C.; Santangelo, R. Alzheimer’s Disease and Gut Microbiota Modifications: The Long Way between Preclinical Studies and Clinical Evidence. *Pharmacol. Res.* **2018**, *129*, 329–336. <https://doi.org/10.1016/j.phrs.2017.12.009>.

36. Cattaneo, A.; Cattane, N.; Galluzzi, S.; Provasi, S.; Lopizzo, N.; Festari, C.; Ferrari, C.; Guerra, U.P.; Paghera, B.; Muscio, C.; et al. Association of Brain Amyloidosis with Pro-Inflammatory Gut Bacterial Taxa and Peripheral Inflammation Markers in Cognitively Impaired Elderly. *Neurobiol. Aging* **2017**, *49*, 60–68. <https://doi.org/10.1016/j.neurobiolaging.2016.08.019>.
37. Kumar, D.K.V.; Choi, S.H.; Washicosky, K.J.; Eimer, W.A.; Tucker, S.; Ghofrani, J.; Lefkowitz, A.; McColl, G.; Goldstein, L.E.; Tanzi, R.E.; et al. Amyloid- $\beta$  Peptide Protects against Microbial Infection in Mouse and Worm Models of Alzheimer's Disease. *Sci. Transl. Med.* **2016**, *8*, 340ra72. <https://doi.org/10.1126/scitranslmed.aaf1059>.
38. Kesika, P.; Suganthy, N.; Sivamaruthi, B.S.; Chaiyasut, C. Role of Gut-Brain Axis, Gut Microbial Composition, and Probiotic Intervention in Alzheimer's Disease. *Life Sci.* **2020**, *264*, 118627. <https://doi.org/10.1016/j.lfs.2020.118627>.
39. Hauss-Wegrzyniak, B.; Vraniak, P.D.; Wenk, G.L. LPS-Induced Neuroinflammatory Effects Do Not Recover with Time. *NeuroReport* **2000**, *11*, 1759–1763. <https://doi.org/10.1097/00001756-200006050-00032>.
40. Kahn, M.S.; Kranjac, D.; Alonzo, C.A.; Haase, J.H.; Cedillos, R.O.; McLinden, K.A.; Boehm, G.W.; Chumley, M.J. Prolonged Elevation in Hippocampal A $\beta$  and Cognitive Deficits Following Repeated Endotoxin Exposure in the Mouse. *Behav. Brain Res.* **2012**, *229*, 176–184. <https://doi.org/10.1016/j.bbr.2012.01.010>.
41. Asti, A.; Gioglio, L. Can a Bacterial Endotoxin Be a Key Factor in the Kinetics of Amyloid Fibril Formation? *J. Alzheimers Dis.* **2014**, *39*, 169–179. <https://doi.org/10.3233/JAD-131394>.
42. Lucas, K.; Maes, M. Role of the Toll Like Receptor (TLR) Radical Cycle in Chronic Inflammation: Possible Treatments Targeting the TLR4 Pathway. *Mol. Neurobiol.* **2013**, *48*, 190–204. <https://doi.org/10.1007/s12035-013-8425-7>.
43. Mohammad, S.; Thiemermann, C. Role of Metabolic Endotoxemia in Systemic Inflammation and Potential Interventions. *Front. Immunol.* **2021**, *11*, 3379. <https://doi.org/10.3389/fimmu.2020.594150>.
44. Maes, M.; Twisk, F.N.M.; Kubera, M.; Ringel, K.; Leunis, J.-C.; Geffard, M. Increased IgA Responses to the LPS of Commensal Bacteria Is Associated with Inflammation and Activation of Cell-Mediated Immunity in Chronic Fatigue Syndrome. *J. Affect. Disord.* **2012**, *136*, 909–917. <https://doi.org/10.1016/j.jad.2011.09.010>.
45. Maes, M.; Mihaylova, I.; Leunis, J.-C. Increased Serum IgA and IgM against LPS of Enterobacteria in Chronic Fatigue Syndrome (CFS): Indication for the Involvement of Gram-Negative Enterobacteria in the Etiology of CFS and for the Presence of an Increased Gut–Intestinal Permeability. *J. Affect. Disord.* **2007**, *99*, 237–240. <https://doi.org/10.1016/j.jad.2006.08.021>.
46. Frost, B.; Diamond, M.I. Prion-like Mechanisms in Neurodegenerative Diseases. *Nat. Rev. Neurosci.* **2010**, *11*, 155–159. <https://doi.org/10.1038/nrn2786>.
47. Emery, D.C.; Shoemark, D.K.; Batstone, T.E.; Waterfall, C.M.; Coghill, J.A.; Cerajewska, T.L.; Davies, M.; West, N.X.; Allen, S.J. 16S rRNA Next Generation Sequencing Analysis Shows Bacteria in Alzheimer's Post-Mortem Brain. *Front. Aging Neurosci.* **2017**, *9*, 195. <https://doi.org/10.3389/fnagi.2017.00195>.
48. Poole, S.; Singhrao, S.K.; Kesavalu, L.; Curtis, M.A.; Crean, S. Determining the Presence of Periodontopathic Virulence Factors in Short-Term Postmortem Alzheimer's Disease Brain Tissue. *J. Alzheimers Dis.* **2013**, *36*, 665–677. <https://doi.org/10.3233/JAD-121918>.
49. Zhao, Y.; Jaber, V.; Lukiw, W.J. Secretory Products of the Human GI Tract Microbiome and Their Potential Impact on Alzheimer's Disease (AD): Detection of Lipopolysaccharide (LPS) in AD Hippocampus. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 318. <https://doi.org/10.3389/fcimb.2017.00318>.
50. Hammond, C.J.; Hallock, L.R.; Howanski, R.J.; Appelt, D.M.; Little, C.S.; Balin, B.J. Immunohistological Detection of Chlamydia Pneumoniae in the Alzheimer's Disease Brain. *BMC Neurosci.* **2010**, *11*, 121. <https://doi.org/10.1186/1471-2202-11-121>.
51. Miklossy, J. Bacterial Amyloid and DNA Are Important Constituents of Senile Plaques: Further Evidence of the Spirochetal and Biofilm Nature of Senile Plaques. *J. Alzheimers Dis.* **2016**, *53*, 1459–1473. <https://doi.org/10.3233/JAD-160451>.
52. Carter, C. Alzheimer's Disease: APP, Gamma Secretase, APOE, CLU, CR1, PICALM, ABCA7, BIN1, CD2AP, CD33, EPHA1, and MS4A2, and Their Relationships with Herpes Simplex, C. Pneumoniae, Other Suspect Pathogens, and the Immune System. *Int. J. Alzheimers Dis.* **2011**, *2011*, 1–34. <https://doi.org/10.4061/2011/501862>.
53. Kountouras, J.; Boziki, M.; Gavalas, E.; Zavos, C.; Deretzi, G.; Grigoriadis, N.; Tsolaki, M.; Chatzopoulos, D.; Katsinelos, P.; Tzilves, D.; et al. Increased Cerebrospinal Fluid Helicobacter Pylori Antibody in Alzheimer's Disease. *Int. J. Neurosci.* **2009**, *119*, 765–777. <https://doi.org/10.1080/00207450902782083>.
54. Shabbir, U.; Arshad, M.S.; Sameen, A.; Oh, D.-H. Crosstalk between Gut and Brain in Alzheimer's Disease: The Role of Gut Microbiota Modulation Strategies. *Nutrients* **2021**, *13*, 690. <https://doi.org/10.3390/nu13020690>.
55. Goyal, D.; Ali, S.A.; Singh, R.K. Emerging Role of Gut Microbiota in Modulation of Neuroinflammation and Neurodegeneration with Emphasis on Alzheimer's Disease. *Prog. Neuro Psychopharmacol. Biol. Psychiatry* **2020**, *106*, 110112. <https://doi.org/10.1016/j.pnpbp.2020.110112>.
56. Zhu, F.; Li, C.; Chu, F.; Tian, X.; Zhu, J. Target Dysbiosis of Gut Microbes as a Future Therapeutic Manipulation in Alzheimer's Disease. *Front. Aging Neurosci.* **2020**, *12*, 544235. <https://doi.org/10.3389/fnagi.2020.544235>.
57. Weiss, G.A.; Hennes, T. Mechanisms and Consequences of Intestinal Dysbiosis. *Cell. Mol. Life Sci.* **2017**, *74*, 2959–2977. <https://doi.org/10.1007/s00018-017-2509-x>.
58. Rinninella, E.; Raoul, P.; Cintoni, M.; Franceschi, F.; Miggiano, G.; Gasbarrini, A.; Mele, M. What Is the Healthy Gut Microbiota Composition? A Changing Ecosystem across Age, Environment, Diet, and Diseases. *Microorganisms* **2019**, *7*, 14. <https://doi.org/10.3390/microorganisms7010014>.
59. Cenit, M.C.; Sanz, Y.; Codoñer-Franch, P. Influence of Gut Microbiota on Neuropsychiatric Disorders. *World J. Gastroenterol.* **2017**, *23*, 5486–5498. <https://doi.org/10.3748/wjg.v23.i30.5486>.



60. Gotkine, M.; Kviatcovsky, D.; Elinav, E. Amyotrophic Lateral Sclerosis and Intestinal Microbiota—toward Establishing Cause and Effect. *Gut Microbes* **2020**, *11*, 1833–1841. <https://doi.org/10.1080/19490976.2020.1767464>.
61. Sun, M.-F.; Shen, Y.-Q. Dysbiosis of Gut Microbiota and Microbial Metabolites in Parkinson’s Disease. *Ageing Res. Rev.* **2018**, *45*, 53–61. <https://doi.org/10.1016/j.arr.2018.04.004>.
62. Gerhardt, S.; Mohajeri, M. Changes of Colonic Bacterial Composition in Parkinson’s Disease and Other Neurodegenerative Diseases. *Nutrients* **2018**, *10*, 708. <https://doi.org/10.3390/nu10060708>.
63. Jiang, H.; Ling, Z.; Zhang, Y.; Mao, H.; Ma, Z.; Yin, Y.; Wang, W.; Tang, W.; Tan, Z.; Shi, J.; et al. Altered Fecal Microbiota Composition in Patients with Major Depressive Disorder. *Brain Behav. Immun.* **2015**, *48*, 186–194. <https://doi.org/10.1016/j.bbi.2015.03.016>.
64. Levy, M.; Kolodziejczyk, A.A.; Thaiss, C.A.; Elinav, E. Dysbiosis and the Immune System. *Nat. Rev. Immunol.* **2017**, *17*, 219–232. <https://doi.org/10.1038/nri.2017.7>.
65. Schwabe, R.F.; Jobin, C. The Microbiome and Cancer. *Nat. Cancer* **2013**, *13*, 800–812. <https://doi.org/10.1038/nrc3610>.
66. Abenavoli, L.; Scarpellini, E.; Colica, C.; Boccuto, L.; Salehi, B.; Sharifi-Rad, J.; Aiello, V.; Romano, B.; de Lorenzo, A.; Izzo, A.A.; et al. Gut Microbiota and Obesity: A Role for Probiotics. *Nutrients* **2019**, *11*, 2690. <https://doi.org/10.3390/nu11112690>.
67. Galicia-Garcia, U.; Benito-Vicente, A.; Jebari, S.; Larrea-Sebal, A.; Siddiqi, H.; Uribe, K.B.; Ostolaza, H.; Martín, C. Pathophysiology of Type 2 Diabetes Mellitus. *Int. J. Mol. Sci.* **2020**, *21*, 6275. <https://doi.org/10.3390/ijms21176275>.
68. Varesi, A.; Deumer, U.-S.; Ananth, S.; Ricevuti, G. The Emerging Role of Gut Microbiota in Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS): Current Evidence and Potential Therapeutic Applications. *J. Clin. Med.* **2021**, *10*, 5077. <https://doi.org/10.3390/jcm10215077>.
69. Ling, Z.; Liu, X.; Jia, X.; Cheng, Y.; Luo, Y.; Yuan, L.; Wang, Y.; Zhao, C.; Guo, S.; Li, L.; et al. Impacts of Infection with Different Toxigenic *Clostridium Difficile* Strains on Faecal Microbiota in Children. *Sci. Rep.* **2014**, *4*, 7485. <https://doi.org/10.1038/srep07485>.
70. Vogt, N.M.; Kerby, R.L.; Dill-McFarland, K.A.; Harding, S.J.; Merluzzi, A.P.; Johnson, S.C.; Carlsson, C.M.; Asthana, S.; Zetterberg, H.; Blennow, K.; et al. Gut Microbiome Alterations in Alzheimer’s Disease. *Sci. Rep.* **2017**, *7*, 1–11. <https://doi.org/10.1038/s41598-017-13601-y>.
71. Haran, J.P.; Bhattarai, S.K.; Foley, S.E.; Dutta, P.; Ward, D. v.; Bucci, V.; McCormick, B.A. Alzheimer’s Disease Microbiome Is Associated with Dysregulation of the Anti-Inflammatory P-Glycoprotein Pathway. *mBio* **2019**, *10*, e00632-19. <https://doi.org/10.1128/mBio.00632-19>.
72. Liu, P.; Wu, L.; Peng, G.; Han, Y.; Tang, R.; Ge, J.; Zhang, L.; Jia, L.; Yue, S.; Zhou, K.; et al. Altered Microbiomes Distinguish Alzheimer’s Disease from Amnesic Mild Cognitive Impairment and Health in a Chinese Cohort. *Brain Behav. Immun.* **2019**, *80*, 633–643. <https://doi.org/10.1016/j.bbi.2019.05.008>.
73. Ling, Z.; Zhu, M.; Yan, X.; Cheng, Y.; Shao, L.; Liu, X.; Jiang, R.; Wu, S. Structural and Functional Dysbiosis of Fecal Microbiota in Chinese Patients with Alzheimer’s Disease. *Front. Cell Dev. Biol.* **2021**, *8*, 1891. <https://doi.org/10.3389/fcell.2020.634069>.
74. Hoffman, J.D.; Parikh, I.; Green, S.J.; Chlipala, G.; Mohny, R.P.; Keaton, M.; Bauer, B.; Hartz, A.M.S.; Lin, A.-L. Age Drives Distortion of Brain Metabolic, Vascular and Cognitive Functions, and the Gut Microbiome. *Front. Aging Neurosci.* **2017**, *9*, 298. <https://doi.org/10.3389/fnagi.2017.00298>.
75. Boyajian, J.L.; Ghebretatios, M.; Schaly, S.; Islam, P.; Prakash, S. Microbiome and Human Aging: Probiotic and Prebiotic Potentials in Longevity, Skin Health and Cellular Senescence. *Nutrients* **2021**, *13*, 4550. <https://doi.org/10.3390/nu13124550>.
76. Zhuang, Z.-Q.; Shen, L.-L.; Li, W.-W.; Fu, X.; Zeng, F.; Gui, L.; Lü, Y.; Cai, M.; Zhu, C.; Tan, Y.-L.; et al. Gut Microbiota Is Altered in Patients with Alzheimer’s Disease. *J. Alzheimers Dis.* **2018**, *63*, 1337–1346. <https://doi.org/10.3233/JAD-180176>.
77. Guo, M.; Peng, J.; Huang, X.; Xiao, L.; Huang, F.; Zuo, Z. Gut Microbiome Features of Chinese Patients Newly Diagnosed with Alzheimer’s Disease or Mild Cognitive Impairment. *J. Alzheimers Dis.* **2021**, *80*, 299–310. <https://doi.org/10.3233/JAD-201040>.
78. He, Y.; Wu, W.; Zheng, H.-M.; Li, P.; McDonald, D.; Sheng, H.-F.; Chen, M.-X.; Chen, Z.-H.; Ji, G.-Y.; Zheng, Z.-D.-X.; et al. Regional Variation Limits Applications of Healthy Gut Microbiome Reference Ranges and Disease Models. *Nat. Med.* **2018**, *24*, 1532–1535. <https://doi.org/10.1038/s41591-018-0164-x>.
79. Leblhuber, F.; Steiner, K.; Geisler, S.; Fuchs, D.; Gostner, J.M. On the Possible Relevance of Bottom-up Pathways in the Pathogenesis of Alzheimer’s Disease. *Curr. Top. Med. Chem.* **2020**, *20*, 1415–1421. <https://doi.org/10.2174/1568026620666200514090359>.
80. Morris, G.; Berk, M.; Carvalho, A.; Caso, J.R.; Sanz, Y.; Walder, K.; Maes, M. The Role of the Microbial Metabolites Including Tryptophan Catabolites and Short Chain Fatty Acids in the Pathophysiology of Immune-Inflammatory and Neuroimmune Disease. *Mol. Neurobiol.* **2017**, *54*, 4432–4451. <https://doi.org/10.1007/s12035-016-0004-2>.
81. Lee, H.-J.; Lee, K.-E.; Kim, J.-K.; Kim, D.-H. Suppression of Gut Dysbiosis by *Bifidobacterium Longum* Alleviates Cognitive Decline in 5XFAD Transgenic and Aged Mice. *Sci. Rep.* **2019**, *9*, 1–12. <https://doi.org/10.1038/s41598-019-48342-7>.
82. Doifode, T.; Giridharan, V.V.; Generoso, J.S.; Bhatti, G.; Collodel, A.; Schulz, P.E.; Forlenza, O.V.; Barichello, T. The Impact of the Microbiota-Gut-Brain Axis on Alzheimer’s Disease Pathophysiology. *Pharmacol. Res.* **2020**, *164*, 105314. <https://doi.org/10.1016/j.phrs.2020.105314>.
83. Parada Venegas, D.; de la Fuente, M.K.; Landskron, G.; González, M.J.; Quera, R.; Dijkstra, G.; Harmsen, H.J.M.; Faber, K.N.; Hermoso, M.A. Short Chain Fatty Acids (SCFAs)-Mediated Gut Epithelial and Immune Regulation and Its Relevance for Inflammatory Bowel Diseases. *Front. Immunol.* **2019**, *10*, 277. <https://doi.org/10.3389/fimmu.2019.00277>.

84. Colombo, A.V.; Sadler, R.K.; Llovera, G.; Singh, V.; Roth, S.; Heindl, S.; Sebastian Monasor, L.; Verhoeven, A.; Peters, F.; Parhizkar, S.; et al. Microbiota-Derived Short Chain Fatty Acids Modulate Microglia and Promote A $\beta$  Plaque Deposition. *eLife* **2021**, *10*, e59826. <https://doi.org/10.7554/eLife.59826>.
85. Morrison, D.J.; Preston, T. Formation of Short Chain Fatty Acids by the Gut Microbiota and Their Impact on Human Metabolism. *Gut Microbes* **2016**, *7*, 189–200. <https://doi.org/10.1080/19490976.2015.1134082>.
86. Wenzel, T.J.; Gates, E.J.; Ranger, A.L.; Klegeris, A. Short-Chain Fatty Acids (SCFAs) Alone or in Combination Regulate Select Immune Functions of Microglia-like Cells. *Mol. Cell. Neurosci.* **2020**, *105*, 103493. <https://doi.org/10.1016/j.mcn.2020.103493>.
87. Köhler, C.; Maes, M.; Slyepchenko, A.; Berk, M.; Solmi, M.; Lanctôt, K.; Carvalho, A. The Gut-Brain Axis, Including the Microbiome, Leaky Gut and Bacterial Translocation: Mechanisms and Pathophysiological Role in Alzheimer's Disease. *Curr. Pharm. Des.* **2016**, *22*, 6152–6166. <https://doi.org/10.2174/1381612822666160907093807>.
88. Fuke, N.; Nagata, N.; Suganuma, H.; Ota, T. Regulation of Gut Microbiota and Metabolic Endotoxemia with Dietary Factors. *Nutrients* **2019**, *11*, 2277. <https://doi.org/10.3390/nu11102277>.
89. Browne, T.C.; McQuillan, K.; McManus, R.M.; O'Reilly, J.-A.; Mills, K.H.G.; Lynch, M.A. IFN- $\gamma$  Production by Amyloid  $\beta$ -Specific Th1 Cells Promotes Microglial Activation and Increases Plaque Burden in a Mouse Model of Alzheimer's Disease. *J. Immunol.* **2013**, *190*, 2241–2251. <https://doi.org/10.4049/jimmunol.1200947>.
90. Bonfili, L.; Cecarini, V.; Gogoi, O.; Gong, C.; Cuccioloni, M.; Angeletti, M.; Rossi, G.; Eleuteri, A.M. Microbiota Modulation as Preventative and Therapeutic Approach in Alzheimer's Disease. *FEBS J.* **2021**, *288*, 2836–2855. <https://doi.org/10.1111/febs.15571>.
91. Gamba, P.; Testa, G.; Sottero, B.; Gargiulo, S.; Poli, G.; Leonarduzzi, G. The Link between Altered Cholesterol Metabolism and Alzheimer's Disease. *Ann. N. Y. Acad. Sci.* **2012**, *1259*, 54–64. <https://doi.org/10.1111/j.1749-6632.2012.06513.x>.
92. Brandscheid, C.; Schuck, F.; Reinhardt, S.; Schäfer, K.-H.; Pietrzik, C.U.; Grimm, M.; Hartmann, T.; Schwartz, A.; Endres, K. Altered Gut Microbiome Composition and Tryptic Activity of the 5xFAD Alzheimer's Mouse Model. *J. Alzheimers Dis.* **2017**, *56*, 775–788. <https://doi.org/10.3233/JAD-160926>.
93. Gao, Q.; Wang, Y.; Wang, X.; Fu, S.; Zhang, X.; Wang, R.-T.; Zhang, X. Decreased Levels of Circulating Trimethylamine N-Oxide Alleviate Cognitive and Pathological Deterioration in Transgenic Mice: A Potential Therapeutic Approach for Alzheimer's Disease. *Aging* **2019**, *11*, 8642–8663. <https://doi.org/10.18632/aging.102352>.
94. Colciaghi, F.; Marcello, E.; Borroni, B.; Zimmermann, M.; Caltagirone, C.; Cattabeni, F.; Padovani, A.; di Luca, M. Platelet APP, ADAM 10 and BACE Alterations in the Early Stages of Alzheimer Disease. *Neurology* **2004**, *62*, 498–501. <https://doi.org/10.1212/01.WNL.0000106953.49802.9C>.
95. Evin, G. Platelets and Alzheimer's Disease: Potential of APP as a Biomarker. *World J. Psychiatry* **2012**, *2*, 102–113. <https://doi.org/10.5498/wjp.v2.i6.102>.
96. Szabo, C. Gaseotransmitters: New Frontiers for Translational Science. *Sci. Transl. Med.* **2010**, *2*, 59ps54. <https://doi.org/10.1126/scitranslmed.3000721>.
97. Oleskin, A.V.; Shenderov, B.A. Neuromodulatory Effects and Targets of the SCFAs and Gasotransmitters Produced by the Human Symbiotic Microbiota. *Microb. Ecol. Heal. Dis.* **2016**, *27*, 634. <https://doi.org/10.3402/mehd.v27.30971>.
98. Mietelska-Porowska, A.; Wojda, U. T Lymphocytes and Inflammatory Mediators in the Interplay between Brain and Blood in Alzheimer's Disease: Potential Pools of New Biomarkers. *J. Immunol. Res.* **2017**, *2017*, 1–17. <https://doi.org/10.1155/2017/4626540>.
99. Milà-Alomà, M.; Suárez-Calvet, M.; Molinuevo, J.L. Latest Advances in Cerebrospinal Fluid and Blood Biomarkers of Alzheimer's Disease. *Ther. Adv. Neurol. Disord.* **2019**, *12*, 1756286419888819. <https://doi.org/10.1177/1756286419888819>.
100. Mullane, K.; Williams, M. Alzheimer's Disease beyond Amyloid: Can the Repetitive Failures of Amyloid-Targeted Therapeutics Inform Future Approaches to Dementia Drug Discovery? *Biochem. Pharmacol.* **2020**, *177*, 113945. <https://doi.org/10.1016/j.bcp.2020.113945>.
101. O'Toole, P.W.; Jeffery, I.B. Gut Microbiota and Aging. *Science* **2015**, *350*, 1214–1215. <https://doi.org/10.1126/science.aac8469>.
102. Magne, F.; Gotteland, M.; Gauthier, L.; Zazueta, A.; Poeso, S.; Navarrete, P.; Balamurugan, R. The Firmicutes/Bacteroidetes Ratio: A Relevant Marker of Gut Dysbiosis in Obese Patients? *Nutrients* **2020**, *12*, 1474. <https://doi.org/10.3390/nu12051474>.
103. Lucidi, L.; Pettorusso, M.; Vellante, F.; di Carlo, F.; Ceci, F.; Santovito, M.C.; di Muzio, I.; Fornaro, M.; Ventriglio, A.; Tomasetti, C.; et al. Gut Microbiota and Bipolar Disorder: An Overview on a Novel Biomarker for Diagnosis and Treatment. *Int. J. Mol. Sci.* **2021**, *22*, 3723. <https://doi.org/10.3390/ijms22073723>.
104. Qian, Y.; Yang, X.; Xu, S.; Huang, P.; Li, B.; Du, J.; He, Y.; Su, B.; Xu, L.-M.; Wang, L.; et al. Gut Metagenomics-Derived Genes as Potential Biomarkers of Parkinson's Disease. *Brain* **2020**, *143*, 2474–2489. <https://doi.org/10.1093/brain/awaa201>.
105. Farrokhi, V.; Nemati, R.; Nichols, F.C.; Yao, X.; Anstadt, E.; Fujiwara, M.; Grady, J.; Wakefield, D.; Castro, W.; Donaldson, J.; et al. Bacterial Lipopeptide, Lipid 654, Is a Microbiome-Associated Biomarker for Multiple Sclerosis. *Clin. Transl. Immunol.* **2013**, *2*, e8. <https://doi.org/10.1038/cti.2013.11>.
106. Yan, Y.; Gao, Y.; Fang, Q.; Zhang, N.; Kumar, G.; Yan, H.; Song, L.; Li, J.; Zhang, Y.; Sun, J.; et al. Inhibition of Rho Kinase by Fasudil Ameliorates Cognition Impairment in APP/PS1 Transgenic Mice via Modulation of Gut Microbiota and Metabolites. *Front. Aging Neurosci.* **2021**, *13*, 649. <https://doi.org/10.3389/fnagi.2021.755164>.
107. Bello-Medina, P.C.; Hernández-Quiroz, F.; Pérez-Morales, M.; González-Franco, D.A.; Cruz-Pauseno, G.; García-Mena, J.; Díaz-Cintra, S.; Pacheco-López, G. Spatial Memory and Gut Microbiota Alterations Are Already Present in Early Adulthood in a Pre-Clinical Transgenic Model of Alzheimer's Disease. *Front. Neurosci.* **2021**, *15*, 595583. <https://doi.org/10.3389/fnins.2021.595583>.

108. Gu, X.; Zhou, J.; Zhou, Y.; Wang, H.; Si, N.; Ren, W.; Zhao, W.; Fan, X.; Gao, W.; Wei, X.; et al. Huanglian Jiedu Decoction Remodels the Periphery Microenvironment to Inhibit Alzheimer's Disease Progression Based on the "Brain-Gut" Axis through Multiple Integrated Omics. *Alzheimers Res. Ther.* **2021**, *13*, 1–18. <https://doi.org/10.1186/s13195-021-00779-7>.
109. Shen, L.; Liu, L.; Ji, H.-F. Alzheimer's Disease Histological and Behavioral Manifestations in Transgenic Mice Correlate with Specific Gut Microbiome State. *J. Alzheimers Dis.* **2017**, *56*, 385–390. <https://doi.org/10.3233/JAD-160884>.
110. Chen, Y.; Fang, L.; Chen, S.; Zhou, H.; Fan, Y.; Lin, L.; Li, J.; Xu, J.; Chen, Y.; Ma, Y.; et al. Gut Microbiome Alterations Precede Cerebral Amyloidosis and Microglial Pathology in a Mouse Model of Alzheimer's Disease. *BioMed Res. Int.* **2020**, *2020*, 1–15. <https://doi.org/10.1155/2020/8456596>.
111. Tan, F.H.P.; Liu, G.; Lau, S.-Y.A.; Jaafar, M.H.; Park, Y.-H.; Azzam, G.; Li, Y.; Liong, M.-T. Lactobacillus Probiotics Improved the Gut Microbiota Profile of a Drosophila Melanogaster Alzheimer's Disease Model and Alleviated Neurodegeneration in the Eye. *Benef. Microbes* **2020**, *11*, 79–89. <https://doi.org/10.3920/BM2019.0086>.
112. Zhang, X.; Wang, Y.; Liu, W.; Wang, T.; Wang, L.; Hao, L.; Ju, M.; Xiao, R. Diet Quality, Gut Microbiota, and MicroRNAs Associated with Mild Cognitive Impairment in Middle-Aged and Elderly Chinese Population. *Am. J. Clin. Nutr.* **2021**, *114*, 429–440. <https://doi.org/10.1093/ajcn/nqab078>.
113. Li, B.; He, Y.; Ma, J.; Huang, P.; Du, J.; Cao, L.; Wang, Y.; Xiao, Q.; Tang, H.; Chen, S. Mild Cognitive Impairment Has Similar Alterations as Alzheimer's Disease in Gut Microbiota. *Alzheimers Dement.* **2019**, *15*, 1357–1366. <https://doi.org/10.1016/j.jalz.2019.07.002>.
114. Vogt, N.M.; Romano, K.A.; Darst, B.F.; Engelman, C.D.; Johnson, S.C.; Carlsson, C.M.; Asthana, S.; Blennow, K.; Zetterberg, H.; Bendlin, B.B.; et al. The Gut Microbiota-Derived Metabolite Trimethylamine N-Oxide Is Elevated in Alzheimer's Disease. *Alzheimers Res. Ther.* **2018**, *10*, 1–8. <https://doi.org/10.1186/s13195-018-0451-2>.
115. Wu, L.; Han, Y.; Zheng, Z.; Peng, G.; Liu, P.; Yue, S.; Zhu, S.; Chen, J.; Lv, H.; Shao, L.; et al. Altered Gut Microbial Metabolites in Amnesic Mild Cognitive Impairment and Alzheimer's Disease: Signals in Host-Microbe Interplay. *Nutrients* **2021**, *13*, 228. <https://doi.org/10.3390/nu13010228>.
116. Goetz, M.E.; Hanfelt, J.J.; John, S.E.; Bergquist, S.H.; Loring, D.W.; Quyyumi, A.; Clifford, G.D.; Vaccarino, V.; Goldstein, F.; Johnson, T.M., II; et al. Rationale and Design of the Emory Healthy Aging and Emory Healthy Brain Studies. *Neuroepidemiology* **2019**, *53*, 187–200. <https://doi.org/10.1159/000501856>.
117. Forloni, G. Alzheimer's Disease: From Basic Science to Precision Medicine Approach. *BMJ Neurol. Open* **2020**, *2*, e000079. <https://doi.org/10.1136/bmjno-2020-000079>.
118. Bouftas, M. A Systematic Review on the Feasibility of Salivary Biomarkers for Alzheimer's Disease. *J. Prev. Alzheimers Dis.* **2020**, *8*, 84–91. <https://doi.org/10.14283/jpad.2020.57>.
119. Olsen, I.; Singhrao, S.K. Low Levels of Salivary Lactoferrin May Affect Oral Dysbiosis and Contribute to Alzheimer's Disease: A Hypothesis. *Med Hypotheses* **2020**, *146*, 110393. <https://doi.org/10.1016/j.mehy.2020.110393>.
120. Floden, A.M.; Sohrabi, M.; Nookala, S.; Cao, J.J.; Combs, C.K. Salivary A $\beta$  Secretion and Altered Oral Microbiome in Mouse Models of AD. *Curr. Alzheimer Res.* **2021**, *17*, 1133–1144. <https://doi.org/10.2174/1567205018666210119151952>.
121. Wu, Y.-F.; Lee, W.-F.; Salamanca, E.; Yao, W.-L.; Su, J.-N.; Wang, S.-Y.; Hu, C.-J.; Chang, W.-J. Oral Microbiota Changes in Elderly Patients, an Indicator of Alzheimer's Disease. *Int. J. Environ. Res. Public Heal.* **2021**, *18*, 4211. <https://doi.org/10.3390/ijerph18084211>.
122. Zhang, Z.; Tan, X.; Sun, X.; Wei, J.; Li, Q.X.; Wu, Z. Isoorientin Affects Markers of Alzheimer's Disease via Effects on the Oral and Gut Microbiota in APP/PS1 Mice. *J. Nutr.* **2022**, *152*, 140–152. <https://doi.org/10.1093/jn/nxab328>.
123. David, L.A.; Maurice, C.F.; Carmody, R.N.; Gootenberg, D.B.; Button, J.E.; Wolfe, B.E.; Ling, A. v.; Devlin, A.S.; Varma, Y.; Fischbach, M.A.; et al. Diet Rapidly and Reproducibly Alters the Human Gut Microbiome. *Nature* **2014**, *505*, 559–563. <https://doi.org/10.1038/nature12820>.
124. Davani-Davari, D.; Negahdaripour, M.; Karimzadeh, I.; Seifan, M.; Mohkam, M.; Masoumi, S.; Berenjian, A.; Ghasemi, Y. Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications. *Foods* **2019**, *8*, 92. <https://doi.org/10.3390/foods8030092>.
125. den Besten, G.; van Eunen, K.; Groen, A.K.; Venema, K.; Reijngoud, D.-J.; Bakker, B.M. The Role of Short-Chain Fatty Acids in the Interplay between Diet, Gut Microbiota, and Host Energy Metabolism. *J. Lipid Res.* **2013**, *54*, 2325–2340. <https://doi.org/10.1194/jlr.R036012>.
126. Rajanala, K.; Kumar, N.; Chamallamudi, M.R. Modulation of Gut-Brain Axis by Probiotics: A Promising Anti-Depressant Approach. *Curr. Neuropharmacol.* **2021**, *19*, 990–1006. <https://doi.org/10.2174/1570159X19666201215142520>.
127. Xu, M.; Mo, X.; Huang, H.; Chen, X.; Liu, H.; Peng, Z.; Chen, L.; Rong, S.; Yang, W.; Xu, S.; et al. Yeast  $\beta$ -Glucan Alleviates Cognitive Deficit by Regulating Gut Microbiota and Metabolites in A $\beta$ 1–42-Induced AD-like Mice. *Int. J. Biol. Macromol.* **2020**, *161*, 258–270. <https://doi.org/10.1016/j.ijbiomac.2020.05.180>.
128. Wang, Y.; Lim, Y.-Y.; He, Z.; Wong, W.-T.; Lai, W.-F. Dietary Phytochemicals That Influence Gut Microbiota: Roles and Actions as Anti-Alzheimer Agents. *Crit. Rev. Food Sci. Nutr.* **2021**, 1–27. <https://doi.org/10.1080/10408398.2021.1882381>.
129. Lee, Y.-S.; Lai, D.-M.; Huang, H.-J.; Lee-Chen, G.-J.; Chang, C.-H.; Hsieh-Li, H.M.; Lee, G.-C. Prebiotic Lactulose Ameliorates the Cognitive Deficit in Alzheimer's Disease Mouse Model through Macroautophagy and Chaperone-Mediated Autophagy Pathways. *J. Agric. Food Chem.* **2021**, *69*, 2422–2437. <https://doi.org/10.1021/acs.jafc.0c07327>.
130. Liu, Q.; Xi, Y.; Wang, Q.; Liu, J.; Li, P.; Meng, X.; Liu, K.; Chen, W.; Liu, X.; Liu, Z. Mannan Oligosaccharide Attenuates Cognitive and Behavioral Disorders in the 5xFAD Alzheimer's Disease Mouse Model via Regulating the Gut Microbiota-Brain Axis. *Brain Behav. Immun.* **2021**, *95*, 330–343. <https://doi.org/10.1016/j.bbi.2021.04.005>.

131. Nishikawa, M.; Brickman, A.M.; Manly, J.J.; Schupf, N.; Mayeux, R.P.; Gu, Y. Association of Dietary Prebiotic Consumption with Reduced Risk of Alzheimer's Disease in a Multiethnic Population. *Curr. Alzheimer Res.* **2021**, *18*, 984–992. <https://doi.org/10.2174/1567205019666211222115142>.
132. Deng, S.; Chen, C.; Lin, H.; Cheng, I.H. The Beneficial Effect of Synbiotics Consumption on Alzheimer's Disease Mouse Model via Reducing Local and Systemic Inflammation. *IUBMB Life* 2021, in press. <https://doi.org/10.1002/iub.2589>.
133. Barbosa, R.S.D.; Vieira-Coelho, M.A. Probiotics and Prebiotics: Focus on Psychiatric Disorders—A Systematic Review. *Nutr. Rev.* **2019**, *78*, 437–450. <https://doi.org/10.1093/nutrit/nuz080>.
134. Xin, Y.; Diling, C.; Jian, Y.; Ting, L.; Guoyan, H.; Hualun, L.; Xiaocui, T.; Guoxiao, L.; Ou, S.; Chaoqun, Z.; et al. Effects of Oligosaccharides from *Morinda Officinalis* on Gut Microbiota and Metabolome of APP/PS1 Transgenic Mice. *Front. Neurol.* **2018**, *9*, 412. <https://doi.org/10.3389/fneur.2018.00412>.
135. Chen, D.; Yang, X.; Yang, J.; Lai, G.; Yong, T.; Tang, X.; Shuai, O.; Zhou, G.; Xie, Y.; Wu, Q. Prebiotic Effect of Fructooligosaccharides from *Morinda Officinalis* on Alzheimer's Disease in Rodent Models by Targeting the Microbiota-Gut-Brain Axis. *Front. Aging Neurosci.* **2017**, *9*, 403. <https://doi.org/10.3389/fnagi.2017.00403>.
136. Chok, K.C.; Ng, K.Y.; Koh, R.Y.; Chye, S.M. Role of the Gut Microbiome in Alzheimer's Disease. *Rev. Neurosci.* **2021**, *32*, 767–789. <https://doi.org/10.1515/revneuro-2020-0122>.
137. Lilly, D.M.; Stillwell, R.H. Probiotics: Growth-Promoting Factors Produced by Microorganisms. *Science* **1965**, *147*, 747–748. <https://doi.org/10.1126/science.147.3659.747>.
138. Martinez, F.A.C.; Balciunas, E.M.; Converti, A.; Cotter, P.D.; de Souza Oliveira, R.P. Bacteriocin Production by *Bifidobacterium* Spp. A Review. *Biotechnol. Adv.* **2013**, *31*, 482–488. <https://doi.org/10.1016/j.biotechadv.2013.01.010>.
139. Collado, M.C.; Hernandez, M.; Sanz, Y. Production of Bacteriocin-Like Inhibitory Compounds by Human Fecal *Bifidobacterium* Strains. *J. Food Prot.* **2005**, *68*, 1034–1040. <https://doi.org/10.4315/0362-028X-68.5.1034>.
140. Collado, M.C.; Gueimonde, M.; Sanz, Y.; Salminen, S. Adhesion Properties and Competitive Pathogen Exclusion Ability of *Bifidobacteria* with Acquired Acid Resistance. *J. Food Prot.* **2006**, *69*, 1675–1679. <https://doi.org/10.4315/0362-028X-69.7.1675>.
141. Bajaj, B.K.; Claes, I.J.J.; Lebeer, S. Functional Mechanisms of Probiotics. *J. Microbiol. Biotechnol. Food Sci.* **2015**, *4*, 321–327. <https://doi.org/10.15414/jmbfs.2015.4.4.321-327>.
142. Liu, G.; Ren, L.; Song, Z.; Wang, C.; Sun, B. Purification and Characteristics of Bifidocin A, a Novel Bacteriocin Produced by *Bifidobacterium* Animals BB04 from Centenarians' Intestine. *Food Control* **2015**, *50*, 889–895. <https://doi.org/10.1016/j.foodcont.2014.10.049>.
143. Thomas, C.M.; Versalovic, J. Probiotics-Host Communication. *Gut Microbes* **2010**, *1*, 148–163. <https://doi.org/10.4161/gmic.1.3.11712>.
144. Vanderpool, C.; Yan, F.; Polk, B.D. Mechanisms of Probiotic Action: Implications for Therapeutic Applications in Inflammatory Bowel Diseases. *Inflamm. Bowel Dis.* **2008**, *14*, 1585–1596. <https://doi.org/10.1002/ibd.20525>.
145. Bermudez-Brito, M.; Plaza-Díaz, J.; Muñoz-Quezada, S.; Gómez-Llorente, C.; Gil, A. Probiotic Mechanisms of Action. *Ann. Nutr. Metab.* **2012**, *61*, 160–174. <https://doi.org/10.1159/000342079>.
146. van Baarlen, P.; Wells, J.M.; Kleerebezem, M. Regulation of Intestinal Homeostasis and Immunity with Probiotic *Lactobacilli*. *Trends Immunol.* **2013**, *34*, 208–215. <https://doi.org/10.1016/j.it.2013.01.005>.
147. la Rosa, F.; Clerici, M.; Ratto, D.; Occhinegro, A.; Licito, A.; Romeo, M.; Iorio, C.; Rossi, P. The Gut-Brain Axis in Alzheimer's Disease and Omega-3. A Critical Overview of Clinical Trials. *Nutrients* **2018**, *10*, 1267. <https://doi.org/10.3390/nu10091267>.
148. Dargahi, N.; Johnson, J.; Donkor, O.; Vasiljevic, T.; Apostolopoulos, V. Immunomodulatory Effects of Probiotics: Can They Be Used to Treat Allergies and Autoimmune Diseases? *Maturitas* **2019**, *119*, 25–38. <https://doi.org/10.1016/j.maturitas.2018.11.002>.
149. Sichetti, M.; de Marco, S.; Pagiotti, R.; Traina, G.; Pietrella, D. Anti-Inflammatory Effect of Multistrain Probiotic Formulation (*L. Rhamnosus*, *B. Lactis*, and *B. Longum*). *Nutrition* **2018**, *53*, 95–102. <https://doi.org/10.1016/j.nut.2018.02.005>.
150. Martinowich, K.; Lu, B. Interaction between BDNF and Serotonin: Role in Mood Disorders. *Neuropsychopharmacology* **2008**, *33*, 73–83. <https://doi.org/10.1038/sj.npp.1301571>.
151. Heldt, S.A.; Stanek, L.; Chhatwal, J.P.; Ressler, K.J. Hippocampus-Specific Deletion of BDNF in Adult Mice Impairs Spatial Memory and Extinction of Aversive Memories. *Mol. Psychiatry* **2007**, *12*, 656–670. <https://doi.org/10.1038/sj.mp.4001957>.
152. Lu, Y.; Christian, K.; Lu, B. BDNF: A Key Regulator for Protein Synthesis-Dependent LTP and Long-Term Memory? *Neurobiol. Learn. Mem.* **2008**, *89*, 312–323. <https://doi.org/10.1016/j.nlm.2007.08.018>.
153. Dinan, T.G.; Stanton, C.; Cryan, J.F. Psychobiotics: A Novel Class of Psychotropic. *Biol. Psychiatry* **2013**, *74*, 720–726. <https://doi.org/10.1016/j.biopsych.2013.05.001>.
154. Distrutti, E.; O'Reilly, J.-A.; McDonald, C.; Cipriani, S.; Renga, B.; Lynch, M.A.; Fiorucci, S. Modulation of Intestinal Microbiota by the Probiotic VSL#3 Resets Brain Gene Expression and Ameliorates the Age-Related Deficit in LTP. *PLoS ONE* **2014**, *9*, e106503. <https://doi.org/10.1371/journal.pone.0106503>.
155. Xie, Z.; Wei, M.; Morgan, T.E.; Fabrizio, P.; Han, D.; Finch, C.E.; Longo, V.D. Peroxynitrite Mediates Neurotoxicity of Amyloid  $\beta$ -Peptide<sub>1-42</sub>- and Lipopolysaccharide-Activated Microglia. *J. Neurosci.* **2002**, *22*, 3484–3492. <https://doi.org/10.1523/JNEUROSCI.22-09-03484.2002>.
156. van Dyke, K. The Possible Role of Peroxynitrite in Alzheimer's Disease: A Simple Hypothesis That Could Be Tested More Thoroughly. *Med. Hypotheses* **1997**, *48*, 375–380. [https://doi.org/10.1016/S0306-9877\(97\)90031-1](https://doi.org/10.1016/S0306-9877(97)90031-1).

157. Bonfili, L.; Cecarini, V.; Cuccioloni, M.; Angeletti, M.; Berardi, S.; Scarpona, S.; Rossi, G.; Eleuteri, A.M. SLAB51 Probiotic Formulation Activates SIRT1 Pathway Promoting Antioxidant and Neuroprotective Effects in an AD Mouse Model. *Mol. Neurobiol.* **2018**, *55*, 7987–8000. <https://doi.org/10.1007/s12035-018-0973-4>.
158. O'Hagan, C.; Li, J. v.; Marchesi, J.R.; Plummer, S.; Garaiova, I.; Good, M.A. Long-Term Multi-Species Lactobacillus and Bifidobacterium Dietary Supplement Enhances Memory and Changes Regional Brain Metabolites in Middle-Aged Rats. *Neurobiol. Learn. Mem.* **2017**, *144*, 36–47. <https://doi.org/10.1016/j.nlm.2017.05.015>.
159. Kobayashi, Y.; Sugahara, H.; Shimada, K.; Mitsuyama, E.; Kuhara, T.; Yasuoka, A.; Kondo, T.; Abe, K.; Xiao, J. Therapeutic Potential of Bifidobacterium Breve Strain A1 for Preventing Cognitive Impairment in Alzheimer's Disease. *Sci. Rep.* **2017**, *7*, 1–10. <https://doi.org/10.1038/s41598-017-13368-2>.
160. Athari Nik Azm, S.; Djazayeri, A.; Safa, M.; Azami, K.; Ahmadvand, B.; Sabbaghziarani, F.; Sharifzadeh, M.; Vafa, M. Lactobacilli and Bifidobacteria Ameliorate Memory and Learning Deficits and Oxidative Stress in  $\beta$ -Amyloid (1–42) Injected Rats. *Appl. Physiol. Nutr. Metab.* **2018**, *43*, 718–726. <https://doi.org/10.1139/apnm-2017-0648>.
161. Akbari, E.; Asemi, Z.; Daneshvar Kakhaki, R.; Bahmani, F.; Kouchaki, E.; Tamtaji, O.R.; Hamidi, G.A.; Salami, M. Effect of Probiotic Supplementation on Cognitive Function and Metabolic Status in Alzheimer's Disease: A Randomized, Double-Blind and Controlled Trial. *Front. Aging Neurosci.* **2016**, *8*, 256. <https://doi.org/10.3389/fnagi.2016.00256>.
162. Den, H.; Dong, X.; Chen, M.; Zou, Z. Efficacy of Probiotics on Cognition, and Biomarkers of Inflammation and Oxidative Stress in Adults with Alzheimer's Disease or Mild Cognitive Impairment—A Meta-Analysis of Randomized Controlled Trials. *Aging* **2020**, *12*, 4010–4039. <https://doi.org/10.18632/aging.102810>.
163. Agahi, A.; Hamidi, G.A.; Daneshvar, R.; Hamdieh, M.; Soheili, M.; Alinaghypour, A.; Esmaeili Taba, S.M.; Salami, M. Does Severity of Alzheimer's Disease Contribute to Its Responsiveness to Modifying Gut Microbiota? A Double Blind Clinical Trial. *Front. Neurol.* **2018**, *9*, 662. <https://doi.org/10.3389/fneur.2018.00662>.
164. Klingbeil, E.; de La Serre, C.B. Microbiota Modulation by Eating Patterns and Diet Composition: Impact on Food Intake. *Am. J. Physiol. Integr. Comp. Physiol.* **2018**, *315*, R1254–R1260. <https://doi.org/10.1152/ajpregu.00037.2018>.
165. Merra, G.; Noce, A.; Marrone, G.; Cintoni, M.; Tarsitano, M.G.; Capacci, A.; de Lorenzo, A. Influence of Mediterranean Diet on Human Gut Microbiota. *Nutrients* **2020**, *13*, 7. <https://doi.org/10.3390/nu13010007>.
166. Duplantier, S.C.; Gardner, C.D. A Critical Review of the Study of Neuroprotective Diets to Reduce Cognitive Decline. *Nutrients* **2021**, *13*, 2264. <https://doi.org/10.3390/nu13072264>.
167. Bartochowski, Z.; Conway, J.; Wallach, Y.; Chakkamparambil, B.; Alakkassery, S.; Grossberg, G.T. Dietary Interventions to Prevent or Delay Alzheimer's Disease: What the Evidence Shows. *Curr. Nutr. Rep.* **2020**, *9*, 210–225. <https://doi.org/10.1007/s13668-020-00333-1>.
168. Gutierrez, L.; Folch, A.; Rojas, M.; Cantero, J.L.; Atienza, M.; Folch, J.; Camins, A.; Ruiz, A.; Papandreou, C.; Bulló, M. Effects of Nutrition on Cognitive Function in Adults with or without Cognitive Impairment: A Systematic Review of Randomized Controlled Clinical Trials. *Nutrients* **2021**, *13*, 3728. <https://doi.org/10.3390/nu13113728>.
169. Limongi, F.; Siviero, P.; Bozanic, A.; Noale, M.; Veronese, N.; Maggi, S. The Effect of Adherence to the Mediterranean Diet on Late-Life Cognitive Disorders: A Systematic Review. *J. Am. Med. Dir. Assoc.* **2020**, *21*, 1402–1409. <https://doi.org/10.1016/j.jamda.2020.08.020>.
170. Kheirouri, S.; Alizadeh, M. MIND Diet and Cognitive Performance in Older Adults: A Systematic Review. *Crit. Rev. Food Sci. Nutr.* **2021**, 1–19. <https://doi.org/10.1080/10408398.2021.1925220>.
171. Lilamand, M.; Mouton-Liger, F.; Paquet, C. Ketogenic Diet Therapy in Alzheimer's Disease: An Updated Review. *Curr. Opin. Clin. Nutr. Metab. Care* **2021**, *24*, 372–378. <https://doi.org/10.1097/MCO.0000000000000759>.
172. Grammatikopoulou, M.G.; Goulis, D.G.; Gkiouras, K.; Theodoridis, X.; Gkouskou, K.K.; Evangelidou, A.; Dardiotis, E.; Bogdanos, D.P. To Keto or Not to Keto? A Systematic Review of Randomized Controlled Trials Assessing the Effects of Ketogenic Therapy on Alzheimer Disease. *Adv. Nutr. Int. Rev. J.* **2020**, *11*, 1583–1602. <https://doi.org/10.1093/advances/nmaa073>.
173. Pavón, S.; Lázaro, E.; Martínez, O.; Amayra, I.; López-Paz, J.F.; Caballero, P.; Al-Rashaida, M.; Luna, P.M.; García, M.; Pérez, M.; et al. Ketogenic Diet and Cognition in Neurological Diseases: A Systematic Review. *Nutr. Rev.* **2020**, *79*, 802–813. <https://doi.org/10.1093/nutrit/nuaa113>.
174. Jensen, N.J.; Wodschow, H.Z.; Nilsson, M.; Rungby, J. Effects of Ketone Bodies on Brain Metabolism and Function in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 8767. <https://doi.org/10.3390/ijms21228767>.
175. Christensen, M.G.; Damsgaard, J.; Fink-Jensen, A. Use of Ketogenic Diets in the Treatment of Central Nervous System Diseases: A Systematic Review. *Nord. J. Psychiatry* **2020**, *75*, 1–8. <https://doi.org/10.1080/08039488.2020.1795924>.
176. Moreira, S.C.; Jansen, A.K.; Silva, F.M. Dietary Interventions and Cognition of Alzheimer's Disease Patients: A Systematic Review of Randomized Controlled Trial. *Dement. Neuropsychol.* **2020**, *14*, 258–282. <https://doi.org/10.1590/1980-57642020dn14-030008>.
177. Zhang, H.; Hardie, L.; Bawajeeh, A.O.; Cade, J. Meat Consumption, Cognitive Function and Disorders: A Systematic Review with Narrative Synthesis and Meta-Analysis. *Nutrients* **2020**, *12*, 1528. <https://doi.org/10.3390/nu12051528>.
178. Dimache, A.M.; Şalaru, D.L.; Sascău, R.; Stătescu, C. The Role of High Triglycerides Level in Predicting Cognitive Impairment: A Review of Current Evidence. *Nutrients* **2021**, *13*, 2118. <https://doi.org/10.3390/nu13062118>.
179. Gkotszamanis, V.; Panagiotakos, D. Dietary Interventions and Cognition: A Systematic Review of Clinical Trials. *Psychiatriki* **2020**, *31*, 248–256. <https://doi.org/10.22365/jpsych.2020.313.248>.

180. el Gaamouch, F.; Liu, K.; Lin, H.; Wu, C.; Wang, J. Development of Grape Polyphenols as Multi-Targeting Strategies for Alzheimer's Disease. *Neurochem. Int.* **2021**, *147*, 105046. <https://doi.org/10.1016/j.neuint.2021.105046>.
181. Colizzi, C. The Protective Effects of Polyphenols on Alzheimer's Disease: A Systematic Review. *Alzheimers Dementia Transl. Res. Clin. Interv.* **2018**, *5*, 184–196. <https://doi.org/10.1016/j.trci.2018.09.002>.
182. Mielech, A.; Puścion-Jakubik, A.; Markiewicz-Żukowska, R.; Socha, K. Vitamins in Alzheimer's Disease—Review of the Latest Reports. *Nutrients* **2020**, *12*, 3458. <https://doi.org/10.3390/nu12113458>.
183. Szczechowiak, K.; Diniz, B.S.; Leszek, J. Diet and Alzheimer's Dementia—Nutritional Approach to Modulate Inflammation. *Pharmacol. Biochem. Behav.* **2019**, *184*, 172743. <https://doi.org/10.1016/j.pbb.2019.172743>.
184. Kosti, R.I.; Kasdagli, M.I.; Kyrozis, A.; Orsini, N.; Lagiou, P.; Taiganidou, F.; Naska, A. Fish Intake, n-3 Fatty Acid Body Status, and Risk of Cognitive Decline: A Systematic Review and a Dose–Response Meta-Analysis of Observational and Experimental Studies. *Nut. Rev.* **2021**, nuab078. <https://doi.org/10.1093/nutrit/nuab078>.
185. Haider, S.; Schwarzingler, A.; Stefanac, S.; Soysal, P.; Smith, L.; Veronese, N.; Dorner, T.E.; Grabovac, I. Nutritional Supplements for Neuropsychiatric Symptoms in People with Dementia: A Systematic Review and Meta-analysis. *Int. J. Geriatr. Psychiatry* **2020**, *35*, 1285–1291. <https://doi.org/10.1002/gps.5407>.
186. Arbo, B.D.; André-Miral, C.; Nasre-Nasser, R.G.; Schimith, L.E.; Santos, M.G.; Costa-Silva, D.; Muccillo-Baisch, A.L.; Hort, M.A. Resveratrol Derivatives as Potential Treatments for Alzheimer's and Parkinson's Disease. *Front. Aging Neurosci.* **2020**, *12*, 103. <https://doi.org/10.3389/fnagi.2020.00103>.
187. Sofi, F.; Macchi, C.; Abbate, R.; Gensini, G.F.; Casini, A. Mediterranean Diet and Health. *BioFactors* **2013**, *39*, 335–342. <https://doi.org/10.1002/biof.1096>.
188. Filippou, C.D.; Tsioufis, C.P.; Thomopoulos, C.G.; Mihas, C.C.; Dimitriadis, K.S.; Sotiropoulou, L.I.; Chrysochoou, C.A.; Nihoyannopoulos, P.I.; Tousoulis, D.M. Dietary Approaches to Stop Hypertension (DASH) Diet and Blood Pressure Reduction in Adults with and without Hypertension: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Adv. Nutr. Int. Rev. J.* **2020**, *11*, 1150–1160. <https://doi.org/10.1093/advances/nmaa041>.
189. Morris, M.C.; Tangney, C.C.; Wang, Y.; Sacks, F.M.; Bennett, D.A.; Aggarwal, N.T. MIND Diet Associated with Reduced Incidence of Alzheimer's Disease. *Alzheimers Dement.* **2015**, *11*, 1007–1014. <https://doi.org/10.1016/j.jalz.2014.11.009>.
190. Martínez-Lapiscina, E.H.; Clavero, P.; Toledo, E.; Estruch, R.; Salas-Salvadó, J.; San Julián, B.; Sanchez-Tainta, A.; Ros, E.; Valls-Pedret, C.; Martínez-Gonzalez, M.Á. Mediterranean Diet Improves Cognition: The PREDIMED-NAVARRA Randomised Trial. *J. Neurol. Neurosurg. Psychiatry* **2013**, *84*, 1318–1325. <https://doi.org/10.1136/jnnp-2012-304792>.
191. Valls-Pedret, C.; Sala-Vila, A.; Serra-Mir, M.; Corella, D.; de la Torre, R.; Martínez-González, M.Á.; Martínez-Lapiscina, E.H.; Fitó, M.; Pérez-Heras, A.; Salas-Salvadó, J.; et al. Mediterranean Diet and Age-Related Cognitive Decline. *JAMA Intern. Med.* **2015**, *175*, 1094–1103. <https://doi.org/10.1001/jamainternmed.2015.1668>.
192. Marseglia, A.; Xu, W.; Fratiglioni, L.; Fabbri, C.; Berendsen, A.A.M.; Bialecka-Debek, A.; Jennings, A.; Gillings, R.; Meunier, N.; Caumon, E.; et al. Effect of the NU-AGE Diet on Cognitive Functioning in Older Adults: A Randomized Controlled Trial. *Front. Physiol.* **2018**, *9*, 349. <https://doi.org/10.3389/fphys.2018.00349>.
193. Knight, A.; Bryan, J.; Wilson, C.; Hodgson, J.; Davis, C.; Murphy, K. The Mediterranean Diet and Cognitive Function among Healthy Older Adults in a 6-Month Randomised Controlled Trial: The MedLey Study. *Nutrients* **2016**, *8*, 579. <https://doi.org/10.3390/nu8090579>.
194. Liu, Y.-H.; Gao, X.; Na, M.; Kris-Etherton, P.M.; Mitchell, D.C.; Jensen, G.L. Dietary Pattern, Diet Quality, and Dementia: A Systematic Review and Meta-Analysis of Prospective Cohort Studies. *J. Alzheimers Dis.* **2020**, *78*, 151–158. <https://doi.org/10.3233/JAD-200499>.
195. Barbaresko, J.; Lellmann, A.W.; Schmidt, A.; Lehmann, A.; Amini, A.M.; Egert, S.; Schlesinger, S.; Nöthlings, U. Dietary Factors and Neurodegenerative Disorders: An Umbrella Review of Meta-Analyses of Prospective Studies. *Adv. Nutr. Int. Rev. J.* **2020**, *11*, 1161–1173. <https://doi.org/10.1093/advances/nmaa053>.
196. Prinelli, F.; Fratiglioni, L.; Kalpouzos, G.; Musicco, M.; Adorni, F.; Johansson, I.; Marseglia, A.; Xu, W. Specific Nutrient Patterns Are Associated with Higher Structural Brain Integrity in Dementia-Free Older Adults. *NeuroImage* **2019**, *199*, 281–288. <https://doi.org/10.1016/j.neuroimage.2019.05.066>.
197. Croll, P.H.; Voortman, T.; Ikram, M.A.; Franco, O.H.; Schoufour, J.D.; Bos, D.; Vernooij, M.W. Better Diet Quality Relates to Larger Brain Tissue Volumes. *Neurology* **2018**, *90*, e2166–e2173. <https://doi.org/10.1212/WNL.0000000000005691>.
198. Kao, Y.-C.; Ho, P.-C.; Tu, Y.-K.; Jou, I.-M.; Tsai, K.-J. Lipids and Alzheimer's Disease. *Int. J. Mol. Sci.* **2020**, *21*, 1505. <https://doi.org/10.3390/ijms21041505>.
199. Rainey-Smith, S.R.; Gu, Y.; Gardener, S.L.; Doecke, J.D.; Villemagne, V.L.; Brown, B.M.; Taddei, K.; Laws, S.M.; Sohrabi, H.R.; Weinborn, M.; et al. Mediterranean Diet Adherence and Rate of Cerebral A $\beta$ -Amyloid Accumulation: Data from the Australian Imaging, Biomarkers and Lifestyle Study of Ageing. *Transl. Psychiatry* **2018**, *8*, 1–7. <https://doi.org/10.1038/s41398-018-0293-5>.
200. Cole, G.M.; Frautschy, S.A. Docosahexaenoic Acid Protects from Amyloid and Dendritic Pathology in an Alzheimer's Disease Mouse Model. *Nutr. Heal.* **2006**, *18*, 249–259. <https://doi.org/10.1177/026010600601800307>.
201. Freund-Levi, Y.; Eriksdotter-Jönhagen, M.; Cederholm, T.; Basun, H.; Faxén-Ingvar, G.; Garlind, A.; Vedin, I.; Vessby, B.; Wahlund, L.-O.; Palmblad, J.  $\omega$ -3 Fatty Acid Treatment in 174 Patients with Mild to Moderate Alzheimer Disease: OmegaAD Study. *Arch. Neurol.* **2006**, *63*, 1402–1408. <https://doi.org/10.1001/archneur.63.10.1402>.

202. Ma, Q.-L.; Teter, B.; Ubeda, O.J.; Morihara, T.; Dhoot, D.; Nyby, M.D.; Tuck, M.L.; Frautschy, S.A.; Cole, G.M. Omega-3 Fatty Acid Docosahexaenoic Acid Increases SorLA/LR11, a Sorting Protein with Reduced Expression in Sporadic Alzheimer's Disease (AD): Relevance to AD Prevention. *J. Neurosci.* **2007**, *27*, 14299–14307. <https://doi.org/10.1523/JNEUROSCI.3593-07.2007>.
203. Wilson, D.M.; Binder, L.I. Free Fatty Acids Stimulate the Polymerization of Tau and Amyloid Beta Peptides. In Vitro Evidence for a Common Effector of Pathogenesis in Alzheimer's Disease. *Am. J. Pathol.* **1997**, *150*, 2181–2195.
204. Snowden, S.G.; Ebshiana, A.A.; Hye, A.; An, Y.; Pletnikova, O.; O'Brien, R.; Troncoso, J.; Legido-Quigley, C.; Thambisetty, M. Association between Fatty Acid Metabolism in the Brain and Alzheimer Disease Neuropathology and Cognitive Performance: A Nontargeted Metabolomic Study. *PLoS Med.* **2017**, *14*, e1002266. <https://doi.org/10.1371/journal.pmed.1002266>.
205. Takechi, R.; Galloway, S.; Paltridge-Gamarallage, M.M.; Lam, V.; Dhaliwal, S.S.; Mamo, J.C. Probiotic Prevents Blood-Brain Barrier Dysfunction in Wild-Type Mice Induced by Saturated Fat or Cholesterol Feeding. *Clin. Exp. Pharmacol. Physiol.* **2012**, *40*, 45–52. <https://doi.org/10.1111/1440-1681.12032>.
206. Bernath, M.M.; Bhattacharyya, S.; Nho, K.; Barupal, D.K.; Fiehn, O.; Baillie, R.; Risacher, S.L.; Arnold, M.; Jacobson, T.; Trojanowski, J.Q.; et al. Serum Triglycerides in Alzheimer Disease. *Neurology* **2020**, *94*, e2088–e2098. <https://doi.org/10.1212/WNL.0000000000009436>.
207. Yan, Y.; Yang, H.; Xie, Y.; Ding, Y.; Kong, D.; Yu, H. Research Progress on Alzheimer's Disease and Resveratrol. *Neurochem. Res.* **2020**, *45*, 989–1006. <https://doi.org/10.1007/s11064-020-03007-0>.
208. Rahman, Md.H.; Akter, R.; Bhattacharya, T.; Abdel-Daim, M.M.; Alkahtani, S.; Arafah, M.W.; Al-Johani, N.S.; Alhoshani, N.M.; Alkeraishan, N.; Alhenaky, A.; et al. Resveratrol and Neuroprotection: Impact and Its Therapeutic Potential in Alzheimer's Disease. *Front. Pharmacol.* **2020**, *11*, 2272. <https://doi.org/10.3389/fphar.2020.619024>.
209. Ułamek-Kozioł, M.; Czuczwar, S.J.; Januszewski, S.; Pluta, R. Ketogenic Diet and Epilepsy. *Nutrients* **2019**, *11*, 2510. <https://doi.org/10.3390/nu11102510>.
210. Włodarek, D. Role of Ketogenic Diets in Neurodegenerative Diseases (Alzheimer's Disease and Parkinson's Disease). *Nutrients* **2019**, *11*, 169. <https://doi.org/10.3390/nu11010169>.
211. Broom, G.M.; Shaw, I.C.; Rucklidge, J.J. The Ketogenic Diet as a Potential Treatment and Prevention Strategy for Alzheimer's Disease. *Nutrition* **2019**, *60*, 118–121. <https://doi.org/10.1016/j.nut.2018.10.003>.
212. Rusek, M.; Pluta, R.; Ułamek-Kozioł, M.; Czuczwar, S.J. Ketogenic Diet in Alzheimer's Disease. *Int. J. Mol. Sci.* **2019**, *20*, 3892. <https://doi.org/10.3390/ijms20163892>.
213. Masino, S.A.; Rho, J.M. Mechanisms of Ketogenic Diet Action. *Epilepsia* **2012**, *51*, 85.
214. Rojas-Morales, P.; Pedraza-Chaverri, J.; Tapia, E. Ketone Bodies, Stress Response, and Redox Homeostasis. *Redox Biol.* **2020**, *29*, 101395. <https://doi.org/10.1016/j.redox.2019.101395>.
215. Kashiwaya, Y.; Bergman, C.; Lee, J.-H.; Wan, R.; King, M.T.; Mughal, M.R.; Okun, E.; Clarke, K.; Mattson, M.P.; Veech, R.L. A Ketone Ester Diet Exhibits Anxiolytic and Cognition-Sparing Properties, and Lessens Amyloid and Tau Pathologies in a Mouse Model of Alzheimer's Disease. *Neurobiol. Aging* **2012**, *34*, 1530–1539. <https://doi.org/10.1016/j.neurobiolaging.2012.11.023>.
216. Yin, J.X.; Maalouf, M.; Han, P.; Zhao, M.; Gao, M.; Dharshaun, T.; Ryan, C.; Whitelegge, J.; Wu, J.; Eisenberg, D.; et al. Ketones Block Amyloid Entry and Improve Cognition in an Alzheimer's Model. *Neurobiol. Aging* **2015**, *39*, 25–37. <https://doi.org/10.1016/j.neurobiolaging.2015.11.018>.
217. Włodarek, D. The Possibility of Use of the Ketogenic Diet and Medium Chain Triglycerides Supplementation in the Support Therapy of Alzheimer Disease. *Curr. Opin. Clin. Nutr. Metab. Care* **2021**, *24*, 385–391. <https://doi.org/10.1097/MCO.0000000000000752>.
218. Taylor, M.K.; Sullivan, D.K.; Mahnken, J.D.; Burns, J.M.; Swerdlow, R.H. Feasibility and Efficacy Data from a Ketogenic Diet Intervention in Alzheimer's Disease. *Alzheimers Dementia: Transl. Res. Clin. Interv.* **2018**, *4*, 28–36. <https://doi.org/10.1016/j.trci.2017.11.002>.
219. Bostock, E.C.S.; Kirkby, K.C.; Taylor, B.V.M. The Current Status of the Ketogenic Diet in Psychiatry. *Front. Psychiatry* **2017**, *8*, 43. <https://doi.org/10.3389/fpsy.2017.00043>.
220. Garcia-Mantrana, I.; Selma-Royo, M.; Alcantara, C.; Collado, M.C. Shifts on Gut Microbiota Associated to Mediterranean Diet Adherence and Specific Dietary Intakes on General Adult Population. *Front. Microbiol.* **2018**, *9*, 890. <https://doi.org/10.3389/fmicb.2018.00890>.
221. Ghosh, T.S.; Rampelli, S.; Jeffery, I.B.; Santoro, A.; Neto, M.; Capri, M.; Giampieri, E.; Jennings, A.; Candela, M.; Turrone, S.; et al. Mediterranean Diet Intervention Alters the Gut Microbiome in Older People Reducing Frailty and Improving Health Status: The NU-AGE 1-Year Dietary Intervention across Five European Countries. *Gut* **2020**, *69*, 1218–1228. <https://doi.org/10.1136/gut-2019-319654>.
222. Moreno-Arribas, M.V.; Bartolomé, B.; Peñalvo, J.L.; Pérez-Matute, P.; Motilva, M.J. Relationship between Wine Consumption, Diet and Microbiome Modulation in Alzheimer's Disease. *Nutrients* **2020**, *12*, 3082. <https://doi.org/10.3390/nu12103082>.
223. Nash, V.; Ranadheera, C.S.; Georgousopoulou, E.N.; Mellor, D.D.; Panagiotakos, D.B.; McKune, A.J.; Kellett, J.; Naumovski, N. The Effects of Grape and Red Wine Polyphenols on Gut Microbiota—A Systematic Review. *Food Res. Int.* **2018**, *113*, 277–287. <https://doi.org/10.1016/j.foodres.2018.07.019>.
224. Ma, D.; Wang, A.C.; Parikh, I.; Green, S.J.; Hoffman, J.D.; Chlipala, G.; Murphy, M.P.; Sokola, B.S.; Bauer, B.; Hartz, A.M.S.; et al. Ketogenic Diet Enhances Neurovascular Function with Altered Gut Microbiome in Young Healthy Mice. *Sci. Rep.* **2018**, *8*, 1–10. <https://doi.org/10.1038/s41598-018-25190-5>.

225. Nagpal, R.; Neth, B.J.; Wang, S.; Craft, S.; Yadav, H. Modified Mediterranean-Ketogenic Diet Modulates Gut Microbiome and Short-Chain Fatty Acids in Association with Alzheimer's Disease Markers in Subjects with Mild Cognitive Impairment. *EBio-Medicine* **2019**, *47*, 529–542. <https://doi.org/10.1016/j.ebiom.2019.08.032>.
226. Park, S.; Zhang, T.; Wu, X.; Yi Qiu, J. Ketone Production by Ketogenic Diet and by Intermittent Fasting Has Different Effects on the Gut Microbiota and Disease Progression in an Alzheimer's Disease Rat Model. *J. Clin. Biochem. Nutr.* **2020**, *67*, 188–198. <https://doi.org/10.3164/jcfn.19-87>.
227. Gupta, S.; Allen-Vercoe, E.; Petrof, E.O. Fecal Microbiota Transplantation: In Perspective. *Ther. Adv. Gastroenterol.* **2015**, *9*, 229–239. <https://doi.org/10.1177/1756283X15607414>.
228. Surawicz, C.M.; Brandt, L.J.; Binion, D.G.; Ananthakrishnan, A.N.; Curry, S.R.; Gilligan, P.H.; McFarland, L. v.; Mellow, M.; Zuckerbraun, B.S. Guidelines for Diagnosis, Treatment, and Prevention of Clostridium Difficile Infections. *Am. J. Gastroenterol.* **2013**, *108*, 478–498. <https://doi.org/10.1038/ajg.2013.4>.
229. Hazan, S. Rapid Improvement in Alzheimer's Disease Symptoms Following Fecal Microbiota Transplantation: A Case Report. *J. Int. Med Res.* **2020**, *48*, 0300060520925930. <https://doi.org/10.1177/0300060520925930>.
230. Park, S.-H.; Lee, J.H.; Shin, J.; Kim, J.-S.; Cha, B.; Lee, S.; Kwon, K.S.; Shin, Y.W.; Choi, S.H. Cognitive Function Improvement after Fecal Microbiota Transplantation in Alzheimer's Dementia Patient: A Case Report. *Curr. Med Res. Opin.* **2021**, *37*, 1739–1744. <https://doi.org/10.1080/03007995.2021.1957807>.
231. Kim, N.; Jeon, S.H.; Ju, I.G.; Gee, M.S.; Do, J.; Oh, M.S.; Lee, J.K. Transplantation of Gut Microbiota Derived from Alzheimer's Disease Mouse Model Impairs Memory Function and Neurogenesis in C57BL/6 Mice. *Brain Behav. Immun.* **2021**, *98*, 357–365. <https://doi.org/10.1016/j.bbi.2021.09.002>.
232. Sun, J.; Xu, J.; Ling, Y.; Wang, F.; Gong, T.; Yang, C.; Ye, S.; Ye, K.; Wei, D.; Song, Z.; et al. Fecal Microbiota Transplantation Alleviated Alzheimer's Disease-like Pathogenesis in APP/PS1 Transgenic Mice. *Transl. Psychiatry* **2019**, *9*, 1–13. <https://doi.org/10.1038/s41398-019-0525-3>.
233. Wang, M.; Cao, J.; Gong, C.; Amakye, W.K.; Yao, M.; Ren, J. Exploring the Microbiota-Alzheimer's Disease Linkage Using Short-Term Antibiotic Treatment Followed by Fecal Microbiota Transplantation. *Brain Behav. Immun.* **2021**, *96*, 227–238. <https://doi.org/10.1016/j.bbi.2021.06.003>.
234. Harach, T.; Marungruang, N.; Duthilleul, N.; Cheatham, V.; Mc Coy, K.D.; Frisoni, G.; Neher, J.J.; Fåk, F.; Jucker, M.; Lasser, T.; et al. Reduction of Abeta Amyloid Pathology in APP/PS1 Transgenic Mice in the Absence of Gut Microbiota. *Sci. Rep.* **2017**, *7*, 41802. <https://doi.org/10.1038/srep41802>.
235. Fujii, Y.; Nguyen, T.T.T.; Fujimura, Y.; Kameya, N.; Nakamura, S.; Arakawa, K.; Morita, H. Fecal Metabolite of a Gnotobiotic Mouse Transplanted with Gut Microbiota from a Patient with Alzheimer's Disease. *Biosci. Biotechnol. Biochem.* **2019**, *83*, 2144–2152. <https://doi.org/10.1080/09168451.2019.1644149>.
236. Zhan, G.; Yang, N.; Li, S.; Huang, N.; Fang, X.; Zhang, J.; Zhu, B.; Yang, L.; Yang, C.; Luo, A. Abnormal Gut Microbiota Composition Contributes to Cognitive Dysfunction in SAMP8 Mice. *Aging* **2018**, *10*, 1257–1267. <https://doi.org/10.18632/aging.101464>.
237. Dodiya, H.B.; Kuntz, T.; Shaik, S.M.; Baufeld, C.; Leibowitz, J.; Zhang, X.; Gottle, N.; Zhang, X.; Butovsky, O.; Gilbert, J.A.; et al. Sex-Specific Effects of Microbiome Perturbations on Cerebral A $\beta$  Amyloidosis and Microglia Phenotypes. *J. Exp. Med.* **2019**, *216*, 1542–1560.
238. Cui, B.; Su, D.; Li, W.; She, X.; Zhang, M.; Wang, R.; Zhai, Q. Effects of Chronic Noise Exposure on the Microbiome-Gut-Brain Axis in Senescence-Accelerated Prone Mice: Implications for Alzheimer's Disease. *J. Neuroinflammation* **2018**, *15*, 190. <https://doi.org/10.1186/s12974-018-1223-4>.
239. Valeri, F.; dos Santos Guilherme, M.; He, F.; Stoye, N.M.; Schwiertz, A.; Endres, K. Impact of the Age of Cecal Material Transfer Donors on Alzheimer's Disease Pathology in 5xFAD Mice. *Microorganisms* **2021**, *9*, 2548. <https://doi.org/10.3390/microorganisms9122548>.
240. Shanahan, F.; Quigley, E.M.M. Manipulation of the Microbiota for Treatment of IBS and IBD—Challenges and Controversies. *Gastroenterology* **2014**, *146*, 1554–1563. <https://doi.org/10.1053/j.gastro.2014.01.050>.
241. Schmulson, M.; Bashashati, M. Fecal Microbiota Transfer for Bowel Disorders: Efficacy or Hype? *Curr. Opin. Pharmacol.* **2018**, *43*, 72–80. <https://doi.org/10.1016/j.coph.2018.08.012>.
242. Evrensel, A.; Ceylan, M.E. Fecal Microbiota Transplantation and Its Usage in Neuropsychiatric Disorders. *Clin. Psychopharmacol. Neurosci.* **2016**, *14*, 231–237. <https://doi.org/10.9758/cpn.2016.14.3.231>.
243. Choi, H.H.; Cho, Y.-S. Fecal Microbiota Transplantation: Current Applications, Effectiveness, and Future Perspectives. *Clin. Endosc.* **2016**, *49*, 257–265. <https://doi.org/10.5946/ce.2015.117>.
244. Tan, P.; Li, X.; Shen, J.; Feng, Q. Fecal Microbiota Transplantation for the Treatment of Inflammatory Bowel Disease: An Update. *Front. Pharmacol.* **2020**, *11*, 1409. <https://doi.org/10.3389/fphar.2020.574533>.