

Validation of the doubly labeled water method using off-axis integrated cavity output spectroscopy and isotope ratio mass spectrometry.

Edward L. Melanson, Tracy Swibas, Wendy M. Kohrt, Vicki A. Catenacci, Seth A. Creasy, Guy Plasqui, Loek Wouters, John R. Speakman, and Elena S.F. Berman

Author Affiliations:

- Division of Endocrinology, Metabolism, and Diabetes (EM, VC, SC) and Division of Geriatric Medicine (EM, TS, WK), Department of Medicine, University of Colorado Anschutz Medical Campus, Aurora, CO;
- Geriatric Research, Education, and Clinical Center, VA Eastern Colorado Health Care System, Denver, CO (EM, WK)
- NUTRIM School of Nutrition and Translational Research in Metabolism, Maastricht University, Maastricht, Netherlands (GP, LW)
- Institute of Biological and Environmental Sciences, Aberdeen University, Aberdeen, United Kingdom and State Key Laboratory of Molecular Developmental Biology, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing, China (JRS),
- Los Gatos Research/ABB, San Jose, CA (EB)

Author's last names: Melanson, Swibas, Kohrt, Catenacci, Creasy, Plasqui, Wouters, Speakman, Berman

Corresponding Author: Edward L. Melanson, Ph.D. (Ed.melanson@ucdenver.edu)
MS 8106, 12801 East 17th Ave, RC1 South RM 7103
University of Colorado Anschutz Medical Campus, Aurora, CO 80045
Phone: (303) 724-0935 FAX: (303) 724-3920

Sources of Support: This work was supported by an NIH Small Business Innovation (SBIR) research Grant (R44 DK093362), as well as support from the Colorado Nutrition and Obesity Research Center (P30 DK048520) and the Colorado Clinical and Translational Science Institute (UL1 RR025780). Dr. Melanson is also supported by resources from the Geriatric Research, Education, and Clinical Center at the Denver VA Medical Center

Clinical Trial Registry: The study was registered on ClinicalTrials.gov (NCT01938794)

Running Head: Doubly-labeled water measurements by OA-ICOS and IRMS

ABSTRACT

1
2
3 When the doubly-labeled water (DLW) method is used to measure total daily energy expenditure
4 (TDEE), isotope measurements are typically performed using isotope ratio mass spectrometry
5 (IRMS). New technologies, such as off-axis integrated cavity output spectroscopy (OA-ICOS)
6 provide comparable isotopic measurements of standard waters and human urine samples, but the
7 accuracy of carbon dioxide production (VCO_2) determined with OA-ICOS has not been
8 demonstrated. We compared simultaneous measurement VCO_2 obtained using whole-room
9 indirect calorimetry (IC) with DLW-based measurements from IRMS and OA-ICOS. 17 subjects
10 (10 female; 22 to 63 yrs.) were studied for 7 consecutive days in the IC. Subjects consumed a
11 dose of 0.25 g $H_2^{18}O$ (98% APE) and 0.14 g 2H_2O (99.8% APE) per kg of total body water, and
12 urine samples were obtained on days 1 and 8 to measure average daily CO_2 production (VCO_2)
13 using OA-ICOS and IRMS. VCO_2 was calculated using both the plateau and intercept methods.
14 There were no differences in VCO_2 measured by OA-ICOS or IRMS compared with IC when the
15 plateau method was used. When the intercept method was used, VCO_2 using OA-ICOS did not
16 differ from IC, but VCO_2 measured using IRMS was significantly lower than IC. Accuracy (~1-
17 5%), precision (~8%), intraclass correlation coefficients ($R=0.87-90$), and root mean squared
18 error (30-40 L/day) of VCO_2 measured by OA-ICOS and IRMS were similar. Both OA-ICOS
19 and IRMS produced measurements of VCO_2 with comparable accuracy and precision when
20 compared to IC.

21

22 **Key Words:** Adult, Humans, Oxygen Isotope, Deuterium, Respiratory Gas Exchange

INTRODUCTION

23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45

The gold-standard for measuring total daily energy expenditure (TDEE) in free-living individuals is the doubly-labeled water (DLW) method, which is based on the principle that differential elimination rates of isotopic labels of hydrogen and oxygen provides a measure of carbon dioxide (CO₂) production, subject to certain limiting assumptions (10, 19). TDEE measured using the DLW method has been shown to have an accuracy in humans of $\pm 1-5\%$ against whole room indirect calorimetry (IC) (5, 8, 15, 17-19, 23). Although the number of DLW studies in humans has increased over time (approximately 100 per year), widespread adoption of the DLW method in humans has been limited by the costs of the isotopic labels, and challenges related to sample collection, preparation, and analysis using isotope ratio mass spectrometry (IRMS).

An alternative approach to IRMS for water isotope analysis is laser absorption spectroscopy. These instruments are less expensive than IRMS (~\$100,000 vs. \$250,000), do not require highly trained technicians for their operation (1), and provide simultaneous measurement of multiple isotopes with less tedious sample preparation (20). There are two commercially-available forms of laser absorption spectroscopy for water isotope analysis, cavity ring-down spectroscopy (CRDS) and Off-Axis Integrated Cavity Output Spectroscopy (OA-ICOS). With CRDS, a laser pulse is trapped in a highly reflective optical cavity. The exponential decay of the light intensity is measured (“ringdown” time) and used to calculate the concentration of the absorbing substance in the gas mixture in the cavity. Although CRDS water isotope analyzers provide accurate and precise measurements of total body water ($0.5 \pm 1\%$) and TDEE ($0.5 \pm 6\%$)

46 compared with IRMS, commercial CRDS analyzers have substantial instrumental memory
47 effects, necessitating both careful considerations for reducing isotopic disparity between
48 measured samples and mathematical correction (21). Furthermore, in the above referenced study
49 CRDS was validated against IRMS, but not against the criterion measure of near continuous
50 respiratory gas exchange.

51
52 The other commercially-available form of laser absorption spectroscopy for water isotopes, OA-
53 ICOS, uses a laser light source that is coupled to an optical cavity in an off-axis fashion. The
54 laser light wavelength is scanned over absorption features of interest, providing a direct
55 measurement of the absorbing substances in the gas mixture (1). As with IRMS and CRDS, OA-
56 ICOS also suffers from memory issues between adjacent samples. However, because the time to
57 measure each sample (100 seconds) with OA-ICOS is relatively short and requires only a small
58 volume of sample per injection (~1000 nL), memory issues can be circumvented using a higher
59 number of injections per sample, negating the need to perform mathematical corrections. We
60 have previously shown this approach to be accurate and precise when compared to IRMS for
61 both measuring isotopic measurements of pure water and of human urine samples at both
62 enriched and natural abundances (1-3). However, the accuracy and precision of measuring daily
63 carbon dioxide production (VCO_2) using the DLW method with samples measured using OA-
64 ICOS by comparison to whole room indirect calorimetry has not yet been determined. Thus, the
65 purpose of this study was to compare measurement of daily carbon dioxide production (VCO_2)
66 L/day in a whole-room indirect calorimeter with VCO_2 measured simultaneously using the
67 doubly-labeled water (DLW) method with the resultant body water samples (urine) analyzed
68 using OA-ICOS. We also compared the accuracy and precision of OA-ICOS to that of IRMS.

69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91

METHODS

Institutional Approval and Ethics - Procedures followed were in accordance with the ethical standards of the Helsinki Declaration of 1975 as revised in 1983. The study was approved by the Colorado Multiple Institutional Review Board on May 2, 2013. The study was registered on ClinicalTrials.gov (NCT01938794) on September 5, 2013. Subject recruitment and enrollment commenced in September, 2013, and the last study visit occurred in February, 2017.

Subjects and screening procedures – Adult volunteers (≥ 18 years) were recruited from the University of Colorado Anschutz Medical Campus (CU-AMC) and local communities. After providing informed, written consent, a Health History and Physical Examination was performed to confirm that volunteers were in a good state of health and that they met criteria for inclusion or exclusion. Primary study exclusion criteria were self-reported smoking or use of smokeless tobacco products, self-reported chronic disease (e.g. heart disease, diabetes, or thyroid disease), or currently pregnant. Body composition was then assessed using whole-body dual-energy x-ray absorptiometry (DXA, Hologic Delphi-W, Hologic, Inc., Bedford, MA). Because of weight limitations of the DXA, volunteers with a body weight greater than 135 kg were also excluded.

Experimental design and study procedures – Subjects were studied for 1 week in the whole-room indirect calorimeter located at the University of Colorado Anschutz Medical Campus. Upon subject arrival on day 1, body weight was measured to ± 0.1 kg and a baseline urine sample was obtained for determination of background abundances of $\delta^2\text{H}$ and $\delta^{18}\text{O}$. Subjects were then

92 given an oral dose of 0.25 g of 98 atom percent (98% APE) ^{18}O labeled water and 0.14 g 99.8%
93 APE ^2H labeled water (Sigma Aldrich) per kg of total body water (estimated as 73% of FFM
94 derived from DXA). The dosing cup was twice rinsed with 30 mL of tap water and consumed to
95 ensure complete dosing. After the dose was provided, subjects entered the room calorimeter to
96 begin the 7 day study. Subjects were instructed to completely void ~1 hour after the dose was
97 delivered. Post-dosing urine samples were obtained 4 (PD4) and 5 hours (PD5) after the DLW
98 dosing. On days 2-7, subjects exited the calorimeter for 1 h each day (0700-0800), during which
99 time body weight was measured and then subjects were permitted to shower. For the entire 7
100 day study, *ad libitum* meals were provided each day at 9 AM, 1 PM, and 6 PM. Subjects were
101 instructed to perform exercise (30 min of treadmill walking at a brisk walking pace) each day to
102 increase TDEE above sedentary levels. On Day 8, subjects exited the calorimeter and end-dose
103 urine and blood samples were obtained at the same time of day as on Day 1 (ED4 and ED5).
104 Approximately 20 ml of each urine sample was immediately pipetted into airtight cryotube and
105 stored at $\sim -10^\circ\text{C}$ until transferred to a -80°C freezer. Duplicate samples remained frozen at -
106 80°C until analysis.

107

108 *Whole room-indirect calorimetry* – Average daily VCO_2 and 24 h EE over the 7 day period were
109 measured using the whole-room indirect calorimeter located at CU-AMC using a previously
110 described indirect calorimetry system (Sable Systems, International, Las Vegas, NV) (13). O_2
111 consumption (VO_2) and VCO_2 were calculated in 1-minute intervals using the flow rate and the
112 differences in CO_2 and O_2 concentrations between entering and exiting air, and minute by minute
113 energy expenditure (EE) was calculated using the equations of Jequier et al. (7). Daily 24 h
114 VCO_2 and EE were obtained by summing minute values over the 23 hour measurement period

115 and extrapolating to 24 h values. The accuracy and precision of the system was tested monthly
116 using propane combustion tests. The average O₂ and CO₂ recoveries during the study were
117 $\geq 97.0\%$. While this study was being performed, we also performed several tests using infusions
118 of nitrogen and CO₂ using high precision mass flow controllers, and those tests yielded an
119 accuracy of the IC within 1% of the expected values (unpublished).

120

121 *OA-ICOS analysis of urine samples* - Previously frozen urine samples were prepared by
122 centrifugation, as previously described (3); no distillation or decolorizing steps were undertaken.
123 The OA-ICOS instrument was calibrated using deionized working standards that had been
124 previously calibrated by OA-ICOS against the VSMOW2 and SLAP2 international standards, as
125 previously described (1, 3). Briefly, centrifuged urine samples were injected into heated (~85
126 °C) stainless steel injection block to produce water vapor, which was then introduced into the
127 OA-ICOS optical cavity. Simultaneous measurements of $\delta^2\text{H}$ and $\delta^{18}\text{O}$ were performed on each
128 individual injection. Isotope range within each run was minimized by grouping samples
129 expected to have similar enrichments (e.g. PD4/PD5, ED4/ED5) and by using working standards
130 that closely bracketed the expected isotope ratios. Samples, working standards, and internal
131 controls were interleaved throughout each analysis to ensure high accuracy by frequent intra-run
132 calibration. For every individual measurement within a run, samples, working standards, and
133 internal controls were injected 8-12 times, depending on the total isotope range of the run (e.g.
134 runs with high enriched samples were injected 12 times,). We have previously shown this
135 approach to produce accurate and precise measurements without memory correction when
136 compared to IRMS (1, 3). 3-5 urine samples were typically included in an individual run which
137 took ~5 to 7 hours to complete. At the conclusion of each OA-ICOS run, the syringe, injector

138 block, tubes, and filters were cleaned as previously described (1). Each sample was analyzed in a
139 duplicate run on a subsequent day (typically within the same week). If the difference between
140 duplicate runs exceeded 2 ‰ for $^2\text{H}:^1\text{H}$ or 1 ‰ for $^{18}\text{O}:^{16}\text{O}$ for a given sample, then that
141 sample was run again and only duplicate values that fell within this range were used.

142
143 Isotopic data from the OA-ICOS analyzer were processed using commercially-available Post
144 Analysis Software (LGR, version 3.1.0.9) as previously described (1, 2). Within each run,
145 working standard measurements were utilized with a cubic spline standardization to calibrate
146 urine sample measurements. Specifically, a cubic spline was fit to all measurements of a single
147 standard throughout the run. For each sample injection, an individual calibration curve was
148 constructed from the splined values of each of the working standards. This approach maximally
149 corrects for any instrument drift over the course of the run. To mitigate the effects of sample to
150 sample memory on the OA-ICOS measurements, several procedures were employed (1, 3).
151 First, to account for memory effects between successive samples, the last 4 injections for each
152 sample were averaged, ignoring the first 4-8 injections. Second, to monitor instrument
153 performance, including memory effects between successive samples, an internal control water of
154 known isotopic composition within the range of the isotope ratios of the working standards was
155 measured periodically within each run. Internal controls were checked against the known values.
156 Runs where the internal controls differed from known values by more than ± 1.0 - 2.0 ‰ per mil
157 (‰) (for low and high-enriched samples) for $\delta^2\text{H}$, or ± 0.3 ‰ for $\delta^{18}\text{O}$ from the known value were
158 repeated. Precision of the urine samples was assessed using these same parameters. Finally, an
159 injection volume (linearity) correction was employed to reduce the effects of different water
160 concentrations (due to syringe volume fluctuations) on the measured isotope ratios. The post-

161 analysis software also identified any individual injections that were outliers (isotope ratio ± 3.0
162 SD within an injection set) and for the presence of any organic contamination using the
163 integrated Spectral Contamination Identifier feature (9). The presence of any outliers also
164 identified samples where memory effects had not been eliminated.

165

166 *IRMS analysis of urine samples* – Frozen urine samples were shipped from University of
167 Colorado Anschutz Medical Campus to Maastricht University using airtight sealed glass vials
168 and kept frozen using dry ice. Samples were transferred to a -80° freezer and remained frozen
169 until analyzed. For the analysis of $^2\text{H}:^1\text{H}$, a 2 ml glass vial containing 300 μl of urine was filled
170 with hydrogen gas and equilibration occurred for 1 day at room temperature with a catalyst (5%
171 platinum-on alumina, 325 mesh; Aldrich Chemical Company Ltd) placed in an insert in the vial.
172 For the analysis of ^{18}O , 300 μl of urine was put in a glass vial, which was then filled with CO_2
173 gas. Equilibration then took place for 4 hours at 40° C. The relative amounts of $^2\text{H}:^1\text{H}$ in
174 hydrogen gas and $^{18}\text{O}:^{16}\text{O}$ in CO_2 were then determined using isotope ratio mass spectrometry
175 (Micromass Optima Dual Inlet mass spectrometer with a Multiprep; Manchester, UK, 1998).
176 Each run contained a total of 60 samples of which 12 were working standards with isotope
177 concentrations that bracketed the expected isotope ratios of the urine samples. Each sample was
178 analyzed in a duplicate run on a subsequent day (typically within the same week).

179

180 *Calculation of CO_2 production (VCO_2) and TDEE* – For both OA-ICOS and IRMS, total body
181 water (TBW) was calculated as the average of the dilution spaces of ^2H and ^{18}O after correction
182 for isotopic exchange with other body pools (14). Deuterium (K_D) and oxygen (K_O) turnover
183 rates were calculated by linear regression of the natural logarithm of isotope enrichment as a

184 function of time. All 4 time points were used in the calculation of K_D and K_O . TBW and VCO_2
 185 were calculated using the plateau and intercept methods (using the average of the PD4 and PD5
 186 enrichments) and the equation A6 of Schoeller et al. (15):

187

$$188 \quad rCO_2(\text{mol/d}) = (N/2.078) * (1.01k_O - 1.041k_D) - 0.0246 * rGF$$

189

190 where 1.01 and 1.04 represent the dilution spaces for deuterium and ^{18}O , respectively, N is the
 191 body water dilution space, and rGF is the rate of gas fractionation estimated as $1.05N(k_O - k_D)$
 192 (5). TDEE from OA-ICOS and IRMS was calculated using the calculated VCO_2 and the equation
 193 of Weir [$TDEE = 3.94 \times VO_2 + 1.1 VCO_2$, where $VO_2 = VCO_2 / RQ$] (22), assuming a
 194 respiratory quotient of 0.86, and averaged over 7 days.

195

196 *Sample Size Justification* - Samples size estimates were based on repeated measures on 15
 197 individuals studied in the room calorimeter located at the University of Colorado Anschutz
 198 Medical Campus (unpublished data). The difference between the two 24 h VCO_2 measurements
 199 was $\sim 12.7 \pm 7.5$ L/day ($\sim 3\%$ of mean values). A total sample of 16 paired measurements was
 200 estimated to achieve $\sim 80\%$ power to detect equivalence in 24 h VCO_2 between IC and either
 201 IRMS or OA-ICOS when the margin of equivalence is ± 7.7 L/day with a 0.05 significance level.

202

203 *Statistics* - Prior to analysis, all data were tested for normality. Differences between IC, OA-
 204 ICOS, and IRMS were determined using a repeated measures ANOVA. Post-hoc comparisons
 205 were performed using Tukey's multiple comparison test. Because our primary objective was to
 206 compare each instrument type to the criterion measure IC, we report only the comparison
 207 between IC and OA-ICOS and IC and IRMS. Level of agreement was evaluated using the

208 difference between the criterion and observed values (percent error, a measure of accuracy), the
209 variance around the accuracy (a measure of precision), intraclass correlation coefficient (a
210 measure of level of agreement), root mean squared error (rMSE, a measure of the magnitude of
211 errors resulting from both bias and variability), and Bland-Altman plots (which provides a
212 measure of bias and limits of agreement, as well as determining whether the error is associated
213 with the magnitude of the criterion measure). The Bland-Altman analyses were performed using
214 the IC as the criterion measure. Associations between subject characteristics and measurement
215 error were determined using the Pearson's correlation coefficient. Significance for all tests was
216 set at $P=0.05$. Analyses were performed using GraphPad Prism (5.03, La Jolla, CA). Data are
217 reported as mean \pm SD.

218

219

RESULTS

220

221 19 subjects participated in the study. One subject withdrew after one day in the calorimeter.
222 Due to technical issues, two days of data were lost on another subject, and this subject was
223 excluded from the analysis. Thus, the final study sample consisted of 17 participants (Table 1).

224

225 Average daily turnover rates of deuterium (k_D /day) and oxygen (k_O /day) determined using OA-
226 ICOS (0.118 ± 0.031 /day and 0.142 ± 0.034 /day, respectively) were nearly identical to those
227 determined using IRMS (0.118 ± 0.032 /day, 0.141 ± 0.033 /day). The individual k_O , k_D , N_O , and
228 N_D data used to perform these calculations is contained in the supplementary data file.

229

230 **Results using the plateau method**

231
232 TBW, fat free mass (FFM), fat mass (FM), and body fat percentage (%fat) measured by DXA,
233 OA-ICOS, and IRMS are shown in Table 2. There were no differences in TBW, FFM, FM, or
234 %fat measured by OA-ICOS or IRMS when compared with DXA. Regardless of approach N_D
235 and N_O were similar (Table 3), and the average dilution space ratios were close to the empirically
236 derived value in adult humans of 1.031 (15).

237
238 There were no significant differences in average VCO_2 measured by OA-ICOS (433.0 ± 72.7
239 L/day) or IRMS (418.3 ± 73.0 L/day) when compared with IC (411.2 ± 62.1 L/day) (Figure 1,
240 Table 1). To demonstrate the effect on calculated TDEE, 24 h EE from IC (calculated using the
241 measured RQ) was compared to TDEE calculated from OA-ICOS and IRMS using the assumed
242 RQ of 0.86, as would be done in a standard DLW study. Mean TDEE measured by OA-ICOS
243 (10.16 ± 1.70 MJ/day) and IRMS (9.91 ± 1.70 MJ/day) did not significantly differ from IC (9.88
244 ± 1.56 MJ/day).

245
246 The accuracy of VCO_2 measured by OA-ICOS (mean % error) and IRMS was 5.4 and 1.7%,
247 respectively (Table 4). The accuracy of OA-ICOS was significantly different from zero (95% CI
248 does not cross zero). However, the size of the 95% CIs around the percent error were similar for
249 OA-ICOS (+1.1 to +9.6 L/day) and IRMS (-2.5 to +5.8 L/day), indicating a similar level of
250 precision. The ICC between OA-ICOS and IC [0.87 (95% CI = 0.67 – 0.95)] was similar to the
251 ICC between IRMS and IC [0.89 (0.72 – 0.96)]. The RMSE was 40.2 L/day for OA-ICOS and
252 31.5 L/day for IRMS. Results of the Bland-Altman analysis are presented in Figure 2. There
253 was a significant bias for OA-ICOS (+21.8 L/day, 95% CI = +3.9 to +39.8 L/day) compared to

254 IC, but not for IRMS (+7.1 L/day, 95% CI = -9.1 to +23.4 L/day). The reduced accuracy and
255 significant bias for OA-ICOS was driven by a single outlier. The Bland-Altman correlations for
256 OA-ICOS and IRMS were not significant indicating no bias with absolute level of VCO_2 . VCO_2
257 for each individual measured by IC, OA-ICOS, and IRMS are shown in Table 1. For most
258 individuals, all three methods produced similar results.

259

260 **Results using the intercept method**

261

262 When the intercept method was used, TBW and FFM estimated using IRMS were significantly
263 lower, and FM and %fat significantly higher compared to DXA ($P < 0.001$) (Table 2). There were
264 no differences in TBW, FFM, FM, and %fat measured by DXA compared with OA-ICOS. N_D
265 and N_O were similar, and the average dilution space ratios were close to the theoretical value in
266 adult humans of 1.031 (15) (Table 3). There was no difference in average VCO_2 measured by
267 OA-ICOS (422.9 ± 70.7 L/day) when compared with IC (411.2 ± 62.1 L/day), but VCO_2
268 measured by IRMS (381.9 ± 69.2 L/day) was significantly different compared with IC (Figure
269 3). Similarly, mean TDEE measured by OA-ICOS (10.40 ± 1.70 MJ/day) was not different than
270 24 h EE. However, mean TDEE measured by IRMS using the intercept method (9.05 ± 1.62
271 MJ/day) was significantly lower than 24 h EE. Individual subject VCO_2 results calculated using
272 the intercept method are presented in the **Supplemental Table 1**.

273

274 As with the plateau method, there was a similar level of agreement when VCO_2 measured using
275 OA-ICOS and IRMS were compared with IC (Table 4). Interestingly, accuracy between OA-
276 ICOS and IC tended to be better using the intercept method, whereas accuracy between IRMS

277 and IC tended to be better using the plateau method. Precision, ICC, and RMSE were similar for
278 OA-ICOS and IRMS using the intercept method. Results of the Bland-Altman analysis are
279 presented in Figures 4. There was a significant bias for IRMS (-29.2 L/day, 95% CI = -44.6 to -
280 13.9 L/day) compared to IC, but not for OA-ICOS (+11.7 L/day, 95% CI = -5.1 to +28.5 L/day).
281 The Bland-Altman correlations between average VCO_2 from IC and both IRMS and OA-ICOS
282 were not significant indicating no bias with absolute level of EE.

283

284 **Additional Analyses**

285

286 To determine if %fat, BMI, or age were contributing factors to differences between IC and IRMS
287 or OA-ICOS, correlations between these variables and the differences in VCO_2 between IC and
288 OA-ICOS and IC and IRMS were determined (using the plateau data). The differences in VCO_2
289 between IC and OA-ICOS were not significantly correlated with %fat ($r=0.41$) or BMI ($r=0.42$),
290 but were positively and significantly ($P<0.05$) associated with age ($r=0.59$). However, this
291 significant correlation was driven solely by one subject (S12, 60 yr. old female) where OA-ICOS
292 substantially overestimated IC (+54 L/day). The differences between IC and IRMS were not
293 significantly correlated with %fat ($r=-0.07$), BMI ($r=-0.20$), or age ($r=0.04$). We also examined
294 the association between the differences in VCO_2 (IC – OA-ICOS, IC – IRMS) with measured
295 RQ. The differences (TDEE – 24 h EE) between IC and OA-ICOS ($r=0.19$) and IRMS ($r=0.46$)
296 were positively but weakly ($P>0.05$) correlated with average daily 24 hr RQ. We performed
297 these same analyses using the intercept data, and results were similar (data not shown).

298

299

DISCUSSION

300
301 Because of the high costs of operation and technical expertise required for operation of IRMS,
302 only a few specialized labs are equipped to perform DLW measurements of TDEE. Although
303 new approaches such as OA-ICOS are available, they have not yet been validated against room
304 calorimetry. We compared VCO_2 calculated using isotopic measurements obtained using OA-
305 ICOS against 24 h VCO_2 measured using whole-room indirect calorimetry as the criterion
306 measure. We also compared VCO_2 calculated using isotopic measurements obtained using IRMS
307 on the same samples to then evaluate if the techniques provide comparable accuracy and
308 precision compared to IC. Mean VCO_2 measured using OA-ICOS did not differ significantly
309 from IC, whether using plateau or intercept calculation approach. Mean VCO_2 measured using
310 IRMS did not differ from IC when the plateau method was used, but was significantly lower than
311 IC when the intercept method was used. Nonetheless, measurements of accuracy (% error),
312 precision (SD of mean % error), ICC, RMSE, and Bland-Altman analyses suggested that level of
313 agreement with IC was similar for both IRMS and OA-ICOS. Thus, results of this study
314 demonstrate that off-axis integrated cavity output spectroscopy provides estimates of VCO_2 from
315 DLW studies in humans that are as accurate and precise as estimates derived from IRMS.
316

317 Initial validation work of the DLW method performed in the 1950's in several small animal
318 species showed that VCO_2 was within ~3% of that measured simultaneously by indirect
319 calorimetry (11, 12). Schoeller and van Santen (16) performed the first validation studies in
320 humans in 1982, and reported that TDEE from the DLW method differed from measured energy
321 intake (adjusted for changes in body composition) by an average of 2%. Subsequent validation
322 studies against near continuous respiratory gas exchange measured over 4-7 days reported

323 precisions of ~1-8% for measuring VCO_2 and TDEE (5, 8, 15, 17, 18, 23). The range of
324 accuracies for both OA-ICOS and IRMS in the current study (Table 4), using both the plateau
325 and intercept method, were similar to these previous studies. Surprisingly, when using the
326 intercept method, we observed a significant difference between mean VCO_2 measured by IC and
327 IRMS, which is not consistent with previous validation studies.

328
329 To more thoroughly compare the IC to OA-ICOS (and IC to IRMS), we performed several
330 statistical tests to assess the levels of agreement between instruments, some of which are more
331 reflective of individual errors. Specifically, both the ICC and RMSE describe how concentrated
332 the data are around the line of best fit (in this case, the line of identity), whereas the Bland-
333 Altman allows identification of systematic differences between two measurements (4). Both the
334 RMSE and Bland-Altman can be also used to identify where measurement errors are driven by
335 the presence of outliers. Because DLW studies are performed on groups of individuals (e.g., to
336 compare differences between groups to determine the effect of some intervention), more weight
337 should be given to tests that are based on mean differences. For example, even though the
338 Bland-Altman test indicated a significant, positive bias in measuring VCO_2 using the plateau
339 method with OA_ICOS (+21.8 L/day), there was no difference in mean VCO_2 measured by OA-
340 ICOS and IC. Based on the current analyses, we conclude that OA-ICOS provides a measure of
341 average daily VCO_2 that is accurate (1% to 5%) and precise (8%) without systematic bias. We
342 also conclude that accuracy, precision, and bias are similar to those observed with IRMS.

343
344 It has been suggested that adiposity and nutritional status affect the dilution space ratio (N_d/N_o)
345 between ^2H and ^{18}O , causing potential errors in VCO_2 when the DLW method is used (6). In that

346 study, it was reported that there was an overestimation of VCO_2 by the DLW method in high fat
347 (HF) diet fed mice compared with measured VCO_2 using continuous measurements with IC.
348 This overestimation occurred in both a diet-induced obesity-prone (DIO) and diet-induced
349 obesity-resistant (DR) groups, suggesting that the overestimation is independent of body fat gain
350 during a HF diet. In the current study, we found no association between either %fat or BMI and
351 the difference in VCO_2 measured with IC and DLW. We also explored the association between
352 measured RQ and the difference in VCO_2 measured with IC and DLW. These associations were
353 also non-significant with both OA-ICOS and IRMS. Although we did not measure energy intake
354 (subjects consumed an *ad libitum* diet), our subjects were weight stable throughout the 7 day
355 study (-0.5 ± 0.8 kg, mean \pm SD), suggesting that individual differences in average 24 hr RQ
356 reflected differences in habitual energy macronutrient intake rather than energy balance. Under
357 this assumption, if VCO_2 is overestimated during consumption of a high fat diet, a negative
358 correlation would be expected when the differences between the DLW and IC VCO_2 are plotted
359 against RQ (with a lower RQ indicative of a higher fat intake). Thus, results of the current study
360 do not support the conclusion that VCO_2 from the DLW method is overestimated during a high-
361 fat diet, but we concede that this can only be determined during studies in which energy and
362 macronutrient intake is highly controlled.

363
364 *Strengths and limitations:* A strength of the current study is the sample size, which is larger
365 (N=17) than previous validation studies performed using near continuous measurements of
366 respiratory gas exchange (N<10) (5, 8, 15, 17, 18, 24). A limitation of the current study, as in all
367 validation studies, is the validity of the criterion measure (IC). However, as described in the
368 Methods section, the room calorimeter system at AMC consistently measures within 1-3% of

369 expected values using gas infusion and propane combustion tests. In addition to costs, OA-ICOS
370 offers several advantages over IRMS including easier sample preparation and reducing the need
371 for highly trained technicians. However, it should be noted that the sample measurement
372 configuration used in the current study (e.g. 8-12 injections per sample, with multiple interleaved
373 measurements of working standards and internal controls) does not increase the throughput
374 compared to IRMS and CRDS. The advantage of this approach is that it negates the need for
375 mathematical correction due to memory effects. Throughput could be increased by reducing the
376 number of injections per sample, but the tradeoff would then be the need to apply mathematical
377 correction for memory effects.

378

379 In conclusion, , mean VCO_2 measured using OA-ICOS did not differ significantly from
380 concurrently measured 24 h VCO_2 using whole room indirect calorimetry, whether using plateau
381 or intercept calculation approach. Furthermore, both OA-ICOS and IRMS produced
382 measurements of VCO_2 with comparable accuracy and precision when compared to whole room
383 indirect calorimetry. Based on these results, we conclude that off-axis integrated cavity output
384 spectroscopy provides a valid and viable alternative to IRMS for measuring TDEE using DLW
385 in humans.

386 **Acknowledgements:** This work was supported with resources and use of facilities from the
387 Geriatric Research, Education, and Clinical Center at the Denver VA Medical Center

388

389 **Conflict of Interest (COI) Statement:** Elena Berman is employed by ABB/Los Gatos Research,
390 the company that manufactures the OA-ICOS analyzer.

391

392

393 **Disclaimer:** The contents do not represent the views of the U.S. Department of Veterans Affairs
394 or the United States Government.

395

396 Table 1. Subject characteristics and individual average total daily carbon dioxide production (VCO₂) measured by measured by IC
 397 and by OA-ICOS and IRMS using the plateau method.

Subject #	Sex	Age (yrs.)	Weight (kg)	BMI (kg/m ²)	VCO ₂ (L/day)		
					IC	OA-ICOS	IRMS
1	F	46	63.0	24.0	310.6	307.1	267.3
2	M	32	82.8	23.9	457.4	456.2	440.2
3	M	43	74.8	25.1	455.8	484.1	487.8
4	M	28	61.0	22.4	346.8	374.9	367.8
5	F	24	93.8	31.9	471.3	474.7	476.9
6	F	60	48.9	19.4	293.4	339.0	334.7
7	F	62	53.3	21.8	349.9	372.8	351.3
8	M	34	91.5	32.2	444.2	458.3	436.9
9	M	40	71.6	23.0	442.8	448.3	390.5
10	F	27	111.6	46.4	514.4	568.2	529.5
11	F	60	95.0	34.8	437.1	560.5	421.6
12	F	63	115.0	42.8	423.1	474.7	453.7
13	F	34	101.4	36.1	433.7	449.7	453.9
14	M	24	73.9	23.0	473.4	450.7	545.9
15	F	30	72.0	28.5	394.0	367.3	391.7
16	F	22	61.7	24.5	353.8	358.4	336.6
17	M	43	69.6	20.8	387.9	415.9	424.4
Mean (SD)		39 (14)	78.8 (19.7)	28.3 (7.9)	411.2 (62.1)	433.0 (72.7)	418.3 (73.0)

398 Table 2. Total body water (TBW), fat free mass (FFM), fat mass (FM), and percent body fat (%Fat) measured
 399 by DXA, OA-ICOS, and IRMS. OA-ICOS and IRMS results are presented for both plateau and intercept
 400 methods. Mean (SD).

	Intercept Method			Plateau Method	
	DXA	OA-ICOS	IRMS	OA-ICOS	IRMS
TBW (kg)	38.3 (7.3)	38.3 (6.7)	35.6 (6.5) ^a	38.3 (6.7)	39.0 (6.7)
FFM (kg)	52.5 (10.0)	52.2 (9.4)	48.8 (8.9) ^a	52.5 (10.0)	53.4 (9.2)
FM (kg)	25.9 (15.8)	26.6 (15.8)	29.9 (16.0) ^a	26.3 (16.0)	25.3 (15.8)
%Fat	31.0 (12.5)	31.9 (11.9)	36.8 (11.3) ^a	31.5 (12.6)	30.2 (12.0)

401 ^a significantly different from DXA

402

403 Table 3. Deuterium (N_D) and oxygen (N_O) dilution spaces and dilution space ratio ($N_D:N_O$) measured by OA-
 404 ICOS and IRMS. Results are presented for both plateau and intercept methods. Mean (SD).

	Intercept Method		Plateau Method	
	OA-ICOS	IRMS	OA-ICOS	IRMS
N_D (kg)	38.0 (6.7)	37.9 (6.9)	38.9 (6.8)	40.4 (6.7)
N_O (kg)	36.8 (6.6)	36.8 (6.7)	37.8 (6.6)	39.0 (6.8)
$N_D:N_O$	1.033 (0.005)	1.030 (0.006)	1.029 (0.0068)	1.037 (0.013)

405

406

407 Table 4. Limits of agreement for CO₂ production (VCO₂) measured by OA-ICOS and IRMS. Results are
 408 presented for both plateau and intercept methods.

	Error (%)	ICC	RMSE
	Mean (95% CI)	(95% CI)	(L/day)
OA-ICOS - Plateau	5.4 (+1.1, +9.6)	0.87 (0.67, 0.95)	40.2
IRMS - Plateau	1.7 (-2.5, +5.8)	0.89 (0.72, 0.96)	31.5
OA-ICOS – Intercept	2.9 (-1.1, +6.9)	0.88 (0.70, 0.90)	33.8
IRMS – Intercept	-7.2 (-11.2, -3.3)	0.90 (0.74, 0.96)	35.9

409 ICC – interclass correlation; RMSE – Root mean Square Error

410

FIGURE LEGENDS

411

412 Figure 1. VCO_2 (Mean \pm SEM) measured by IC and by OA-ICOS and IRMS using the plateau
413 method.

414

415 Figure 2. Bland-Altman plots of OA-ICOS (A) and IRMS (B) using the plateau method vs. the
416 criterion measure IC.

417

418 Figure 3. VCO_2 (Mean \pm SEM) measured by IC and by OA-ICOS and IRMS using the intercept
419 method. * Significantly different than IC.

420

421 Figure 4. Bland-Altman plots of OA-ICOS (A) and IRMS (B) using the intercept method vs. the
422 criterion measure IC.

423

424

REFERENCES

- 425 1. Berman ES, Fortson SL, Snaith SP, Gupta M, Baer DS, Chery I, Blanc S, Melanson EL,
426 Thomson PJ, and Speakman JR. Direct analysis of delta²H and delta¹⁸O in natural and
427 enriched human urine using laser-based, off-axis integrated cavity output spectroscopy.
428 *Analytical chemistry* 84: 9768-9773, 2012.
- 429 2. Berman ES, Levin NE, Landais A, Li S, and Owano T. Measurement of delta¹⁸O,
430 delta¹⁷O, and ¹⁷O-excess in Water by Off-Axis Integrated Cavity Output Spectroscopy
431 and Isotope Ratio Mass Spectrometry. *Analytical chemistry* 2013.
- 432 3. Berman ES, Melanson EL, Swibas T, Snaith SP, and Speakman JR. Inter- and
433 intraindividual correlations of background abundances of (²)H, (¹⁸)O and (¹⁷)O in
434 human urine and implications for DLW measurements. *European journal of clinical*
435 *nutrition* 69: 1091-1098, 2015.
- 436 4. Bland JM, and Altman DG. Statistical methods for assessing agreement between two
437 methods of clinical measurement. *Lancet* 1: 307-310, 1986.
- 438 5. Coward WA, and Prentice AM. Isotope method for the measurement of carbon dioxide
439 production rate in man. *The American journal of clinical nutrition* 41: 659-663, 1985.
- 440 6. Guidotti S, Meijer HA, and van Dijk G. Validity of the doubly labeled water method for
441 estimating CO₂ production in mice under different nutritional conditions. *American*
442 *journal of physiology Endocrinology and metabolism* 305: E317-324, 2013.
- 443 7. Jequier E, Acheson K, and Schutz Y. Assessment of energy expenditure and fuel
444 utilization in man. *Annual review of nutrition* 7: 187-208, 1987.
- 445 8. Klein PD, James WP, Wong WW, Irving CS, Murgatroyd PR, Cabrera M, Dallosso HM,
446 Klein ER, and Nichols BL. Calorimetric validation of the doubly-labelled water method

- 447 for determination of energy expenditure in man. *Human nutrition Clinical nutrition* 38:
448 95-106, 1984.
- 449 9. Leen JB, Berman ESF, Liebson L, and Gupta M. Spectral contaminant identifier for off-
450 axis integrated cavity output spectroscopy measurements of liquid water isotopes. *Rev Sci*
451 *Instrum* 83: 2012.
- 452 10. Lifson N. Theory of use of the turnover rates of body water for measuring energy and
453 material balance. *J Theor Biol* 12: 46-74, 1966.
- 454 11. Lifson N, Gordon GB, and Mc CR. Measurement of total carbon dioxide production by
455 means of D₂O¹⁸. *J Appl Physiol* 7: 704-710, 1955.
- 456 12. McClintock R, and Lifson N. Measurement of basal and total metabolism in hereditarily
457 obese-hyperglycemic mice. *The American journal of physiology* 193: 495-498, 1958.
- 458 13. Melanson EL, Ingebrigtsen JP, Bergouignan A, Ohkawara K, Kohrt WM, and Lighton
459 JR. A new approach for flow-through respirometry measurements in humans. *American*
460 *journal of physiology Regulatory, integrative and comparative physiology* 298: R1571-
461 1579, 2010.
- 462 14. Racette SB, Schoeller DA, Luke AH, Shay K, Hnilicka J, and Kushner RF. Relative
463 dilution spaces of ²H- and ¹⁸O-labeled water in humans. *The American journal of*
464 *physiology* 267: E585-590, 1994.
- 465 15. Schoeller DA, Ravussin E, Schutz Y, Acheson KJ, Baertschi P, and Jequier E. Energy
466 expenditure by doubly labeled water: validation in humans and proposed calculation. *The*
467 *American journal of physiology* 250: R823-830, 1986.

- 468 16. Schoeller DA, and van Santen E. Measurement of energy expenditure in humans by
469 doubly labeled water method. *Journal of applied physiology: respiratory, environmental*
470 *and exercise physiology* 53: 955-959, 1982.
- 471 17. Schoeller DA, and Webb P. Five-day comparison of the doubly labeled water method
472 with respiratory gas exchange. *The American journal of clinical nutrition* 40: 153-158,
473 1984.
- 474 18. Seale JL, Conway JM, and Canary JJ. Seven-day validation of doubly labeled water
475 method using indirect room calorimetry. *Journal of applied physiology* 74: 402-409,
476 1993.
- 477 19. Speakman JR. *Doubly labelled water: Theory and Practice*. London: Chapman Press,
478 1997.
- 479 20. Steig EJ, Gkinis V, Schauer AJ, Schoenemann SW, Samek K, Hoffnagle J, Dennis KJ,
480 and Tan SM. Calibrated high-precision O-17-excess measurements using cavity ring-
481 down spectroscopy with laser-current-tuned cavity resonance. *Atmos Meas Tech* 7: 2421-
482 2435, 2014.
- 483 21. Thorsen T, Shriver T, Racine N, Richman BA, and Schoeller DA. Doubly labeled water
484 analysis using cavity ring-down spectroscopy. *Rapid communications in mass*
485 *spectrometry : RCM* 25: 3-8, 2011.
- 486 22. Weir JB. New methods for calculating metabolic rate with special reference to protein
487 metabolism. 1949. *Nutrition* 6: 213-221, 1990.
- 488 23. Westerterp KR, Brouns F, Saris WH, and ten Hoor F. Comparison of doubly labeled
489 water with respirometry at low- and high-activity levels. *Journal of applied physiology*
490 65: 53-56, 1988.

491 24. Westerterp KR, Lafeber HN, Sulkers EJ, and Sauer PJ. Comparison of short term indirect
492 calorimetry and doubly labeled water method for the assessment of energy expenditure in
493 preterm infants. *Biology of the neonate* 60: 75-82, 1991.

494

495

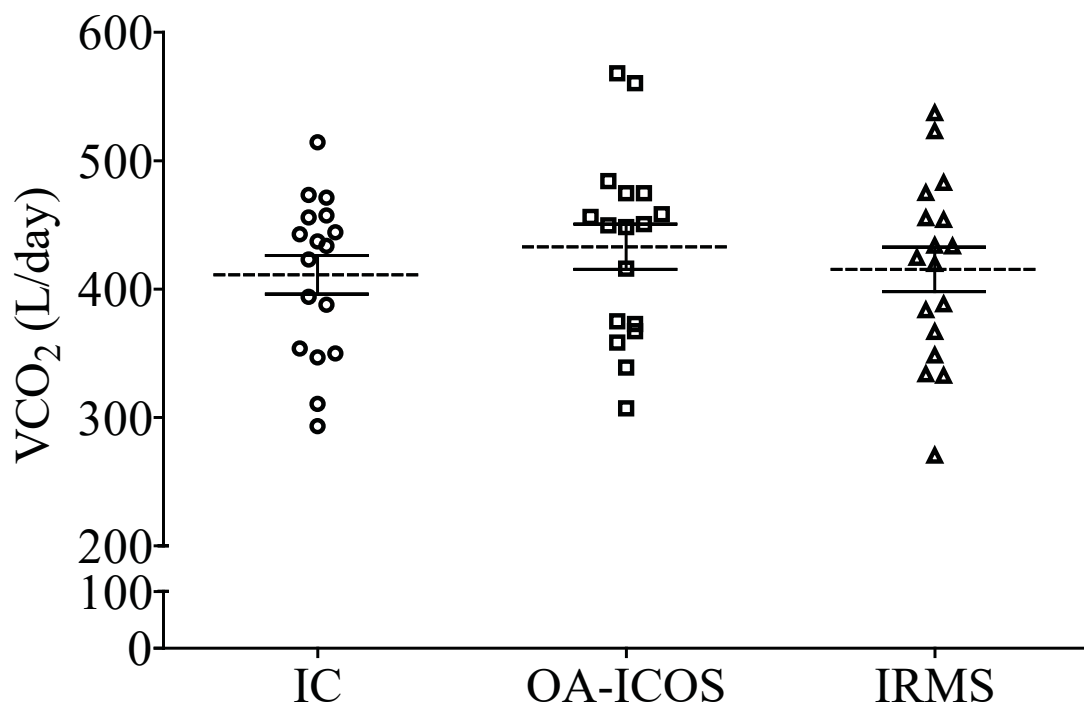


Figure 1

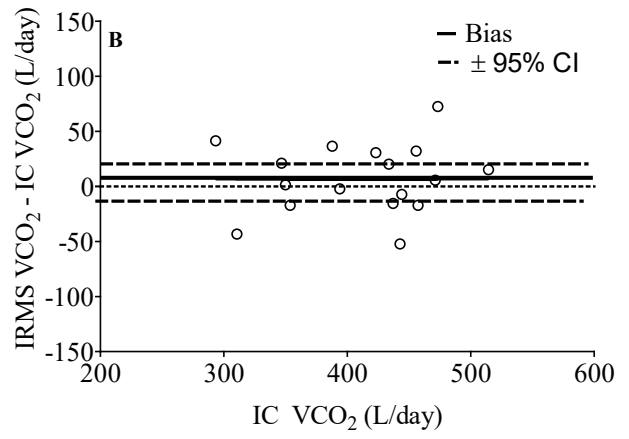
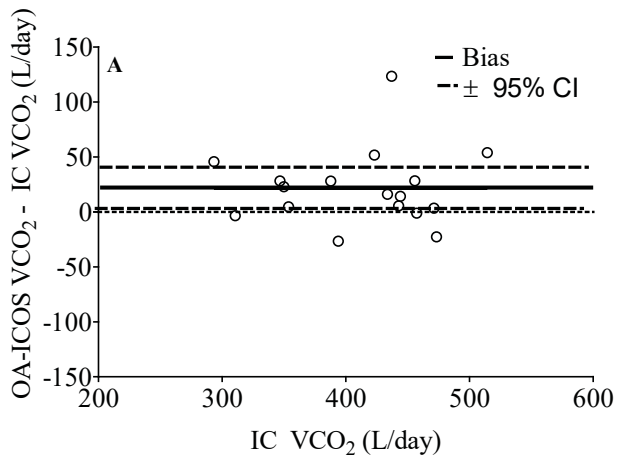


Figure 2

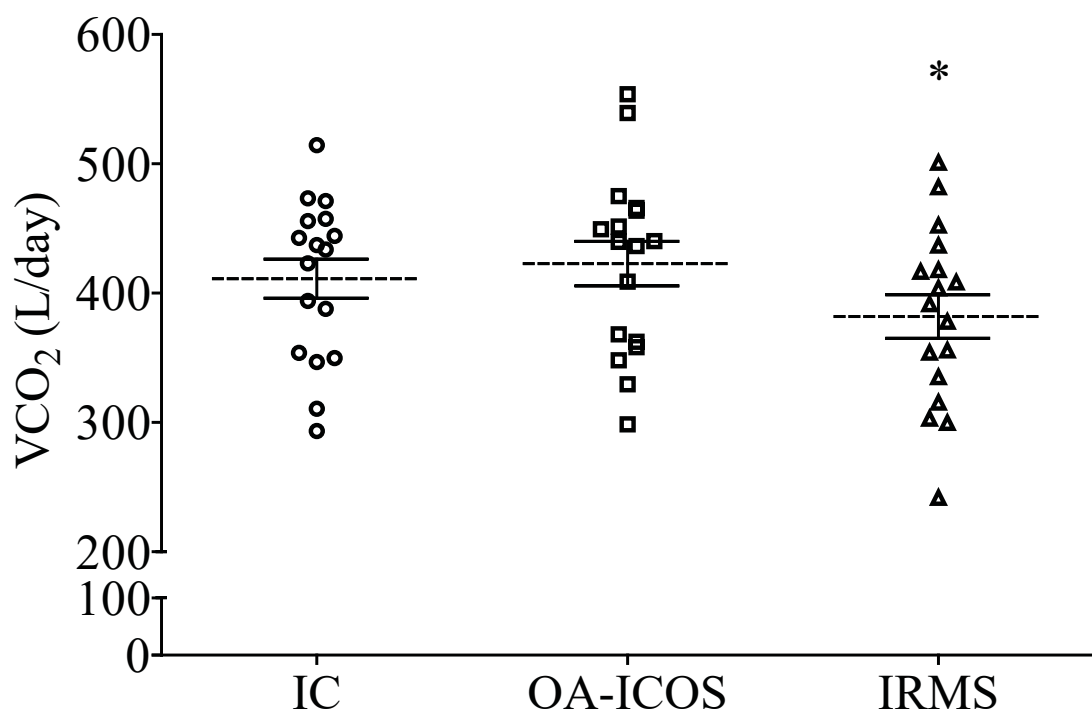


Figure 3

Figure 3

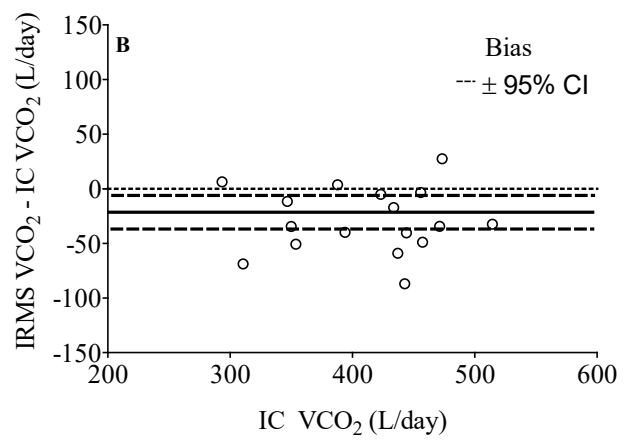
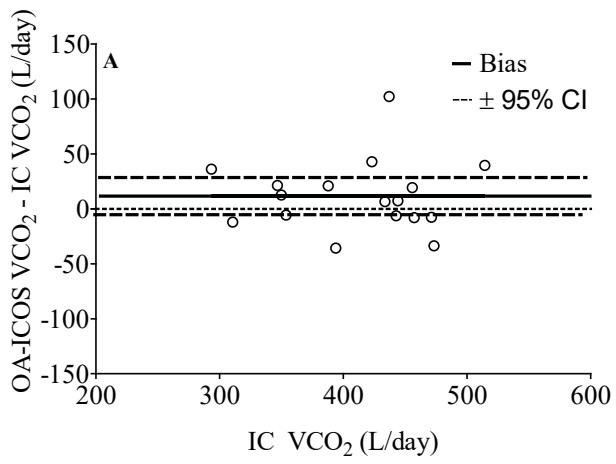


Figure 4