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## Leukocyte Integrins: Role in Leukocyte Recruitment and as Therapeutic Targets in Inflammatory Disease

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# Leukocyte Integrins: Role in Leukocyte Recruitment and as Therapeutic Targets in Inflammatory Disease

## Abstract

Infection or sterile inflammation triggers site-specific attraction of leukocytes. Leukocyte recruitment is a process comprising several steps orchestrated by adhesion molecules, chemokines, cytokines and endogenous regulatory molecules. Distinct adhesive interactions between endothelial cells and leukocytes and signalling mechanisms contribute to the temporal and spatial fine-tuning of the leukocyte adhesion cascade. Central players in the leukocyte adhesion cascade include the leukocyte adhesion receptors of the  $\beta$ 2-integrin family, such as the  $\alpha$ L $\beta$ 2 and  $\alpha$ M $\beta$ 2 integrins, or of the  $\beta$ 1-integrin family, such as the  $\alpha$ 4 $\beta$ 1- integrin. Given the central involvement of leukocyte recruitment in different inflammatory and autoimmune diseases, the leukocyte adhesion cascade in general, and leukocyte integrins in particular, represent key therapeutic targets. In this context, the present review focuses on the role of leukocyte integrins in the leukocyte adhesion cascade. Experimental evidence that has implicated leukocyte integrins as targets in animal models of inflammatory disorders, such as experimental autoimmune encephalomyelitis, psoriasis, inflammatory bone loss and inflammatory bowel disease as well as preclinical and clinical therapeutic applications of antibodies that target leukocyte integrins in various inflammatory disorders are presented. Finally, we review recent findings on endogenous inhibitors that modify leukocyte integrin function, which could emerge as promising therapeutic targets.

## Keywords

integrin, leukocyte adhesion, natalizumab, efalizumab, vedolizumab, Del-1

## Disciplines

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## Leukocyte integrins: Role in leukocyte recruitment and as therapeutic targets in inflammatory disease

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### Abstract

Infection or sterile inflammation triggers site-specific attraction of leukocytes. Leukocyte recruitment is a process comprising several steps orchestrated by adhesion molecules, chemokines, cytokines and endogenous regulatory molecules. Distinct adhesive interactions between endothelial cells and leukocytes and signalling mechanisms contribute to the temporal and spatial fine-tuning of the leukocyte adhesion cascade. Central players in the leukocyte adhesion cascade include the leukocyte adhesion receptors of the  $\beta 2$ -integrin family, such as the  $\alpha L\beta 2$  and  $\alpha M\beta 2$  integrins, or of the  $\beta 1$ -integrin family, such as the  $\alpha 4\beta 1$ - integrin. Given the central involvement of leukocyte recruitment in different inflammatory and autoimmune diseases, the leukocyte adhesion cascade in general, and leukocyte integrins in particular, represent key therapeutic targets. In this context, the present review focuses on the role of leukocyte integrins in the leukocyte adhesion cascade. Experimental evidence that has implicated leukocyte integrins as targets in animal models of inflammatory disorders, such as experimental autoimmune encephalomyelitis, psoriasis, inflammatory bone loss and inflammatory bowel disease as well as preclinical and clinical therapeutic applications of antibodies that target leukocyte integrins in various inflammatory disorders are presented. Finally, we review recent findings on endogenous inhibitors that modify leukocyte integrin function, which could emerge as promising therapeutic targets.

### Keywords

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## Introduction

Leukocyte trafficking to sites of injury or infection is tightly regulated by the leukocyte adhesion cascade. The cascade starts with selectin-dependent leukocyte rolling, followed by chemokine-induced leukocyte activation and leukocyte slow rolling, mediated by the collaboration between selectin and integrins and the interactions thereof with their respective ligands. Slow rolling paves the way for the integrin-dependent steps comprising firm leukocyte adhesion/arrest, adhesion strengthening and leukocyte crawling on the endothelium, that finally enable the transendothelial migration of leukocytes mostly through endothelial junctions to the subendothelium (Figure 1) (Ley et al., 2007; Chavakis, 2012). The present review will focus particularly on the function of integrins in the leukocyte adhesion cascade, endogenous factors and molecular mechanisms contributing to the regulation of integrin activity, animal models, in which the role of integrins in the context of leukocyte recruitment has been studied, as well as therapeutic approaches targeting leukocyte integrins in inflammatory diseases.

## Integrins; an overview

Integrins are a family of adhesion molecules that exist as transmembrane heterodimers, consisting of one  $\alpha$ - and one  $\beta$ -subunit (Hogg et al., 2011; Moser et al., 2009). In mammals, eighteen  $\alpha$  and eight  $\beta$  subunits have been described, forming 24 different integrin heterodimers (Herter and Zarbock, 2013; Hynes, 2002).

Activated integrins participate in mediating cell adhesion; the strength of integrin-mediated cell adhesion is defined as integrin avidity (Carman and Springer, 2003). Both integrin affinity and integrin valency contribute to integrin avidity (Carman and Springer, 2003). Affinity represents the strength of the individual bond between a single integrin and its ligand and is regulated by the conformational status of integrin subunits, whereas integrin valency is mediated by the clustering of integrin receptors on the cell surface, thereby bringing together several individual bonds (Carman and Springer, 2003). In the absence of activating signals, integrins have an inactive, bent conformation (Askari et al., 2009). Upon activation by different signals, e.g., deriving from chemokines, or PSGL-1 ligation, integrins undergo major conformational changes, which results in exposure of their ligand-binding site in the extracellular domains (Arnaout et al., 2005; Hogg et al., 2011, Moser et al., 2009; Tiwari et al., 2011; Wegener et al., 2007). Another feature of the process of integrin activation is its strong dependency on the presence of divalent cations, especially  $Mn^{2+}$  and  $Mg^{2+}$  ions, which control the conformational changes of the integrin molecules (Gailit and Ruoslahti, 1988; reviewed in Arnaout et al., 2002; Dransfield et al., 1992; Tiwari et al., 2011).

In the present review, we will focus on leukocyte integrins, in particular,  $\beta 2$ -integrins as well as integrin  $\alpha 4\beta 1$  (very late antigen (VLA)-4; CD49d/CD29) and integrin  $\alpha 4\beta 7$  (LPAM-1), which play a role in leukocyte recruitment and in inflammatory disorders. The  $\beta 2$ -integrins consist of a common  $\beta$  subunit (CD18) that associates with 4 different  $\alpha$  subunits. They comprise the  $\alpha L\beta 2$ -integrin (lymphocyte function-associated antigen 1 [LFA-1]; CD11a/CD18) and the  $\alpha M\beta 2$ -integrin (macrophage-1 antigen [Mac-1] also designated complement

receptor 3 [CR3]; CD11b/CD18), which are the most crucial  $\beta$ 2-integrins for leukocyte recruitment, as well as the  $\alpha$ X $\beta$ 2 (CD11c/CD18; p150,95; CR4) and the  $\alpha$ D $\beta$ 2 (CD11d/CD18) integrins (Chavakis, 2012; Moser et al., 2009). LFA-1 is expressed by neutrophils, monocytes and lymphocytes, whereas Mac-1 is found mainly on neutrophils and monocytes and VLA-4 is expressed on monocytes and T lymphocytes (Chan et al., 2001; Hyun et al., 2009; Chavakis, 2012; Moser et al., 2009).  $\alpha$ X $\beta$ 2 is present on macrophages and dendritic cells (DCs) (O'Doherty et al., 1994; Wu et al., 2009), and  $\alpha$ D $\beta$ 2 is expressed on monocytes/macrophages, especially foam cells, which are macrophages found in atherosclerotic lesions (Van der Vieren et al., 1995).

## The leukocyte adhesion cascade

The leukocyte adhesion cascade is initiated with leukocyte capture from the blood stream and rolling on the luminal surface of endothelial cells (Kunkel et al., 1998). Rolling serves as a break for circulating leukocytes and is mediated by E-, P- and L-selectins (Kansas, 1996, Kuwano et al., 2010). E-selectin and P-selectin are expressed in endothelial cells. L-selectin is constitutively expressed on most leukocytes (McEver, 2002) and plays a significant role in the migration of naïve and central memory T cells to lymph nodes (von Andrian and Mempel, 2003). Upon inflammation, the abundance of P- and E-selectin on the luminal endothelial cell surface is enhanced. Elevated surface expression of endothelial selectins is mediated either by exocytosis mechanisms in the case of P-selectin, which is released from cytosolic vesicles (designated Weibel-Palade bodies) or by enhanced transcriptional gene expression of E-selectin (McEver, 2002). On the other hand, leukocyte activation results in rapid shedding of L-selectin from their surface (Kishimoto et al., 1989), which contributes to the modulation of leukocyte rolling velocity (Hafezi-Moghadam and Ley, 1999).

Selectins interact with the P-selectin glycoprotein ligand-1 (PSGL-1) (McEver and Cummings, 1997) as well as other glycosylated ligands, such as CD44 and E-selectin ligand 1 (Hidalgo et al., 2007, reviewed in Zarbock et al., 2011). Leukocyte rolling under flow conditions is mediated by the rapid formation and dissociation of bonds between selectins and their ligands (Alon et al., 1995). These transient selectin-dependent adhesive interactions result in a progressive reduction in the rolling velocity of leukocytes, enabling cells to receive signals deriving from chemokines present on the endothelial cell surface; these signals activate integrins, thus allowing subsequent steps of the cascade to take over (Zarbock et al., 2011).

The interaction between selectins and their ligands leads to intracellular signaling cascades, which cooperate with chemokine-derived signals, resulting in the activation of integrins and thus firm leukocyte adhesion (Zarbock et al., 2007a; Yago et al., 2010; Zarbock et al., 2011). For instance, signalling pathways triggered by E-selectin are mediated by the Src family kinases Fgr, Hck and Lyn (Yago et al., 2010; Zarbock et al., 2008) also involving the adaptor proteins DAP12 and FcR $\gamma$  and the spleen tyrosine kinase (Zarbock et al., 2008; Urzainqui et al., 2002). This pathway results in activation of phospholipase C- $\gamma$ 2 (Mueller et al., 2010; Yago et al., 2010; Zarbock et al., 2011), culminating in the activation of the small GTPase RAS-related protein 1 (Rap1) (Stadtman et al., 2011), which can promote integrin

affinity (Chavakis, 2012). Thus, intracellular signalling induced by PSGL-1 and other selectin ligands is considered a major driver of the leukocyte adhesion cascade as it transforms a rolling/selectin-induced signal into activation of leukocyte integrins, thereby initiating the later steps of the cascade.

Upon activation, integrins mediate firm leukocyte adhesion and crawling. The  $\alpha$ 4-integrins VLA-4 and  $\alpha$ 4 $\beta$ 7 as well as the  $\beta$ 2-integrins are the most important integrins involved in leukocyte recruitment (Chavakis, 2012; Berlin et al., 1993) interacting with endothelial ligands/counter-receptors.

Inflammatory activation of the endothelium triggers the production or upregulation of several molecules promoting leukocyte recruitment: a) endothelial cells secrete a variety of chemokines, which are deposited on their apical surface and induce 'inside-out' signalling leukocyte integrin activation, and b) they induce or upregulate cell adhesion molecules on their luminal cell surface, which function as counter-receptors of integrins on leukocytes, thereby mediating leukocyte adhesion (reviewed in Ley et al., 2007; Chavakis et al., 2009; Chavakis, 2012).

Chemokines act *via* their G-protein-coupled receptors and induce signalling cascades called 'inside-out' signalling, which lead to the activation of the extracellular domains of integrins (reviewed in Chavakis et al., 2009; Ley et al., 2007). Chemokine-induced inside-out signalling comprises several inter-dependent pathways, including a) activation of phospholipase C, which leads to intracellular  $\text{Ca}^{2+}$  flux from the endoplasmic reticulum and generation of inositol-1,4,5-trisphosphate (InsP3) and diacylglycerol (DAG) (Zarbock et al., 2011), b) activation of small GTPases, such as Rap1 (RAS-related protein 1) by guanine-nucleotide-exchange factors (Shimonaka et al., 2003; Lafuente and Boussiotis, 2006; Chavakis et al., 2009; Gahmberg et al., 2009) and c) interaction of intracellular proteins, such as talin-1, kindlin-3, cytohesin-1 and 14-3-3-family members, with the cytoplasmic tail of integrins. The binding of the latter proteins to the  $\beta$  subunit of e.g. LFA-1 results in separation of the two cytoplasmic tails thereby inducing and sustaining the conformational changes in the extracellular domain (reviewed in Alon and Feigelson, 2012, Hogg et al., 2011; Moser et al., 2009; Choi et al., 2008a; Chavakis et al., 2009; Kinashi, 2005; Fagerholm et al., 2002; Kolanus et al., 1996; Gahmberg et al., 2009). Besides chemokines and PSGL-1-ligation, Toll-like receptors also induce integrin activation (Harokopakis et al., 2006; Harokopakis and Hajishengallis, 2005). Recently, Toll like receptor 2 (TLR2)- and Toll like receptor 5 (TLR5)-ligation was shown to rapidly activate integrin-dependent leukocyte adhesion to immobilized intercellular cell-adhesion molecule 1 (ICAM-1) or fibronectin through activation of a pathway requiring Rac1, NADPH oxidase 2-mediated reactive oxygen species production and activation of Rap1-GTPase (Chung et al., 2014). Furthermore, TLRs activate *via* Ras the PI3K isoform p100 $\gamma$ , which then promotes activation of the  $\alpha$ 4 $\beta$ 1-integrin (Schmid et al., 2011). The distinct triggering signals and pathways involved in 'inside-out' signalling ensure a great diversity in integrin activation and thereby stimulation of inflammatory cell recruitment under different inflammatory conditions (Hyduk et al., 2007; Kinashi, 2005; Lafuente and Boussiotis, 2006; Shamri et al., 2005; Wegener et al., 2007 and reviewed in Chavakis et al., 2009; Hogg et al., 2011; Ley et al., 2007).

Upon activation, integrins bind to their ligands, mediating slow rolling, leukocyte adhesion and crawling and participate in transendothelial migration (Ley et al., 2007). Moreover, leukocyte integrins may participate in other functions such as immune synapse formation or phagocytosis (Dupuy and Caron, 2008; Springer and Dustin, 2012). Integrin activation cooperates with selectins to mediate slow rolling. LFA-1 or Mac-1-deficient mice both show significantly increased leukocyte rolling velocities under inflammatory conditions, indicating that  $\beta$ 2-integrins contribute to “slowing-down” of rolling neutrophils (Dunne et al., 2002).

VLA-4 binds to the vascular cell-adhesion molecule (VCAM)-1 and autotaxin (Kanda et al., 2008; Gahmberg et al., 2009), while LFA-1 and Mac-1 interact with ICAM-1 and ICAM-2 (reviewed in Springer, 1994; Chavakis, 2012; Chavakis et al., 2009; Gahmberg et al., 2009). Mac-1 is a very promiscuous receptor interacting with numerous other ligands. For instance, it binds to iC3b, thereby promoting complement-dependent phagocytosis by macrophages (Micklethorn and Sim, 1985; Dupuy and Caron, 2008). Mac-1 also interacts with fibrinogen (Altieri et al., 1990), which was shown to be of importance for bacterial elimination by leukocytes (Flick et al., 2004). Furthermore it binds heparin (Diamond et al., 1995), elastase (Cai and Wright, 1996) and other proteolytic enzymes, such as kininogen components, plasminogen, fragments thereof, urokinase or its receptor (Chavakis et al., 1999; Chavakis et al., 2001; Chavakis et al., 2005; Pluskota et al., 2003; Wei et al., 1996; Simon et al., 1996), thereby orchestrating cell surface-associated proteolytic activity. Mac-1 was also demonstrated to interact with the receptor for advanced glycation end products (RAGE) (Orlova et al., 2007; Chavakis et al., 2003a; Frommhold et al., 2010), an interaction that could be relevant in diabetes-associated vascular inflammation (Yamamoto and Yamamoto, 2013). It also interacts with membrane glycoprotein GPIb $\alpha$  (Ehlers et al., 2003; Chavakis et al., 2003b; Simon et al., 2000) and Junctional Adhesion Molecule-C (JAM-C) on platelets (Santoso et al., 2002), thereby mediating leukocyte-platelet interactions. Finally,  $\alpha$ 4 $\beta$ 7 on lymphocytes interacts with the mucosal vascular addressin cell-adhesion molecule 1 (MAdCAM1) (Berlin et al., 1993; and reviewed in Springer, 1994; Ley et al., 2007; Chavakis et al., 2009).

Leukocyte-platelet cross-talks play also an important role in promoting leukocyte adhesion and leukocyte recruitment (Langer and Chavakis, 2009). Upon vascular injury, platelets adhere to the vascular endothelium where they stimulate leukocytes indirectly by the release of chemokines and inflammatory cytokines (von Hundelshausen and Weber, 2007; Zarbock et al., 2007b), as well as directly through cell-to-cell contact (Zarbock et al., 2007b). For instance, platelets secrete CC-chemokine ligand 5 (CCL5 or RANTES), CXC-chemokine ligand 4 (CXCL4) and CXCL5, thereby triggering monocyte arrest (Huo et al., 2003). Furthermore, platelet P-selectin interacts with PSGL-1 on leukocytes (Moore et al., 1992; Larsen et al., 1989), while GPIb, JAM-C and platelet-associated fibrinogen all bind to leukocyte Mac-1 (Ehlers et al., 2003; Santoso et al., 2002; Weber and Springer, 1997). These interactions contribute to the formation of direct platelet-leukocyte cell-to-cell interactions (Massberg et al., 2002; Massberg et al., 2005; Burger and Wagner, 2003; Zarbock, 2007b; Santoso et al., 2002; Simon et al., 2000). The significance of leukocyte-platelet interactions in vascular inflammation is evidenced by their critical role in the formation of atherosclerotic lesions (Massberg et al., 2002; Huo et al., 2003).

Ligand binding of integrins triggers signalling pathways designated ‘outside-in signalling’, which lead to strengthening of leukocyte adhesion and subsequent spreading. Ligation of  $\beta$ 2-integrin induces the activation of the Src proteins Hck and Fgr (Giagulli et al., 2006) and phosphorylation of spleen tyrosine kinase (Mocsai et al., 2006; reviewed in Hogg et al., 2011). In T cells, proteins involved in outside-in signalling are the Lck and  $\zeta$ -chain-associated protein kinase of 70kDa (Evans et al., 2011). During this step, integrins associate intracellularly with cytoskeletal proteins; for instance  $\alpha$ 4 $\beta$ 1-integrin interacts with paxillin (Liu et al., 1999), which contributes to the stability of activated integrin conformation (Goldfinger et al., 2003; Kinbara et al., 2003). Moreover leukocyte integrins associate with further cytoskeletal proteins, such as vinculin or actinin (Mace et al., 2010; Stanley et al., 2008); this association facilitates formation of focal adhesions and further integrin-dependent steps (Kinbara et al., 2003). Following firm arrest, leukocytes crawl on the endothelial cell surface in a manner dependent on  $\beta$ 2-integrins, a process also named locomotion (Schenkel et al., 2004; Phillipson et al., 2006) that allows leukocytes to identify an appropriate site to migrate through the endothelial cell monolayer.

Transmigration through venular walls is the final step in the process of leukocyte emigration into inflamed tissues. Leukocytes mainly transmigrate paracellularly at interendothelial junctions, although transcellular migration through the endothelial cell body also rarely takes place (Nourshargh et al., 2010; Woodfin et al., 2011; Choi et al., 2009).

The critical role of LFA-1 and Mac-1 during transmigration has long been established (Smith et al., 1989; Henderson et al., 2001). Shaw *et al.* have shown that the distribution of LFA-1 in transmigrating neutrophils rapidly changes to form a ring-like cluster at the neutrophil-endothelial junctions, while endothelial ICAM-1 co-localizes with LFA-1 in these clusters (Shaw et al., 2004). ICAM-2 also mediates neutrophil transmigration, due to its interaction with Mac-1 and LFA-1 (Staunton et al., 1989; Xie et al., 1995), as shown by genetic or antibody-mediated inactivation of ICAM-2 (Huang et al., 2006). Furthermore, there is ample evidence that integrins interact with JAM proteins in the process of transmigration. LFA-1 was shown to bind to JAM-A, while Mac-1 interacts with JAM-C; these interactions may promote leukocyte diapedesis (Chavakis et al., 2004; Lamagna et al., 2005; Keiper et al., 2005; Orlova and Chavakis, 2007; Ostermann et al., 2002). The important role of JAM-C in leukocyte transmigration *in vivo* has been shown in a study by Woodfin *et al.*, linking the reverse transendothelial migration with reduced expression of JAM-C at the endothelial cell junctions (Woodfin et al., 2011). Furthermore, JAM-A was recently demonstrated to promote monocyte recruitment to the arterial wall during atherosclerosis (Schmitt et al., 2014).

Numerous other endothelial molecules, primarily located at junctions, such as platelet/endothelial-cell adhesion molecule 1 (PECAM-1, CD31), CD99 and endothelial cell selective adhesion molecule are also involved in leukocyte transmigration (Ley et al., 2007; Nourshargh et al., 2010; Muller, 2003; Muller, 2014; Schenkel et al., 2002; Wegmann et al., 2006). Interestingly, ICAM-2, JAM-A and PECAM-1 play a rather sequential role during leukocyte transmigration (Woodfin et al., 2009). Endothelial PECAM-1 promotes leukocyte transmigration by undergoing homophilic interactions with PECAM-1 on monocytes (Mamdouh et al., 2003; Muller et al., 2003), through which it can up-regulate  $\alpha$ 6 $\beta$ 1-integrin



(Dangerfield et al., 2002); in addition, PECAM-1 also undergoes a heterophilic interaction with CD177, which may contribute to transmigration (Sachs et al., 2007).

CD99 through homophilic interactions (Muller et al., 2003; Schenkel et al., 2002) and CD99L2 (Bixel et al., 2010), also promote leukocyte transmigration (Schenkel et al., 2002; Bixel et al., 2004; Bixel, 2010). Finally, VE-cadherin, the most important gatekeeper of vascular endothelial integrity may be transiently distributed away from the junctional regions during diapedesis thus permitting trans-endothelial leukocyte migration, whereas phosphorylation of VE-cadherin also plays a regulatory role in the process of diapedesis (Shaw et al., 2001; Vestweber et al., 2009; Wessel et al., 2014).

## Leukocyte integrins as targets in inflammatory disease models

Leukocyte recruitment into an inflamed tissue is a cardinal hallmark in several inflammatory diseases. Infiltrating leukocytes induce, promote and perpetuate inflammation, whereas their clearance is critical for the resolution of the inflammatory process. Due to their critical role in leukocyte recruitment, leukocyte integrins have emerged as therapeutic targets in several inflammatory diseases (Yonekawa and Harlan, 2005). In the following section, we review the importance of leukocyte integrins in different animal models of inflammatory disorders, in which integrin targeting has been proven to be beneficial (Figure 2).

### Multiple sclerosis

Multiple sclerosis (MS) is an autoimmune disease of the central nervous system (CNS) characterized by demyelination. The clinical course ranges from relapsing-remitting to progressive forms and can lead to severe disability and mortality (Bhat and Steinman, 2009; Confavreux et al., 2000; Confavreux and Vukusic, 2006; Lassmann et al., 2012). The rodent model of MS is experimental autoimmune encephalomyelitis (EAE); pathology in this model (and most likely in MS as well) is triggered by infiltration of autoreactive T cells, which are targeted against antigens of the myelin sheath through the blood-brain barrier (BBB) into CNS. This initiates a strong neuroinflammatory response, involving BBB disruption (Minagar and Alexander, 2003) and recruitment of further immune cells (Bhat and Steinman, 2009; Korn, 2008). Thus, leukocyte recruitment is a critical event in the pathogenesis of EAE and MS (Goverman, 2009; Popescu and Lucchinetti, 2012; Simmons et al., 2013).

The role of  $\alpha 4\beta 1$ -integrin in leukocyte recruitment to the inflamed CNS in the course of EAE has been extensively investigated (Sheremata et al., 2005). The identification of this integrin as a major player in EAE and MS resulted in the development of natalizumab, a monoclonal antibody against  $\alpha 4$ , which has been approved for the treatment of patients with MS (Weinstock-Guttman, 2013). Initial studies demonstrated that leukocyte adherence to the inflamed endothelium in the CNS was mediated by the  $\alpha 4\beta 1$ -integrin / VCAM-1 interaction; consistently, blockade of  $\alpha 4\beta 1$ -integrin prevented disease development in rats (Yednock et al., 1992) and mice (Baron et al., 1993). By using a model of adoptive transfer in EAE, Baron *et al.* further demonstrated that only antigen-specific T cells expressing  $\alpha 4\beta 1$ -integrin were able to migrate into the brain parenchyma and that blocking  $\alpha 4$ -integrin with antibodies preferentially inhibited the recruitment of these cells to the inflamed CNS

(Baron et al., 1993). The effect of  $\alpha 4$  antibodies was mediated by blockade of  $\alpha 4\beta 1$ - rather than  $\alpha 4\beta 7$ -integrin, as clarified by inhibition studies with antibodies against  $\alpha 4\beta 7$  heterodimer or  $\beta 7$ -integrin (Engelhardt et al., 1998), whereas endothelial overexpression of the  $\alpha 4\beta 7$ -integrin counter-receptor MAdCAM-1 did not affect EAE severity (Döring et al., 2011). Intravital microscopy approaches underlined the importance of  $\alpha 4$ -integrin and its collaboration with P-selectin for the adhesion of leukocytes to the brain endothelium (Kerfoot and Kubes, 2002). Additional *in vitro* and *in vivo* studies, using intravital microscopy, further supported the involvement of  $\alpha 4\beta 1$ -integrin binding to VCAM-1 in the adhesion of T cells to the vasculature of the white matter (Laschinger and Engelhardt, 2000; Vajkoczy et al., 2001). Blocking of  $\alpha 4\beta 1$ -integrin with natalizumab was shown to inhibit firm adhesion of human T cells to the inflamed BBB in mice with EAE, without affecting the rolling or initial contact of these cells (Coisne et al., 2009). Moreover,  $\alpha 4\beta 1$ -integrin was shown to mediate initial recruitment of encephalitogenic T cells as well as the secondary infiltration of inflammatory cells, thereby being involved in initiating and sustaining CNS inflammation (Brocke et al., 1999). Experiments with cell-specific deletion of  $\beta 1$ -integrin demonstrated that T cells, rather than myeloid cells, were dependent on  $\alpha 4\beta 1$ -integrin for their migration to the inflamed CNS, supporting the notion that the therapeutic effect of antibodies against  $\alpha 4$  is largely mediated by the inhibition of T cell extravasation (Bauer et al., 2009). However,  $\alpha 4\beta 1$ -integrin is involved in the recruitment of additional immune cells in the inflamed CNS, including immature DCs (Jain et al., 2010) and Natural Killer (NK) cells (Gan et al., 2012). The different observations from rodent EAE establishing a major role for  $\alpha 4\beta 1$ -integrin in disease pathogenesis resulted in the clinical use of  $\alpha 4\beta 1$ -integrin inhibition as a therapeutic principle in MS, which will be discussed in the following section.

Several studies investigated whether inhibition of  $\beta 2$ -integrins or their ligands could be of potential benefit in EAE and MS. Using antibodies against CD11a and CD11b, Gordon *et al.* demonstrated that targeting these  $\beta 2$ -integrins could block or delay the onset of EAE (Gordon et al., 1995). Dugger *et al.* performed transfer of CD11a<sup>-/-</sup> T cells to wild type mice and demonstrated that these cells, even though they infiltrate lymph nodes and CNS, caused less severe disease than wild-type controls. On the other hand, transfer of wild type encephalitogenic T cells to CD11a<sup>-/-</sup> mice worsened the outcome of EAE, thereby indicating that CD11a may have both effector and suppressor functions in EAE pathogenesis (Dugger et al., 2009). Specifically, these seemingly contradictory findings could be attributed to the fact that LFA-1 is involved in different functions besides T cell trafficking, including functioning as a co-stimulatory molecule (Van Severter et al., 1991) or as a molecule that regulates the function of regulatory T cells (Treg) (Tran et al., 2009). For instance, LFA-1 deficiency resulted in an altered balance between effector T cells and Tregs in favor of the former, which led to increased severity of EAE in CD11a-deficient mice (Gültner et al., 2010). Based on the fact that both Th1 lymphocytes and Th17 lymphocytes contribute to EAE development (Jäger et al., 2009; Kebir et al., 2007; Kleinschek et al., 2007), the involvement of LFA-1 in the recruitment of these separate lymphocyte populations into the CNS was addressed. The infiltration of lymphocytes to the draining lymph node following immunization as well as the development of Th17 cells were significantly impaired in LFA-1<sup>-/-</sup> mice, thereby resulting in reduced disease severity (Wang et al., 2007). Rothhammer *et al.* further demonstrated that LFA-1 is also important

for Th17 trafficking to the inflamed CNS. In this study, it was reported that Th1 and Th17 cells engage different integrins for their migration to the CNS. Blockade of  $\alpha 4$ -integrin protected from EAE induced by adoptive transfer of Th1 but not of Th17 lymphocytes. The latter entered the CNS in an  $\alpha 4$  integrin-independent manner. In contrast, the infiltration of Th17 cells into the brain parenchyma was mediated by LFA-1. Thus, Th17 cell migration to the CNS requires LFA-1, whereas Th1 cells preferentially use  $\alpha 4$ -integrin for this process (Rothhammer et al., 2011; Glatigny et al., 2011). With regards to the role of LFA-1 in the migration of T lymphocytes across the BBB, Laschinger *et al.* demonstrated that LFA-1 participates in the transendothelial migration but not in endothelial adhesion (Laschinger et al., 2002). The effect of inhibiting the LFA-1 ligand ICAM-1 in the course of EAE was also studied. Even though studies using inhibitory antibodies against ICAM-1 have shown contradictory results (Archelos et al., 1993; Morrissey et al., 1996; Willenborg et al., 1993), EAE development was significantly attenuated in ICAM-1<sup>-/-</sup> mice (Bullard et al., 2007b), as well as by targeting ICAM-1 with a staphylococcal-derived anti-adhesive protein (Xie et al., 2006). Besides CD11a, deficiency of CD11b (Bullard et al., 2005) or CD11c (Bullard et al., 2007a) has also been demonstrated to ameliorate development of EAE. Moreover, the CD11b-mediated interaction of leukocytes with platelets contributes to inflammatory cell recruitment to the CNS in the course of EAE pathogenesis. Inhibition of the leukocyte-platelet interaction by blocking the binding of Mac-1 to platelet GPIb resulted in significant amelioration of disease severity (Langer et al., 2012).

## Psoriasis

Psoriasis is a chronic inflammatory disease of the skin, which is characterized by erythematous scales; recent evidence has suggested an important role of IL-17 derived from different cells as a pathogenic component in psoriasis (Krueger et al., 2012; Leonardi et al., 2012; Martin et al., 2013; Pantelyushin et al., 2012). Th1, Th17 cells,  $\gamma\delta$  T cells, innate lymphocytes and DCs are abundant inflammatory cell populations in psoriatic lesions (Chamian and Krueger, 2004; Chamian et al., 2005; Lowes et al., 2008; Pantelyushin et al., 2012). Treatment of patients with efalizumab, a monoclonal antibody that targets CD11a resulted in significant reduction of TNF and iNOS-expression and of DC numbers in lesions (Lowes et al., 2005). In a psoriatic model engaging human skin-SCID (severe combined immunodeficiency) mouse chimeras, treatment with efalizumab limited skin inflammation in the transplant (Zeigler et al., 2001; Stenderup et al., 2011). Besides LFA-1 blockade, Schön *et al.* demonstrated that Mac-1 blockade ameliorates skin manifestations in the flaky skin mice (*fsn*<sup>-/-</sup> mice) that develop psoriasiform skin lesions. Histological analysis revealed that treatment with an antibody against CD11b resulted in decreased neutrophil and T cell infiltration. This finding implies Mac-1 on neutrophils in the inflammatory process that characterizes at least this model of psoriasis (Schön et al., 2000). The effect of Mac-1 blockade was also confirmed in another murine model of experimental psoriasiform dermatitis (Leon et al., 2006). One of the Mac-1 counter-receptors on the endothelium is RAGE (Chavakis et al., 2003a; Orlova et al., 2007; Frommhold et al., 2010). Interestingly, RAGE serves also as the receptor for S100A7, also designated psoriasin, which is overexpressed in psoriatic lesions and exerts a strong chemotactic activity for neutrophils (Wolf et al., 2008; Wolf et al., 2010). Whether blockade of the interaction of RAGE with its

counter-receptor Mac-1, or its S100 ligands could be therapeutically relevant in psoriasis is worth assessing.

In addition to  $\beta 2$ -integrins LFA-1 and Mac-1, blocking of  $\alpha 1\beta 1$ -integrin (VLA-1), which serves as a collagen receptor, inhibited the infiltration of epidermal T cells in a xenotransplantation psoriasis model. This study, moreover, established that the presence of  $\alpha 1\beta 1$ -integrin in epidermal effector T cells was associated with the expression of high levels of interferon- $\gamma$ . Thus,  $\alpha 1\beta 1$ -integrin may represent a potential therapeutic target in psoriasis (Conrad et al., 2007).

Interestingly the  $\beta 2$ -integrin was demonstrated to play a regulatory role in the development of psoriasis as well. Mice bearing a hypomorphic CD18 mutation (CD18<sup>hyp</sup>PL/J mice), resulting in decreased CD18 expression, develop psoriasiform dermatitis (Peters et al., 2006), featured by increased accumulation of CD4+ and  $\gamma\delta$  T cells (Gatzka et al., 2013) and decreased presence of regulatory T cells (Singh et al., 2013) at the skin lesions.  $\beta 2$ -integrin deficiency has been previously associated with dysregulation of IL-17 production, due to defective neutrophil recruitment to peripheral tissues and subsequently reduced clearance of apoptotic neutrophils by macrophages (Stark et al., 2005; Moutsopoulos et al., 2014), as discussed in the following section. The resulting reduction of IL-23 production by macrophages leads to  $\gamma\delta$  T cell activation, which then produce IL-17 (Stark et al., 2005). This mechanism could be a possible explanation for the aforementioned findings in mice with decreased CD18 expression.

### Inflammatory bone loss

Inflammatory cell recruitment is a common denominator of several disorders involving bone loss, with rheumatoid arthritis (RA) being the most prominent one. Synovial inflammation in RA results from infiltrating neutrophils, T cells, B cells, plasma cells, DCs and macrophages. Inflammatory cells together with osteoclasts present in the synovial membrane act synergistically in the destruction of the adjacent bone (Choy, 2012). Presentation of arthritis-specific antigens by DCs, macrophages and B cells to CD4+ T cells is considered an important trigger in disease development (Choy, 2012).

The role of leukocyte integrins in RA has been addressed in several animal models. In the rat model of adjuvant arthritis, T cell infiltration was shown to be independent of the  $\beta 2$ -integrins LFA-1 and Mac-1, whereas a partial inhibition of neutrophil recruitment was observed when LFA-1 was inhibited. However, upon concomitant blockade of both LFA-1 and Mac-1, neutrophil recruitment was abolished (Issekutz and Issekutz, 1993). In the same model, T cell recruitment was shown to depend on  $\alpha 4\beta 1$ -integrin (Issekutz and Issekutz, 1991); this integrin mediated infiltration of adoptively transferred arthritogenic T cells to the synovium (Issekutz et al., 2003). Furthermore, combined blockade of LFA-1 and VLA-4-integrins was required for the complete inhibition of monocyte infiltration (Issekutz and Issekutz, 1995).

The involvement of  $\beta 2$ -integrins in the development of murine arthritis was studied in the model of K/B xN serum transfer arthritis. Watts *et al.* demonstrated that mice deficient in the common  $\beta 2$  subunit were protected from arthritis development. Additionally, CD11a

deficient mice did not develop arthritis, whereas no effect was observed in CD11b deficient mice, suggesting that LFA-1 but not Mac-1 contributes to disease induction. LFA-1 was required not only for arthritis development but also for maintenance of synovial inflammation, as shown by disease amelioration after administration of anti-LFA-1 antibody in mice with already established arthritis, thereby suggesting a therapeutic potential for LFA-1 targeting in RA (Watts et al., 2005).

LFA-1 is also involved in bone loss in the context of inflammatory arthritis. Barck *et al.* studied the effect of LFA-1 blockade in the formation of bone erosions in the murine model of collagen induced arthritis. Using 3-dimensional micro-CT of mouse paws to evaluate joint cortical bone volume, they demonstrated that treatment with anti-LFA-1 antibody ameliorated both the clinical course and the radiographic findings (Barck et al., 2004).

Furthermore,  $\alpha 2\beta 1$  integrin, a receptor for collagen type I, was shown to promote arthritis in the antigen-induced arthritis (AIA) and the human TNF-transgenic mouse model. Mice deficient in  $\alpha 2\beta 1$  showed reduced cartilage erosions and joint pathology, as compared to wild-type mice (Peters et al., 2012).

A link between inflammatory bone loss and LFA-1 has been recently demonstrated in periodontitis. LFA-1 deficiency was associated with severe periodontal bone loss and lack of neutrophil recruitment in the periodontal tissue in both mice and men with type I leukocyte adhesion deficiency (LAD-I). This was attributed to dysregulated overproduction of IL-17 as a result of a defective “neurostat”-feedback mechanism in mice and individuals with defective leukocyte recruitment (Moutsopoulos et al., 2014). Specifically, when neutrophils cannot transmigrate to peripheral tissues (as occurs in LAD-I), the inhibitory signals for IL-23 expression by phagocytes (normally associated with apoptotic cell efferocytosis) are abrogated leading to T cell overexpression of IL-17. IL-17 in turn promotes granulopoiesis through G-CSF, consistent with blood neutrophilia in LAD-I patients and relevant mouse models (Stark et al., 2005). Interestingly, topical inhibition of IL-17 blocked inflammatory bone loss in mouse models mimicking LAD-I-associated periodontitis (Moutsopoulos et al., 2014).

### Inflammatory bowel disease

Inflammatory bowel diseases (IBD), namely Crohn’s disease (CD) and ulcerative colitis (UC), are chronic disorders of the gastrointestinal tract. Advances in the understanding of the pathogenesis of intestinal inflammation enabled the development of specific therapies, with monoclonal antibodies e.g. targeting TNF (Kontoyiannis et al., 2002; Dassopoulos et al., 2013).

Studies with animal models of IBD revealed the critical involvement of leukocyte integrins in intestinal inflammation, which enabled the development of targeted therapies for the treatment of CD and UC, as discussed below. Early studies demonstrated the central role of  $\beta 7$ -integrins for the formation of gut-associated lymphoid tissue (GALT), which includes Peyer’s patches, lamina propria lymphocytes and intra-epithelial lymphocytes of the intestine (Koboziev et al., 2010; Gorfu et al., 2009). Mice deficient in the  $\beta 7$ -integrin subunit have hypoplastic GALT, which was further attributed to defective T cell recruitment

(Wagner et al., 1996). Furthermore, antibodies targeting  $\alpha 4$  integrin, an epitope present on the  $\alpha 4\beta 7$ -integrin or the  $\beta 7$  subunit significantly attenuated T cell homing to Peyer's patches, without affecting homing to peripheral lymph nodes. In the same study, it was also shown that the mucosal addressin MAdCAM-1 mediates T cell infiltration in mucosal lymphoid tissue (Hamann et al., 1994). Using  $\beta 7^{-/-}$  mice, Kunkel et al. confirmed the involvement of  $\beta 7$ -integrin in adhesion of leukocytes to Peyer's patch endothelial venules (Kunkel et al., 1998). Treatment with anti- $\beta 7$  and anti-MAdCAM-1 antibodies was shown to ameliorate experimental murine chronic colitis, by affecting T cell recruitment to the colonic mucosa and mesenteric lymph nodes (Picarella et al., 1997). The involvement of  $\beta 7$ -integrins in the development of mouse colitis was also shown in the SAMP1/YitFc model of inflammatory Crohn's disease-like ileitis. In this model,  $\beta 7$ -integrin blockade was efficient to significantly suppress inflammation, only when combined with L-selectin inhibition (Rivera-Nieves et al., 2005). In the same model,  $\beta 7$ -integrin deficiency ameliorated colitis by inhibiting B cell homing to mesenteric lymph nodes on top of blocking T cell infiltration (Gorfu et al., 2010). Inhibition of  $\alpha E\beta 7$ -integrin reduced mucosal inflammation in  $IL-2^{-/-}$  mice immunized with TNP-OVA, even when mice were treated after the establishment of colitis (Ludviksson et al., 1999). However,  $\beta 7$ -deficiency in mice did not significantly ameliorate colitis course (Sydora et al., 2002). On the other hand, Apostolaki *et al.* have shown the dependency of T cell recruitment to intestinal mucosa on  $\beta 7$ -integrins in a TNF-dependent model of Crohn's disease; moreover,  $\beta 7$ -deficiency completely abrogated colitis development (Apostolaki et al., 2008). Furthermore, a recent study by Villablanca *et al.* demonstrated that mice deficient in  $\beta 7$ -integrin in the innate immune compartment show increased susceptibility to T cell mediated colitis. The authors proposed that the reduced migration of  $\beta 7$ -deficient tolerogenic mononuclear phagocytes into gut mucosa contributes to this phenotype (Villablanca et al., 2014).

Several studies suggested a role of  $\beta 2$ -integrins in experimental colitis. In the rat model of 2,4,6-trinitrobenzene sulfonic acid-induced colitis, administration of antibodies against Mac-1 resulted in significant inhibition of inflammatory cell recruitment, which was associated with a decreased mucosal damage (Palmen et al., 1995). By transferring  $CD11a^{-/-}$   $CD4^{+}CD25^{-}$  T cells to  $RAG^{-/-}$  recipients, Pavlick *et al.* demonstrated that expression of LFA-1 on T cells is required for disease induction and for the infiltration of  $CD4^{+}$  T cells to mesenteric lymph nodes and to intestinal lamina propria (Pavlick et al., 2006). Using the same approach  $CD18$  on T cells was identified to promote colitis induction (Ostanin et al., 2007).

The identified role of leukocyte integrins in models of IBD paved the way for the development of targeted therapeutic agents, as discussed in the following section.

## Leukocyte integrin blockade in clinical use

As a result of the large body of evidence regarding the central role of leukocyte integrins in animal models of inflammatory diseases, different leukocyte integrin targeting strategies have reached clinical use (Yonekawa and Harlan, 2005) (Table 1).

## Beta2-integrins

Efalizumab (Raptiva, anti-CD11a, Genentech) is a humanized monoclonal IgG1 antibody that is directed against the alpha chain (CD11a) of the  $\beta$ 2-integrin LFA-1, that blocks the interaction between leukocyte LFA-1 and ICAM-1 (Dedrick et al., 2002). Phase III clinical trials demonstrated that efalizumab administration results in the amelioration of dermatologic features of psoriasis (Gordon et al., 2003; Lebwohl et al., 2003, Leonardi et al., 2005), which can be attributed to reduction of inflammatory cell recruitment and reversal of keratinocyte hyperplasia at psoriatic lesions (Reich et al., 2012; Gottlieb et al., 2002). Despite having beneficial effect on the skin manifestations, it was not effective in the treatment of psoriatic arthritis (Papp et al., 2007), while several reports suggest that treatment with efalizumab might even result in new-onset severe arthritis (Viguier et al., 2008; Myers et al., 2006). Despite its effectiveness, it was withdrawn in 2009 due to the high risk of John Cunningham (JC) polyomavirus reactivation and development of progressive multifocal leukoencephalopathy (PML) in patients under long-term efalizumab administration (Weger, 2010).

## Alpha4-integrins

Natalizumab (Antegren, Tysabri; Elan Pharma Ltd., Letchworth, UK and Biogen-Idec, Durham, NC, USA) is a recombinant humanised monoclonal IgG4 antibody against the  $\alpha$ 4-integrin subunit, blocking both  $\alpha$ 4 $\beta$ 1- and  $\alpha$ 4 $\beta$ 7-integrins. It prevents the migration of leukocytes across the blood–brain barrier into the CNS by blocking the interaction between  $\alpha$ 4 $\beta$ 1-integrin and VCAM-1 (Rudick and Sandrock, 2004; Ransohoff, 2007; Rudick et al., 2013); moreover, it inhibits the interaction of  $\alpha$ 4 $\beta$ 7-integrin with endothelial MAdCAM-1 and thereby homing of lymphocytes to gastrointestinal lymphoid tissue (Hamann et al., 1994). Studies in patients with MS have shown that natalizumab administration significantly reduced the numbers of total leukocytes, CD4+ T cells, CD8+ T cells, B cells and plasma cells in the cerebrospinal fluid (Stüve et al., 2006). A significant reduction in the number of CD209+ antigen presenting DCs in cerebral perivascular spaces and decreased expression of major histocompatibility complex class II was also observed in brain autopsies from a patient with PML that had received natalizumab (del Pilar Martin et al., 2008). These effects are localized to the CNS, as natalizumab did not reduce the numbers of CD4+ and CD8+ T cells in the periphery (Kivisäkk et al., 2009).

After successfully completing phase III clinical trials (Polman et al., 2006, Rudick et al., 2006), natalizumab was approved as the first targeted therapy for the treatment of relapsing-remitting multiple sclerosis. Natalizumab is associated with increased risk for reactivation of latent JC virus infection and development of PML (Bloomgren et al., 2012); however, a recently published study confirmed the beneficial role of natalizumab in stabilizing disease activity and diminishing the risk of relapses in patients with relapsing-remitting multiple sclerosis (O'Connor et al., 2014). Phase II and III studies have also shown that natalizumab is effective as an induction and maintenance therapy in patients with Crohn's disease (Ghosh et al., 2003; Sandborn et al., 2005; Targan et al., 2007). However, due to the risk for PML development it is approved in the United States under a restricted distribution program (Chen et al., 2013).

Furthermore, antibodies targeting the interaction between  $\alpha 4\beta 7$ -integrin and MAdCAM have been developed. Vedolizumab and etrolizumab target  $\alpha 4\beta 7$ -integrin, whereas PF-00547659 targets MAdCAM (Bamias et al, 2013). The selective blockade of  $\alpha 4\beta 7$ -integrin leaving  $\alpha 4\beta 1$  unaffected should minimize the risks associated with the inhibition of leukocyte recruitment to the CNS. Vedolizumab (Millennium Pharmaceuticals) is a humanized IgG1 monoclonal antibody that is effective as an induction and maintenance therapy for moderate to severe UC and Crohn's disease, as well as in patients with moderate to severe Crohn's disease that did not previously respond to other treatments (Feagan et al., 2013; Sandborn et al., 2013; Sands et al., 2014). Vedolizumab was recently approved by the FDA for the treatment of Crohn's disease.

Etrolizumab is a humanised monoclonal antibody that selectively binds the  $\beta 7$ -subunit of both the  $\alpha 4\beta 7$  and  $\alpha E\beta 7$ -integrin heterodimers, antagonizing both  $\alpha 4\beta 7$ -MAdCAM-1-mediated lymphocyte recruitment and the  $\alpha E\beta 7$ -E-cadherin interaction (Lin and Mahadevan, 2014), which mediates adhesion of  $\alpha E\beta 7$  T cells to epithelial cells (Cepek et al, 1993; Hadley and Higgins, 2014). In a recently published phase II clinical trial, etrolizumab was shown to have a beneficial effect as an induction therapy in the treatment of patients with moderate to severe IBD (Vermeire et al., 2014).

Despite the high efficacy of integrin-targeted therapies in inflammatory diseases, the relatively frequent occurrence of severe adverse events precludes the administration of such agents to a wider spectrum of patients with inflammatory diseases.

## Endogenous inhibitors of leukocyte recruitment

Besides the aforementioned antibodies and inhibitors that have been developed to therapeutically target integrins and block leukocyte recruitment, the recent discovery of endogenous negative regulators of the leukocyte adhesion cascade may pave the way for the development of novel, more efficient strategies to therapeutically target leukocyte adhesion in inflammatory and autoimmune pathologies (Chavakis, 2012; Hajishengallis and Chavakis, 2013). We will therefore summarize here recent knowledge on endogenous inhibitors of leukocyte recruitment with a special focus on the integrin inhibitor Developmental endothelial locus-1 (Del-1).

### Del-1: an endogenous anti-inflammatory factor blocking leukocyte $\beta 2$ -integrins

Del-1, also designated epidermal growth factor (EGF)-like repeats and discoidin-I-like domains 3 (EDIL3) is a glycoprotein with three EGF-like repeats and two C-terminal discoidin I-like domains; the latter can interact with phosphatidylserine and phospholipids (Dasgupta et al., 2012; Chavakis et al., 2009). It is expressed and secreted by endothelial cells and can associate with the extracellular matrix as well as the endothelial cell surface (Chavakis et al., 2009). Association with the endothelial cell surface is mediated by binding to surface proteoglycans or to integrin  $\alpha v\beta 3$  by means of an Arg-Gly-Asp motif on the second EGF-like repeat (Chavakis et al., 2009; Hidai et al., 2007). Although expressed by endothelial cells, it is not present in all tissues and organs. Its expression is highest in the brain, followed by the lung, the adrenal or the gingiva, whereas it is hardly expressed in the liver, thereby pointing to the diversity and heterogeneity of the endothelium of different



tissues / organs (Choi et al., 2008b; Kanczkowski et al., 2013). We identified Del-1 to act as a negative regulator of the leukocyte integrin LFA-1. Del-1 binds to LFA-1 integrin and antagonizes binding of the latter to ICAM-1, thereby blocking LFA-1-dependent leukocyte adhesion onto the vascular endothelium (Choi et al., 2008b). Besides LFA-1, Del-1 was recently shown to interact with Mac-1 as well, thereby blocking the binding of Mac-1 with complement component iC3b. This results in a Del-1-mediated inhibition of complement/Mac-1-dependent phagocytosis, an action that could also contribute to the anti-inflammatory functions of Del-1 (Mitroulis et al., 2014). The role of Del-1 as a negative regulator of leukocyte recruitment stands in keeping with its highest expression levels in immune privileged organs, such as the brain or the eye. Del-1-deficient mice displayed elevated LFA-1-dependent neutrophil recruitment in LPS-induced acute lung injury and in the dorsal skinfold chamber model. Injection of soluble Del-1 could ameliorate inflammatory cell recruitment to the lung or the peritoneum in acute models of inflammation (Choi et al., 2008b). Additionally, Del-1-deficiency was recently shown to enhance bleomycin-induced lung fibrosis (Kang et al., 2014) and systemic inflammation-related adrenal dysfunction (Kanczkowski et al., 2013). Thus, Del-1 might represent an interesting novel approach for the treatment of inflammatory lung diseases.

Interestingly, Del-1 expression in endothelial cells is down-regulated by inflammatory factors, such as LPS, TNF or IL-17A but not IL-17F (Choi et al., 2008b; Eskan et al., 2012; Shin et al., 2013), which suggests that inflammatory stimuli do not only elevate expression of integrin ligands ICAM-1 and VCAM-1 on the endothelium, but at the same time reduce the levels of the integrin inhibitor Del-1; both actions function synergistically to promote leukocyte adhesion. The IL-17-mediated down-regulation of endothelial Del-1 expression promoted neutrophil recruitment and inflammatory bone loss in the context of periodontitis, which is a chronic neutrophil-dependent disease (Eskan et al., 2012). Del-1 expression is diminished in the gingival tissue of old mice, correlating with hallmarks of periodontitis, such as excessive neutrophil recruitment and IL-17-dependent inflammatory bone loss (Eskan et al., 2012). Consistent with this, Del-1-deficient mice develop spontaneous periodontitis at a young age, accompanied by heavy neutrophil infiltration in the gingiva in a manner dependent on LFA-1. The periodontitis due to Del-1 deficiency was reversed in mice with Del-1/LFA-1- or Del-1/IL-17-receptor-double deficiency. Interestingly, local periodontal treatment with soluble Del-1 in old mice inhibited IL-17 production, LFA-1-dependent neutrophil infiltration, and associated bone loss (Eskan et al., 2012). Analysis of biopsies from patients with periodontitis revealed decreased levels of Del-1 and enhanced levels of IL-17 in the diseased tissue, as compared to the adjacent healthy tissue, thereby suggesting that the inflammatory IL-17/Del-1-loop is operative in human periodontitis as well (Eskan et al., 2012). These findings suggest that Del-1 and IL-17 are reciprocally cross-regulated and that Del-1 confers a self-regulated homeostatic control mechanism of neutrophil recruitment, being constantly 'on' to prevent chronic inflammation, but being turned 'off' when neutrophil recruitment is needed. Additionally, local application of Del-1 seems a promising therapeutic strategy to prevent chronic inflammatory bone loss in the context of periodontitis (Eskan et al., 2012).

More recently, Del-1 was also shown to prevent IL-17-dependent neuroinflammation in the course of EAE. Specifically, Del-1 expression was reduced in both chronic active MS

lesions and in the inflamed spinal cord of mice subjected to EAE. Del-1-deficiency resulted in enhanced EAE disease severity, accompanied by elevated IL-17-dependent neuro-inflammation. Interestingly, administration of soluble Del-1 ameliorated progression of relapsing-remitting EAE (Choi et al., 2014 in press). Together, application of Del-1 may represent a promising therapeutic approach in IL-17-dependent inflammatory disorders.

### Further endogenous inhibitors of inflammatory cell recruitment

Further endogenous inhibitors of leukocyte recruitment have been identified. Pentraxin-3 (PTX-3) is a member of the long pentraxin family of soluble pattern-recognition molecules. PTX-3 expression in myeloid cells is upregulated in response to TLR signalling and inflammatory cytokines. In neutrophils, PTX-3 is released from intracellular granules upon inflammation (Maina et al., 2009). PTX-3 reduces neutrophil rolling by binding to P-selectin, thereby inhibiting the interaction of P-selectin with its leukocyte counter-receptor, PSGL-1 (Hajishengallis and Chavakis, 2013; Inforzato et al., 2012; Deban et al., 2010). In accordance with the inhibitory effect of PTX-3 on leukocyte rolling, PTX-3-deficient mice display increased leukocyte recruitment in a model of acute lung injury (Deban et al., 2010; Hajishengallis and Chavakis, 2013).

Growth Differentiation Factor (GDF)-15 is a member of the transforming growth factor (TGF)- $\beta$  superfamily. Its tissue expression is increased upon sterile inflammation during ischemic injury and it acts as an inhibitor of leukocyte extravasation limiting myocardial tissue damage (Kempf et al., 2006). Moreover it was shown that coronary artery ligation in Gdf15-deficient mice led to enhanced recruitment of neutrophils into the infarcted myocardium and an increased incidence of cardiac rupture, while, infusion of recombinant GDF-15 repressed neutrophil recruitment. The regulatory role of GDF-15 on leukocyte recruitment is mediated by activation of Cdc42 GTPase and inhibition of Rap1 GTPase, thus blocking chemokine-induced activation of  $\beta$ 2-integrins (Kempf et al., 2011). It remains to be addressed in more detail, whether factors, such as PTX-3 or GDF-15 could be therapeutically engaged in preclinical and clinical inflammatory settings.

Additional endogenous molecules have been described that influence leukocyte recruitment. Galectin-1 has been demonstrated to down-regulate neutrophil expression of PSGL-1, CD11b and L-selectin induced by platelet-activated factor *in vitro* and neutrophil adhesion to endothelial cells under flow. In turn, increased leukocyte adhesion was observed in the cremaster model in galectin-1 deficient mice, using intravital microscopy (Cooper et al., 2008, Herter and Zarbock, 2013).

Annexin 1, a  $\text{Ca}^{2+}$  and phospholipid binding protein, is another potent endogenous regulator of inflammation (Gerke et al., 2005). Annexin 1 is abundant in neutrophils and it is released upon neutrophil activation (Perretti and Flower, 2004). Several peptides deriving from its N-terminal domain potentially inhibit neutrophil recruitment (Perretti et al., 2002). Upon exposure to the cell surface, annexin 1 interacts with the formyl peptide receptor (FPR), the high affinity receptor of fMLP, as well as the lipoxin A4 receptor (ALX or FPR-like receptor) (El Kebir et al., 2008; Perretti and Flower, 2004; Perretti et al., 2002; Perretti, 2003). Glucocorticoids promote annexin 1 synthesis, and annexin 1 is thereby thought to mediate many of the anti-inflammatory effects of these steroids (Perretti and Flower, 2004).

Its important role in the regulation of neutrophil recruitment was demonstrated in mice deficient for annexin 1, which display augmented and prolonged inflammatory reaction and partial resistance to the anti-inflammatory effect of dexamethasone, during acute or chronic inflammation (Hannon et al., 2003; Perretti and Flower, 2004). Combined treatment with aspirin-triggered lipoxins and annexin-1-derived peptides significantly limits neutrophil infiltration (Perretti et al., 2002).

## Conclusion

Leukocyte integrins constitute major players in inflammatory cell recruitment into sites of infection and/or tissue injury. Preclinical studies in animal models of inflammation have indicated that integrin blockade or deficiency may significantly modulate disease initiation and/or progression. These findings led to the development of antibodies against leukocyte integrins, which are used in clinical practise for the treatment of patients with severe inflammatory disorders. However, patients treated with such agents may experience immunosuppression, and rarely, may suffer from severe infections, including PML. The identification of endogenous inhibitors, which homeostatically regulate leukocyte recruitment to inflammatory sites, may have fewer side effects and thus may prove beneficial as a therapeutic alternative in the treatment of inflammatory syndromes.

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## Abbreviations

<b>(BBB)</b>	Blood-brain barrier
<b>(CCL)</b>	CC-chemokine ligand
<b>(CAMs)</b>	cell adhesion molecules
<b>(CNS)</b>	central nervous system
<b>(CD)</b>	Crohn's disease
<b>(CXCL)</b>	CXC-chemokine ligand
<b>(DCs)</b>	dendritic cells
<b>(Del-1)</b>	Developmental endothelial locus-1
<b>(EGF)</b>	epidermal growth factor
<b>(EAE)</b>	experimental autoimmune encephalomyelitis
<b>(GDF)</b>	Growth Differentiation Factor
<b>(GALT)</b>	gut-associated lymphoid tissue
<b>(IBD)</b>	inflammatory bowel diseases

<b>(LAD-I)</b>	I leukocyte adhesion deficiency
<b>(ICAM-1)</b>	intercellular cell-adhesion molecule 1
<b>(JAM)</b>	Junctional Adhesion Molecule
<b>(LFA-1)</b>	lymphocyte function-associated antigen 1
<b>(Mac-1)</b>	macrophage-1 antigen
<b>(MAdCAM1)</b>	mucosal vascular addressin cell-adhesion molecule 1
<b>(MS)</b>	multiple sclerosis
<b>(PTX-3)</b>	pentraxin-3
<b>(PECAM-1)</b>	platelet/endothelial-cell adhesion molecule 1
<b>(PML)</b>	progressive multifocal leukoencephalopathy
<b>(PSGL-1)</b>	P-selectin glycoprotein ligand-1
<b>(RAGE)</b>	receptor advanced glycation end products
<b>(Treg)</b>	regulatory T cells
<b>(RA)</b>	rheumatoid arthritis
<b>(TLRs)</b>	Toll-like receptors
<b>(UC)</b>	ulcerative colitis
<b>(VCAM)</b>	vascular cell-adhesion molecule
<b>(VLA)</b>	very late antigen

## References

- Alon R, Feigelson SW. Chemokine-triggered leukocyte arrest: force-regulated bi-directional integrin activation in quantal adhesive contacts. *Curr Opin Cell Biol.* 2012; 24:670–676. [PubMed: 22770729]
- Alon R, Hammer DA, Springer TA. Lifetime of the P-selectin-carbohydrate bond and its response to tensile force in hydrodynamic flow. *Nature.* 1995; 374:539–542. [PubMed: 7535385]
- Altieri DC, Agbanyo FR, Plescia J, Ginsberg MH, Edgington TS, Plow EF. A unique recognition site mediates the interaction of fibrinogen with the leukocyte integrin Mac-1 (CD11b/CD18). *J Biol Chem.* 1990; 265:12119–12122. [PubMed: 1973686]
- Apostolaki M, Manoloukos M, Roulis M, Wurbel MA, Müller W, Papadakis KA, et al. Role of beta7 integrin and the chemokine/chemokine receptor pair CCL25/CCR9 in modeled TNF-dependent Crohn's disease. *Gastroenterology.* 2008; 134:2025–2035. [PubMed: 18439426]
- Archelos JJ, Jung S, Maurer M, Schmied M, Lassmann H, Tamatani T, et al. Inhibition of experimental autoimmune encephalomyelitis by an antibody to the intercellular adhesion molecule ICAM-1. *Ann Neurol.* 1993; 34:145–154. [PubMed: 7687838]
- Arnaout MA, Mahalingam B, Xiong JP. Integrin structure, allostery, and bidirectional signaling. *Annu Rev Cell Dev Biol.* 2005; 21:381–410. [PubMed: 16212500]
- Arnaout MA, Goodman SL, Xiong JP. Coming to grips with integrin binding to ligands. *Curr Opin Cell Biol.* 2002; 14:641–651. [PubMed: 12231361]
- Askari JA, Buckley PA, Mould AP, Humphries MJ. Linking integrin conformation to function. *J Cell Sci.* 2009; 122:165–70. [PubMed: 19118208]

- Bamias G, Clark DJ, Rivera-Nieves J. Leukocyte traffic blockade as a therapeutic strategy in inflammatory bowel disease. *Curr Drug Targets*. 2013; 14:1490–1500. [PubMed: 23621509]
- Barck KH, Lee WP, Diehl LJ, Ross J, Gribling P, Zhang Y, et al. Quantification of cortical bone loss and repair for therapeutic evaluation in collagen-induced arthritis, by micro-computed tomography and automated image analysis. *Arthritis Rheum*. 2004; 50:3377–3386. [PubMed: 15476252]
- Baron JL, Madri JA, Ruddle NH, Hashim G, Janeway CA Jr. Surface expression of  $\alpha 4$  integrin by CD4 T cells is required for their entry into brain parenchyma. *J Exp Med*. 1993; 177:57–68. [PubMed: 7678116]
- Bauer M, Brakebusch C, Coisne C, Sixt M, Wekerle H, Engelhardt B, et al. B1 integrins differentially control extravasation of inflammatory cell subsets into the CNS during autoimmunity. *Proc Natl Acad Sci U S A*. 2009; 106:1920–1925. [PubMed: 19179279]
- Berlin C, Berg EL, Briskin MJ, Andrew DP, Kilshaw PJ, Holzmann B, et al. Alpha 4 beta 7 integrin mediates lymphocyte binding to the mucosal vascular addressin MAdCAM-1. *Cell*. 1993; 74:185–195. [PubMed: 7687523]
- Bhat R, Steinman L. Innate and adaptive autoimmunity directed to the central nervous system. *Neuron*. 2009; 64:123–132. [PubMed: 19840554]
- Bixel G, Kloep S, Butz S, Petri B, Engelhardt B, Vestweber D. Mouse CD99 participates in T-cell recruitment into inflamed skin. *Blood*. 2004; 104:3205–3213. [PubMed: 15280198]
- Bixel MG, Li H, Petri B, Khandoga AG, Khandoga A, Zarbock A, et al. CD99 and CD99L2 act at the same site as, but independently of, PECAM-1 during leukocyte diapedesis. *Blood*. 2010; 116:1172–1184. [PubMed: 20479283]
- Bloomgren G, Richman S, Hotermans C, Subramanyam M, Goelz S, Natarajan A, et al. Risk of natalizumab-associated progressive multifocal leukoencephalopathy. *N Engl J Med*. 2012; 366:1870–1880. [PubMed: 22591293]
- Brocke S, Piercy C, Steinman L, Weissman IL, Veromaa T. Antibodies to CD44 and integrin  $\alpha 4$ , but not L-selectin, prevent central nervous system inflammation and experimental encephalomyelitis by blocking secondary leukocyte recruitment. *Proc Natl Acad Sci U S A*. 1999; 96:6896–6901. [PubMed: 10359810]
- Bullard DC, Hu X, Adams JE, Schoeb TR, Barnum SR. p150/95 (CD11c/CD18) expression is required for the development of experimental autoimmune encephalomyelitis. *Am J Pathol*. 2007a; 170:2001–2008. [PubMed: 17525267]
- Bullard DC, Hu X, Schoeb TR, Collins RG, Beaudet AL, Barnum SR. Intercellular adhesion molecule-1 expression is required on multiple cell types for the development of experimental autoimmune encephalomyelitis. *J Immunol*. 2007b; 178:851–857. [PubMed: 17202346]
- Bullard DC, Hu X, Schoeb TR, Axtell RC, Raman C, Barnum SR. Critical requirement of CD11b (Mac-1) on T cells and accessory cells for development of experimental autoimmune encephalomyelitis. *J Immunol*. 2005; 175:6327–6333. [PubMed: 16272284]
- Burger PC, Wagner DD. Platelet P-selectin facilitates atherosclerotic lesion development. *Blood*. 2003; 101:2661–2666. [PubMed: 12480714]
- Cai TQ, Wright SD. Human leukocyte elastase is an endogenous ligand for the integrin CR3 (CD11b/CD18, Mac-1,  $\alpha M \beta 2$ ) and modulates polymorphonuclear leukocyte adhesion. *J Exp Med*. 1996; 184:1213–1223. [PubMed: 8879192]
- Carman CV, Springer TA. Integrin avidity regulation: are changes in affinity and conformation underemphasized? *Curr Opin Cell Biol*. 2003; 15:547–556. [PubMed: 14519389]
- Cepek KL, Parker CM, Madara JL, Brenner MB. Integrin  $\alpha E \beta 7$  mediates adhesion of T lymphocytes to epithelial cells. *J Immunol*. 1993; 150:3459–3470. [PubMed: 8468482]
- Chamian F, Krueger JG. Psoriasis vulgaris: an interplay of T lymphocytes, dendritic cells, and inflammatory cytokines in pathogenesis. *Curr Opin Rheumatol*. 2004; 16:331–337. [PubMed: 15201593]
- Chamian F, Lowes MA, Lin SL, Lee E, Kikuchi T, Gilleaudeau P, et al. Alefacept reduces infiltrating T cells, activated dendritic cells, and inflammatory genes in psoriasis vulgaris. *Proc Natl Acad Sci U S A*. 2005; 102:2075–2080. [PubMed: 15671179]

- Chan JR, Hyduk SJ, Cybulsky MI. Chemoattractants induce a rapid and transient upregulation of monocyte alpha4 integrin affinity for vascular cell adhesion molecule 1 which mediates arrest: an early step in the process of emigration. *J Exp Med*. 2001; 193:1149–1158. [PubMed: 11369786]
- Chavakis E, Choi EY, Chavakis T. Novel aspects in the regulation of the leukocyte adhesion cascade. *Thromb Haemost*. 2009; 102:191–197. [PubMed: 19652868]
- Chavakis T, Kanse SM, Pixley RA, May AE, Isordia-Salas I, Colman RW, et al. Regulation of leukocyte recruitment by polypeptides derived from high molecular weight kininogen. *FASEB J*. 2001; 15:2365–2376. [PubMed: 11689462]
- Chavakis T, Bierhaus A, Al-Fakhri N, Schneider D, Witte S, Linn T, et al. The pattern recognition receptor (RAGE) is a counterreceptor for leukocyte integrins: a novel pathway for inflammatory cell recruitment. *J Exp Med*. 2003a; 198:1507–1515. [PubMed: 14623906]
- Chavakis T, Santoso S, Clemetson KJ, Sachs UJ, Isordia-Salas I, Pixley RA, et al. High molecular weight kininogen regulates platelet-leukocyte interactions by bridging Mac-1 and glycoprotein Ib. *J Biol Chem*. 2003b; 278:45375–45381. [PubMed: 12952972]
- Chavakis T, May AE, Preissner KT, Kanse SM. Molecular mechanisms of zinc-dependent leukocyte adhesion involving the urokinase receptor and beta2-integrins. *Blood*. 1999; 93:2976–2983. [PubMed: 10216093]
- Chavakis T. Leukocyte recruitment in inflammation and novel endogenous negative regulators thereof. *Eur J Clin Invest*. 2012; 42:686–691. [PubMed: 22577952]
- Chavakis T, Athanasopoulos A, Rhee JS, Orlova V, Schmidt-Wöll T, Bierhaus A, et al. Angiostatin is a novel anti-inflammatory factor by inhibiting leukocyte recruitment. *Blood*. 2005; 105:1036–1043. [PubMed: 15383457]
- Chen CH, Kularatna G, Stone CD, Gutierrez AM, Dassopoulos T. Clinical experience of natalizumab in Crohn's disease patients in a restricted distribution program. *Ann Gastroenterol*. 2013; 26:189–190. [PubMed: 24714228]
- Choi EY, Lim HJ, Neuwirth A, Economopoulou M, Chatzigeorgiou A, Chung KJ, et al. Developmental endothelial locus-1 (Del-1) is a homeostatic factor in the central nervous system limiting neuroinflammation and demyelination. *Mol Psychiatry*. 2014 2014, in press.
- Choi EY, Santoso S, Chavakis T. Mechanisms of neutrophil transendothelial migration. *Front Biosci*. 2009; 14:1596–1605.
- Choi EY, Orlova VV, Fagerholm SC, Nurmi SM, Zhang L, Ballantyne CM, et al. Regulation of LFA-1-dependent inflammatory cell recruitment by Cbl-b and 14-3-3 proteins. *Blood*. 2008a; 111:3607–3614. [PubMed: 18239087]
- Choi EY, Chavakis E, Czabanka MA, Langer HF, Fraemohs L, Economopoulou M, et al. Del-1, an endogenous leukocyte-endothelial adhesion inhibitor, limits inflammatory cell recruitment. *Science*. 2008b; 322:1101–1104. [PubMed: 19008446]
- Choy E. Understanding the dynamics: pathways involved in the pathogenesis of rheumatoid arthritis. *Rheumatology (Oxford)*. 2012; 51(Suppl 5):v3–11. [PubMed: 22718924]
- Chung KJ, Mitroulis I, Wiessner JR, Zheng YY, Siegert G, Sperandio M, et al. A novel pathway of rapid TLR-triggered activation of integrin-dependent leukocyte adhesion that requires Rap1 GTPase. *Mol Biol Cell*. 2014 pii: mbc.E14-04-0867.
- Coisne C, Mao W, Engelhardt B. Cutting edge: Natalizumab blocks adhesion but not initial contact of human T cells to the blood-brain barrier in vivo in an animal model of multiple sclerosis. *J Immunol*. 2009; 182:5909–5913. [PubMed: 19414741]
- Confavreux C, Vukusic S, Moreau T, Adeleine P. Relapses and progression of disability in multiple sclerosis. *N Engl J Med*. 2000; 343:1430–1438. [PubMed: 11078767]
- Confavreux C, Vukusic S. Natural history of multiple sclerosis: a unifying concept. *Brain*. 2006; 129:606–616. [PubMed: 16415308]
- Conrad C, Boyman O, Tonel G, Tun-Kyi A, Laggner U, de Fougères A, et al. Alpha1beta1 integrin is crucial for accumulation of epidermal T cells and the development of psoriasis. *Nat Med*. 2007; 13:836–842. [PubMed: 17603494]
- Cooper D, Norling LV, Perretti M. Novel insights into the inhibitory effects of Galectin-1 on neutrophil recruitment under flow. *J Leukoc Biol*. 2008; 83:1459–1466. [PubMed: 18372340]

- Dangerfield J, Larbi KY, Huang MT, Dewar A, Nourshargh S. PECAM-1 (CD31) homophilic interaction up-regulates alpha6beta1 on transmigrated neutrophils in vivo and plays a functional role in the ability of alpha6 integrins to mediate leukocyte migration through the perivascular basement membrane. *J Exp Med*. 2002; 196:1201–1211. [PubMed: 12417630]
- Dasgupta SK, Le A, Chavakis T, Rumbaut RE, Thiagarajan P. Developmental endothelial locus-1 (Del-1) mediates clearance of platelet microparticles by the endothelium. *Circulation*. 2012; 125:1664–1672. [PubMed: 22388320]
- Dassopoulos T, Sultan S, Falck-Ytter YT, Inadomi JM, Hanauer SB. American Gastroenterological Association Institute technical review on the use of thiopurines, methotrexate, and anti-TNF- $\alpha$  biologic drugs for the induction and maintenance of remission in inflammatory Crohn's disease. *Gastroenterology*. 2013; 145:1464–1478. [PubMed: 24267475]
- Deban L, Russo RC, Sironi M, Moalli F, Scanziani M, Zambelli V, et al. Regulation of leukocyte recruitment by the long pentraxin PTX3. *Nat Immunol*. 2010; 11:328–334. [PubMed: 20208538]
- Dedrick RL, Walicke P, Garovoy M. Anti-adhesion antibodies efalizumab, a humanized anti-CD11a monoclonal antibody. *Transpl Immunol*. 2002; 9:181–186. [PubMed: 12180828]
- del Pilar Martin M, Cravens PD, Winger R, Frohman EM, Racke MK, Eagar TN, et al. Decrease in the numbers of dendritic cells and CD4 T cells in cerebral perivascular spaces due to natalizumab. *Arch Neurol*. 2008; 65:1596–1603. [PubMed: 18852339]
- Diamond MS, Alon R, Parkos CA, Quinn MT, Springer TA. Heparin is an adhesive ligand for the leukocyte integrin Mac-1 (CD11b/CD18). *J Cell Biol*. 1995; 130:1473–1482. [PubMed: 7559767]
- Döring A, Pfeiffer F, Meier M, Dehouck B, et al. TET inducible expression of the  $\alpha 4\beta 7$ -integrin ligand MAdCAM-1 on the blood-brain barrier does not influence the immunopathogenesis of experimental autoimmune encephalomyelitis. *Eur J Immunol*. 2011; 41:813–821. [PubMed: 21341265]
- Dransfield I, Cabanas C, Craig A, Hogg N. Divalent cation regulation of the function of the leukocyte integrin LFA-1. *J Cell Biol*. 1992; 116:219–226. [PubMed: 1346139]
- Dugger KJ, Zinn KR, Weaver C, Bullard DC, Barnum SR. Effector and suppressor roles for LFA-1 during the development of experimental autoimmune encephalomyelitis. *J Neuroimmunol*. 2009; 206:22–27. [PubMed: 19010554]
- Dunne JL, Ballantyne CM, Beaudet AL, Ley K. Control of leukocyte rolling velocity in TNF- $\alpha$ -induced inflammation by LFA-1 and Mac-1. *Blood*. 2002; 99:336–341. [PubMed: 11756189]
- Dupuy AG, Caron E. Integrin-dependent phagocytosis: spreading from microadhesion to new concepts. *J Cell Sci*. 2008; 121:1773–1783. [PubMed: 18492791]
- Ehlers R, Ustinov V, Chen Z, Zhang X, Rao R, Luscinskas FW, et al. Targeting platelet-leukocyte interactions: identification of the integrin Mac-1 binding site for the platelet counter receptor glycoprotein Iba1. *J Exp Med*. 2003; 198:1077–1088. [PubMed: 14530377]
- El Kebir D, Jozsef L, Filep JG. Opposing regulation of neutrophil apoptosis through the formyl peptide receptor-like 1/lipoxin A4 receptor: implications for resolution of inflammation. *J Leukoc Biol*. 2008; 84:600–606. [PubMed: 18495783]
- Engelhardt B, Laschinger M, Schulz M, Samulowitz U, Vestweber D, Hoch G. The development of experimental autoimmune encephalomyelitis in the mouse requires  $\alpha 4$ -integrin but not  $\alpha 4\beta 7$ -integrin. *J Clin Invest*. 1998; 102:2096–2105. [PubMed: 9854045]
- Eskan MA, Jotwani R, Abe T, Chmellar J, Lim JH, Liang S, et al. The leukocyte integrin antagonist Del-1 inhibits IL-17-mediated inflammatory bone loss. *Nat Immunol*. 2012; 13:465–473. [PubMed: 22447028]
- Evans R, Lellouch AC, Svensson L, McDowall A, Hogg N. The integrin LFA-1 signals through ZAP-70 to regulate expression of high-affinity LFA-1 on T lymphocytes. *Blood*. 2011; 117:3331–3342. [PubMed: 21200022]
- Fagerholm S, Morrice N, Gahmberg CG, Cohen P. Phosphorylation of the cytoplasmic domain of the integrin CD18 chain by protein kinase C isoforms in leukocytes. *J Biol Chem*. 2002; 277:1728–1738. [PubMed: 11700305]
- Feagan BG, Rutgeerts P, Sands BE, Hanauer S, Colombel JF, Sandborn WJ, et al. Vedolizumab as induction and maintenance therapy for ulcerative colitis. *N Engl J Med*. 2013; 369:699–710. [PubMed: 23964932]

- Flick MJ, Du X, Witte DP, Jirousková M, Soloviev DA, Busuttill SJ, et al. Leukocyte engagement of fibrin(ogen) via the integrin receptor  $\alpha$ 5 $\beta$ 1/Mac-1 is critical for host inflammatory response in vivo. *J Clin Invest*. 2004; 113:1596–1606. [PubMed: 15173886]
- Frommhold D, Kamphues A, Hepper I, Pruenster M, Lukic IK, Socher I, et al. RAGE and ICAM-1 cooperate in mediating leukocyte recruitment during acute inflammation in vivo. *Blood*. 2010; 116:841–849. [PubMed: 20407037]
- Gahmberg CG, Fagerholm SC, Nurmi SM, Chavakis T, Marchesan S, Grönholm M. Regulation of integrin activity and signalling. *Biochim Biophys Acta*. 2009; 1790:431–44. [PubMed: 19289150]
- Gailit J, Ruoslahti E. Regulation of the fibronectin receptor affinity by divalent cations. *J Biol Chem*. 1988; 263:12927–12932. [PubMed: 2458338]
- Gan Y, Liu R, Wu W, Bompreszi R, Shi FD. Antibody to  $\alpha$ 4 integrin suppresses natural killer cells infiltration in central nervous system in experimental autoimmune encephalomyelitis. *J Neuroimmunol*. 2012; 247:9–15. [PubMed: 22503411]
- Gatzka M, Hainzl A, Peters T, Singh K, Tasdogan A, Wlaschek M, et al. Reduction of CD18 promotes expansion of inflammatory  $\gamma\delta$  T cells collaborating with CD4+ T cells in chronic murine psoriasisform dermatitis. *J Immunol*. 2013; 191:5477–5488. [PubMed: 24190659]
- Gerke V, Creutz CE, Moss SE. Annexins: linking Ca<sup>2+</sup> signalling to membrane dynamics. *Nat Rev Mol Cell Biol*. 2005; 6:449–461. [PubMed: 15928709]
- Ghosh S, Goldin E, Gordon FH, Malchow HA, Rask-Madsen J, Rutgeerts P, et al. Natalizumab for active Crohn's disease. *N Engl J Med*. 2003; 348:24–32. [PubMed: 12510039]
- Giagulli C, Ottoboni L, Cavegion E, Rossi B, Lowell C, Constantin G, et al. The Src family kinases Hck and Fgr are dispensable for inside-out, chemoattractant-induced signaling regulating beta 2 integrin affinity and valency in neutrophils, but are required for beta 2 integrin-mediated outside-in signaling involved in sustained adhesion. *J Immunol*. 2006; 177:604–611. [PubMed: 16785558]
- Glatigny S, Duhon R, Oukka M, Bettelli E. Cutting edge: loss of  $\alpha$ 4 integrin expression differentially affects the homing of Th1 and Th17 cells. *J Immunol*. 2011; 187:6176–6179. [PubMed: 22084440]
- Goldfinger LE, Han J, Kiosses WB, Howe AK, Ginsberg MH. Spatial restriction of alpha4 integrin phosphorylation regulates lamellipodial stability and alpha4beta1-dependent cell migration. *J Cell Biol*. 2003; 162:731–741. [PubMed: 12913113]
- Gordon KB, Papp KA, Hamilton TK, Walicke PA, Dummer W, Li N, et al. Efalizumab for patients with moderate to severe plaque psoriasis: a randomized controlled trial. *JAMA*. 2003; 290:3073–3080. [PubMed: 14679270]
- Gordon EJ, Myers KJ, Dougherty JP, Rosen H, Ron Y. Both anti-CD11a (LFA-1) and anti-CD11b (MAC-1) therapy delay the onset and diminish the severity of experimental autoimmune encephalomyelitis. *J Neuroimmunol*. 1995; 62:153–160. [PubMed: 7499503]
- Gorfu G, Rivera-Nieves J, Hoang S, Abbott DW, Arbenz-Smith K, Azar DW, et al. Beta7 integrin deficiency suppresses B cell homing and attenuates chronic ileitis in SAMPI/YitFc mice. *J Immunol*. 2010; 185:5561–5568. [PubMed: 20926792]
- Gorfu G, Rivera-Nieves J, Ley K. Role of beta7 integrins in intestinal lymphocyte homing and retention. *Curr Mol Med*. 2009; 9:836–850. [PubMed: 19860663]
- Gottlieb AB, Krueger JG, Wittkowski K, Dedrick R, Walicke PA, Garovoy M. Psoriasis as a model for T-cell-mediated disease: immunobiologic and clinical effects of treatment with multiple doses of efalizumab, an anti-CD11a antibody. *Arch Dermatol*. 2002; 138:591–600. [PubMed: 12020219]
- Goverman J. Autoimmune T cell responses in the central nervous system. *Nat Rev Immunol*. 2009; 9:393–407. [PubMed: 19444307]
- Gültner S, Kuhlmann T, Hesse A, Weber JP, Riemer C, Baier M, et al. Reduced Treg frequency in LFA-1 deficient mice allows enhanced T effector differentiation and pathology in EAE. *Eur J Immunol*. 2010; 40:3403–3412. [PubMed: 21108463]
- Hadley GA, Higgins JM. Integrin  $\alpha$ 5 $\beta$ 1: Molecular Features and Functional Significance in the Immune System. *Adv Exp Med Biol*. 2014; 819:97–110. [PubMed: 25023170]
- Hafezi-Moghadam A, Ley K. Relevance of L-selectin shedding for leukocyte rolling in vivo. *J Exp Med*. 1999; 189:939–948. [PubMed: 10075977]



- Hajishengallis G, Chavakis T. Endogenous modulators of inflammatory cell recruitment. *Trends Immunol.* 2013; 34:1–6. [PubMed: 22951309]
- Hamann A, Andrew DP, Jablonski-Westrich D, Holzmann B, Butcher EC. Role of alpha4-integrins in lymphocyte homing to mucosal tissues in vivo. *J Immunol.* 1994; 152:3282–3293. [PubMed: 7511642]
- Hannon R, Croxtall JD, Getting SJ, Roviezzo F, Yona S, Paul-Clark MJ, et al. Aberrant inflammation and resistance to glucocorticoids in annexin 1<sup>-/-</sup> mouse. *FASEB J.* 2003; 17:253–255. [PubMed: 12475898]
- Harokopakis E, Albzreh MH, Martin MH, Hajishengallis G. TLR2 transmodulates monocyte adhesion and transmigration via Rac1- and PI3K-mediated inside-out signaling in response to *Porphyromonas gingivalis* fimbriae. *J Immunol.* 2006; 176:7645–7656. [PubMed: 16751412]
- Harokopakis E, Hajishengallis G. Integrin activation by bacterial fimbriae through a pathway involving CD14, Toll-like receptor 2, and phosphatidylinositol-3-kinase. *Eur J Immunol.* 2005; 35:1201–1210. [PubMed: 15739163]
- Henderson RB, Lim LH, Tessier PA, Gavins FN, Mathies M, Perretti M, et al. The use of lymphocyte function-associated antigen (LFA)-1-deficient mice to determine the role of LFA-1, Mac-1, and alpha4 integrin in the inflammatory response of neutrophils. *J Exp Med.* 2001; 194:219–226. [PubMed: 11457896]
- Herter J, Zarbock A. Integrin regulation during leukocyte recruitment. *J Immunol.* 2013; 190:4451–4457. [PubMed: 23606722]
- Hidai C, Kawana M, Kitano H, Kokubun S. Discoidin domain of Del1 protein contributes to its deposition in the extracellular matrix. *Cell Tissue Res.* 2007; 330:83–95. [PubMed: 17701220]
- Hidalgo A, Peired AJ, Wild MK, Vestweber D, Frenette PS. Complete identification of E-selectin ligands on neutrophils reveals distinct functions of PSGL-1, ESL-1, and CD44. *Immunity.* 2007; 26:477–489. [PubMed: 17442598]
- Hogg N, Patzak I, Willenbrock F. The insider's guide to leukocyte integrin signalling and function. *Nat Rev Immunol.* 2011; 11:416–426. [PubMed: 21597477]
- Huang MT, Larbi KY, Scheiermann C, Woodfin A, Gerwin N, Haskard DO, et al. ICAM-2 mediates neutrophil transmigration in vivo: evidence for stimulus specificity and a role in PECAM-1-independent transmigration. *Blood.* 2006; 107:4721–4727. [PubMed: 16469869]
- Huo Y, Schober A, Forlow SB, Smith DF, Hyman MC, Jung S, et al. Circulating activated platelets exacerbate atherosclerosis in mice deficient in apolipoprotein E. *Nat Med.* 2003; 9:61–67. [PubMed: 12483207]
- Hyduk SJ, Chan JR, Duffy ST, Chen M, Peterson MD, Waddell TK, et al. Phospholipase C, calcium, and calmodulin are critical for alpha4beta1 integrin affinity up-regulation and monocyte arrest triggered by chemoattractants. *Blood.* 2007; 109:176–184. [PubMed: 16960156]
- Hynes RO. Integrins: bidirectional, allosteric signaling machines. *Cell.* 2002; 110:673–687. [PubMed: 12297042]
- Hyun YM, Chung HL, McGrath JL, Waugh RE, Kim M. Activated integrin VLA-4 localizes to the lamellipodia and mediates T cell migration on VCAM-1. *J Immunol.* 2009; 183:359–369. [PubMed: 19542447]
- Inforzato A, Reading PC, Barbati E, Bottazzi B, Garlanda C, Mantovani A. The “sweet” side of a long pentraxin: how glycosylation affects PTX3 functions in innate immunity and inflammation. *Front Immunol.* 2012; 3:407. [PubMed: 23316195]
- Issekutz AC, Issekutz TB. Monocyte migration to arthritis in the rat utilizes both CD11/CD18 and very late activation antigen 4 integrin mechanisms. *J Exp Med.* 1995; 181:1197–1203. [PubMed: 7532681]
- Issekutz AC, Issekutz TB. A major portion of polymorphonuclear leukocyte and T lymphocyte migration to arthritic joints in the rat is via LFA-1/MAC-1-independent mechanisms. *Clin Immunol Immunopathol.* 1993; 67:257–263. [PubMed: 8500273]
- Issekutz AC, Issekutz TB. T lymphocyte migration to arthritic joints and dermal inflammation in the rat: differing migration patterns and the involvement of VLA-4. *Clin Immunol Immunopathol.* 1991; 61:436–447. [PubMed: 1934631]

- Issekutz AC, Nakazato S, Issekutz TB. Differential roles of VLA-4(CD49d/CD29) and LFA-1(CD11a/CD18) integrins and E- and P-selectin during developing and established active or adoptively transferred adjuvant arthritis in the rat. *Immunol Cell Biol.* 2003; 81:397–408. [PubMed: 12969328]
- Jäger A, Dardalhon V, Sobel RA, Bettelli E, Kuchroo VK. Th1, Th17, and Th9 effector cells induce experimental autoimmune encephalomyelitis with different pathological phenotypes. *J Immunol.* 2009; 183:7169–7177. [PubMed: 19890056]
- Jain P, Coisne C, Enzmann G, Rottapel R, Engelhardt B. Alpha4beta1 integrin mediates the recruitment of immature dendritic cells across the blood-brain barrier during experimental autoimmune encephalomyelitis. *J Immunol.* 2010; 184:7196–7206. [PubMed: 20483748]
- Kanczkowski W, Chatzigeorgiou A, Grossklaus S, Sprott D, Bornstein SR, Chavakis T. Role of the endothelial-derived endogenous anti-inflammatory factor Del-1 in inflammation-mediated adrenal gland dysfunction. *Endocrinology.* 2013; 154:1181–1189. [PubMed: 23364949]
- Kanda H, Newton R, Klein R, Morita Y, Gunn MD, Rosen SD. Autotaxin, an ectoenzyme that produces lysophosphatidic acid, promotes the entry of lymphocytes into secondary lymphoid organs. *Nat Immunol.* 2008; 9:415–423. [PubMed: 18327261]
- Kang YY, Kim DY, Lee SH, Choi EY. Deficiency of developmental endothelial locus-1 (Del-1) aggravates bleomycin-induced pulmonary fibrosis in mice. *Biochem Biophys Res Commun.* 2014; 445:369–374. [PubMed: 24525119]
- Kansas GS. Selectins and their ligands: current concepts and controversies. *Blood.* 1996; 88:3259–3287. [PubMed: 8896391]
- Kanczkowski W, Chatzigeorgiou A, Grossklaus S, Sprott D, Bornstein SR, Chavakis T. Role of the endothelial-derived endogenous anti-inflammatory factor Del-1 in inflammation-mediated adrenal gland dysfunction. *Endocrinology.* 2013; 154:1181–1189. [PubMed: 23364949]
- Kebir H, Kreymborg K, Ifergan I, Dodelet-Devillers A, Cayrol R, Bernard M, et al. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nat Med.* 2007; 13:1173–1175. [PubMed: 17828272]
- Keiper T, Santoso S, Nawroth PP, Orlova V, Chavakis T. The role of junctional adhesion molecules in cell-cell interactions. *Histol Histopathol.* 2005; 20:197–203. [PubMed: 15578438]
- Kempf T, Eden M, Strelau J, Naguib M, Willenbockel C, Tongers J, et al. The transforming growth factor-beta superfamily member growth-differentiation factor-15 protects the heart from ischemia/reperfusion injury. *Circ Res.* 2006; 98:351–360. [PubMed: 16397141]
- Kempf T, Zarbock A, Widera C, Butz S, Stadtmann A, Rossaint J, et al. GDF-15 is an inhibitor of leukocyte integrin activation required for survival after myocardial infarction in mice. *Nat Med.* 2011; 17:581–588. [PubMed: 21516086]
- Kerfoot SM, Kubes P. Overlapping roles of P-selectin and alpha 4 integrin to recruit leukocytes to the central nervous system in experimental autoimmune encephalomyelitis. *J Immunol.* 2002; 169:1000–1006. [PubMed: 12097407]
- Kinashi T. Intracellular signalling controlling integrin activation in lymphocytes. *Nat Rev Immunol.* 2005; 5:546–559. [PubMed: 15965491]
- Kinbara K, Goldfinger LE, Hansen M, Chou FL, Ginsberg MH. Ras GTPases: integrins' friends or foes? *Nat Rev Mol Cell Biol.* 2003; 4:767–776. [PubMed: 14570053]
- Kishimoto TK, Jutila MA, Berg EL, Butcher EC. Neutrophil Mac-1 and MEL-14 adhesion proteins inversely regulated by chemotactic factors. *Science.* 1989; 245:1238–1241. [PubMed: 2551036]
- Kivisäkk P, Healy BC, Viglietta V, Quintana FJ, Hootstein MA, Weiner HL, et al. Natalizumab treatment is associated with peripheral sequestration of proinflammatory T cells. *Neurology.* 2009; 72:1922–1930. [PubMed: 19487650]
- Kleinschek MA, Owyang AM, Joyce-Shaikh B, Langrish CL, Chen Y, Gorman DM, et al. IL-25 regulates Th17 function in autoimmune inflammation. *J Exp Med.* 2007; 204:161–170. [PubMed: 17200411]
- Koboziev I, Karlsson F, Grisham MB. Gut-associated lymphoid tissue, T cell trafficking, and chronic intestinal inflammation. *Ann N Y Acad Sci.* 2010; 1207(Suppl 1):E86–93. [PubMed: 20961311]

- Kolanus W, Nagel W, Schiller B, Zeitlmann L, Godar S, Stockinger H, et al. Alpha L beta 2 integrin/LFA-1 binding to ICAM-1 induced by cytohesin-1, a cytoplasmic regulatory molecule. *Cell*. 1996; 86:233–242. [PubMed: 8706128]
- Kontoyiannis D, Boulougouris G, Manoloukos M, Armaka M, Apostolaki M, Pizarro T, et al. Genetic dissection of the cellular pathways and signaling mechanisms in modeled tumor necrosis factor-induced Crohn's-like inflammatory bowel disease. *J Exp Med*. 2002; 196:1563–1574. [PubMed: 12486099]
- Korn T. Pathophysiology of multiple sclerosis. *J Neurol*. 2008; 255(Suppl 6):2–6. [PubMed: 19300953]
- Krueger JG, Fretzin S, Suárez-Fariñas M, Haslett PA, Phipps KM, Cameron GS, et al. IL-17A is essential for cell activation and inflammatory gene circuits in subjects with psoriasis. *J Allergy Clin Immunol*. 2012; 130:145–154. [PubMed: 22677045]
- Kunkel EJ, Chomas JE, Ley K. Role of primary and secondary capture for leukocyte accumulation in vivo. *Circ Res*. 1998; 82:30–38. [PubMed: 9440702]
- Kunkel EJ, Ramos CL, Steeber DA, Muller W, Wagner N, Tedder TF, et al. The roles of L-selectin,  $\beta$  7 integrins and P-selectin in leukocyte rolling and adhesion in high endothelial venules of Peyer's patches. *J Immunol*. 1998; 161:2449–2456. [PubMed: 9725243]
- Kuwano Y, Spelten O, Zhang H, Ley K, Zarbock A. Rolling on E- or P-selectin induces the extended but not high-affinity conformation of LFA-1 in neutrophils. *Blood*. 2010; 116:617–624. [PubMed: 20445017]
- Lafuente E, Boussiotis VA. Rap1 regulation of RIAM and cell adhesion. *Methods Enzymol*. 2006; 407:345–358. [PubMed: 16757337]
- Lamagna C, Meda P, Mandicourt G, Brown J, Gilbert RJ, Jones EY, et al. Dual interaction of JAM-C with JAM-B and alpha(M)beta2 integrin: function in junctional complexes and leukocyte adhesion. *Mol Biol Cell*. 2005; 16:4992–5003. [PubMed: 16093349]
- Langer HF, Chavakis T. Leukocyte-endothelial interactions in inflammation. *J Cell Mol Med*. 2009; 13:1211–1220. [PubMed: 19538472]
- Langer HF, Choi EY, Zhou H, Schleicher R, Chung KJ, Tang Z, et al. Platelets contribute to the pathogenesis of experimental autoimmune encephalomyelitis. *Circ Res*. 2012; 110:1202–1210. [PubMed: 22456181]
- Larsen E, Celi A, Gilbert GE, Furie BC, Erban JK, Bonfanti R, et al. PADGEM protein: a receptor that mediates the interaction of activated platelets with neutrophils and monocytes. *Cell*. 1989; 59:305–312. [PubMed: 2478294]
- Laschinger M, Engelhardt B. Interaction of alpha4-integrin with VCAM-1 is involved in adhesion of encephalitogenic T cell blasts to brain endothelium but not in their transendothelial migration in vitro. *J Neuroimmunol*. 2000; 102:32–43. [PubMed: 10626664]
- Laschinger M, Vajkoczy P, Engelhardt B. Encephalitogenic T cells use LFA-1 for transendothelial migration but not during capture and initial adhesion strengthening in healthy spinal cord microvessels in vivo. *Eur J Immunol*. 2002; 32:3598–3606. [PubMed: 12516546]
- Lassmann H, van Horssen J, Mahad D. Progressive multiple sclerosis: pathology and pathogenesis. *Nat Rev Neurol*. 2012; 8:647–656. [PubMed: 23007702]
- Lebwohl M, Tyring SK, Hamilton TK, Toth D, Glazer S, Tawfik NH, et al. A novel targeted T-cell modulator, efalizumab, for plaque psoriasis. *N Engl J Med*. 2003; 349:2004–2013. [PubMed: 14627785]
- Leon F, Contractor N, Fuss I, Marth T, Lahey E, Iwaki S, et al. Antibodies to complement receptor 3 treat established inflammation in murine models of colitis and a novel model of psoriasiform dermatitis. *J Immunol*. 2006; 177:6974–6982. [PubMed: 17082612]
- Leonardi C, Matheson R, Zachariae C, Cameron G, Li L, Edson-Heredia E, et al. Anti-interleukin-17 monoclonal antibody ixekizumab in chronic plaque psoriasis. *N Engl J Med*. 2012; 366:1190–1199. [PubMed: 22455413]
- Leonardi CL, Papp KA, Gordon KB, Menter A, Feldman SR, Caro I, et al. Extended efalizumab therapy improves chronic plaque psoriasis: results from a randomized phase III trial. *J Am Acad Dermatol*. 2005; 52:425–433. [PubMed: 15761420]

- Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat Rev Immunol.* 2007; 7:678–689. [PubMed: 17717539]
- Lin KK, Mahadevan U. Etrolizumab: anti- $\beta$ 7-a novel therapy for ulcerative colitis. *Gastroenterology.* 2014; 146:307–309. [PubMed: 24269566]
- Liu S, Thomas SM, Woodside DG, Rose DM, Kiosses WB, Pfaff M, et al. Binding of paxillin to  $\alpha$ 4 integrins modifies integrin-dependent biological responses. *Nature.* 1999; 402:676–681. [PubMed: 10604475]
- Lowes MA, Kikuchi T, Fuentes-Duculan J, Cardinale I, Zaba LC, Haider AS, et al. Psoriasis vulgaris lesions contain discrete populations of Th1 and Th17 T cells. *J Invest Dermatol.* 2008; 128:1207–1211. [PubMed: 18200064]
- Lowes MA, Chamian F, Abello MV, Fuentes-Duculan J, Lin SL, Nussbaum R, et al. Increase in TNF-alpha and inducible nitric oxide synthase-expressing dendritic cells in psoriasis and reduction with efalizumab (anti-CD11a). *Proc Natl Acad Sci U S A.* 2005; 102:19057–19062. [PubMed: 16380428]
- Ludviksson BR, Strober W, Nishikomori R, Hasan SK, Ehrhardt RO. Administration of mAb against  $\alpha$ E $\beta$ 7 prevents and ameliorates immunization-induced colitis in IL-2<sup>-/-</sup> mice. *J Immunol.* 1999; 162:4975. [PubMed: 10202045]
- Mace EM, Zhang J, Siminovitch KA, Takei F. Elucidation of the integrin LFA-1-mediated signaling pathway of actin polarization in natural killer cells. *Blood.* 2010; 116:1272–1279. [PubMed: 20472831]
- Maina V, Cotena A, Doni A, Nebuloni M, Pasqualini F, Milner CM, et al. Coregulation in human leukocytes of the long pentraxin PTX3 and TSG-6. *J Leukoc Biol.* 2009; 86:123–132. [PubMed: 19389798]
- Mamdouh Z, Chen X, Pierini LM, Maxfield FR, Muller WA. Targeted recycling of PECAM from endothelial surface-connected compartments during diapedesis. *Nature.* 2003; 421:748–53. [PubMed: 12610627]
- Martin DA, Towne JE, Kricorian G, Klekotka P, Gudjonsson JE, Krueger JG, et al. The emerging role of IL-17 in the pathogenesis of psoriasis: preclinical and clinical findings. *J Invest Dermatol.* 2013; 133:17–26. [PubMed: 22673731]
- Massberg S, Brand K, Grüner S, Page S, Müller E, Müller I, et al. A critical role of platelet adhesion in the initiation of atherosclerotic lesion formation. *J Exp Med.* 2002; 196:887–896. [PubMed: 12370251]
- Massberg S, Schurzinger K, Lorenz M, Konrad I, Schulz C, Plesnila N, et al. Platelet adhesion via glycoprotein IIb integrin is critical for atheroprotection and focal cerebral ischemia: an in vivo study in mice lacking glycoprotein IIb. *Circulation.* 2005; 112:1180–1188. [PubMed: 16103235]
- McEver RP. Selectins: lectins that initiate cell adhesion under flow. *Curr Opin Cell Biol.* 2002; 14:581–586. [PubMed: 12231353]
- McEver RP, Cummings RD. Role of PSGL-1 binding to selectins in leukocyte recruitment. *J Clin Invest.* 1997; 100:S97–103. [PubMed: 9413410]
- Micklem KJ, Sim RB. Isolation of complement-fragment-iC3b-binding proteins by affinity chromatography. *Biochem J.* 1985; 231:233–236. [PubMed: 4062888]
- Minagar A, Alexander JS. Blood-brain barrier disruption in multiple sclerosis. *Mult Scler.* 2003; 9:540–549. [PubMed: 14664465]
- Mitroulis I, Kang YY, Gahmberg CG, Siebert G, Hajishengallis G, Chavakis T, et al. Developmental endothelial locus-1 attenuates complement-dependent phagocytosis through inhibition of Mac-1-integrin. *Thromb Haemost.* 2014; 111:1004–1006. [PubMed: 24352615]
- Mocsai A, Abram CL, Jakus Z, Hu Y, Lanier LL, Lowell CA. Integrin signaling in neutrophils and macrophages uses adaptors containing immunoreceptor tyrosine-based activation motifs. *Nat Immunol.* 2006; 7:1326–1333. [PubMed: 17086186]
- Moore KL, Stults NL, Diaz S, Smith DF, Cummings RD, Varki A, et al. Identification of a specific glycoprotein ligand for P-selectin (CD62) on myeloid cells. *J Cell Biol.* 1992; 118:445–456. [PubMed: 1378449]

- Morrissey SP, Deichmann R, Syha J, Simonis C, Zettl U, Archelos JJ, et al. Partial inhibition of AT-EAE by an antibody to ICAM-1: clinico-histological and MRI studies. *J Neuroimmunol.* 1996; 69:85–93. [PubMed: 8823379]
- Moser M, Legate KR, Zent R, Fässler R. The tail of integrins, talin, and kindlins. *Science.* 2009; 324:895–899. [PubMed: 19443776]
- Moutsopoulos NM, Konkel J, Sarmadi M, Eskan MA, Wild T, Dutzan N, et al. Defective neutrophil recruitment in leukocyte adhesion deficiency type I disease causes local IL-17-driven inflammatory bone loss. *Sci Transl Med.* 2014; 6:229ra40.
- Mueller H, Stadtmann A, Van Aken H, Hirsch E, Wang D, Ley K, et al. Tyrosine kinase Btk regulates E-selectin-mediated integrin activation and neutrophil recruitment by controlling phospholipase C (PLC)  $\gamma$ 2 and PI3K pathways. *Blood.* 2010; 115:3118–3127. [PubMed: 20167705]
- Muller WA. Leukocyte-endothelial-cell interactions in leukocyte transmigration and the inflammatory response. *Trends Immunol.* 2003; 24:327–334. [PubMed: 12810109]
- Muller WA. How endothelial cells regulate transmigration of leukocytes in the inflammatory response. *Am J Pathol.* 2014; 184:886–896. [PubMed: 24655376]
- Myers WA, Najarian D, Gottlieb AB. New-onset, debilitating arthritis in psoriasis patients receiving efalizumab. *J Dermatolog Treat.* 2006; 17:353–354. [PubMed: 17853308]
- Nourshargh S, Hordijk PL, Sixt M. Breaching multiple barriers: leukocyte motility through venular walls and the interstitium. *Nat Rev Mol Cell Biol.* 2010; 11:366–378. [PubMed: 20414258]
- O'Connor P, Goodman A, Kappos L, Lublin F, Polman C, Rudick RA, et al. Long-term safety and effectiveness of natalizumab redosing and treatment in the STRATA MS Study. *Neurology.* 2014; 83:78–86. [PubMed: 24898925]
- O'Doherty U, Peng M, Gezelter S, Swiggard WJ, Betjes M, Bhardwaj N, et al. Human blood contains two subsets of dendritic cells, one immunologically mature and the other immature. *Immunology.* 1994; 82:487–493. [PubMed: 7525461]
- Orlova VV, Choi EY, Xie C, Chavakis E, Bierhaus A, Ihanus E, et al. A novel pathway of HMGB1-mediated inflammatory cell recruitment that requires Mac-1-integrin. *EMBO J.* 2007; 26:1129–1139. [PubMed: 17268551]
- Orlova VV, Chavakis T. Regulation of vascular endothelial permeability by junctional adhesion molecules (JAM). *Thromb Haemost.* 2007; 98:327–332. [PubMed: 17721614]
- Ostanin DV, Furr KL, Pavlick KP, Gray L, Kevil CG, Shukla D, et al. T cell-associated CD18 but not CD62L, ICAM-1, or PSGL-1 is required for the induction of chronic colitis. *Am J Physiol Gastrointest Liver Physiol.* 2007; 292:G1706–14.
- Ostermann G, Weber KS, Zerneck A, Schröder A, Weber C. JAM-1 is a ligand of the beta(2) integrin LFA-1 involved in transendothelial migration of leukocytes. *Nat Immunol.* 2002; 3:151–158. [PubMed: 11812992]
- Palmen MJ, Dijkstra CD, van der Ende MB, Peña AS, van Rees EP. Anti-CD11b/CD18 antibodies reduce inflammation in acute colitis in rats. *Clin Exp Immunol.* 1995; 101:351–356. [PubMed: 7648720]
- Pantelyushin S, Haak S, Ingold B, Kulig P, Heppner FL, Navarini AA, et al. Ror $\gamma$ t+ innate lymphocytes and  $\gamma\delta$  T cells initiate psoriasiform plaque formation in mice. *J Clin Invest.* 2012; 122:2252–2256. [PubMed: 22546855]
- Papp KA, Caro I, Leung HM, Garovoy M, Mease PJ. Efalizumab for the treatment of psoriatic arthritis. *J Cutan Med Surg.* 2007; 11:57–66. [PubMed: 17374316]
- Pavlick KP, Ostanin DV, Furr KL, Laroux FS, Brown CM, Gray L, et al. Role of T-cell-associated lymphocyte function-associated antigen-1 in the pathogenesis of experimental colitis. *Int Immunol.* 2006; 18:389–398. [PubMed: 16415103]
- Perretti M, Chiang N, La M, Fierro IM, Marullo S, Getting SJ, et al. Endogenous lipid- and peptide-derived anti-inflammatory pathways generated with glucocorticoid and aspirin treatment activate the lipoxin A4 receptor. *Nat Med.* 2002; 8:1296–1302. [PubMed: 12368905]
- Perretti M. The annexin 1 receptor(s): is the plot unravelling? *Trends Pharmacol Sci.* 2003; 24:574–579. [PubMed: 14607080]

- Perretti M, Flower RJ. Annexin 1 and the biology of the neutrophil. *J Leukoc Biol.* 2004; 76:25–29. [PubMed: 14966195]
- Peters T, Sindrilaru A, Wang H, Oreshkova T, Renkl AC, Kess D, et al. CD18 in monogenic and polygenic inflammatory processes of the skin. *J Investig Dermatol Symp Proc.* 2006; 11:7–15.
- Phillipson M, Heit B, Colarusso P, Liu L, Ballantyne CM, Kubes P. Intraluminal crawling of neutrophils to emigration sites: a molecularly distinct process from adhesion in the recruitment cascade. *J Exp Med.* 2006; 203:2569–2595. [PubMed: 17116736]
- Picarella D, Hurlbut P, Rottman J, Shi X, Butcher E, Ringler DJ. Monoclonal antibodies specific for  $\beta 7$  integrin and mucosal addressin cell adhesion molecule-1 (MAdCAM-1) reduce inflammation in the colon of SCID mice reconstituted with CD45RBhigh CD4+ T cells. *J Immunol.* 1997; 158:2099–2106. [PubMed: 9036954]
- Pluskota E, Soloviev DA, Plow EF. Convergence of the adhesive and fibrinolytic systems: recognition of urokinase by integrin alpha Mbeta 2 as well as by the urokinase receptor regulates cell adhesion and migration. *Blood.* 2003; 101:1582–1590. [PubMed: 12393547]
- Polman CH, O'Connor PW, Havrdova E, Hutchinson M, Kappos L, Miller DH, et al. A randomized, placebo-controlled trial of Natalizumab for relapsing multiple sclerosis. *N Engl J Med.* 2006; 354:899–910. [PubMed: 16510744]
- Popescu BF, Lucchinetti CF. Pathology of demyelinating diseases. *Ann Rev Pathol.* 2012; 7:185–217. [PubMed: 22313379]
- Ransohoff RM. Natalizumab for multiple sclerosis. *N Engl J Med.* 2007; 356:2622–2629. [PubMed: 17582072]
- Reich K, Burden AD, Eaton JN, Hawkins NS. Efficacy of biologics in the treatment of moderate to severe psoriasis: a network meta-analysis of randomized controlled trials. *Br J Dermatol.* 2012; 166:179–188. [PubMed: 21910698]
- Rivera-Nieves J, Olson T, Bamias G, Bruce A, Solga M, Knight RF, et al. L-selectin, alpha4 $\beta$ 1, and alpha4 $\beta$ 7 integrins participate in CD4+ T cell recruitment to chronically inflamed small intestine. *Immunol.* 2005; 174:2343–2352.
- Rothhammer V, Heink S, Petermann F, Srivastava R, Claussen MC, Hemmer B, et al. Th17 lymphocytes traffic to the central nervous system independently of alpha4 integrin expression during EAE. *J Exp Med.* 2011; 208:2465–2476. [PubMed: 22025301]
- Rudick RA, Sandrock A. Natalizumab: alpha4-integrin antagonist selective adhesion molecule inhibitors for MS. *Expert Rev Neurother.* 2004; 4:571–580. [PubMed: 15853576]
- Rudick R, Polman C, Clifford D, Miller D, Steinman L. Natalizumab: bench to bedside and beyond. *JAMA Neurol.* 2013; 70:172–182. [PubMed: 23128399]
- Rudick RA, Stuart WH, Calabresi PA, Confavreux C, Galetta SL, Radue EW, et al. Natalizumab plus interferon beta-1a for relapsing multiple sclerosis. *N Engl J Med.* 2006; 354:911–923. [PubMed: 16510745]
- Sachs UJ, Andrei-Selmer CL, Maniar A, Weiss T, Paddock C, Orlova VV, et al. The neutrophil-specific antigen CD177 is a counter-receptor for platelet endothelial cell adhesion molecule-1 (CD31). *J Biol Chem.* 2007; 282:23603–23612. [PubMed: 17580308]
- Sandborn WJ, Colombel JF, Enns R, Feagan BG, Hanauer SB, Lawrance IC, et al. Natalizumab induction and maintenance therapy for Crohn's disease. *N Engl J Med.* 2005; 353:1912–1925. [PubMed: 16267322]
- Sandborn WJ, Feagan BG, Rutgeerts P, Hanauer S, Colombel JF, Sands BE, et al. Vedolizumab as induction and maintenance therapy for Crohn's disease. *N Engl J Med.* 2013; 369:711–721. [PubMed: 23964933]
- Sands BE, Feagan BG, Rutgeerts P, Colombel JF, Sandborn WJ, Sy R, et al. Effects of Vedolizumab induction therapy for patients with Crohn's disease in whom Tumor Necrosis Factor antagonist treatment had failed. *Gastroenterology.* 2014; 147:618–627. [PubMed: 24859203]
- Santoso S, Sachs UJ, Kroll H, Linder M, Ruf A, Preissner KT, et al. The junctional adhesion molecule 3 (JAM-3) on human platelets is a counterreceptor for the leukocyte integrin Mac-1. *J Exp Med.* 2002; 196:679–691. [PubMed: 12208882]

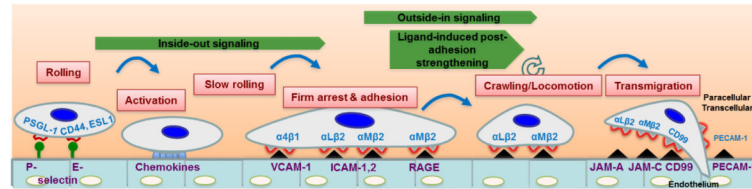
- Schenkel AR, Mamdouh Z, Chen X, Liebman RM, Muller WA. CD99 plays a major role in the migration of monocytes through endothelial junctions. *Nat Immunol.* 2002; 3:143–150. [PubMed: 11812991]
- Schenkel AR, Mamdouh Z, Muller WA. Locomotion of monocytes on endothelium is a critical step during extravasation. *Nat Immunol.* 2004; 5:393–400. [PubMed: 15021878]
- Schmid MC, Avraamides CJ, Dippold H,C, Franco I, Foubert P, Ellies LG, et al. Receptor tyrosine kinases and TLR/IL1Rs unexpectedly activate myeloid cell PI3kgamma, a single convergent point promoting tumor inflammation and progression. *Cancer Cell.* 2011; 19:715–727. [PubMed: 21665146]
- Schmitt MM, Megens RT, Zerneck A, Bidzhekov K, van den Akker NM, Rademakers T, et al. Endothelial junctional adhesion molecule-a guides monocytes into flow-dependent predilection sites of atherosclerosis. *Circulation.* 2014; 129:66–76. [PubMed: 24065611]
- Schön M, Denzer D, Kubitzka RC, Ruzicka T, Schön MP. Critical role of neutrophils for the generation of psoriasiform skin lesions in flaky skin mice. *J Invest Dermatol.* 2000; 114:976–983. [PubMed: 10771480]
- Shamri R, Grabovsky V, Gauguet JM, Feigelson S, Manevich E, Kolanus W, et al. Lymphocyte arrest requires instantaneous induction of an extended LFA-1 conformation mediated by endothelium-bound chemokines. *Nat Immunol.* 2005; 6:497–506. [PubMed: 15834409]
- Shaw SK, Bamba PS, Perkins BN, Luscinskas FW. Real-time imaging of vascular endothelial-cadherin during leukocyte transmigration across endothelium. *J Immunol.* 2001; 167:2323–2330. [PubMed: 11490021]
- Shaw SK, Ma S, Kim MB, Rao RM, Hartman CU, Froio RM, et al. Coordinated redistribution of leukocyte LFA-1 and endothelial cell ICAM-1 accompany neutrophil transmigration. *J Exp Med.* 2004; 200:1571–1580. [PubMed: 15611287]
- Sheremata WA, Minagar A, Alexander JS, Vollmer T. The role of alpha-4 integrin in the aetiology of multiple sclerosis: current knowledge and therapeutic implications. *CNS Drugs.* 2005; 19:909–922. [PubMed: 16268663]
- Shimonaka M, Katagiri K, Nakayama T, Fujita N, Tsuruo T, Yoshie O, et al. Rap1 translates chemokine signals to integrin activation, cell polarization, and motility across vascular endothelium under flow. *J Cell Biol.* 2003; 161:417–27. [PubMed: 12707305]
- Shin J, Hosur KB, Pyaram K, Jotwani R, Liang S, Chavakis T, et al. Expression and function of the homeostatic molecule Del-1 in endothelial cells and the periodontal tissue. *Clin Dev Immunol.* 2013; 2013:617809. [PubMed: 24416060]
- Simmons SB, Pierson ER, Lee SY, Goverman JM. Modeling the heterogeneity of multiple sclerosis in animals. *Trends Immunol.* 2013; 34:410–422. [PubMed: 23707039]
- Simon DI, Rao NK, Xu H., Wei, Y. Majdic O, Ronne E, et al. Mac-1 (CD11b/CD18) and the urokinase receptor (CD87) form a functional unit on monocytic cells. *Blood.* 1996; 88:3185–3194. [PubMed: 8874219]
- Simon DI, Chen Z, Xu H, Li CQ, Dong Jf, McIntire LV, et al. Platelet glycoprotein ibalpha is a counterreceptor for the leukocyte integrin Mac-1 (CD11b/CD18). *J Exp Med.* 2000; 192:193–204. [PubMed: 10899906]
- Singh K, Gatzka M, Peters T, Borkner L, Hainzl A, Wang H, et al. Reduced CD18 levels drive regulatory T cell conversion into Th17 cells in the CD18hypo PL/J mouse model of psoriasis. *J Immunol.* 2013; 190:2544–53. [PubMed: 23418628]
- Smith CW, Marlin SD, Rothlein R, Toman C, Anderson DC. Cooperative interactions of LFA-1 and Mac-1 with intercellular adhesion molecule-1 in facilitating adherence and transendothelial migration of human neutrophils in vitro. *J Clin Invest.* 1989; 83:2008–2017. [PubMed: 2566624]
- Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell.* 1994; 76:301–314. [PubMed: 7507411]
- Springer TA, Dustin ML. Integrin inside-out signaling and the immunological synapse. *Curr Opin Cell Biol.* 2012; 24:107–115. [PubMed: 22129583]
- Stadtmann A, Brinkhaus L, Mueller H, Rossaint J, Bolomini-Vittori M, Bergmeier W, et al. Rap1a activation by CalDAG-GEFI and p38 MAPK is involved in E-selectin-dependent slow leukocyte rolling. *Eur J Immunol.* 2011; 41:2074–2085. [PubMed: 21480213]

- Stanley P, Smith A, McDowall A, Nicol A, Zicha D, Hogg N. Intermediate-affinity LFA-1 binds alpha-actinin-1 to control migration at the leading edge of the T cell. *EMBO J*. 2008; 27:62–75. [PubMed: 18079697]
- Stark MA, Huo Y, Burcin TL, Morris MA, Olson TS, Ley K. Phagocytosis of apoptotic neutrophils regulates granulopoiesis via IL-23 and IL-17. *Immunity*. 2005; 22:285–294. [PubMed: 15780986]
- Staunton DE, Dustin ML, Springer TA. Functional cloning of ICAM-2, a cell adhesion ligand for LFA-1 homologous to ICAM-1. *Nature*. 1989; 33:61–64. [PubMed: 2497351]
- Stenderup K, Rosada C, Dam TN, Salerno E, Belinka BA, Kachlany SC. Resolution of psoriasis by a leukocyte-targeting bacterial protein in a humanized mouse model. *J Invest Dermatol*. 2011; 131:2033–2039. [PubMed: 21654835]
- Stüve O, Marra CM, Jerome KR, Cook L, Cravens PD, Cepok S, et al. Immune surveillance in multiple sclerosis patients treated with natalizumab. *Ann Neurol*. 2005; 59:743–747. [PubMed: 16634029]
- Sydora BC, Wagner N, Lohler J, Yakoub G, Kronenberg M, Muller W, et al.  $\beta 7$  Integrin expression is not required for the localization of T cells to the intestine and colitis pathogenesis. *Clin Exp Immunol*. 2002; 129:35–42. [PubMed: 12100020]
- Targan SR, Feagan BG, Fedorak RN, Lashner BA, Panaccione R, Present DH, et al. Natalizumab for the treatment of active Crohn's disease: results of the ENCORE Trial. *Gastroenterology*. 2007; 132:1672–1683. [PubMed: 17484865]
- Tiwari S, Askari JA, Humphries MJ, Bulleid NJ. Divalent cations regulate the folding and activation status of integrins during their intracellular trafficking. *J Cell Sci*. 2011; 124:1672–1680. [PubMed: 21511727]
- Tran DQ, Glass DD, Uzel G, Darnell DA, Spalding C, Holland SM, et al. Analysis of adhesion molecules, target cells, and role of IL-2 in human FOXP3+ regulatory T cell suppressor function. *J Immunol*. 2009; 182:2929–2938. [PubMed: 19234188]
- Urzainqui A, Serrador JM, Viedma F, Yáñez-Mó M, Rodríguez A, Corbí AL, et al. ITAM-based interaction of ERM proteins with Syk mediates signaling by the leukocyte adhesion receptor PSGL-1. *Immunity*. 2002; 17:401–412. [PubMed: 12387735]
- Vajkoczy P, Laschinger M, Engelhardt B. Alpha4-integrin-VCAM-1 binding mediates G protein-independent capture of encephalitogenic T cell blasts to CNS white matter microvessels. *J Clin Invest*. 2001; 108:557–565. [PubMed: 11518729]
- Van der Vieren M, Le Trong H, Wood CL, Moore PF, St John T, Staunton DE, et al. A novel leukointegrin, alpha d beta 2, binds preferentially to ICAM-3. *Immunity*. 1995; 3:683–690. [PubMed: 8777714]
- Van Seventer GA, Shimizu Y, Horgan KJ, Luce GE, Webb D, Shaw S. Remote T cell co-stimulation via LFA-1/ICAM-1 and CD2/LFA-3: demonstration with immobilized ligand/mAb and implication in monocyte-mediated co-stimulation. *Eur J Immunol*. 1991; 21:1711–1718. [PubMed: 1711977]
- Vermeire S, O'Byrne S, Keir M, Williams M, Lu TT, Mansfield JC, et al. Etrolizumab as induction therapy for ulcerative colitis: a randomised, controlled, phase 2 trial. *Lancet*. 2014; 384:309–318. [PubMed: 24814090]
- Vestweber D, Winderlich M, Cagna G, Nottebaum AF. Cell adhesion dynamics at endothelial junctions: VE-cadherin as a major player. *Trends Cell Biol*. 2009; 19:8–15. [PubMed: 19010680]
- Villablanca EJ, De Calisto J, Torregrosa Paredes P, Cassani B, Nguyen DD, Gabrielsson S, et al.  $\beta 7$  integrins are required to give rise to intestinal mononuclear phagocytes with tolerogenic potential. *Gut*. 2014; 63:1431–1440. [PubMed: 24030488]
- Viguier M, Richette P, Aubin F, Beylot-Barry M, Lahfa M, Bedane C, et al. Onset of psoriatic arthritis in patients treated with efalizumab for moderate to severe psoriasis. *Arthritis Rheum*. 2008; 58:1796–1802. [PubMed: 18512820]
- von Andrian UH, Mempel TR. Homing and cellular traffic in lymph nodes. *Nat Rev Immunol*. 2003; 3:867–878. [PubMed: 14668803]
- von Hundelshausen P, Weber C. Platelets as immune cells: bridging inflammation and cardiovascular disease. *Circ Res*. 2007; 100:27–40. [PubMed: 17204662]



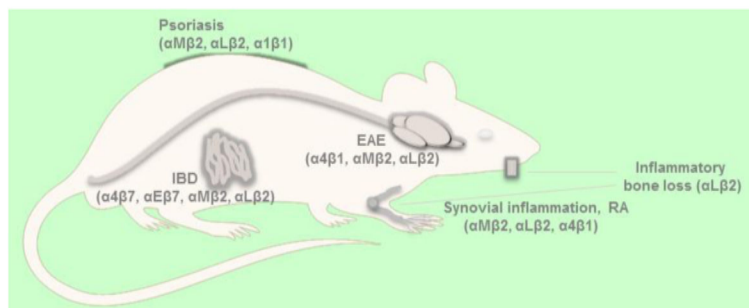
- Wagner N, Lohler J, Kunkel EJ, Ley K, Leung E, Krissansen G, et al. Critical role for  $\beta$ 7-integrins in formation of the gut-associated lymphoid tissue. *Nature*. 1996; 382:366–370. [PubMed: 8684468]
- Wang Y, Kai H, Chang F, Shibata K, Tahara-Hanaoka S, Honda S, et al. A critical role of LFA-1 in the development of Th17 cells and induction of experimental autoimmune encephalomyelitis. *Biochem Biophys Res Commun*. 2007; 353:857–862. [PubMed: 17207459]
- Watts GM, Beurskens FJ, Martin-Padura I, Ballantyne CM, Klickstein LB, Brenner MB, et al. Manifestations of inflammatory arthritis are critically dependent on LFA-1. *J Immunol*. 2005; 174:3668–3675. [PubMed: 15749905]
- Weber C, Springer TA. Neutrophil accumulation on activated, surface-adherent platelets in flow is mediated by interaction of Mac-1 with fibrinogen bound to  $\alpha$ IIb $\beta$ 3 and stimulated by platelet-activating factor. *J Clin Invest*. 1997; 100:2085–2093. [PubMed: 9329974]
- Wei Y, Lukashev M, Simon DI, Bodary SC, Rosenberg S, Doyle MV, et al. Regulation of integrin function by the urokinase receptor. *Science*. 1996; 13:1551–1555. [PubMed: 8703217]
- Wegener KL, Partridge AW, Han J, Pickford AR, Liddington RC, Ginsberg MH, et al. Structural basis of integrin activation by talin. *Cell*. 2007; 128:171–182. [PubMed: 17218263]
- Weger W. Current status and new developments in the treatment of psoriasis and psoriatic arthritis with biological agents. *Br J Pharmacol*. 2010; 160:810–820. [PubMed: 20590580]
- Weinstock-Guttman B. An update on new and emerging therapies for relapsing-remitting multiple sclerosis. *Am J Manag Care*. 2013; 19:s343–354. [PubMed: 24494635]
- Wegmann F, Petri B, Khandoga AG, Moser C, Khandoga A, Volkery S, et al. ESAM supports neutrophil extravasation, activation of Rho, and VEGF-induced vascular permeability. *J Exp Med*. 2006; 203:1671–1677. [PubMed: 16818677]
- Wessel F, Winderlich M, Holm M, Frye M, Rivera-Galdos R, Vockel M, et al. Leukocyte extravasation and vascular permeability are each controlled in vivo by different tyrosine residues of VE-cadherin. *Nat Immunol*. 2014; 15:223–230. [PubMed: 24487320]
- Willenborg DO, Simmons RD, Tamatani T, Miyasaka M. ICAM-1-dependent pathway is not critically involved in the inflammatory process of autoimmune encephalomyelitis or in cytokine-induced inflammation of the central nervous system. *J Neuroimmunol*. 1993; 45:147–154. [PubMed: 7687251]
- Wolf R, Howard OM, Dong HF, Voscopoulos C, Boeshans K, Winston J, et al. Chemotactic activity of S100A7 (Psoriasin) is mediated by the receptor for advanced glycation end products and potentiates inflammation with highly homologous but functionally distinct S100A15. *J Immunol*. 2008; 181:1499–1506. [PubMed: 18606705]
- Wolf R, Mascia F, Dharamsi A, Howard OM, Cataisson C, Bliskovski V, et al. Gene from a psoriasis susceptibility locus primes the skin for inflammation. *Sci Transl Med*. 2010; 2:61ra90.
- Woodfin A, Voisin MB, Imhof BA, Dejana E, Engelhardt B, Nourshargh S. Endothelial cell activation leads to neutrophil transmigration as supported by the sequential roles of ICAM-2, JAM-A, and PECAM-1. *Blood*. 2009; 113:6246–6257. [PubMed: 19211506]
- Woodfin A, Voisin MB, Beyrau M, Colom B, Caille D, Diapouli FM, et al. The junctional adhesion molecule JAM-C regulates polarized transendothelial migration of neutrophils in vivo. *Nat Immunol*. 2011; 12:761–769. [PubMed: 21706006]
- Wu H, Gower RM, Wang H, Perrard XY, Ma R, Bullard DC, et al. Functional role of CD11c+ monocytes in atherogenesis associated with hypercholesterolemia. *Circulation*. 2009; 119:2708–2717. [PubMed: 19433759]
- Xie C, Alcaide P, Geisbrecht BV, Schneider D, Herrmann M, Preissner KT, et al. Suppression of experimental autoimmune encephalomyelitis by extracellular adherence protein of *Staphylococcus aureus*. *J Exp Med*. 2006; 203:985–994. [PubMed: 16585266]
- Xie J, Li R, Kotovuori P, Vermot-Desroches C, Wijdenes J, Arnaout MA, et al. Intercellular adhesion molecule-2 (CD102) binds to the leukocyte integrin CD11b/CD18 through the A domain. *J Immunol*. 1995; 15:3619–3628. [PubMed: 7561061]
- Yago T, Shao B, Miner JJ, Yao L, Klopocki AG, Maeda K, et al. E-selectin engages PSGL-1 and CD44 through a common signaling pathway to induce integrin  $\alpha$ L $\beta$ 2-mediated slow leukocyte rolling. *Blood*. 2010; 116:485–494. [PubMed: 20299514]

- Yamamoto Y, Yamamoto H. RAGE-Mediated Inflammation, Type 2 Diabetes, and Diabetic Vascular Complication. *Front Endocrinol.* 2013; 4:105.
- Yednock TA, Cannon C, Fritz LC, Sanchez-Madrid F, Steinman L, Karin N. Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha4beta1 integrin. *Nature.* 1992; 356:63–66. [PubMed: 1538783]
- Yonekawa K, Harlan JM. Targeting leukocyte integrins in human diseases. *J Leukoc Biol.* 2005; 77:129–140. [PubMed: 15548573]
- Zarbock A, Lowell CA, Ley K. Spleen tyrosine kinase Syk is necessary for E-selectin-induced alpha(L)beta(2) integrin-mediated rolling on intercellular adhesion molecule-1. *Immunity.* 2007a; 26:773–783. [PubMed: 17543554]
- Zarbock A, Polanowska-Grabowska RK, Ley K. Platelet-neutrophil-interactions: linking hemostasis and inflammation. *Blood Rev.* 2007b; 21:99–111. [PubMed: 16987572]
- Zarbock A, Abram CL, Hundt M, Altman A, Lowell CA, Ley K. PSGL-1 engagement by E-selectin signals through Src kinase Fgr and ITAM adapters DAP12 and FcR gamma to induce slow leukocyte rolling. *J Exp Med.* 2008; 205:2339–2347. [PubMed: 18794338]
- Zarbock A, Ley K, McEver RP, Hidalgo A. Leukocyte ligands for endothelial selectins: specialized glycoconjugates that mediate rolling and signaling under flow. *Blood.* 2011; 118:6743–6751. [PubMed: 22021370]
- Zeigler M, Chi Y, Tumas DB, Bodary S, Tang H, Varani J. Anti-CD11a ameliorates disease in the human psoriatic skin-SCID mouse transplant model: comparison of antibody to CD11a with Cyclosporin A and clobetasol propionate. *Lab Invest.* 2001; 81:1253–1261. [PubMed: 11555673]



**Figure 1. The multistep model of leukocyte recruitment**

The sequential steps of the leukocyte adhesion cascade and the adhesive interactions between endothelium and leukocytes are shown. The cascade is initiated with leukocyte capturing and rolling on the endothelium, followed by chemokine-induced leukocyte activation, slow rolling, firm leukocyte adhesion/arrest, adhesion strengthening induced by integrin ligation, crawling and leukocyte transmigration. Essential molecular players involved in the adhesive processes include: selectins and their glycoprotein ligands, chemokines and their receptors, integrins and adhesion receptors of the immunoglobulin-superfamily.  $\alpha 4$ - and  $\beta 2$ -integrins play a critical role in the course of the cascade. Integrin activation occurs upon chemokine triggered signaling (inside-out signaling) in cooperation with selectin-activated pathways. Activated integrins contribute to slow rolling, firm adhesion, crawling and to transendothelial migration.



**Figure 2. Leukocyte integrin targeting in animal inflammatory models**

The role of the  $\beta 2$  integrins  $\alpha M\beta 2$  and  $\alpha L\beta 2$  has been studied in several animal models of inflammatory disorders. In autoimmune encephalomyelitis (EAE), a rodent model resembling multiple sclerosis, either treatment with an antibody against CD11b or CD11a or deficiency in one of these integrins leads to reduced EAE disease severity. Administration of anti-CD11b or anti-LFA-1 antibodies limits skin manifestations and inflammation in psoriatic models. In addition, deficiency in the  $\beta 2$  subunit CD18 protects animals from arthritis development and combined inhibition of CD11b and CD11a leads to significantly decreased neutrophil recruitment. CD11a blockade has also been linked to decreased bone loss in inflammatory arthritis models, whereas mice lacking CD11a demonstrate bone loss in a periodontitis model. Regarding the inflammatory bowel diseases (IBD), CD11b inhibition leads to reduced inflammatory cell infiltration and less mucosal damage in a model of colitis.

In addition,  $\beta 1$  integrins serve as targets for intervention in inflammatory disease models. Blockade of  $\alpha 4\beta 1$  prevents disease development in an EAE model. Furthermore, T cell recruitment to the synovium is dependent on  $\alpha 4\beta 1$ -integrin as shown in a model of arthritis. Besides  $\alpha 4\beta 1$ , the inhibition of  $\alpha 1\beta 1$ -integrin has been shown to be beneficial by inhibiting disease development in a psoriatic model. In addition,  $\alpha 4\beta 7$  blockade attenuates T cell homing to Peyer's patches, a main component of gut-associated lymphoid tissue (GALT). Moreover, treatment with antibodies against either  $\alpha E\beta 7$  or  $\beta 7$  reduces inflammation in a model of colitis.

**Table 1**

Therapeutic targeting of leukocyte integrins in clinical use.

Disease	Integrin	Drug	Target	Treatment
Multiple Sclerosis	$\alpha 4\beta 1$	Natalizumab	humanized monoclonal IgG4 antibody against the $\alpha 4$ integrin subunit	Approved for the treatment of relapsing-remitting multiple sclerosis
Psoriasis	$\alpha L\beta 2$	Efalizumab	humanized monoclonal IgG1 antibody against the alpha chain (CD11a) of $\alpha L\beta 2$ integrin	Effective in the treatment of psoriasis (withdrawn in 2009)
Inflammatory Bowel Disease	$\alpha 4\beta 7, \alpha E\beta 7$	Natalizumab	humanized monoclonal IgG4 antibody against the $\alpha 4$ integrin subunit	Approved for the treatment of medium to severe Crohn's disease under restrictive distribution program (USA)
		Vedolizumab	humanized IgG1 monoclonal antibody against $\alpha 4\beta 7$ -integrin	Approved for the treatment of moderate to severe Crohn's disease and ulcerative colitis
		Etrolizumab	humanized IgG1 monoclonal antibody against the $\beta 7$ subunit of $\alpha 4\beta 7$ - and $\alpha E\beta 7$ -integrins	Effective in the treatment of moderate to severe ulcerative colitis (clinical trials)