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Ethnodermatological use of medicinal plants in India: From ayurvedic formulations to clinical perspectives – A review

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ABSTRACT

Ethnopharmacological relevance: Traditional knowledge is a particular form of practice or skill set that was developed in ancient times and was sustained through generations via the passing of knowledge, essentially confined within a specific tribe, local people, or family lineages. Ethnodermatological use of medicinal plants in India is still a subject to conduct more studies to see if there is chemical, microbiological, and/or clinical evidence, from a scientific perspective, of their effectiveness for those skin disorders. Thus, this review can be the basis for further studies and may provide targets for drug development.

Aim of the study: We compile and emphasize the most important part of ethnodermatology, namely, traditional knowledge of medicinal plants and their applications for several skin diseases in India. We also include a brief review and explanation on dermatology in Ayurvedic and Unani medicine. We review the pharmacological activity of extracts derived from some of the most cited plants against problem skin diseases as well.

Materials and methods: Different kinds of key phrases such as “Indian traditional ethnodermatology”, “ethnodermatology”, “ethnobotany”, “skin diseases”, “Ayurveda dermatology”, “pharmacological activity” were searched in online search servers/databases such as Google Scholar (<https://scholar.google.com/>), ResearchGate (<https://www.researchgate.net/>), PubMed (<https://pubmed.ncbi.nlm.nih.gov/>), NISCAIR Online Periodicals Repository (NOPR) (<http://nopr.niscair.res.in/>). Based upon the analyses of data obtained from 178 articles, we formulated several important findings which are a summary shown in Tables. A total of 119 records of plants' uses have been found across India against 39 skin diseases. These are depicted with their localities of report, parts used, and preparation and administration methods against particular skin diseases.

Results: The knowledge and utilisation of herbal medicine in the Indian subcontinent has great potential to treat different kinds of human skin disorders. The administration of extracts from most of the plant species used is topical and few only are administered orally. We also investigated the pharmacological activity of the extracts of the most cited plants against mice, bacterial and fungal pathogens, and human cells.

Conclusions: Complementary therapy for dermatological problems and treatment remains the main option for millions of people in the Indian subcontinent. This review on the practices of ethnobotanical dermatology in India confirms the belief that their analysis will accelerate the discovery of new, effective therapeutic agents for skin diseases. However, more studies and clinical evidence are still required to determine if the identified species may contribute to skin condition treatment, particularly in atopic eczema. Today, ethnodermatology is a well-accepted international discipline and many new practices have been initiated in numerous countries. We hope this article will further accelerate the development of this area to identify a new generation of natural human

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skin treatments that will help meet the growing consumer demand for safe, sustainable, and natural treatments. In this context, research on plants utilised in ethnodermatology in India and elsewhere should be intensified.

1. Introduction

The complementary or traditional medicinal system along with the presence of several cultural and socio-religious practices plays a major role in healthcare throughout the Indian subcontinent. Complementary and Alternative Medicine (CAM) in the field of dermatology covers a wide variety of diagnosis and treatment methods that complement conventional dermatology practices by drawing on an extended knowledge base that includes CAM together with the latest research findings (Tirant et al., 2018). CAM mainly focuses on treating the human body as a whole in the hope that this approach will benefit the dermatological treatment (Tirant et al., 2018). As reported by the World Health Organization (WHO), approximately 80% of the human population relies on traditional botanical medicines worldwide. Around 40,000 to 70,000 medicinal plant species are utilised across the world as traditional medicines (Verpoorte et al., 2006). Currently, the world trade in medicinal plants and derived products is evaluated at US\$ 100 billion with an annual growth rate of 15% (Ahmad Khan and Ahmad, 2018).

Significantly, approximately 25% of contemporary pharmaceuticals are also derived directly or indirectly from plants, highlighting the solid basis for plant-derived medicines (Anand et al., 2019). Plants, due to their sessile nature have evolved the capacity to generate astounding chemical cornucopia (Anand et al., 2019). Furthermore, these plant-based natural products are chemically complex, typically consisting of multiple chiral centers. Thus, these molecules are difficult to chemically synthesise in the laboratory and often even where this is possible, the process is not commercially viable. It is the structural complexity of these plant-derived molecules that has directly contributed to their wide-ranging therapeutic activities (Anand et al., 2019, 2020; Banerjee et al., 2021; Das et al., 2021; Datta et al., 2021; Mohammed et al., 2021; Shoshan-Barmatz et al., 2021). Therefore, plants are routinely the only source of these medically significant biomolecules (Halder et al., 2021; Khare et al., 2021; Tandon et al., 2021).

India is one of the seventeen mega biodiversity centers in the world that also includes four biodiversity hotspots, namely, Western Ghats-Sri Lanka, Indo-Burma, Sundaland, and Himalaya (Fig. 1) (Venkataraman and Sivaperuman, 2018). It has been reported by the Botanical Survey of India (BSI) that out of the 45,000 plant species listed, at least 30,000 (two-thirds) are of high interest because of potential therapeutic potential (Nautiyal et al., 2015). Local people utilise some of these plants as herbal medicines, which are effective against a number of diseases. The application of traditional medicines for skin disorders is a well-established and commonly deployed practice in India (Yadav et al., 2012). Modern healthcare facilities are routinely not available in rural areas. Thus, traditional medicines are typically employed to address healthcare issues (Routh and Bhowmik, 1999). Overall, 84 species of medicinal plants are employed in dermatology throughout India. Specific plant species were found to be used in four Indian states with the majority being administered in Tamil Nadu (24), Karnataka (17), Assam, and Uttar Pradesh (nine plant species in each) (Fig. 1). The ready availability of herbal plants, their lower cost, and minimal or absence of side effects has generated a growing demand for the application of these natural sources as treatments for various skin diseases (Abdelouahab and Heard, 2008). In India, even where contemporary medical systems are accessible, a large portion of the population still seeks the benefits of different types of alternative medications, for example, the Ayurvedic or Unani systems (Routh and Bhowmik, 1999; Dutta et al., 2021).

Ethnodermatology is a section of ethnomedicine and ethnobiology related to the diagnosis and treatment of various skin diseases/infections, skincare, and beautification in ethnic populations (Idu et al.,

2011). The importance of the skin to the overall physiology of the human body is easily discernible due to the serious systemic disorders that appear after complex and severe damage to the skin by burns or generalized dermatitis (Idu et al., 2011). The skin barrier serves to limit passive water loss from the body, limits the absorption of chemicals from the environment, and prevents microbial infections (Vaughn et al., 2018). Key research has shown that natural plant-derived ingredients possess a rich potential source of cutaneous protection (Abdelouahab and Heard, 2008). In India, many herbal traditional medicines have been reported for treating several skin disorders: acne, bruises, and burns. Moreover, medicinal plants are most often used against cuts, wounds, skin diseases (generally), ringworm, eczema, leprosy, scabies, etc. (Fig. 2). Tribal and local people routinely use plant organs including leaves, fruit, seeds, and roots for the care and protection of human skin, its beautification, or treatment of associated diseases (Fig. 3) (Idu et al., 2011). Even *Cannabis indica* has demonstrated anti-inflammatory, anti-pruritic, anti-ageing, and anticancer properties by various mechanisms, including interacting with the newly found endocannabinoid system of the skin and thereby providing a promising alternative to traditional treatments (Sheriff et al., 2020). Moreover, cosmetic companies are increasingly applying basic research findings in the chemical and biological sciences to generate more sophisticated products in this area. In Europe only, the cosmetics industry has a €65 billion market size and employs a workforce of half a million indirectly or indirectly (Rinaldi, 2008). However, no comprehensive compilation of the ethno-dermatological reports have been attempted so far. In addition, besides some sparse evidence, the clinical relevance of such widespread traditional practices has not been described either.

The aim of the study is to compile and emphasize the most important part of ethnodermatology, namely, traditional knowledge of medicinal plants and their applications for several skin diseases in India. We also include a brief review and explanation on dermatology in Ayurvedic and Unani medicine. We review the pharmacological activity of extracts derived from some of the most cited plants against problem skin diseases as well.

2. Methodology

Different kinds of key phrases such as “Indian traditional ethnodermatology”, “ethnodermatology”, “ethnobotany”, “skin diseases”, “Ayurveda dermatology”, “pharmacological activity” were searched in online search servers/databases such as Google Scholar (<https://scholar.google.com/>), ResearchGate (<https://www.researchgate.net/>), PubMed (<https://pubmed.ncbi.nlm.nih.gov/>), NISCAIR Online Periodicals Repository (NOPR) (<http://nopr.niscair.res.in/>) thus the appropriate databases and keywords were used. Only articles published in English language from 1975 to December 2020 were included for this study. A total number of 178 studies are included in the study from 213 published papers screened initially. Based upon the data obtained from different online search engines, we formulated several important findings. Table 1 presents plant species in traditional medicine against dermatological diseases in India, Table 2 presents the pharmacological activity of medicinal plants and their preclinical evidence, Table 3 presents clinical evidence of plant use in humans, Table 4 presents Ayurvedic formulations in dermatology, Table 5 presents herbal drugs and cosmetic products used in dermatology, and Table 6 presents bacterial pathogens responsible for the skin diseases in human body and related symptoms. Species names were checked against The Plant List 1.1 (“The Plant List,” 2013), and family names follow the Angiosperm Phylogeny Group IV (Chase et al., 2016).

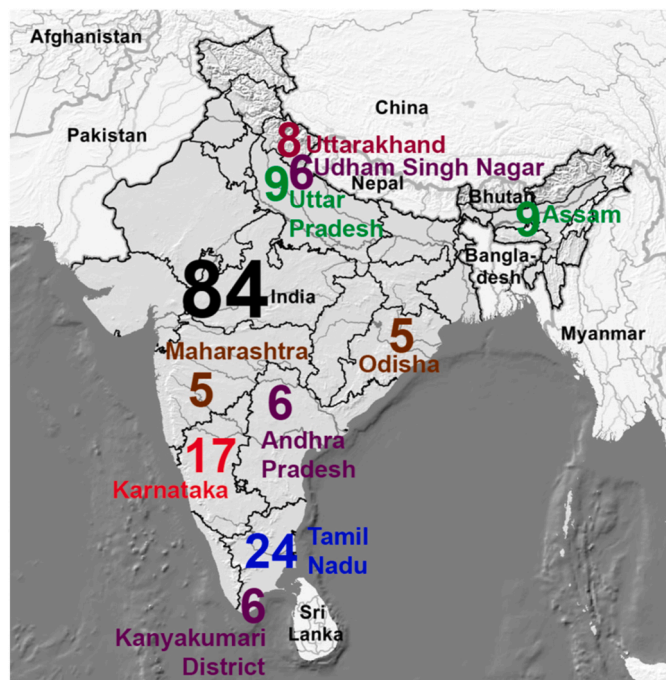


Fig. 1. Number of plant species reported from specific Indian states.

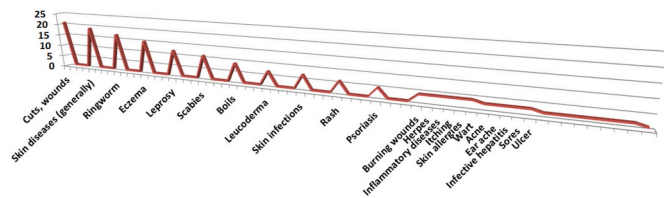


Fig. 2. Number of plant species reported deployed for dermatological problems in India. Y axis: number of plant species reported.

2.1. Dermatology in ayurvedic medicine

Ayurveda, translated as the science of life in Sanskrit, is deep rooted in South Asian history and culture. In India, detailed reports on different dermatologic treatments are outlined in Ayurveda texts. Causes of various dermatoses include dietary practice, daily activities, climatic conditions, or sexual activity. In the Ayurveda, “Rasayana drugs” are considered essential in the treatment of dermatological disorders (Tirant et al., 2018). The “Rasayana drugs” are immunomodulatory, adaptogenic, antioxidant, nootropic, and antistress (Tirant et al., 2018). According to Ayurveda, an analysis of prakruthi (nature) and local skin pathology (which is called Sthaneeya-vikruthi in Sanskrit) is mandatory before prescribing. Namely, prakruthi is considered important for long-term health control, and it has less utility for immediate short-term control. In this context, vikruthi is more important for short-term symptom management in skin diseases.

According to Ayurveda, there are three distinctive doshas or energies (vatha, pitta, and kapha), believed to circulate in the body to govern physical and mental activity. This means complex patterns of different symptoms and signs producing typical “constellations” of clinical signs and symptoms and only about 18 diseases are referred to in Ayurveda under the chapter kustha (meaning skin disease) (Narahari et al., 2013). Ayurvedic medicine has referred to the explanation of skin diseases with pathogenesis, etiology, clinical features, and effective treatment. Stubborn skin diseases, including leprosy, are triggered by inconsistency within several doshas. From consumption of contradictory liquids and food, fasting, excessive exercise, raw or undercooked food, drinking cold

water after prolonged exposure to the sun, and early meal consumption before digesting the former, exceeding emetic therapy procedures, laxative treatment, medical enema, and inhalation therapy are responsible for these diseases. In Ayurvedic medicine for skin disease treatment, many preparations are used including medicinal plants (Fig. 4, Fig. 5): tinctures, ointments, emulsions, pastes, pills, suppositories, patches, lotions, liniments, oils, and bougie. Furthermore, chemical elements and inorganic compounds like copper, gold, lead, mercury, silver, sulfur, tin, zinc, zinc carbonate, alum, borax, and iron sulfate are also mentioned (Routh and Bhowmik, 1999). For acne, rubbing lemon is the best home remedy. The decoction of some plants such as *Justicia adhatoda* L. (= *Adhatoda vasica* Nees, Acanthaceae) (Vasak), *Acorus calamus* L. (Acoraceae) (Vaka), *Trichosanthes dioica* Roxb. (Cucurbitaceae) (Patola), *Azadirachta indica* A. Juss. (Meliaceae) (Neem), *Catunaregam spinosa* (Thunb.) Tirveng. (*Randia dumetorum* (Retz.) Poir., Rubiaceae) (Madana) with honey, while a paste made from some plants such as *Curcuma longa* L. (Zingiberaceae), *Senna alata* (L.) Roxb (Fabaceae), *Cynodon dactylon* (L.). Pers. (Poaceae), *Lawsonia inermis* L. (= *Lawsonia alba* Lam., Lythraceae), *Terminalia chebula* Retz. (Combretaceae) is used for treating several skin diseases such as pruritus, urticaria, eczema, cut wounds, acne, psoriasis, dermatitis, dermatophytosis, syphilis and scabies (Figs. 2 and 4) (Routh and Bhowmik, 1999) (Gilca et al., 2018). For example, it was found that an aqueous extract of *T. chebula* showed inhibitory effects higher than those measured in ethanol extracts (Fig. 5) (Vonshak et al., 2003). However, it should be also remembered that psoriasis, for example, is not a one disease from the Ayurvedic perspective as there are different vikruthis and prakruthis that can lead to the same end result of psoriasis. This case complicates the issue of controlling psoriasis.

2.2. Plants used in the treatment of skin disorders in India

In Bahraich of Uttar Pradesh, Tripathi and Srivastava (2010) found that a total of 11 medicinal plant species such as *Acalypha indica* L., *Phyllanthus emblica* L. (= *Embolica officinalis* Gaertn.), *Jatropha curcas* L., *Euphorbia thymifolia* Burm among others belonging to the Euphorbiaceae (Fig. 6, Fig. 7) family are used as home remedies for various skin diseases like eczema, leprosy, itching, wounds, ringworm, warts and scabies (Fig. 2) (Tirant et al., 2018). In Uttar Pradesh, 92 species of medicinal plants were cited, and the most common and most frequently used medicinal plants in the treatment of dermatological health problems were *Azadirachta indica* A. Juss, *Melia azedarach* L. (both from Meliaceae), and *Curcuma longa* L. (Zingiberaceae) (Kumar et al., 2013). In the Coastal Karnataka (Fig. 8), Bhandary and Chandrashekar (2011) reported 25 of 57 plant species, e.g., *Abrus precatorius* L. (Fabaceae), *Bridelia retusa* (L.) Spreng (Phyllanthaceae), *Rothea serrata* (L.). Steane & Mabb (= *Clerodendrum serratum* (L.). Moon, Lamiaceae) that carry antiviral and other antimicrobial activities (Bhandary and Chandrashekar, 2011). Moreover, 16 plant species such as *Azadirachta indica* A. Juss., *Eclipta prostrata* (L.) L. (Asteraceae), *Celastrus paniculatus* Willd. (Celastraceae) have anti-inflammatory activity. The latter species showed also antifungal inhibitory activity (Vonshak et al., 2003). Ten species like *Acorus calamus* L., *Curcuma longa* L., *Centella asiatica* (L.) Urb. (Apiaceae) are attributed to wound/ulcer healing properties and six species such as *Cyclea peltata* Hook. f. & Thomson (Menispermaceae), *Centella asiatica* (L.) Urb., *Hemidesmus indicus* (L.) R. Br. ex Schult. (Apocynaceae) have cooling properties and *Citrus limon* (L.) Osbeck (Rutaceae) has the antiseptic property (Bhandary and Chandrashekar, 2011). In addition, this review reports the successful use of *Areca catechu* L. (Arecaceae), *Aristolochia indica* L. (Aristolochiaceae), *Coscinium fenestratum* Colebr (Menispermaceae), *Rauvolfia serpentina* (L.) Benth. ex Kurz (Apocynaceae), *Curcuma longa* L. for treating herpes (Bhandary and Chandrashekar, 2011). Saikia et al. (2006) documented 85 plant species belonging to 49 families for their therapeutic use in skin treatment in Assam (Fig. 8). Some of these medicinal plants are *Allium sativum* L. (Amaryllidaceae), *Justicia adhatoda* L., *Swertia chirata* Buch.-Ham. ex C.

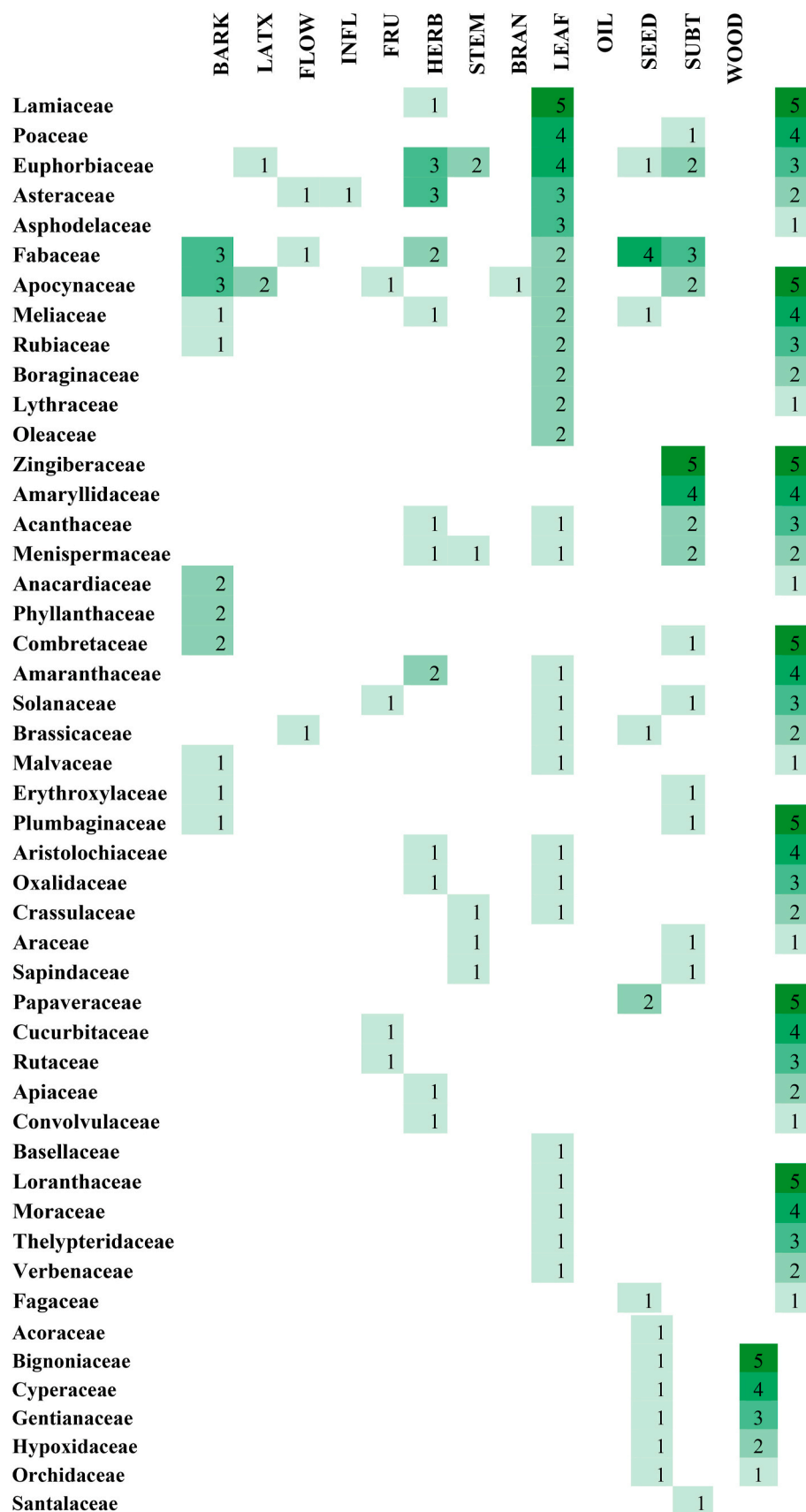


Fig. 3. Quantification of deployment in Indian ethnodermatology by plant parts and botanical family. BARK, LATX, FLOW, flowers; INFL, inflorescences; FRU, fruits; HERB, (= aerial parts); STEM, BRAN, branches; LEAF, OIL, SEED, SUBT, subterranean parts; WOOD.

Table 1
Traditional medicine against dermatological diseases in India.

| Plant name | Family | Locality (state) | Plant parts | Preparation methods | Administration modes | Used against | Reference |
|--|----------------|--|-------------------------|-------------------------|-------------------------------|--|-------------------------------------|
| <i>Acalypha indica</i> L. | Euphorbiaceae | Nanpara Tehsil, Bahraich, Uttar Pradesh | Leaves | Mixed with salt, paste | Topical | Eczema of hands, sole on legs, burning area, ringworm | Tripathi and Srivastava (2010) |
| <i>Croton bonplandianus</i> Baill. | Euphorbiaceae | Nanpara Tehsil, Bahraich, Uttar Pradesh | Latex Leaves Stem | Paste Juice | Topical Topical Topical | Scabies, sores Cuts, wounds Ringworm | Tripathi and Srivastava (2010) |
| <i>Jatropha curcas</i> L. | Euphorbiaceae | Nanpara Tehsil, Bahraich, Uttar Pradesh | Seed | Oil | Topical | Leucoderma, sores, pimple | Tripathi and Srivastava (2010) |
| <i>Ricinus communis</i> L. | Euphorbiaceae | Nanpara Tehsil, Bahraich, Uttar Pradesh | Root | Decoction | Topical | Skin diseases, wart, wounds | Tripathi and Srivastava (2010) |
| | | | Fresh leaves | Paste | Topical | Wounds, carbuncle | |
| <i>Citrus limon</i> (L.) Osbeck | Rutaceae | Coastal Karnataka | Fruit | Paste | Oral | Anticeptic | Bhandary and Chandrashekar (2011) |
| <i>Eclipta prostrata</i> (L.) L. | Asteraceae | Coastal Karnataka | Whole plant | Decoction | Oral | Herpes, infective hepatitis | Bhandary and Chandrashekar (2011) |
| <i>Cosciniium fenestratum</i> Colebr. | Menispermaceae | Coastal Karnataka | Stem | Decoction | Oral | Herpes, infective hepatitis | Bhandary and Chandrashekar (2011) |
| <i>Croton persimilis</i> Müll. Arg. (= <i>Croton roxburghii</i> N.P. Balakr., illeg. name) | Euphorbiaceae | Coastal Karnataka | Root | Decoction | Oral | Herpes, infective hepatitis | Bhandary and Chandrashekar (2011) |
| <i>Bambusa vulgaris</i> Schrad. | Poaceae | Perambalur, Tamil Nadu | Leaves | Decoction | Baby bath | Skin rashes | Balamurugan et al. (2019) |
| <i>Calotropis procera</i> (Aiton) Dryand. | Apocynaceae | Perambalur, Tamil Nadu | Latex | Decoction | Oral | Eczema, abdominal cramps, ringworms, snake bites and wound healing | Balamurugan et al. (2019) |
| <i>Allium cepa</i> L. | Amaryllidaceae | Perambalur, Tamil Nadu | Bulb | Cooking oil | Topical ear drop | Eczema, ear infection | Balamurugan et al. (2019) |
| | | Ganjam District, Orissa | Bulb | Mixed with mustered oil | Topical | Inflammatory swelling | Mishra (2011) |
| <i>Aloe vera</i> (L.) Burm.f. | Asphodelaceae | Medline, Embase, Cochrane Library and CISCOM India | Leaves | Juice | Topical | Severe and prolonged allergic dermatitis | Ernst (2000) |
| | | Bidar district, Karnataka | Leaf, gel | Extraction | Topical | Acne, burning wounds, UV induced damage | Haniadka et al. (2013) |
| | | East India | Leaf | Juice | Topical | Scars, lesions, acne vulgaris | Prashantkumar and Vidyasagar (2008) |
| <i>Santalum album</i> L. | Santalaceae | East India | Heartwood | Oil | Topical | Acne, psoriasis, eczema, common warts, molluscum contagiosum | Moy and Levenson (2017) |
| <i>Curcuma longa</i> L. | Zingiberaceae | Jadavpur, Kolkata, West Bengal | Rhizome | Extraction | Oral | Premature skin wrinkle, excessive melanin secretion, hyperpigmentation | Biswas et al. (2016) |
| | | India | Rhizome | Gel preparation extract | Topical | Psoriasis | Baliga et al. (2013) |
| <i>Azadirachta indica</i> A. Juss. | Meliaceae | India | Whole part | Paste | Topical | Scabies, chronic ulcer | Bedi and Shenefelt (2002) |
| | | Tripura, India | Leaf, bark | Paste | Topical | Leprosy, skin infection, chicken pox | Sen et al. (2011) |
| | | Jalgon District, Maharashtra | Seed | Paste | Topical | Rheumatism, skin diseases | (D. L. Jain et al., 2010) |
| <i>Calotropis procera</i> (Aiton) Dryand. | Apocynaceae | India | Root | Paste | Topical | Wart | Bedi and Shenefelt (2002) |
| <i>Calendula officinalis</i> L. | Asteraceae | Tamil Nadu, India | Flower heads | Extraction | Topical | Acne | Kumar et al. (2005) |
| <i>Terminalia arjuna</i> (Roxb. ex DC.) Wight & Arn. | Combretaceae | Haridwar, India | Stem bark | Decoction | Oral | Skin disease, skin itching | Bharati and Kumar (2014) |
| <i>Brassica nigra</i> (L.) K. Koch | Brassicaceae | India | Leaf, flower | Aqueous extract | Topical | Leprosy | (A. Gupta et al., 2010) |
| <i>Desmodium oojeinense</i> (Roxb.) H. Ohashi (= <i>Ougeinia oojeinensis</i> (Roxb.) Hochr.) | Fabaceae | India | Bark | Powder | Topical | Wound healing | (A. Gupta et al., 2010) |
| | Araceae | Kerala, India | Stem | Extraction | Topical | | |

(continued on next page)

Table 1 (continued)

| Plant name | Family | Locality (state) | Plant parts | Preparation methods | Administration modes | Used against | Reference |
|--|------------------|---|-------------|---|----------------------|---|--------------------------------------|
| <i>Rhaphidophora pertusa</i> (Roxb.) Schott | | | | | | Skin diseases, ulcer, body inflammation | Shanavaskhan et al. (2012) |
| <i>Calotropis gigantea</i> (L.) Dryand. | Apocynaceae | Uttara Kannada district, Karnataka, India | Leaf, latex | Juice | Topical | Scabies, skin infection | Bhandary et al. (1995) |
| <i>Dendrophthoe falcata</i> (L.f.) Ettingsh. | Loranthaceae | Uttara Kannada district, Karnataka, India | Leaf | Paste | Topical | Skin ulcers | Bhandary et al. (1995) |
| <i>Neolamarckia cadamba</i> (Roxb.) Bosser | Rubiaceae | Uttara Kannada district, Karnataka, India | Stem bark | Decoction | Topical | Skin diseases | Bhandary et al. (1995) |
| <i>Curculigo orchioides</i> Gaertn. | Hypoxidaceae | Sonbhadra district, Uttar Pradesh, India | Root | Paste | Topical | Itching, skin diseases | Singh et al. (2002) |
| <i>Lansea coromandelica</i> (Houtt.) Merr. | Anacardiaceae | Sonbhadra district, Uttar Pradesh, India | Bark | Lotion | Topical | Astringent, leprosy | Singh et al. (2002) |
| <i>Pongamia pinnata</i> (L.) Pierre | Fabaceae | Gujarat, India | Seed | Oil | Topical | Burning area | Bedi (1978) |
| <i>Lepidagathis incurva</i> Buch.-Ham. ex D. Don | Acanthaceae | Northeast India | Whole plant | Decoction | Orally | Measles | Begum and Nath (2000) |
| <i>Rhinacanthus nasutus</i> (L.) Kurz | Acanthaceae | Northeast India | Root | pound with 6–7 match tips, mix with vaselline | Topical | Ringworm | Begum and Nath (2000) |
| <i>Centella asiatica</i> (L.) Urb. | Apiaceae | Northeast India | Whole plant | Grind | Topical | Syphilis | Begum and Nath (2000) |
| <i>Excoecaria oppositifolia</i> var. <i>crenulata</i> (Wight) Chakrab. & M.Gangop. (= <i>Excoecaria crenulata</i> Wight) | Euphorbiaceae | Madurai District, Tamil Nadu, India | Stem | Paste | Topical | Affected area of skin | Ignacimuthu et al. (2006) |
| <i>Allium sativum</i> L. | Amaryllidaceae | Assam, India | Bulb | Pieces was crushed | Topical, oral | Leprosy, ringworm, scabies | Ignacimuthu et al. (2006) |
| <i>Basella alba</i> L. (= <i>Basella rubra</i> L.) | Basellaceae | Assam, India | Leaves | Extracted juice | Topical | Urticaria | Ignacimuthu et al. (2006) |
| <i>Bischofia javanica</i> Blume | Phyllanthaceae | Assam, India | Bark | Crush | Topical | Insect bite | Saikia et al. (2006) |
| <i>Datura stramonium</i> L. | Solanaceae | Assam, India | Leaves | Paste, juice | Topical | Eczema, pediculosis | Saikia et al. (2006) |
| <i>Dodonaea viscosa</i> (L.) Jacq. | Sapindaceae | Jalgon District, Maharashtra | Fruit | Juice | Topical | Skin disorder, ulcer | (D. L. Jain et al., 2010) |
| <i>Dodonaea viscosa</i> (L.) Jacq. | Sapindaceae | Villupuram district, Tamil Nadu, India | Stem, root | Decoction | Topical | Skin infections, skin rashes, sore throat, dermatitis | Sankaranarayanan et al. (2010) |
| <i>Rhinacanthus nasutus</i> (L.) Kurz | Acanthaceae | Villupuram district, Tamil Nadu, India | Root | Powder | Topical | Ringworm, skin diseases, chronic wound | Sankaranarayanan et al. (2010) |
| <i>Cissampelos pareira</i> L. | Menispermaceae | Villupuram district, Tamil Nadu, India | Root | Paste | Topical | Pruritus, skin disorder | Sankaranarayanan et al. (2010) |
| <i>Anacardium occidentale</i> L. | Anacardiaceae | Kanyakumari District, Southern India | Bark | Powder | Oral | Leprosy | Kingston et al. (2009) |
| <i>Crinum viviparum</i> (Lam.) R.Ansari & V. J.Nair (= <i>Crinum defixum</i> Ker Gawl.) | Amaryllidaceae | Kanyakumari District, Southern India | Bulb | Paste | Oral | Tinea cruris | Kingston et al. (2009) |
| <i>Wrightia tinctoria</i> R. Br. | Apocynaceae | Kanyakumari District, Southern India | Leaves | Pounded leaves with coconut oil | Applied twice a day | Psoriasis | Kingston et al. (2009) |
| | | Andhra Pradesh, India | Stem bark | Extract | Topical | Skin diseases | Kahleel Basha and Sudarshanam (2010) |
| <i>Caesalpinia crista</i> L. | Fabaceae | Kannada District, Karnataka, India | Seed | Paste | Oral | Scabies, skin allergies | Harsha et al. (2003) |
| <i>Aristolochia indica</i> L. | Aristolochiaceae | Kannada District, Karnataka, India | Leaf | Juice | Topical | Wart | Harsha et al. (2003) |
| <i>Globba marantina</i> L. | Zingiberaceae | Eastern Ghats, India | Root | Fresh rhizomes crushed, mixed with Pongamia seed oil and paste applied on white spots | Topical | Leukoderma | Jeevan Ram et al. (2004) |
| | Oxalidaceae | Tamil Nadu, India | Leaves | Paste | Topical | | |

(continued on next page)

Table 1 (continued)

| Plant name | Family | Locality (state) | Plant parts | Preparation methods | Administration modes | Used against | Reference |
|---|------------------|--|-----------------|------------------------|--------------------------------|--------------------------------------|-------------------------------------|
| <i>Biophytum sensitivum</i> (L.) DC. | | | | | | Scabies, skin rashes, eczema | Chendurpandy et al. (2010) |
| <i>Cissampelos pareira</i> L. | Menispermaceae | Tamil Nadu, India | Whole plant | Paste | Topical | Ringworm infection, insect bites | Chendurpandy et al. (2010) |
| <i>Rauvolfia serpentina</i> (L.) Benth. ex Kurz | Apocynaceae | Tamil Nadu, India | Root | Oil extract | Topical | Leprosy, scabies, ringworm infection | Chendurpandy et al. (2010) |
| <i>Canthium coromandelicum</i> (Burm. f.) Alston (= <i>Canthium parviflorum</i> Lam.) | Rubiaceae | Kanyakumari district, Tamil Nadu, India | Leaves | Paste | Externally applied twice a day | Ringworm, Scabies | Chendurpandy et al. (2010) |
| <i>Scleria lithosperma</i> (L.) Sw. | Cyperaceae | Tamil Nadu, India | Rhizome | Paste | Topical | Eczema, leucoderma, scabies | Chendurpandy et al. (2010) |
| <i>Cuscuta reflexa</i> Roxb. | Convolvulaceae | Bihar, India | Whole plant | Paste | Topical | Leucoderma, eczema | Upadhyay et al. (1998) |
| <i>Acorus calamus</i> L. | Acoraceae | Arunachal Pradesh | Rhizome | Paste | Topical | Scabies, cut, injuries | Saklani and Jain (1989) |
| <i>Ageratum conyzoides</i> (L.) L. | Asteraceae | Assam, Meghalaya, Nagaland, Manipur | Leaves | Paste | Topical | Bites, cuts, wounds | Saklani and Jain (1989) |
| <i>Aphanamixis polystachya</i> (Wall.) R. Parker | Meliaceae | Purulia District, West Bengal | Whole plant | Infusion | Topical | Wormy skin sores | Dey et al. (2012) |
| <i>Ocimum tenuiflorum</i> L. (= <i>Ocimum sanctum</i> L.) | Lamiaceae | Nagaland | Leaves | Paste | Topical | Skin disease | Rao and Jamir (1982) |
| <i>Tamarindus indica</i> L. | Fabaceae | Bidar District, Karnataka | Leaf | Paste | Topical | Swelling on hands and legs | Prashantkumar and Vidyasagar (2008) |
| <i>Nyctanthes arbor-tristis</i> L. | Oleaceae | Purulia District, West Bengal | Leaf | Fresh/raw | Oral | Smoothen rough skin | Dey et al. (2012) |
| <i>Exacum tetragonum</i> Roxb. | Gentianaceae | Purulia District, West Bengal | Root | Paste | Topical | Sores of leech bites | Dey et al. (2012) |
| <i>Abrus precatorius</i> L. | Fabaceae | Assam | Root | Paste | Topical | Leucoderma | Buragohain and Konwar (2007) |
| <i>Justicia adhatoda</i> L. (= <i>Adhatoda zeylanica</i> Medik.) | Acanthaceae | Chandauli District, Uttar Pradesh, India | Root | Paste with water | Topical | Dandruff | Poonam and Singh (2009) |
| <i>Curcuma aromatica</i> Salisb. | Zingiberaceae | Assam | Leaf | Paste | Topical | Abscesses | Buragohain and Konwar (2007) |
| <i>Bombax ceiba</i> L. | Malvaceae | Assam | Rhizome | Paste | Topical | Scabies, ringworm | Buragohain and Konwar (2007) |
| <i>Mallotus philippensis</i> (Lam.) Müll. Arg. | Euphorbiaceae | Mysore and Coorg District, Karnataka | Bark | Paste | Topical | Cattle wounds | Kshirsagar and Singh (2001) |
| <i>Argemone mexicana</i> L. | Papaveraceae | Chandauli District, Uttar Pradesh, India | Leaves | Paste | Topical | Skin diseases | Singh and Singh (2009) |
| <i>Tabernaemontana undulata</i> Vahl | Apocynaceae | Terai Arc Landscape, India | Seed | Oil | Topical | Skin infection, leprosy | Poonam and Singh (2009) |
| <i>Arisaema jacquemontii</i> Blume | Araceae | Niyamgiri hills, Odisha, India | Seed | Paste | Topical | Eczema | Kumar et al. (2012) |
| <i>Woodfordia fruticosa</i> (L.) Kurz | Lythraceae | Rajasthan, India | Bark, branch | Extraction | Chewed | Syphilis | Sharma and Kumar (2011) |
| <i>Christella dentata</i> (Forssk.) Brownsey & Jermy | Thelypteridaceae | West Himalaya, India | Bulb | Paste | Topical | Ringworm, skin disease | Pant and Samant (2010) |
| <i>Euphorbia hirta</i> L. | Euphorbiaceae | Betul District, Madhya Pradesh, India | Leaf | Paste | Topical | Skin diseases | Dahare and Jain (2010) |
| <i>Pongamia pinnata</i> (L.) Pierre | Fabaceae | Niyamgiri hills, Odisha, India | Leaf | Paste with coconut oil | Topical | Skin infection | Kumar et al. (2012) |
| <i>Erythroxylum monogynum</i> var. <i>caffrum</i> Eyles | Erythroxylaceae | Niyamgiri hills, Odisha, India | Whole plant | Paste | Topical | Cuts | Kumar et al. (2012) |
| <i>Anisochilus carnosus</i> (L. f.) Wall. | Lamiaceae | Niyamgiri hills, Odisha, India | Whole plant | Paste | Topical | Eczema, ringworm, skin diseases | Jain (2003) |
| <i>Cassia fistula</i> L. | Fabaceae | Niyamgiri hills, Odisha, India | Seed | Oil | Topical | Eczema, skin infection | Kumar et al. (2012) |
| | | Nellore District, Andhra Pradesh, India | Root, stem bark | Juice | Topical | Affected skin | Savithamma et al. (2012) |
| | | Odisha | Leaf | Crush | Topical | Itching | Saxena and Dutta (1975) |
| | | | | Extract | Topical | | |

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Table 1 (continued)

| Plant name | Family | Locality (state) | Plant parts | Preparation methods | Administration modes | Used against | Reference |
|--|------------------|---|----------------------|---|--|---|-----------------------------------|
| | | Jalgon District, Maharashtra | Leaves, flower, bark | | | Ringworm, skin infection, astringent | (D. L. Jain et al., 2010) |
| <i>Sonchus arvensis</i> L. | Asteraceae | Jalgon District, Maharashtra | Whole plant | Paste | Topical | Leprosy, white spot of skin, ringworm | (D. L. Jain et al., 2010) |
| <i>Streblus asper</i> Lour. | Moraceae | Erode District, Tamil Nadu | Leaf | Paste | Topical | Measles like swellings on the skin | Revathi and Parimelazhagan (2010) |
| <i>Lantana camara</i> L. | Verbenaceae | Erode District, Tamil Nadu | Leaf | Paste | Topical | Wounds | Revathi and Parimelazhagan (2010) |
| <i>Terminalia bellirica</i> (Gaertn.) Roxb. | Combretaceae | Erode District, Tamil Nadu | Bark, root | Extract | Topical | Unnecessary peelings on the skin | Revathi and Parimelazhagan (2010) |
| <i>Indigofera aspalathoides</i> DC. | Fabaceae | Erode District, Tamil Nadu | Whole plant | Ash of the whole plant was added with coconut oil | Topical | Psoriasis | Revathi and Parimelazhagan (2010) |
| <i>Bauhinia variegata</i> L. | Fabaceae | India | Bark | Extract | Topical | Leprosy, skin diseases | Jain (2003) |
| <i>Momordica charantia</i> L. | Cucurbitaceae | India | Fruit | Juice | Topical | Eczema | Jain (2003) |
| <i>Tectona grandis</i> L. f. | Lamiaceae | Palamalai hills, Coimbatore, Tamil Nadu | Leaf | Paste | Oral | Ulcer, skin diseases | Umapiya et al. (2011) |
| <i>Aristolochia bracteolata</i> Lam. | Aristolochiaceae | Palamalai hills, Coimbatore, Tamil Nadu | Whole plant | Decoction | Topical | Boils, skin disease | Umapiya et al. (2011) |
| <i>Bryophyllum pinnatum</i> (Lam.) Oken (= <i>Kalanchoe pinnata</i> (Lam.) Pers.) | Crassulaceae | Arunachal Pradesh, India | Leaf, stem | Crush | Topical | Skin burn | Namsa et al. (2009) |
| <i>Bidens pilosa</i> L. | Asteraceae | Arunachal Pradesh, India | Leaf | Paste | Topical | Cut, wound | Namsa et al. (2009) |
| <i>Alpinia galanga</i> (L.) Willd. | Zingiberaceae | Arunachal Pradesh, India | Rhizome | Rhizome powder mixed with young leaf paste of <i>Euphorbia nerifolia</i> L. | Topical | Local inflammation, skin allergy caused by insect bites or microbes | Namsa et al. (2009) |
| <i>Oxalis corniculata</i> L. (= <i>Oxalis corniculata</i> subsp. <i>albicans</i> (Kunth) Lourteig) | Oxalidaceae | Chittoor District, Andhra Pradesh, India | Whole plant | Juice | Gently rubbed on the skin for 2–5 days and also it can be taken orally for a month | Skin allergies | Jyothi et al. (2010) |
| <i>Plumbago zeylanica</i> L. | Plumbaginaceae | Chittoor District, Andhra Pradesh, India | Root, bark | Diluted paste | Applied on the skin for a period of 3–4 weeks | Leucoderma, ringworm | Jyothi et al. (2010) |
| <i>Tinospora sinensis</i> (Lour.) Merr. (= <i>Tinospora cordifolia</i> (Willd.) Miers) | Menispermaceae | Chittoor District, Andhra Pradesh, India | Root | Paste | Topical | Leprosy | Jyothi et al. (2010) |
| <i>Cullen corylifolium</i> (L.) Medik. (= <i>Psoralea corylifolia</i> L.) | Fabaceae | Chittoor District, Andhra Pradesh, India | Seed | Powder | Topical | Leucoderma, psoriasis, leprosy, inflammatory diseases of skin | Jyothi et al. (2010) |
| <i>Raphanus raphanistrum</i> subsp. <i>sativus</i> (L.) Domin (= <i>Raphanus sativus</i> L.) | Brassicaceae | Valsad district, Gujarat | Seed | Paste with little ghee | Topical | Fungal infection | Shah et al. (2011) |
| <i>Solanum viarum</i> Dunal | Solanaceae | Morigaon district, Assam, India | Root | Paste | Topical | Leprosy | Deka and Deka (2007) |
| <i>Randia dumetorum</i> (Retz.) Poir. | Rubiaceae | Central Western Ghats, Karnataka, India | Leaf | Paste | Topical | Skin ailments | Bhat et al. (2014) |
| <i>Cynodon dactylon</i> (L.) Pers. | Poaceae | South Travancore, Kanyakumari district, Tamil Nadu, India | Leaf | Mixed with coconut oil; paste | Topical | Skin ailments | Jeeva et al. (2007) |
| <i>Lawsonia inermis</i> L. (= <i>Lawsonia alba</i> Lam.) | Lythraceae | Hyderabad, Karnataka, India | Leaf | Paste | Topical | Heal cuts and wounds | Policepatel and Manikrao (2013) |
| <i>Bridelia retusa</i> (L.) Spreng. | Phyllanthaceae | Coastal region, Karnataka, India | Stem bark | Paste | Topical | Herpes | Bhandary and Chandrashekar (2011) |
| <i>Euphorbia thymifolia</i> Burm. | Euphorbiaceae | low hill region, Uttarakhand, India | Whole plant | Paste | Topical | Heal itching | Sharma et al. (2014) |
| <i>Tridax procumbens</i> (L.) L. | Asteraceae | Sub-Himalayan tract, India | Leaves | Paste | Topical | Cuts, wound | Sharma et al. (2014) |
| | Malvaceae | | Leaves | Paste | Topical | Boils | Sharma et al. (2014) |

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Table 1 (continued)

| Plant name | Family | Locality (state) | Plant parts | Preparation methods | Administration modes | Used against | Reference |
|--|----------------|---|----------------|---|----------------------|---|------------------------------------|
| <i>Sida cordata</i> (Burm. f.) Borss. Waalk. | | Udham Singh Nagar, Uttarakhand, India | | | | | |
| <i>Millettia extensa</i> (Benth.) Baker | Fabaceae | Udham Singh Nagar, Uttarakhand, India | Root | Paste | Topical | Boils, sores | Sharma et al. (2014) |
| <i>Caesulia axillaris</i> Roxb. | Asteraceae | Udham Singh Nagar, Uttarakhand, India | Flower | Paste | Topical | Cuts, wound | Sharma et al. (2014) |
| <i>Vanda tessellata</i> (Roxb.) Hook. ex G. Don | Orchidaceae | Udham Singh Nagar, Uttarakhand, India | Root | Paste | Topical | Boils, carbuncle | Sharma et al. (2014) |
| <i>Ehretia laevis</i> Roxb. | Boraginaceae | Udham Singh Nagar, Uttarakhand, India | Leaves | Paste | Topical | Wound | Sharma et al. (2014) |
| <i>Eulaliopsis binata</i> (Retz.) C.E. Hubb. | Poaceae | Udham Singh Nagar, Uttarakhand, India | Leaves | Juice | Topical | Wound | Sharma et al. (2014) |
| <i>Rotheca serrata</i> (L.) Steane & Mabb. (= <i>Clerodendrum serratum</i> (L.) Moon) | Lamiaceae | Dudhwa National Park, Lakhimpur Kheri District, Uttar Pradesh, India | Leaves | Mixed with mustard oil, paste | Topical | Eye-lid inflammation | Kumar and Bharati (2014) |
| <i>Cyclea peltata</i> Hook. f. & Thomson | Menispermaceae | Uttara Kannada district, Karnataka, India | Leaves | Paste | Topical | Boils | Bhat et al. (2014) |
| <i>Vitex trifolia</i> L. | Lamiaceae | Thodu hills, Kerala, India | Plant | Juice/decoction | Oral and topical | Wound | Xavier et al. (2014) |
| <i>Jasminum grandiflorum</i> L. | Oleaceae | Central Western Ghats, Karnataka, India | Leaves | Paste | Topical | Boils, eczema | Bhat et al. (2014) |
| <i>Quercus infectoria</i> G. Olivier | Fagaceae | South Travancore, Kanyakumari district, Tamil Nadu, India | Seed | Mixed with seeds of <i>Terminalia chebula</i> (Gaertn) Roxb. | Topical | Rash | Jeeva et al. (2007) |
| <i>Verbena officinalis</i> L. | Verbenaceae | Tarai region, Uttarakhand, India | Leaves | Crushed | Topical | Boils, cuts, wounds | Pandey et al. (2012) |
| <i>Cymbopogon citratus</i> (DC.) Stapf | Poaceae | Wayanad district, Kerala, India | Root, leaf | Paste, oil | Topical | Sprain and toothache | Silja et al. (2008) |
| <i>Millingtonia hortensis</i> L. f. | Bignoniaceae | Mayabunder, Andaman, India | Root | Paste | Topical | Skin disease, fever and pain | Chander et al. (2015) |
| <i>Mimosa pudica</i> L. | Fabaceae | Central Western Ghats, Karnataka, India | Whole Plant | Paste | Topical | Dermatitis, ringworm infection, eczema | Bhat et al. (2014) |
| <i>Amaranthus spinosus</i> L. | Amaranthaceae | Hyderabad, Karnataka, India | Whole Plant | Juice | Topical | Skin allergy | Policepatel and Manikrao (2013) |
| <i>Achyranthes aspera</i> L. | Amaranthaceae | Central Western Ghats, Karnataka, India | Plant, leaf | Paste | Topical | Boil, eczema | Bhat et al. (2014) |
| <i>Nerium oleander</i> L. (= <i>Nerium indicum</i> Mill.) | Apocynaceae | Pachamalai Hills, Tamil Nadu, India | Stem bark | Juice, mixed with gingly oil | Topical | Ear ache | Bhaskar and Samant (2012) |
| <i>Nerium oleander</i> L. (= <i>Nerium indicum</i> Mill.) | Apocynaceae | Kalavai, Vellore district, Tamil Nadu, India | Fruit | Paste | Topical | Ear pain, rash | Natarajan et al. (2013) |
| <i>Heliotropium indicum</i> L. | Boraginaceae | Kancheepuram district, Tamil Nadu, India | Leaf | Paste | Topical | Wounds and skin affections | Muthu et al. (2006) |
| <i>Vitex negundo</i> L. | Lamiaceae | Bhandara district, Maharashtra, India | Leaf | Mixed with a number of herbs; juice | Topical | Boils | (R. Gupta et al., 2010) |

B. Clarke [unresolved name, Gentianaceae], *Vitex negundo* L. (Lamiaceae), etc. (Saikia et al., 2006). In Tamil Nadu, SE India, Chendurpandy et al. (2010) investigated 88 plants that cure 15 different types of skin diseases; a maximum of 46 plants are used to treat ringworm infection, followed by 41 species for scabies, 24 for eczema, nine for leprosy, seven for cuts and wounds, five plants each for dandruff and skin inflammation, three for alopecia, two plants each for healing the cracked foot, cysts, polyps, acne and prickle heat (Fig. 2) (Chendurpandy et al., 2010). These 88 plants belong to 52 families such as Celastraceae, Asteraceae, Euphorbiaceae, Fabaceae, Apocynaceae, Aristolochiaceae, Cucurbitaceae, Malvaceae, Rubiaceae, Zingiberaceae

(Figs. 6 and 7) and some common plants in this study are *Azadirachta indica* A. Juss., *Curcuma aromatica* Salisb (Zingiberaceae), *Aloe vera* (L.) Burm. f. (Asphodelaceae), *Canthium coromandelicum* (Burm. f.) Alston (= *Canthium parviflorum* Lam., Rubiaceae), *Heliotropium indicum* L. (Boraginaceae), etc. (Chendurpandy et al., 2010). Upadhyay et al. (1998) reported a total of 54 medicinal plants from 34 families for curing skin diseases such as eczema, itching, cuts, burns, leucoderma, etc. in Bihar, eastern India. In this study, most of the common families used are Fabaceae, Lamiaceae, Euphorbiaceae (Figs. 6 and 7), and some of the medicinal plants are *Cuscuta reflexa* Roxb. (Convolvulaceae), *Bombax ceiba* L. (Malvaceae), *Datura metel* L. (Solanaceae), *Indigofera tinctoria* L.,

Table 2
Pharmacological/preclinical evidence.

| Plant species and family name | Pharmacological activity | Extract/fractions/plant parts | In-vitro/in-vivo/ex-vivo assays/models | Reference |
|--|-----------------------------------|---|---|------------------------------|
| <i>Symplocos racemosa</i> Roxb. (Sympllocaceae) | Antimicrobial activity | Ethanol extract of bark | Disc diffusion method against <i>Propionibacterium acnes</i> and <i>Staphylococcus epidermidis</i> | Kumar et al. (2007) |
| <i>Ammannia baccifera</i> L. (Lythraceae) | Antimicrobial activity | Ethanol extract of root | Disc diffusion method against <i>Propionibacterium acnes</i> and <i>Staphylococcus epidermidis</i> | Kumar et al. (2007) |
| <i>Coccinia grandis</i> (L.) Voigt (= <i>Coccinia indica</i> Wight and Arn.) (Cucurbitaceae) | Anti-inflammatory activity | Fruit juice powder | Carrageenin and histamine induced paw oedema | Deokate and Khadabadi (2012) |
| <i>Azadirachta indica</i> A. Juss. (Meliaceae) | Anti-inflammatory activity | Chloroform extract of stem bark | Carrageenin induced paw oedema in rat and mouse ear | Biswas et al. (2002) |
| | Antiulcer activity | Leaf aqueous extract | Rat exposed in restraint | Biswas et al. (2002) |
| | Anti-inflammatory activity | Ethanol extract | Wister albino rats | Sharma et al. (2010) |
| | Wound healing activity | Pure neem oil and neem ointment | Incised and gap wounds in bovine calves | Bhardwaj and Sharma (1997) |
| <i>Nelumbo nucifera</i> Gaertn. (Nelumbonaceae) | Anti-inflammatory activity | Methanol extract | Oedema in rat paw caused by Carrageenan and serotonin | Paudel and Panth (2015) |
| | | Ethanol extract | Proliferation of PMBC induced by phytohemagglutinin | Paudel and Panth (2015) |
| <i>Citrus limon</i> (L.) Osbeck (Rutaceae) | Antimicrobial activity | Whole plant extract | Antimicrobial susceptibility assays | Otang and Afolayan (2016) |
| | Antioxidant activity | Whole plant extract | Assay of DPPH scavenging activity | Otang and Afolayan (2016) |
| <i>Aloe vera</i> (L.) Burm.f. (Asphodelaceae) | Antimicrobial/antifungal activity | Methanol and aqueous Extract | Reducing power assay | Yebpella et al. (2011) |
| <i>Cissus quadrangularis</i> L. (Vitaceae) | Antimicrobial activity | Methanol and aqueous Extract | 10% DMSO | Shah (2011) |
| <i>Celosia argentea</i> L. (Amaranthaceae) | Wound healing activity | Methanol extract | <i>S. aureus</i> , <i>E. coli</i> , <i>P. aeruginosa</i> | |
| | Wound healing activity | Alcoholic extract of flowers | Rat burn wound | Priya et al. (2004) |
| <i>Ixora coccinea</i> L. (Rubiaceae) | Wound healing activity | Alcoholic extract of flowers | Dead space rat | Nayak et al. (1999) |
| <i>Centella asiatica</i> (L.) Urb. (Apiaceae) | Wound healing activity | Ethanol extract | Incision, excision, and dead space | Suguna et al. (1996) |
| <i>Curcuma aromatica</i> Salisb. (Zingiberaceae) | Anti-inflammatory activity | Aqueous and alcoholic extracts | Mice | Sikha et al. (2015) |
| | Wound healing activity | Powdered rhizome | Rabbits | |
| | Repellent activity | Ethanol extract | Mosquito repellent activity | |
| | Anti-melanogenic activity | Plant extract | UVA-induced cellular oxidant formation and depletion of CAT and GPx activities and GSH content | |
| <i>Rubia cordifolia</i> L. (= <i>Rubia cordifolia</i> var. <i>affinis</i> Kurz) (Rubiaceae) | Wound healing activity | Root extract | Swiss albino mice | Karodi et al. (2009) |
| <i>Curcuma longa</i> L. (Zingiberaceae) | Anti-inflammatory activity | Rhizome extract | TNF induced NF-κB activation | Krup et al. (2013) |
| | Anti-allergic activity | Rhizome extract | 48/80-induced rat peritoneal mast cell (RPMC) degranulation, 48/80-induced systemic anaphylaxis | |
| | Anti-dermatophytic activity | Rhizome powder | Cow's urine | |
| <i>Biophytum sensitivum</i> (L.) DC. (= <i>Biophytum sensitivum</i> var. <i>assamicum</i> Edgew. & Hook. f.) (Oxalidaceae) | Anti-inflammatory activity | Methanol extract of aerial parts and roots | Carrageenin-induced rat paw oedema model | Jiny Varghese et al. (2010) |
| <i>Neolamarckia cadamba</i> (Roxb.) Bosser (= <i>Anthocephalus cadamba</i> (Roxb.) Miq.) (Rubiaceae) | Wound healing activity | Air-dried and powdered drug | Inbred house wistar rats, excision wound model, incision wound model | Sanjay et al. (2007) |
| <i>Lagenaria siceraria</i> (Molina) Standl. (Cucurbitaceae) | Anti-microbial activity | Methanol extracts of the leaves, seeds, and fruit-flesh | Agar-well diffusion method | Prajapati et al. (2010) |
| <i>Crocus sativus</i> L. (Iridaceae) | Antitumor activity | Ethanol extracts | Swiss albino mice | Vijaya Bhargava (2011) |
| <i>Ocimum tenuiflorum</i> L. (= <i>Ocimum sanctum</i> L.) (Lamiaceae) | Anti-inflammatory activity | Methanol extract | Acute (carrageenan-induced pedal oedema) and chronic (croton oil induced granuloma and exudate formation) inflammations in rats | Pandey and Madhuri (2010) |
| <i>Euphorbia hirta</i> L. (Euphorbiaceae) | Anti-inflammatory activity | n-hexane extract of aerial parts | Phorbol acetate-induced ear inflammation in mice | Kumar et al. (2010) |
| <i>Fumaria indica</i> (Hausskn.) Pugsley (Papaveraceae) | Antifungal activity | Alkaloid | Spore germination of some plant pathogenic fungi (<i>Collectotrichum</i> sp., <i>C. gloeosporioides</i> , <i>C. falcatum</i> , <i>Curvularia maculans</i> , <i>Curvularia lunata</i> , <i>Erysiphe cichoracearum</i>) | Gupta et al. (2012) |
| | Anti-inflammatory activity | Plant extract | Acute and chronic cotton models of inflammation in experimental animals | |
| <i>Eclipta prostrata</i> (L.) L. (= <i>Eclipta alba</i> (L.) Hassk.) (Asteraceae) | Antimicrobial activity | Methanol extract | Agar well diffusion method, DMSO | Dalal et al. (2010) |
| <i>Acorus calamus</i> L. (Acoraceae) | Wound healing activity | Ethanol extract of leaves | Rats of either sex | Sharma et al. (2014) |
| | Antioxidant activity | | DPPH assay | |

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Table 2 (continued)

| Plant species and family name | Pharmacological activity | Extract/fractions/plant parts | In-vitro/in-vivo/ex-vivo assays/models | Reference |
|--|--|---|---|--|
| <i>Ageratum conyzoides</i> (L.) L. (Asteraceae) | Wound healing activity Antioxidant activity | Methanol extract of leaf and rhizome Petroleum ether, chloroform, methanolic and aqueous extract Whole plant methanol extract and essential oil | Wistar albino rats of either sex (150–250 g) DPPH assay, Lipid peroxidases activity, FRAP assay | Sharma et al. (2014) |
| <i>Brassica juncea</i> (L.) Czern. (Brassicaceae) | Wound healing activity | Ethanol and aqueous extract | Male wistar albino rats (180–200g) | Sharma et al. (2014) |
| <i>Cassia fistula</i> L. (Fabaceae) | Wound healing activity Anti-inflammatory activity | Methanolic and aqueous extract of bark Extract of leaves | Wistar albino rats (170–200 g) of either sex Carrageenan, histamine and dextran induced paw oedema assays in rats | Sharma et al. (2014) Danish et al. (2011) |
| <i>Acalypha indica</i> L. (Euphorbiaceae) | Wound healing activity | Alcoholic extract of whole plant | Excision and incision rat models | Ayyanar and Ignacimuthu (2009) |
| <i>Anacardium occidentale</i> L. (Anacardiaceae) | Antiulcerogenic activity, acute toxicity | Ethanol extract of leaves | Wounded rats | Ayyanar and Ignacimuthu (2009) |
| <i>Chromolaena odorata</i> (L.) R.M.King & H.Rob. (= <i>Eupatorium odoratum</i> L.) (Asteraceae) | Anti-inflammatory activity | Aqueous extract of whole plant | Carrageenan induced oedema, cotton pellet granuloma and formalin induced oedema methods in rats | Ayyanar and Ignacimuthu (2009) |
| <i>Pongamia pinnata</i> (L.) Pierre (Fabaceae) | Wound healing activity Anti-inflammatory activity, ulcerogenic effect | Aqueous extract of the leaves Ethanol extract of leaves | Wounded rat models Acute, subacute and chronic models of inflammation in rats | Ayyanar and Ignacimuthu (2009) |
| <i>Vitex altissima</i> L.f. (Lamiaceae) | Wound healing activity | Leaf extracts | Excision, incision and dead space wounded rat models | Ayyanar and Ignacimuthu (2009) |
| <i>Cedrus deodara</i> (Roxb. ex D. Don) G. Don (Pinaceae) | Anti-inflammatory activity | Volatile oil extract of wood | Carrageenan induced rat paw oedema method | Chaudhary et al. (2011) |
| <i>Argemone mexicana</i> L. (Papaveraceae) | Wound healing activity | Petroleum ether and butanol fractions of ethanol extract | Excision wound albino rat model | Brahmachari et al. (2013) |
| <i>Ficus carica</i> L. (Moraceae) | Anti-inflammatory activity | Petroleum ether (pee), chloroform and ethanol extracts of leaves | Carrageenan-induced rat paw oedema | Badgujar et al. (2014) |
| <i>Hydnocarpus anthelminthicus</i> Pierre ex Laness. (Achariaceae) | Wound healing activity | Aqueous methanol extract | STZ-induced diabetic male ICR mice | Sahoo et al. (2014) |
| <i>Cleome viscosa</i> L. (Cleomaceae) | Anti-inflammatory activity | Methanol extract | Carageenin, histamine, and dextran induced rat paw oedema | Mali (2010) |
| <i>Rauvolfia tetraphylla</i> L. (Apocynaceae) | Anti-inflammatory activity | Different extracts of root bark | Carrageenan induced acute inflammation in rats | Iqbal et al. (2013) |
| <i>Trichosanthes dioica</i> Roxb. (Cucurbitaceae) | Wound healing activity | Methanolic fruit extract | Excision and incision wound model in rat | Shivhare et al. (2010) |
| <i>Senna alata</i> (L.) Roxb. (= <i>Cassia alata</i> L.) (Fabaceae) | Anti-fungal activity | Leaf extract mixed with soap | Superficial skin infections caused by <i>Tinea versicolor</i> and <i>Tinea corporis</i> in human | Oladele et al. (2010) |
| <i>Senna alata</i> (L.) Roxb. (Fabaceae) | Anti-bacterial | Crude leaf extract | Growth inhibition of infection causing bacteria | Mensah Donkor (2016) |
| <i>Cynodon dactylon</i> (L.) Pers. (Poaceae) | Wound healing activity | Aqueous plant extract | Acute and dermatological toxicity study in Wister rat model | Biswas et al. (2017) |
| <i>Achyranthes aspera</i> L. (Amaranthaceae) | Wound healing activity | Methanolic leaf extract | Excision and incision wound model in albino rat | Fikru et al. (2012) |
| <i>Nerium oleander</i> L. (Apocynaceae) | Wound healing activity | <i>aloe vera</i> based plant extract (nae-8®) | Thermal injury-induced alterations and DNA repair mechanism in rat model | Gul Akgun (2017) |
| <i>Mimosa pudica</i> L. (Fabaceae) | Anti-dermatophilosis activity | Polyphenol rich seed extract | Topical emulgel based skin parameter study in human subjects | Ijaz et al. (2019) |
| <i>Cymbopogon citratus</i> (DC.) Stapf (Poaceae) | Mosquito repellent activity | Essential oil | Relative protection study against <i>Anopheles arabiensis</i> mosquitoes | Solomon et al. (2012) |
| <i>Couroupita guianensis</i> Aubl. (Lecythidaceae) | Skin care activity | Hydroalcoholic leaf extracts | Stimulation of human skin fibroblast (HSF) proliferation and UV-absorption study <i>in-vitro</i> | Martnez et al. (2012) |
| <i>Mucuna pruriens</i> (L.) DC. (Fabaceae) | Skin care activity | Plant extract | Sympathetic skin vasoconstrictions and axon reflex study | Kosteletzky et al. (2009) |
| <i>Hibiscus syriacus</i> L. (Malvaceae) | Chronic UVB induced skin damage protection | Dietary enzyme-treated flowers | Skin hydration and collagen synthesis studied in normal human dermal fibroblasts (NHDFs) <i>in vitro</i> and hairless mice <i>in vivo</i> | Yang et al. (2019) |
| <i>Vitex trifolia</i> L. (Lamiaceae) | Wound healing activity | Ethanol leaf extracts | Excision, incision and dead space wound models. in rats | Manjunatha et al. (2007) |
| <i>Vitex doniana</i> Sweet (Lamiaceae) | Cutaneous wound healing activity | Ethanol plant extracts | Excision, incision and models in rats, planimetry and histological analysis. | Amegbor et al. (2012) |
| <i>Hibiscus syriacus</i> L. (Malvaceae) | Hair growth promoting activity | petroleum ether, benzene, chloroform, methanol, and water extracts of leaves | Hair growth initiation assay in albino mice | Punasiya et al. (2014) |
| <i>Vitex altissima</i> L. (Lamiaceae) | Wound healing activity | Methanolic leaf extract | Excision, incision and dead space wounded rat models | Manjunatha et al. (2007) |
| <i>Couroupita guianensis</i> Aubl. (Lecythidaceae) | Anti-bacterial activity | Alcoholic leaf extract | Resistance model against nosocomial infection induced bacteria | Roumy et al. (2015) |

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Table 2 (continued)

| Plant species and family name | Pharmacological activity | Extract/fractions/plant parts | In-vitro/in-vivo/ex-vivo assays/models | Reference |
|--|--------------------------|--|--|------------------------|
| <i>Heliotropium indicum</i> L. (Boraginaceae) | Wound healing activity | Petroleum ether, chloroform, methanol, and aqueous leaf extracts | Excision, incision and dead space wounded rat models | Dash and Murthy (2011) |

etc. (Upadhyay et al., 1998). Ninety plant species from 86 genera and 48 families were used by the Tharu community (in district Udham Singh Nagar, Uttarakhand) in the treatment of different skin diseases, namely, body infection (2 species (spp.)), boils (32 spp.), burns (4 spp.), chilblains (2 spp.), cracked heels (2 spp.), cuts (18 spp.), dandruff (3 spp.), eczema (10 spp.), hair fall (2 spp.), itching (7 spp.), leprosy (11 spp.), leucoderma (4 spp.), ringworm (5 spp.), toes infection (2 spp.), and wounds (38 spp.) (Fig. 2). According to the use value (UV), the plant species most preferred to cure cutaneous diseases was *Ricinus communis* L. (Euphorbiaceae) followed by *Azadirachta indica* A. Juss., *Ageratum conyzoides* (L.) L. (Asteraceae), *Allium cepa* L. (Amaryllidaceae), and *Tridax procumbens* (L.) L. (Asteraceae) (Sharma et al., 2014). The medicinal plants used for the treatment of skin-related disorders in the northeastern part of India, including the states of Arunachal Pradesh (Fig. 8), Manipur, Meghalaya, Mizoram, Nagaland (Fig. 8), and Tripura were reported by Begum and Nath (2000) (Begum and Nath, 2000). In this study, they documented 275 plant species belonging to 93 plant families, and the most common plants used were *Lepidagathis incurva* Buch.-Ham. ex D. Don (Acanthaceae), *Achyranthes aspera* L. (Amaranthaceae), *Nerium oleander* L. (= *Nerium indicum* Mill., Apocynaceae), *Centella asiatica* (L.) Urb., etc. (Begum and Nath, 2000).

Ethnopharmacological approaches and information on lesser-known Indian plants used in the treatment of cuts, wounds, and burns have been compiled (Kumar et al., 2007) (Singh and Lal, 2008). Here, 51 plant species were indicated with 31% of the plants used to heal wounds, 29% for cuts and 10% for burns, and 22% for cuts and wounds (Fig. 2). An additional 64 Indian plants were reported with wound healing activity.

2.3. Pharmacological activities in preclinical and clinical evidences and cellular studies

Depending on dermatology ethnomedicinal use, plants have been confirmed by several pharmacological, clinical, and preclinical studies. The following section provides a comprehensive overview of the many experiments undertaken to test the bioactivity of plant extracts, fractions, and compound derivatives in *in vitro*, *ex-vivo*, and *in vivo* tests for some specific skin disorders. Tables 2 and 3 show the pharmacological investigations. Biological activities of plant extracts are anti-inflammatory, antifungal, wound healing, antiulcer, and antioxidant (Fig. 2). Most of the plant extracts are prepared using ethanol or methanol. Skin health is maintained by directly acting on the various layers and cells of the skin involved in the ageing process of skin dysfunction, as well as in the pathogenesis of the disease (Tirant et al., 2018).

Leprosy: Hansen's disease (HD) or leprosy is an infectious chronic disease caused by slow-growing bacteria, *Mycobacterium leprae* or *M. lepromatosis*. The serious disease mainly affects the skin, the upper respiratory tract (mucous surfaces), peripheral nerves, and eyes. The symptoms are skin lesions, nerve damage, red or light-colored skin patches with weakness in hands and feet, numbness, and reduced sensation. It is transmitted via droplets from the nose and mouth, during close and frequent contact with untreated cases. Leprosy is cured by multidrug therapy (MDT). Deka and Deka (2007) reported many of the medicinal plants used to treat leprosy such as leaves or root paste of *Amaranthus spinosus* L. (Amaranthaceae), and *Withania somnifera* (L.) Dunal (Solanaceae), bark decoction of *Cassia fistula* L. (Fabaceae), whole plant decoction of *Centella asiatica* (L.) Urb., root paste of *Solanum vilarum* Dunal (Solanaceae), or fruit paste of *Momordica charantia* L.

(Cucurbitaceae) (Deka and Deka, 2007; Paul et al., 2021). Joseph et al. (2013) documented that *Mimosa pudica* L. (Fabaceae) also helps to treat leprosy (Joseph et al., 2013). The antimicrobial activity of *M. pudica* methanol extract was checked against *Aspergillus fumigatus*, *Citrobacter diversens*, and *Klebsiella pneumoniae* at different concentrations of 50, 100 and 200 µg/disc. The anti-inflammatory effect of *M. pudica* leaf ethanol extract at doses of 200 and 400 mg/kg was tested for significant inhibition of carrageenan paw oedema and found that the analgesic effect was more pronounced with the acetic acid writhing model than with the tail-flick model (Joseph et al., 2013).

Kumar et al. (2014) studied the extraction of the stem bark of *Millingtonia hortensis* L. f. (Bignoniaceae) used to treat Dapsone resistance in leprosy (Kumar et al., 2014). The antibacterial activity of the extract of *M. hortensis* was examined by Well diffusion method. In this method, *Escherichia coli* was used as the test organism, four wells of 10 mm in diameter were made in all agar plates, and approximately 0.3 ml of plant extract of varying concentrations was added into the well with a sterilized dropper pipette and the kanamycin disc was added at the center of the plates which were then incubated at 37 °C for 20 h, thus the diameter of the inhibition zone was recorded to determine the antimicrobial activity (Kumar et al., 2014). In this study also, GC-MS analysis was performed on the methanolic and ethanolic extracts of *M. hortensis*.

Singh et al. (2013) studied the anti-inflammatory effect of methanol extract (400 mg/kg) of the *Cassia* species (Fabaceae), which showed a maximum setback of oedema of 53.57%, 40.33%, 31.37% and 29.15% at the end of 3 h with serotonin-induced, carrageenin, dextran, and histamine rat paw oedema and the extract exhibited a 48.13% reduction in granuloma weight in the granuloma pouch in rats by using the chronic test. The antiulcer activity of the methanolic seed extract of *Cassia* species was explored using a pylorus ligation model (75% protection) and 70.31% protection in ulcers induced by indomethacin in Wistar albino rats (Shivjeet et al., 2013). The antipyretic activity of the root extract of *Calotropis gigantea* (L.) Dryand. (Apocynaceae) concluded yeast-induced fever and TAB (Typhoid) vaccine-induced pyrexia in rats and rabbits (Singh et al., 2014).

Ringworm: Ringworm is a fungal infection of the skin or scalp, also known as dermatophytosis, dermatophyte infection, or tinea. Three various kinds of fungi can cause ringworm: *Epidermophyton* sp., *Microsporum* sp., and *Trichophyton* sp. These fungi live in the soil in the form of spores. The infection initially appears as red spots on the affected areas of the skin and later spreads to other body parts. It can affect the scalp, groin, chin, feet, nails, or other areas. Singh et al. (2015) reported *Lawsonia inermis* L. was used as a treatment of ringworm, it has anti-inflammatory properties concluded by the extracted compounds, i. e., methyl naphthalene carboxylate and 1,5-Diphenylpent-3-en-1-ynes (obtained from leaves and stems) used in an *in-vitro* bioassay by measuring elastase release and superoxide production in human neutrophils (IC50 range was 1.58–1.80 µg/ml) and the crude ethanol leaf extract along with butanol, chloroform, and water fractions exhibited against the anti-inflammatory model of rat (Singh et al., 2015). Furthermore, this study documented the wound healing effect of *L. inermis* ethanol leaf extract on male Wistar rats in excision and dead space models and excision, incision and dead space models in Sprague Dawley rats (Singh et al., 2015). Ndhkala et al. (2015) studied *Achyranthes aspera* L. for treating ringworm. The antifungal activity of the methanol extracts from *A. aspera* leaves against *Candida albicans* (ATCC10231), a diploid yeast fungus, was assessed by means of a micordilution test. Its anthelmintic activity was also assessed against

Table 3
Clinical evidence and confirmation in cellular research.

| Plant species and family name | Extract/plant parts/fractions | Mode of action in human | References |
|--|---|---|---|
| <i>Calendula officinalis</i> L. (Asteraceae) | Aqueous extract of flowers | Stimulates the phagocytosis of human granulocytes, interferes neutrophil radical oxygen species (ROS) and radical nitrogen species (RNS) generation, particularly nitric oxide, a reduced pro-inflammatory marker including TNF - α , IL - 1 β , (IL - 6), interferon gamma (IFN - γ), c - reactive protein (CRP), and cyclooxygenase - 2 (COX - 2) | (Tirant et al., 2018) (Kodiyan and Amber, 2015) |
| | Aqueous extract of flowers, water fraction | Normal human dermal fibroblasts (HDF) cell proliferation and migration | Dinda et al. (2016) |
| | Isolated calendula oil from calendula flowers | SPF of sunscreen formulations [calculated as the ratio of the minimal erythema dose (MED) of sunscreen-protected skin to the MED of unprotected skin] | Mishra et al. (2012) |
| <i>Centella asiatica</i> (L.) Urb. (Apiaceae) | Ethanol extract of whole plant | Human monolayer cell culture, activates the smad pathway, increases collagen synthesis in human dermal fibroblasts cell, affects extracellular matrix proteins deposition | (Tirant et al., 2018) (Gohil et al., 2010) (Seevaratnam et al., 2012) |
| <i>Aloe vera</i> (L.) Burm.f. (Asphodelaceae) | Ethanol extract | Synthesis of hyaluronic acid and dermatan sulfate in the granulation tissue, fibroblast's activity and proliferation resulting in significantly increased collagen synthesis, Glucosaminan and gibberellin interact with growth factor receptors on the fibroblast | Tirant et al. (2018) |
| <i>Phyllanthus emblica</i> L. (Phyllanthaceae) | Ethanol extract | Human colon cancer cell lines, human AML leukemic cells | Joseph and Justin Raj (2010) |
| | Aqueous extract of dried fruit | Chromium-induced oxidative damage through decreased GSH and GPx activity in macrophages, inhibition of matrix metalloproteinase levels, MMP-1, hyaluronidase activities, promotion of pro-collagen content UV-B induced skin photo ageing in fibroblasts by MTT assay, marked up-regulation of cytokine (il-10) concentration, efficient reduction of cytokine (tnf- α and il-1 β) levels | (Tirant et al., 2018) (Dang et al., 2011) |
| <i>Terminalia chebula</i> Retz. (Combretaceae) | Extract of fruits | Enhances concentration of melatonin in the pineal gland and the levels of cytokines, increases concentration of antioxidant enzymes, GSH, T and B cell | Tirant et al. (2018) |
| <i>Cullen corylifolium</i> (L.) Medik. (= <i>Psoralea corylifolia</i> L.) (Fabaceae) | Aqueous extract | Human cell lines, human liver-derived HepG2 cells, keratinocytes replication in psoriasis, inhibition on IL-6-induced STAT3 activation and phosphorylation | (Tirant et al., 2018) (Khushboo et al., 2010) |
| <i>Azadirachta indica</i> A. Juss. (Meliaceae) | Aqueous extract of bark | Human serum | Biswas et al. (2002) |
| | Aqueous extract of leaves | IgM and IgG levels along with increased titer of anti-ovalbumin antibody | |
| | Extract of leaves | Certain human fungi, including <i>Trichophyton</i> sp., <i>Epidermophyton</i> sp., <i>Microsporium</i> sp., <i>Trichosporon</i> sp., <i>Geotricum</i> sp. and <i>Candida</i> sp. | |
| <i>Curcuma longa</i> L. (Zingiberaceae) | Extracts of rhizomes | H ₂ O ₂ -induced damage in human keratinocytes and fibroblasts and in NG 108-15 cells, human erythrocyte by oxidizing oxyhaemoglobin | Chattopadhyay et al. (2004) |
| <i>Datura metel</i> L. (Solanaceae) | Methanol extract of flowers | Human lung carcinoma cells, human colorectal adenocarcinoma cells | Al-Snafi (2017) |
| <i>Calotropis gigantea</i> (L.) Dryand. (Apocynaceae) | Extract of latex | Human fibrinogen, human blood sample, human blood clot and plasma clot | Rajesh et al. (2005) |
| <i>Ocimum tenuiflorum</i> L. (= <i>Ocimum sanctum</i> L.) (Lamiaceae) | Ethanol extract | Human keratinocyte cell lines, human A549 lung cancer cells, human fibrosarcoma | Baliga et al. (2013) |
| <i>Hibiscus syriacus</i> L. (Malvaceae) | Ethanol cell culture extract | cells, cultured human lymphocytes Stimulated fibronectin and collagen synthesis in human HaCaT keratinocytes and Human Dermal Fibroblasts (HDF) | Di Martino et al. (2017) |

Caenorhabditis elegans var. Bristol (N2), a free-living nematode, using the rapid colorimetric microdilution test with modifications to obtain the minimum lethal concentration (MLC) values (Ndhala et al., 2015). Ibrahim and Osman (1995) reported the leaves of *Senna alata* (L.) Roxb (= *Cassia alata* L.). have a laxative effect and are also used against ringworms. These plant extracts were utilised for antimicrobial activity test by the diffusion method, with 1 ml of 105 bacterial cells inoculum or 4 × 10⁵ yeast cells or fungal spores per plate and discs with concentration 500 mg/ml were incubated at 30 °C for three days for fungi, or at 37 °C overnight for bacteria (Ibrahim and Osman, 1995). *In vitro* results showed that the extract had a high level of activity against dermatophytic fungi but a low level of activity against non-dermatophytic fungi.

Acne: Acne is the most common disease among all skin problems and has three major forms: *acne vulgaris*, *acne conglobata*, and *acne rosacea*. This is a skin condition that occurs when hair follicles become clogged with oil and dead skin cells. It often causes blackheads, whiteheads, or pimples, appearing on the face, forehead, chest, upper back, and arms. Manvitha and Bidya (2014) reported that *Cymbopogon citratus* (DC.) Stapf (Poaceae) (lemongrass) helps improve the skin by reducing acne and pimples and acts as a muscle and tissue toner (Karkala Manvitha, 2014). The ethanol extracts of the leaves of lemongrass were explored to find potential antimicrobial properties against *Staphylococcus aureus*

and both flavonoids and tannins are responsible for this activity as well (Karkala Manvitha, 2014). Kumar et al. (2007) documented the anti-bacterial screening of 12 Indian medicinal plants against acne-causing bacteria (Kumar et al., 2007). In this study, 12 medicinal ethanol plant extracts such as 300 g *Verbena officinalis* L. (Verbenaceae, roots, 20.2% w/w), *Quercus infectoria* G. Olivier (Fagaceae, fruits, 19.1% w/w), *Berberis aristata* DC. (Berberidaceae, roots, 25.4% w/w), *Cocos nucifera* (Arecaceae, seeds, 19.9% w/w), *Couroupita guianensis* Aubl. (Lecythidaceae, roots, 16.9% w/w), *Jasminum grandiflorum* L. (Oleaceae, flowers, 17.3% w/w), *Mucuna pruriens* (L.) DC. (Fabaceae, seeds, 20.1% w/w), *Symplocos racemosa* Roxb. (Symplocaceae, barks, 19.5% w/w), *Tectona grandis* L. f. (Lamiaceae, roots, 17.7% w/w), *Hibiscus syriacus* L. (Malvaceae, roots, 18.4% w/w), *Ammannia baccifera* L. (Lythraceae, roots, 15.6% w/w), and *Vitex trifolia* L. (Lamiaceae, roots, 12.5% w/w) were tested for antimicrobial activity against *Propionibacterium acnes* and *Staphylococcus epidermidis* by disc diffusion method. This method showed that only seven medicinal plants (*Ammannia baccifera*, *Berberis aristata*, *Couroupita guianensis*, *Hibiscus syriacus*, *Mucuna pruriens*, *Quercus infectoria*, *Symplocos racemosa*) prevent the growth of *Propionibacterium acnes* (Kumar et al., 2007).

Eczema: A particular type of skin inflammatory reaction where there is erythema (reddening), oedema (swelling), and papules (bumps) on

Table 4
Ayurvedic formulations in dermatology.

| Ayurvedic formulation | Ingredients | Used as/against | Reference |
|------------------------------|---|--|----------------------------|
| Navayasa Rasayana Leha | <i>Phyllanthus emblica</i> L. (= <i>Emblca officinalis</i> Gaertn.) (Phyllanthaceae), <i>Terminalia bellirica</i> (Gaertn.) Roxb. (Combretaceae), <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Embelia ribes</i> Burm.f. (Primulaceae), <i>Plumbago zeylanica</i> L. (Plumbaginaceae), <i>Semecarpus anacardium</i> L.f. [unresolved] (Anacardiaceae), <i>Cullen corylifolium</i> (L.) Medik. (= <i>Psoralea corylifolia</i> L.) (Fabaceae), Iron oxide, <i>Eclipta prostrata</i> (L.) L. (Asteraceae) | Anti-inflammatory, immunomodulatory activity, | Tirant et al. (2018) |
| Medhya Rasayana | <i>Acorus calamus</i> L. (Acoraceae), <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Nardostachys jatamansi</i> (D. Don) DC. (Caprifoliaceae), <i>Celastrus paniculatus</i> Willd. (Celastraceae), <i>Glycyrrhiza glabra</i> L. (Fabaceae), <i>Semecarpus anacardium</i> L.f. [unresolved] (Anacardiaceae), <i>Tinospora sinensis</i> (Lour.) Merr. (= <i>Tinospora cordifolia</i> (Willd.) Miers) (Menispermaceae), <i>Bacopa monnieri</i> (L.) Wettst. (Plantaginaceae), <i>Convolvulus prostratus</i> Forssk. (= <i>Convolvulus pluricaulis</i> Choisy) (Convolvulaceae) | Antiproliferative, antioxidant | Tirant et al. (2018) |
| Divya Kayakalpa Taila | <i>Azadirachta indica</i> A. Juss. (Meliaceae), <i>Berberis aristata</i> DC. (Berberidaceae), <i>Senna tora</i> (L.) Roxb. (= <i>Cassia tora</i> L.) (Fabaceae), <i>Cedrus deodara</i> (Roxb. ex D. Don) G. Don (Pinaceae), <i>Curcuma longa</i> L. (Zingiberaceae), <i>Phyllanthus emblica</i> L. (= <i>Emblca officinalis</i> Gaertn.) (Phyllanthaceae), <i>Leucas cephalotes</i> (Roth) Spreng. (Lamiaceae), <i>Nigella sativa</i> L. (Ranunculaceae), <i>Picrorhiza kurroo</i> Royle ex Benth. [unresolved] (Plantaginaceae), <i>Paullinia pinnata</i> L. (Sapindaceae), <i>Cullen corylifolium</i> (L.) Medik. (= <i>Psoralea corylifolia</i> L.) (Fabaceae), <i>Pterocarpus santalinus</i> L.f. (Fabaceae), <i>Rubia cordifolia</i> L. (Rubiaceae), <i>Sapindus trifoliatus</i> L. (Sapindaceae), <i>Smilax ornata</i> Lem. (Smilacaceae), <i>Sesamum indicum</i> L. (Pedaliaceae), <i>Solanum indicum</i> L. (Solanaceae), <i>Swertia chirata</i> Buch.-Ham. ex Wall. [unresolved name] (Gentianaceae), <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Tinospora sinensis</i> (Lour.) Merr. (= <i>Tinospora cordifolia</i> (Willd.) Miers) (Menispermaceae) | Skin diseases like ring worm, itching, sun burning, eczema, leucoderma, psoriasis, urticaria, skin allergy | (B. Aggarwal et al., 2011) |
| Divya Kesa Taila | <i>Abrus precatorius</i> L. (Fabaceae), <i>Bacopa monnieri</i> (L.) Wettst. (Plantaginaceae), <i>Berberis aristata</i> DC. (Berberidaceae), <i>Callicarpa macrophylla</i> Vahl (Lamiaceae), <i>Cyperus rotundus</i> L. (Cyperaceae), <i>Eclipta prostrata</i> (L.) L. (Asteraceae), <i>Phyllanthus emblica</i> L. (= <i>Emblca officinalis</i> Gaertn.) (Phyllanthaceae), <i>Fagonia chilensis</i> Hook. & Arn. (= <i>Fagonia cretica</i> var. <i>aspera</i> (Gay) Engl.) (Zygophyllaceae), <i>Indigofera tinctoria</i> L. (Fabaceae), <i>Mesua ferrea</i> L. (Calophyllaceae), <i>Nardostachys jatamansi</i> (D. Don) DC. (Caprifoliaceae), <i>Nelumbo nucifera</i> Gaertn. (Nelumbonaceae), <i>Onosma echioides</i> L. (Boraginaceae), <i>Pandanus tectorius</i> Parkinson ex Du Roi (Pandanaeae), <i>Pterocarpus santalinus</i> L.f. (Fabaceae), <i>Sida cordifolia</i> L. (Malvaceae), <i>Symplocos paniculata</i> (Thunb.) Miq. (= <i>Symplocos crataegoides</i> Buch.-Ham. ex D. Don) (Symplocaceae) | Hair fall, dandruff, alopecia, premature graying of hair | (B. Aggarwal et al., 2011) |
| Divya Kanti Lep | <i>Areca catechu</i> L. (Arecaceae), <i>Cinnamomum camphora</i> (L.) J. Presl (Lauraceae), <i>Curcuma amada</i> Roxb. (Zingiberaceae), <i>Curcuma longa</i> L. (Zingiberaceae), <i>Myristica fragrans</i> Houtt. (Myristicaceae), <i>Rubia cordifolia</i> L. (Rubiaceae), <i>Santalum album</i> L. (Santalaceae), <i>Valeriana wallichii</i> DC. (Caprifoliaceae) | Pimples, acne, wrinkles on face | (B. Aggarwal et al., 2011) |
| Vipadikahara ghrita taila | <i>Leptadenia reticulata</i> (Retz.) Wight & Arn. (Apocynaceae), <i>Rubia cordifolia</i> L. (Rubiaceae), <i>Berberis aristata</i> DC. (Berberidaceae), <i>Mallotus philippensis</i> (Lam.) Müll. Arg. (Euphorbiaceae), <i>Shorea robusta</i> Gaertn. (Dipterocarpaceae), <i>Sesamum indicum</i> L. (Pedaliaceae), Cow's milk and ghee (clarified butter) | Skin lesions | Hewageegana et al. (2013) |
| Aragwadharistam | <i>Cassia fistula</i> L. (Fabaceae), <i>Embelia ribes</i> Burm.f. (Primulaceae), <i>Saccharum officinarum</i> L. (Poaceae), honey, <i>Phyllanthus emblica</i> L. (Phyllanthaceae), <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Operculina turpethum</i> (L.) Silva Manso (Convolvulaceae), <i>Elettaria cardamomum</i> (L.) Maton (Zingiberaceae), <i>Syzygium aromaticum</i> (L.) Merr. & L.M. Perry (Myrtaceae), <i>Piper nigrum</i> L. (Piperaceae) | Skin disorders, ulcers | Mohammad et al. (2019) |
| Chyawanprash | <i>Phyllanthus emblica</i> L. (= <i>Emblca officinalis</i> Gaertn.) (Phyllanthaceae), <i>Justicia adhatoda</i> L. (= <i>Adhatoda vasica</i> Nees) (Acanthaceae), <i>Aegle marmelos</i> (L.) Correa (Rutaceae), <i>Asparagus racemosus</i> Willd. (Asparagaceae), <i>Boerhaavia diffusa</i> Brandege [unresolved] (Nyctaginaceae), <i>Cinnamomum tamala</i> (Buch.-Ham.) T. Nees & Eberm. (Lauraceae), <i>Cinnamomum verum</i> J.Presl (= <i>Cinnamomum zeylanicum</i> Blume) (Lauraceae), <i>Dendrobium plicatile</i> Lindl. (= <i>Desmotrichum fimbriatum</i> Blume) (Orchidaceae), <i>Desmodium gangeticum</i> (L.) DC. (Fabaceae), <i>Elettaria cardamomum</i> (L.) Maton (Zingiberaceae), <i>Gmelina arborea</i> Roxb. (Lamiaceae), <i>Hedychium spicatum</i> Sm. (Zingiberaceae), <i>Jacquemontia paniculata</i> (Burm.f.) Hallier f. (= <i>Ipomoea paniculata</i> Burm.f.) (Convolvulaceae), <i>Mesua ferrea</i> L. (Calophyllaceae), <i>Nymphaea nouchali</i> Burm.f. (= <i>Nymphaea cyanea</i> Roxb. ex G. Don) (Nymphaeaceae), <i>Vigna trilobata</i> (L.) Verdc. (= <i>Phaseolus trilobus</i> Aiton) (Fabaceae), <i>Phyllanthus niruri</i> L. (Phyllanthaceae), <i>Piper longum</i> L. (Piperaceae), <i>Premna serratifolia</i> L. (= <i>Premna integrifolia</i> L.) (Lamiaceae), <i>Oroxylum indicum</i> (L.) Kurz (Bignoniaceae), <i>Pterocarpus santalinus</i> L.f. (Fabaceae), <i>Rhus succedanea</i> L. (Anacardiaceae), <i>Solanum virginianum</i> L. (= <i>Solanum xanthocarpum</i> Schrad. & H. Wendl.) (Solanaceae), <i>Solanum dulcamara</i> L. (Solanaceae), <i>Solanum indicum</i> L. [unresolved] (Solanaceae), <i>Stereospermum chelonoides</i> (L.f.) DC. (= <i>Stereospermum suaveolens</i> (Roxb.) DC. (Bignoniaceae), <i>Teramnus labialis</i> (L.f.) Spreng. (Fabaceae), <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Tinospora crispa</i> (L.) Hook. f. & Thomson (Menispermaceae), | Antioxidant, skin photoaging, UV irradiation | Takauji et al. (2016) |

(continued on next page)

Table 4 (continued)

| Ayurvedic formulation | Ingredients | Used as/against | Reference |
|-------------------------|---|--|---------------------------|
| Chopchiniyadi Churna | <i>Uraria lagopodoides</i> (L.) DC. (= <i>Uraria lagopoides</i> (L.) DC) (Fabaceae), <i>Vitis vinifera</i> L. (Vitaceae) and <i>Withania somnifera</i> (L.) Dunal (Solanaceae) <i>Smilax china</i> L. (Smilacaceae) (roots), <i>Piper longum</i> L. (Piperaceae) (fruits, roots), <i>Syzygium aromaticum</i> (L.) Merr. & L.M. Perry (Myrtaceae) (flower buds), <i>Piper nigrum</i> L. (Piperaceae) (fruits), <i>Anacyclus pyrethrum</i> (L.) Lag. (Asteraceae) (roots), <i>Zingiber officinale</i> Roscoe (Zingiberaceae) (rhizomes), <i>Embelia ribes</i> Burm.f. (Primulaceae) (fruits), <i>Hyoscyamus niger</i> L. (Solanaceae) (seeds), <i>Cinnamomum verum</i> J.Presl (= <i>Cinnamomum zeylanicum</i> Blume) (Lauraceae) (bark) | Anthelmintic, skin diseases, malignant ulcers, syphilis | (S. Jain et al., 2010) |
| Triphala | <i>Phyllanthus emblica</i> L. (= <i>Emblia officinalis</i> Gaertn.) (Phyllanthaceae), <i>Terminalia bellirica</i> (Gaertn.) Roxb. (Combretaceae), <i>Terminalia chebula</i> Retz. (Combretaceae) | Antioxidant, increase the resistance of the body against any infection | Ponnusankar et al. (2011) |
| Nimba arishta | <i>Azadirachta indica</i> A. Juss. (Meliaceae), <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Terminalia bellirica</i> (Gaertn.) Roxb. (Combretaceae), <i>Phyllanthus emblica</i> L. (Phyllanthaceae), <i>Woodfordia fruticosa</i> (L.) Kurz (Lythraceae) | Skin diseases, inflammation, immunomodulatory activity | Kroes et al. (1993) |
| Thiostanin | <i>Terminalia chebula</i> Retz. (Combretaceae), <i>Terminalia bellirica</i> (Gaertn.) Roxb. (Combretaceae), <i>Phyllanthus emblica</i> L. (= <i>Emblia officinalis</i> Gaertn.) (Phyllanthaceae), <i>Commiphora mukul</i> (Hook. ex Stocks) Engl. (Burseraceae), <i>Plumbago zeylanica</i> L. (Plumbaginaceae), <i>Picrorhiza kurroo</i> Royle ex Benth. [unresolved] (Plantaginaceae) | Anti-acne drug | Ghosh et al. (2011) |
| Takezama | <i>Azadirachta indica</i> A. Juss. (Meliaceae), <i>Phyllanthus emblica</i> L. (Phyllanthaceae), <i>Rubia cordifolia</i> L. (Rubiaceae), <i>Curcuma longa</i> L. (Zingiberaceae), <i>Berberis aristata</i> DC. (Berberidaceae), <i>Glycyrrhiza glabra</i> L. (Fabaceae) | Eczema, anti-inflammatory | Saple et al. (2007) |
| Finbid | <i>Piper nigrum</i> L. (Piperaceae), <i>Hemidesmus indicus</i> (L.) R. Br. ex Schult. (Apocynaceae), <i>Rubia cordifolia</i> L. (Rubiaceae), <i>Glycyrrhiza glabra</i> L. (Fabaceae), <i>Nardostachys jatamansi</i> (D. Don) DC. (Caprifoliaceae), <i>Mesua ferrea</i> L. (Calophyllaceae), <i>Saussurea costus</i> (Falc.) Lipsch. (Asteraceae), <i>Piper cubeba</i> L.f. [unresolved] (Piperaceae), <i>Withania somnifera</i> (L.) Dunal (Solanaceae), <i>Commiphora wightii</i> (Arn.) Bhandari (Burseraceae), detoxified hydragyrum, purified sulfur, purified yellow arsenic tri-sulphide, purified <i>Aconitum ferox</i> Wall. ex Ser. (Ranunculaceae), purified Soda Bi Boras, <i>Curcuma longa</i> L. (Zingiberaceae), <i>Phyllanthus emblica</i> L. (= <i>Emblia officinalis</i> Gaertn.) (Phyllanthaceae), <i>Terminalia bellirica</i> (Gaertn.) Roxb. (Combretaceae), <i>Terminalia chebula</i> Retz. (Combretaceae) | Auto-immune skin disorders, fungal infection, psoriasis | Khan (2016) |

the skin, followed by lichenification (thickening) and scaling of the skin. Eczema usually causes itching and burning of the skin. Eczema, also known as atopic dermatitis, is a condition where patches of skin become inflamed, itchy, red, cracked, and rough. Karodi et al. (2009) reported that *Rubia cordifolia* L. (Rubiaceae) is popular for its medicinal uses in eczema (Karodi et al., 2009). In this study, the alcohol extract and the *R. cordifolia* hydrogel were tested to evaluate the wound healing activity in an excision wound model in mice. The mice were divided into five groups of six animals each, group I–V was treated with gels with various extract concentrations (0.2%, 0.5%, 1%), once a day for 15 days, so as a result of the gel effect in terms of wound contractility, wound closure, wound surface reduction, tissue regeneration at the wound site, and histopathological features were significant in treated mice (Karodi et al., 2009). Mukhopadhyay et al. (2018), studied *Eclipta prostrata* (L.) L. (= *Eclipta alba* (L.) Hassk.), which is also thought to help treat eczema. *E. prostrata* shoot extract showed antibacterial activity against *E. coli*, *Staphylococcus aureus*, and anti-inflammatory effect on albino mice (Mukhopadhyay et al., 2018). Priyadarshi and Ram (2018) reported *Carica papaya* L. (Caricaceae) for the treatment of eczema. In this study, the three extracts (n-hexane, ethyl acetate, and ethanol extract) of *C. papaya* exhibited significant analgesic activity at all three dose levels (0.175, 0.35 and 0.70 mg/kg bw orally) compared to aspirin (standard drug) on a mouse model having acetic acid-induced pain (Priyadarshi and Ram, 2018).

Psoriasis: Psoriasis is a chronic inflammatory skin disease – cells build up and form scales and itchy, dry patches, mostly appearing on the scalp, elbows, knees, and lower back. It is an auto-immune problem. Schafer et al. (2010) demonstrated the broad anti-inflammatory activity of apremilast (a particular type of inhibitor used against psoriasis and psoriatic arthritis, marketed with the brand name Otezia) inhibited the production of multiple inflammatory mediators, including CXCL9, TNF- α , IFN- γ (monokines induced by MIG or IFN- γ), CXCL10 (IFN- γ

induced protein of IP-10 or 10 kDa), IL-2, IL-12, IL-23, (MIP)-1 α (macrophage inflammatory protein), (MCP)-1 (monocyte chemo-attractant protein) and GM-CSF (granulocyte macrophage-colony stimulating factor) from PBMC (Schafer et al., 2010). In this study, in particular, the inhibition of IL-12, IL-23, and TNF- α production and the apremilast ability to inhibit psoriasis lesions *in vivo* suggests that this constituent might be a useful remedy in the treatment of psoriasis through a multifaceted mechanism (Schafer et al., 2010).

Schonthaler et al. (2009) investigated systemic VEGF (vascular endothelial growth factor) inhibition reducing psoriasis-like dermatitis. In this study, RT-PCR analyses was performed in the anti-VEGF-treated separated mouse epidermis and for VEGF in double knockout (DKO) mice administered with IgG, RNA levels were significantly reduced and for chemotactic proteins [S100A8 (also known as myeloid-related protein-8) and S100A9 (myeloid-related protein-14)] as well. RNA levels were reduced significantly in epidermal samples of anti-VEGF-treated mice (Schonthaler et al., 2009). Chen et al. (2017) demonstrated that Quercetin (QC) had anti-psoriasis effects in imiquimod (IMQ) induced mice, which is characterized by modulating many distinct signaling pathways, including the MAPK (Mitogen-activated protein kinase) signaling, the NF- κ B pathway and the Nrf-2 pathway, so QC has significant potential value for the treatment of psoriasis and improving anti-oxidant and anti-inflammatory activity (Chen et al., 2017).

2.4. Medicinal plants in other countries and their negative healing effects

In Pakistan, the Manoer Valley, which is located about 320 km from the border of India, 48 plant species have been used in the treatment of skin disorders (Rahman et al., 2018). *Achyranthes aspera* L. (Amaranthaceae) is used in India for wound healing as well as for the treatment of eczema and wound healing in Pakistan. *Cedrus deodara* (Roxb. ex D. Don) G. Don (Pinaceae) is used in India as anti-inflammatory agent

Table 5
Herbal drugs and cosmetics products in dermatology.

| Manufacturer | Product | Ingredients | Used as/against |
|---------------|-------------------------|--|--|
| Oziva | Anti-ageing food | Guava powder (<i>Psidium guajava</i> L., Myrtaceae), Sea Buckthorn powder (<i>Elaeagnus rhamnoides</i> (L.) A. Nelson = <i>Hippophae rhamnoides</i> L., Elaeagnaceae), Lemon powder (<i>Citrus limon</i> (L.) Osbeck, Rutaceae), Rose Petal powder (<i>Rosa</i> sp., Rosaceae), Orange Peel powder (<i>Citrus sinensis</i> (L.) Osbeck, Rutaceae), Spinach powder (<i>Spinacia oleracea</i> L., Amaranthaceae), Pink salt, Spirulina extract (<i>Arthrospira</i> sp., Microcoleaceae [Cyanobacteria]), Barley Grass powder (<i>Hordeum vulgare</i> L., Poaceae), Amla powder (<i>Phyllanthus emblica</i> L., Phyllanthaceae), Holy Basil powder (<i>Ocimum tenuiflorum</i> L. = <i>Ocimum sanctum</i> L., Lamiaceae), Pomegranate powder (<i>Punica granatum</i> L., Lythraceae), Strawberry powder (<i>Fragaria</i> × <i>ananassa</i> (Duchesne ex Weston) Duchesne ex Rozier, Rosaceae), Acai [Açai] Berry powder (<i>Euterpe oleracea</i> Mart., Arecaceae), Ginger powder (<i>Zingiber officinale</i> Roscoe, Zingiberaceae), Green Tea extract (<i>Camellia sinensis</i> (L.) Kuntze, Theaceae), Cinnamon extract (<i>Cinnamomum</i> sp., Lauraceae), Acerola Cherry extract standardized to 17% Vitamin C (<i>Malpighia emarginata</i> DC., Malpighiaceae), Sesbania extract standardized to 0.5% Biotin (<i>Sesbania</i> sp., Fabaceae), Bamboo shoot extract standardized to 70% Silica (<i>Bambusa</i> sp., Poaceae) | Skin repair and regeneration, protects from UV damage |
| Khadi Natural | Face gold massage cream | Purified water, Sunflower oil (<i>Helianthus annuus</i> L., Asteraceae), Sheabutter (<i>Vitellaria paradoxa</i> C.F. Gaertn. (= <i>Butyrospermum parkii</i> (G. Don) Kotschy), Sapotaceae), Rose extract (<i>Rosa centifolia</i> L., Rosaceae), Olive oil (<i>Olea europaea</i> L., Oleaceae), Kokum butter (<i>Garcinia indica</i> (Thouars) Choisy, Clusiaceae), Wheatgerm oil (<i>Triticum aestivum</i> L. = <i>Triticum vulgare</i> Vill., Poaceae), Almond oil (<i>Prunus dulcis</i> (Mill.) D.A. Webb (= <i>Prunus amygdalus</i> var. <i>dulcis</i> (Borkh. ex DC.) Koehne), Rosaceae), Aloe Vera ext (<i>Aloe vera</i> (L.) Burm.f. (= <i>Aloe barbadensis</i> Mill.), Asphodelaceae), Licorice ext (<i>Glycyrrhiza glabra</i> L., Fabaceae), Beeswax (<i>cera alba</i>), Apple ext (<i>Malus domestica</i> Borkh. = <i>Pyrus malus</i> L., Rosaceae), Chamomile ext (<i>Matricaria chamomilla</i> L. = <i>Matricaria recutita</i> L., Asteraceae), Mulberry ext (<i>Morus alba</i> L., Moraceae, leaf), Ginkgo Biloba ext (<i>Ginkgo biloba</i> L., Ginkgoaceae), Rosemary ext (<i>Rosmarinus officinalis</i> L., Lamiaceae), Jojoba oil (<i>Simmondsia chinensis</i> (Link) C.K. Schneid., Simmondsiaceae), Grapeseed oil (<i>Vitis vinifera</i> L., Vitaceae), Sandalwood oil (<i>Santalum album</i> L., Santalaceae), Swarna Bhasma (Gold), Base Q.S. | Anti-ageing, reduce dead skin cells, skin fairness and glowing |
| | Facial massage gel | Rose extract (<i>Rosa</i> sp., Rosaceae), Geranium oil (<i>Pelargonium graveolens</i> L'Hér., Geraniaceae), Ashwagandha extract (<i>Withania somnifera</i> (L.) Dunal, Solanaceae), Demineralized water, E.D.T.A, Propyl Glycol, Glycerine, Stabilized Aloe vera Gel (<i>Aloe vera</i> (L.) Burm.f., Asphodelaceae), Carbomer, Kathon Cg, Sodium Hydroxide, Xylitylglucoside, Anhydroxylitol, Xylitol, Aloe vera extract, Liquorice extract (<i>Glycyrrhiza glabra</i> L., Fabaceae), Fragrance, Color | Soothing skin |
| | Anti-wrinkle cream | <i>Crocus sativus</i> L., Iridaceae (Kumkuma), <i>Hydnocarpus wightianus</i> Blume, Achariaceae (Chaulmoogra), Wheat germ oil (<i>Triticum aestivum</i> L., Poaceae), <i>Prunus dulcis</i> (Mill.) D. A. Webb (= <i>Prunus amygdalus</i> Batsch), Rosaceae (Vatadha), <i>Santalum album</i> L., Santalaceae (Chandana), <i>Carica papaya</i> L., Caricaceae (Chirbhita), Shea butter (<i>Vitellaria paradoxa</i> C.F. Gaertn. (= <i>Butyrospermum parkii</i> (G. Don) Kotschy), Sapotaceae), Cocoa butter (<i>Theobroma cacao</i> L., Malvaceae) | Reduce fine lines, wrinkles, remove dark spots, protect the skin from sun damage |
| | Antiacne cream | Purified water, Sunflower oil (<i>Helianthus annuus</i> L., Asteraceae), neem extract (<i>Azadirachta indica</i> A. Juss., Meliaceae), Aloe vera extract (<i>Aloe vera</i> (L.) Burm.f. (= <i>Aloe barbadensis</i> Mill., Asphodelaceae), olive oil (<i>Olea europaea</i> L., Oleaceae), Shea butter (<i>Vitellaria paradoxa</i> C.F. Gaertn. (= <i>Butyrospermum parkii</i> (G. Don) Kotschy), Sapotaceae), Wheatgerm oil (<i>Triticum aestivum</i> L. = <i>Triticum vulgare</i> Vill., Poaceae), Almond oil (<i>Prunus dulcis</i> (Mill.) D.A. Webb (= <i>Prunus amygdalus</i> var. <i>dulcis</i> (Borkh. ex DC.) Koehne), Rosaceae), Calendula extract (<i>Calendula officinalis</i> L., Asteraceae, flower), basil extract (<i>Ocimum basilicum</i> L., Lamiaceae), clove extract (<i>Syzygium aromaticum</i> (L.) Merr. & L.M.Perry = <i>Eugenia caryophyllus</i> (Spreng.) Bullock & S.G. Harrison, Myrtaceae), <i>Lavandula angustifolia</i> Mill. (= <i>Lavandula officinalis</i> Chaix, Lamiaceae) extract, <i>Symphytum officinale</i> L. (Boraginaceae) extract, <i>Cinnamomum verum</i> J.Presl (= <i>Cinnamomum zeylanicum</i> Blume, Lauraceae) extract, <i>Garcinia indica</i> (Thouars) Choisy (Clusiaceae), Jojoba oil (<i>Simmondsia chinensis</i> (Link) C.K. Schneid., Simmondsiaceae), grapeseed oil (<i>Vitis vinifera</i> L., Vitaceae), Tumeric extract (<i>Curcuma longa</i> L., Zingiberaceae), Teatree oil (<i>Camellia sinensis</i> (L.) Kuntze, Theaceae) | Acne, pimple, skin irritation |
| | Moisturizer cream | Purified Water, Sunflower Oil (<i>Helianthus annuus</i> L., Asteraceae), Sheabutter (<i>Vitellaria paradoxa</i> C.F. Gaertn. (= <i>Butyrospermum parkii</i> (G. Don) Kotschy), Sapotaceae), Beeswax (<i>cera alba</i>), Aloe vera Ext (<i>Aloe vera</i> (L.) Burm.f. (= <i>Aloe barbadensis</i> Mill., Asphodelaceae), Mulberry Ext (<i>Morus alba</i> L., Moraceae, Root), Olive Oil (<i>Olea europaea</i> L., Oleaceae), Kokum Butter (<i>Garcinia indica</i> (Thouars) Choisy, Clusiaceae), Almond Oil (<i>Prunus dulcis</i> (Mill.) D.A. Webb (= <i>Prunus amygdalus</i> var. <i>dulcis</i> (Borkh. ex DC.) Koehne), Rosaceae), Wheatgerm Oil (<i>Triticum aestivum</i> L. = <i>Triticum vulgare</i> Vill., Poaceae), Pineapple Ext (<i>Ananas comosus</i> (L.) Merr. = <i>Ananas sativus</i> Schult. & Schult.f., Bromeliaceae), Kastoori Methi (<i>Trigonella foenum-graecum</i> L., Fabaceae), Lavender Oil (<i>Lavandula angustifolia</i> Mill., Lamiaceae), Grapeseed Oil (<i>Vitis vinifera</i> L., Vitaceae), Basil Ext (<i>Ocimum basilicum</i> L., Lamiaceae), Saffron Ext (<i>Crocus sativus</i> L., Iridaceae), Sandalwood Oil (<i>Santalum album</i> L., Santalaceae) | Skin fairness, remove dead skin cells |
| Himalaya | Nourishing skin cream | Water (aqua), mineral oil, glycerin, cetyl alcohol, cetearyl ethylhexanoate, glyceryl stearate se, cetostearyl alcohol, triethanolamine, <i>Aloe vera</i> (L.) Burm.f. (= <i>Aloe barbadensis</i> Mill., Asphodelaceae) (aloe vera) leaf extract, glyceryl stearate & peg 100 stearate, phenoxyethanol, carbomer, <i>Pterocarpus marsupium</i> Roxb. (Fabaceae) (indian kino tree) wood extract, fragrance, <i>Withania somnifera</i> (L.) Dunal (Solanaceae) | Skin care |

(continued on next page)

Table 5 (continued)

| Manufacturer | Product | Ingredients | Used as/against |
|----------------------------|-----------------------------|---|--|
| | Face wash | (ashwagandha) root extract, <i>Centella asiatica</i> (L.) Urb. (Apiaceae) (gotu kola) extract, methylparaben, propylparaben, disodium EDTA. | |
| VLCC Natural Sciences | Active D pigmentation cream | Water (aqua), ammonium lauryl sulfate, <i>Azadirachta indica</i> A. Juss. (= <i>Melia azadirachta</i> L., Meliaceae) (neem) leaf extract, cocamidopropyl betaine, sodium cocoyl glutamate & disodium cocoyl glutamate, glycerin, acrylates/c10-30 alkyl acrylate crosspolymer, <i>Curcuma longa</i> L. (Zingiberaceae) (turmeric) root extract, sodium hydroxide, phenoxyethanol, fragrance, methylchloroisothiazolinone & methylisothiazolinone, sodium metabisulfite, citric acid, disodium edta, tocopheryl acetate | Skin impurities, dead skin, excess oil |
| COSMAX | Skin tiger cica gel cream | Jojoba oil (<i>Simmondsia chinensis</i> (Link) C.K. Schneid., Simmondsiaceae), Carrot extract (<i>Daucus carota</i> subsp. <i>sativus</i> (Hoffm.) Arcang., Apiaceae), Chamomile oil (<i>Matricaria chamomilla</i> L., Asteraceae), Licorice extract (<i>Glycyrrhiza glabra</i> L., Fabaceae), <i>Aloe vera</i> (L.) Burm.f. (Asphodelaceae) | Reduces pigmentation, lightens spots |
| | Anti-acne gel | <i>Centella asiatica</i> (L.) Urb. (Apiaceae) extract (68%), Dipropylene Glycol, Glycerin, Butylene Glycol, Cyclopentasiloxane, Alcohol Denat., Water, Polyacrylamide, 1,2-Hexanediol, Pentylene Glycol, <i>Citrus × aurantium</i> L. (Rutaceae) Bergamia (Bergamot) fruit oil, <i>Citrus limon</i> (L.) Osbeck (Rutaceae) (Lemon) Peel Oil, <i>Lavandula angustifolia</i> Mill. (Lamiaceae) (Lavender) oil, <i>Cymbopogon schoenanthus</i> (L.) Spreng. (Poaceae) Oil, <i>Cananga odorata</i> (Lam.) Hook.f. & Thomson (Annonaceae) flower oil, <i>Citrus aurantium</i> var. <i>dulcis</i> Hayne (Rutaceae) (Orange) peel oil, <i>Eucalyptus globulus</i> Labill. (Myrtaceae) leaf extract, Dimethicone, Cyclohexasiloxane, C13-14 Isoparaffin, Caprylyl Glycol, Dimethicone, Laureth-7, Hydrogenated Lecithin, Ethylhexylglycerin, Dimethicone/Vinyl Dimethicone Crosspolymer, Stearic Acid, Glyceryl Acrylate/Acrylic Acid Copolymer, Panthenol, Tocopheryl Acetate, Ceramide AP, Ceramide AS, Ceramide EOP, Ceramide NP, Ceramide NS, Cholesterol, Phytosphingosine, Propanediol, Sodium Hyaluronate Crosspolymer, Hydrolyzed Glycosaminoglycans, Sodium Hyaluronate, Benzyl Glycol, Hydrolyzed Hyaluronic Acid, Madecassoside, Asiaticoside, Asiatic Acid, Madecassic Acid, Hyaluronic Acid, Raspberry Ketone, Cetearyl Alcohol, Disodium EDTA | Sensitive, acne prone, and itchy skin |
| Biotique Advanced Ayurveda | Anti-acne face pack | Chinai ghas (<i>Gelidium amansii</i> (J.V.Lamouroux) J.V.Lamouroux, Gelidiaceae [Rhodophyta]), Kuda (<i>Holarrhena pubescens</i> Wall. ex G. Don = <i>Holarrhena antidysenterica</i> (Roth) Wall. ex A. DC., Apocynaceae), Ghrit kumari (<i>Aloe vera</i> (L.) Burm. f. = <i>Aloe vera</i> var. <i>chinensis</i> (Steud. ex Baker) Baker = <i>Aloe vera</i> 'indica', Asphodelaceae), Kikkar gaund (<i>Acacia nilotica</i> (L.) Delile = <i>Acacia arabica</i> (Lam.) Willd., Fabaceae), Samundir Kai (<i>Gelidium amansii</i> (J.V.Lamouroux) J.V.Lamouroux, Gelidiaceae [Rhodophyta]), Himalayan Water Q.S. | Acne prone skin |
| | Smooth skin lotion | Jaiphal (<i>Myristica fragrans</i> Houtt., Myristicaceae), Pippli (<i>Piper longum</i> L., Piperaceae), Haldi (<i>Curcuma longa</i> L., Zingiberaceae), Chandan (<i>Santalum album</i> L., Santalaceae), Geru (Red ochre), Multani mitti (Fuller's earth), Kikkar gaund (<i>Acacia nilotica</i> (L.) Delile = <i>Acacia arabica</i> (Lam.) Willd., Fabaceae), Himalayan Water Q.S. | Skin redness, skin irritation |
| | Sunscreen | Mandukaparni Panchang (<i>Centella asiatica</i> (L.) Urb., Apiaceae) extract, Surajmukhi Tail (Sunflower oil, <i>Helianthus annuus</i> L., Asteraceae), Sarso Tail (mustard oil, <i>Brassica</i> sp., Brassicaceae), Ashwagandha (<i>Withania somnifera</i> (L.) Dunal, Solanaceae) root extract, multi leaves extract, Mahua (<i>Madhuca longifolia</i> (J.Koenig ex L.) J.F.Macbr., Sapotaceae) seed extract, Lotion base Q.S. | Dry and dehydrated skin care |
| Manuka honey | Moisturizing cream | Ghritkumari (<i>Aloe vera</i> (L.) Burm.f. = <i>Aloe barbadensis</i> Mill., Asphodelaceae), Methi (<i>Trigonella foenum-graecum</i> , Fabaceae), Madhu (Honey), Ankurit gehun (<i>Triticum aestivum</i> L. = <i>Triticum vulgare</i> Vill., Poaceae), Kusumbhi (<i>Carthamus tinctorius</i> L., Asteraceae), Lotion base Q.S. | Nourishes dry skin |
| | Antiseptic liquid | <i>Aloe vera</i> (L.) Burm.f. = <i>Aloe barbadensis</i> Mill. (Asphodelaceae) leaf juice, Emulsifying Wax NF, Caprylic/Capric Triglyceride, Glycerol, <i>Leptospermum scoparium</i> J.R.Forst. & G. Forst. (Myrtaceae) (Manuka) Mel, <i>Cocos nucifera</i> L. (Arecaceae) (Coconut) Oil, <i>Theobroma cacao</i> L. (Malvaceae) (Cocoa) Seed Butter, <i>Theobroma cacao</i> (Cocoa) Seed Butter Deodorized, Carbomer, Tocopheryl Acetate, Cellulose Gum, Sodium Hyaluronate, Tapioca Natural, Titanium Dioxide, <i>Daucus carota</i> subsp. <i>sativus</i> (Hoffm.) Arcang. (Apiaceae) (Carrot) Seed Oil, DL-Panthenol, <i>Avena sativa</i> L. (Poaceae) (Oat) kernel flour, <i>Vitellaria paradoxa</i> C.F. Gaertn. = <i>Butyrospermum parkii</i> (G. Don) Kotschy, Sapotaceae (Shea) Butter, <i>Olea europaea</i> L., Oleaceae (Olive) Fruit Oil, (Hemp) Oil, Sodium Hydroxide, L-ascorbic acid, <i>Plumeria alba</i> L. (Apocynaceae) (Frangipani) Flower extract, Oat Amino Acids, Cehami (<i>Centipeda cunninghamii</i> (DC.) A. Braun & Asch., Asteraceae), MSM (MethylSulfonylMethane), Gluconolactone (and) Sodium Benzoate, Phenoxyethanol & Ethylhexylglycerin | Eczema, dry and itchy skin |
| Boroline | Moisturizing cream | Neem (<i>Azadirachta indica</i> A. Juss. (Meliaceae), Turmeric (<i>Curcuma longa</i> L. (Zingiberaceae), Marigold, <i>Aloe vera</i> (L.) Burm.f. (Asphodelaceae), Cetrimide (<i>Centipeda cunninghamii</i> (DC.) A. Braun & Asch., Asteraceae), Chlorhexidine Gluconate | Prickly heat, itches and skin rashes |
| Cetaphil | Brightening skin serum | Purified Water, Polyglycerylmethacrylate & Propylene Glycol, Light Liquid Paraffin, Dicaprylyl Ether, PEG-5 Glyceryl Stearate, Glycerin, Dimethicone and Dimethicone, Cetyl Alcohol, Refined Almond Oil, Benzyl Alcohol, Tocopheryl Acetate, Acrylates/C10-30 Alkyl Acrylate, Disodium Edetate, Sodium Hydroxide, ALL-RAC-Alpha Tocopherol | Skin allergies |
| Ilana | Moisturizing cream | <i>Citrus limon</i> (L.) Osbeck (Rutaceae) (lemon), <i>Carica papaya</i> L., Caricaceae (papaya), <i>Pelargonium graveolens</i> L'Hér. (Geraniaceae) (geranium), Wheat Germ Oil (Natural Vit E) (<i>Triticum aestivum</i> L. = <i>Triticum vulgare</i> Vill., Poaceae) | Hyperpigmentation and dark circles |
| Belif | | Water, Glycerin, Cyclohexasiloxane, Hydrogenated Polydecene, Dipropylene Glycol, <i>Macadamia ternifolia</i> F. Muell. (Proteaceae) Seed Oil, 1,2-Hexanediol, Bis-PEG-18 Methyl Ether Dimethyl Silane, Triethylhexanoin, Stearyl Alcohol, Glyceryl Stearate, Pentaerythrityl Tetraethylhexanoate, PEG-40 Stearate, <i>Saccharomyces/Viscum album</i> L., Santalaceae (Mistletoe) Ferment Extract, <i>Lactobacillus/Soybean</i> Ferment Extract | Dryness, loss of firmness and elasticity |

(continued on next page)

Table 5 (continued)

| Manufacturer | Product | Ingredients | Used as/against |
|--------------|------------------|--|----------------------|
| | | (<i>Glycine max</i> (L.) Merr., Fabaceae), <i>Saccharomyces/Imperata cylindrica</i> (L.) Raesch., Poaceae Root Ferment Extract, Cetearyl Alcohol, Cetearyl Oliviate, Sorbitan Oliviate, Hydrogenated Lecithin, Sorbitan Stearate, Stearic Acid, PEG-100 Stearate, Dimethicone, Dimethicone/Vinyl Dimethicone Crosspolymer, <i>Citrus trifoliata</i> L. = <i>Poncirus trifoliata</i> (L.) Raf., Rutaceae Fruit Extract, Panthenol, <i>Avena sativa</i> L. (Poaceae) (Oat) Kernel Extract, <i>Calendula officinalis</i> L., Asteraceae Flower Extract, <i>Nepeta cataria</i> L., Lamiaceae Extract, <i>Rubus idaeus</i> L., Rosaceae (Raspberry) Leaf Extract, <i>Baptisia tinctoria</i> (L.) Vent., Fabaceae Root Extract, <i>Stellaria media</i> (L.) Vill., Caryophyllaceae (Chickweed) Extract, <i>Alchemilla xanthochlora</i> Rothm. = <i>Alchemilla vulgaris</i> auct., Rosaceae Leaf Extract, <i>Viola tricolor</i> L., Violaceae Extract, <i>Rosa × damascena</i> Herrm., Rosaceae Flower Extract, <i>Filipendula ulmaria</i> (L.) Maxim. = <i>Spiraea ulmaria</i> L., Rosaceae Flower Extract, <i>Althaea officinalis</i> L., Malvaceae Root Extract, <i>Symphytum officinale</i> L. (Boraginaceae) Leaf Extract, <i>Trifolium pratense</i> L., Fabaceae (Clover) Flower Extract, <i>Achillea millefolium</i> L., Asteraceae flower Extract, <i>Euphrasia officinalis</i> L. [unresolved], Orobanchaceae Extract, <i>Thuja occidentalis</i> L., Cupressaceae Leaf Extract, <i>Menyanthes trifoliata</i> L., Menyanthaceae Leaf Extract, <i>Salix alba</i> L., Salicaceae (Willow) Bark Extract, <i>Chelidonium majus</i> L., Papaveraceae Extract, Urea, Hydroxyethylpiperazine Ethane Sulfonic Acid, Glycosyl Trehalose, Hydrogenated Starch Hydrolysate, Acrylates/C10-30 Alkyl Acrylate Crosspolymer, Carbomer, Xanthan Gum, Trisodium EDTA, Tromethamine, Frgrance, Citronellol, Limonene, Napiers Original Formula, Napiers Moisture Formula, Fragrances of Natural Origin. | |
| Just Herbs | Anti-blemish gel | Indian madder (<i>Rubia cordifolia</i> L., Rubiaceae) ext. 0.5%, cinnamon (<i>Cinnamomum verum</i> J.Presl = <i>Cinnamomum zeylanicum</i> Blume, Lauraceae) ext. 0.2%, fumitory (<i>Fumaria officinalis</i> L., Papaveraceae) ext. 0.2%, chiraunji (<i>Buchanania cochinchinensis</i> (Lour.) M.R. Almeida = <i>Buchanania lanzan</i> Spreng., Anacardiaceae) ext. 0.2%, nutgrass (<i>Cyperus rotundus</i> L., Cyperaceae) ext.0.1%, nutmeg (<i>Myristica fragrans</i> Houutt., Myristicaceae) ext. 0.1%, holy basil (<i>Ocimum tenuiflorum</i> L. = <i>Ocimum sanctum</i> L., Lamiaceae) ext. 0.2%, lemon (<i>Citrus medica</i> L., Rutaceae) peel ext. 1%, tomato (<i>Lycopersicon esculentum</i> L., Solanaceae) ext. 1%, purified water (aqua), cetearyl olivate, aloe vera (<i>Aloe vera</i> (L.) Burm.f. = <i>Aloe barbadensis</i> Mill., Asphodelaceae) leaf juice, vegetable glycerine, cold pressed grapeseed (<i>Vitis vinifera</i> L., Vitaceae) oil, cold pressed jojoba (<i>Simmondsia chinensis</i> (Link) C.K. Schneid., Simmondsiaceae) oil, octyldodecanol, sorbitan olivate, caprylyl glycol, potassium sorbate, gluconolactone, sodium gluconate, ethylhexylglycerin, natural preservative blend of herbs and essential oils. | Hyper pigmented skin |

and in Pakistan against hair loss and psoriasis.

In India, it is common to use traditional drugs to treat dermatological conditions. It was found that most of the used healing plant species came from traditional healers, while those that were not readily available were bought on the market or collected directly in the wild. According to this, it was concluded that the management practice of dermatological healthcare in many research areas depends largely on wild-growing species of medicinal plants (Kumar et al., 2013; Dutta et al., 2021). Kumar et al. (2007) found, however, that the pharmacological research on Indian medicinal plants is very limited and a large number of plants used in tribal and folklore with enormous potential remains to add scientific information, complement or enrich local knowledge, and even advise locals on some possible side effects of plant use that may arise from clinical and pharmacobotanical studies, among others (Fig. 2) (Kumar et al., 2007).

However, these plants have been tested for generations within traditional medicine contexts and typically do not show significant side effects. Although, plant-induced cutaneous reactions can occur and hence this should provide the basis for the creation of a phytovigilance programme and reevaluation of how traditional medicine is utilised in the general population (Niang et al., 2015). It was found that some plant species used in Indian ethnodermatology can have also negative healing effects, e.g., *Jatropha curcas* L. (Euphorbiaceae), *Calotropis procera* (Aiton) Dryand. (Apocynaceae), *Lawsonia inermis* L. (Lythraceae) were found to induce contact eczema. *Momordica charantia* L. (Cucurbitaceae), *Azadirachta indica* A. Juss. (Meliaceae), and *Anacardium occidentale* L. (Anacardiaceae) were found to induce systemic eczema. *Vitellaria paradoxa* C.F. Gaertn. (= *Butyrospermum parkii* (G.Don) Kotschy), Sapotaceae was found to induce erythrodermic syndrome and erythrodermic psoriasis. The active ingredients in medicinal plants and herbs can often be as potent as many drugs made by pharmaceutical companies. This can cause unforeseen complications when combining active ingredients (Olisova et al., 2018).

Sharma et al. (2014) concluded that further preclinical, clinical, and toxicological studies on poorly developed plant species are necessary to see if there are chemical, microbiological, and/or clinical evidences, from a scientific perspective, of their effectiveness for those health disorders (Sharma et al., 2014). For example, the plants reported by Tharu community, namely, *Caesulia axillaris* Roxb. (Asteraceae), *Ehretia laevis* Roxb. (Boraginaceae), *Eulaliopsis binata* (Retz.) C.E. Hubb. (Poaceae), *Milletia extensa* (Benth.) Baker (Fabaceae), *Sida cordata* (Burm. f.) Bors. Waalk. (Malvaceae), and *Vanda tessellata* (Roxb.) Hook. ex G. Don (Orchidaceae) are advised to assess their potential for developing new medicines or skin care products (Sharma et al., 2014).

2.5. Sunscreen and transepidermal water loss (TEWL)

Modern pharmacological research showed that the extracts of safflower (known in India as Kusumbhi (*Carthamus tinctorius* L., Asteraceae)) have several physiological functions such as anticoagulant, vasodilating, antioxidant, melanin suppressive, immunosuppressive and antitumor (Lin et al., 2018). For this reason, it is a component of the sunscreen (Table 5) as usually the antioxidant activity presents high potential as a UVB sunscreen agent (Mukherjee et al., 2011) (Abdul Karim et al., 2014).

Topical application of soybean oil extracts (*Glycine max* (L.) Merr., Fabaceae) has been shown to reduce Transepidermal water loss (TEWL) of the skin of the forearm (Lin et al., 2018), thus *Lactobacillus*/Soybean ferment extract is an ingredient of moisturizing cream produced in India (Table 5). Soybean with pot marigold (*Calendula officinalis*), green tea (*Camellia sinensis*), turmeric (*Curcuma longa*), liquorice (*Glycyrrhiza glabra*), common chamomile (*Matricaria chamomilla*), *Punica granatum*, and *Vitis vinifera* are polyphenolic plants applied in cosmetology and dermatology as having anti-inflammatory, antiaging, skin-whitening and antimicrobial properties, and estrogen-like effects as well (Ratz-Lyko et al., 2015). Thus, these plants are also listed in the

Table 6
List of bacterial pathogens responsible for skin diseases in human and related symptoms.

| SN | Disease | Bacterial pathogen | Signs and symptoms | Area |
|-----|-----------------------|--|---|--|
| 1. | Cellulitis | <i>Streptococcus pyogenes</i> | Painful, swollen and red infection | Occurs most often on the legs |
| 2. | Folliculitis | <i>Staphylococcus aureus</i> | Red, swollen bumps | Hair follicles |
| 3. | Impetigo | <i>Staphylococcus aureus</i> , <i>Streptococcus pyogenes</i> | Large blisters, yellow, crusted appearance | Face, arms, and legs |
| 4. | Boils | <i>Staphylococcus aureus</i> | Red, tender bumps | Around hair follicle |
| 5. | Erysipelas | <i>Streptococcus pyogenes</i> | Large, raised red patches on the skin | Face, legs and nasal passages |
| 6. | Carbuncles | <i>Staphylococcus aureus</i> | Hair follicle, small scrape, or puncture | Skin surface, throat, and nasal passages |
| 7. | Leprosy | <i>Mycobacterium leprae</i> | Discolored patches of skin, usually flat | Skin ulcers, nerve damage, and muscle weakness |
| 8. | Ecthyma | <i>Staphylococcus aureus</i> | Lymph nodes become swollen and painful | Buttocks, thighs, legs, ankle and feet |
| 9. | Erythrasma | <i>Corynebacterium minutissimum</i> | Skin lesions with pink patches and wrinkling that become red, then brown and scaly. | Armpits, groin, or between the toes |
| 10. | Necrotising fasciitis | <i>Staphylococcus aureus</i> , <i>Haemophilus</i> , <i>Vibrio</i> sp., <i>Escherichia coli</i> , <i>Bacteroides fragilis</i> | Soft tissue and fascia | Any part of the body |
| 11. | Spa pool folliculitis | <i>Pseudomonas aeruginosa</i> | Scattered small red itchy or tender bumps, itchy follicular papules and pustules | Any part of the body |
| 12. | Thermal burn wounds | <i>Pseudomonas aeruginosa</i> | Blister, red skin, swelling | Populate beneath on protected layer |
| 13. | Chronic leg ulcers | <i>Pseudomonas</i> spp. | Malodorous greenish superficial crust | Defect in the skin below the level of knee |
| 14. | Ecthyma gangrenosum | <i>Pseudomonas aeruginosa</i> | Blisters, necrotic ulcers | Perineum and under arm pit |

ethnodermatology of the Indian subcontinent.

3. Discussion

Various plant parts are used to treat dermatologic conditions in India: rhizomes, root, stem, flowers, fruit, bark, seed, seed oil, buds, latex, and whole plant parts (Fig. 3). Preparation methods and administration differ significantly. Topical administration takes the form of more complex preparations: decoctions, infusions, ointments, oils, poultices, tars, tinctures, unguents, or in a dried state (powder or ash) (Gilca et al., 2018). This review presents an addition to the existing knowledge of home remedies that are in current practice for the treatment of skin diseases in India. Most traditional medicines were prepared using water as a medium. The method of administration was topical and confined to the affected body part but also orally (in some cases). Analysis of how to use herbal preparations orally in diseases such as acne, carbuncles, leprosy, measles, urticaria, scabies, leucoderma, ringworm and dry skin conditions indicate that healers are perfectly

familiar with the systemic course of the malady. In addition to pure herbal preparations, in some cases, the medicine was given along with milk, curd, ghee (clarified butter), coconut oil, or honey, for an enhanced impact (Manohar, 2012). For example, in the case of acne, Winkelman (2018) reported that aromatherapy, plant ingredients, and essential oils (plant extracts) are also important as potential agents in acne treatment (Winkelman, 2018), confirmed by bioactivity studies, preliminary evidence and small pilot clinical trials conducted outside of North America, predominantly in young adults. Sandalwood oil (*Santalum album* L., Santalaceae) is used as a remedy in many Asian countries for the treatment of inflammatory and skin eruptions as it possesses antimicrobial activity against *Staphylococcus aureus*, *S. epidermidis*, and *P. acnes* in concentrations of 0.06% and lower. In India, sandalwood oil is also an ingredient of face gold massage cream and moisturizer cream (Table 5). Sandalwood oil products were generally well tolerated, but burning, dryness, and stinging were reported as the most common treatment-related complaints (Winkelman, 2018). In Europe, however, other native (or other more popular cultivated) plants are used against acne, e.g., stinging nettle leaves (*Urtica dioica* L., Urticaceae), walnut husk (*Juglans regia* L., Juglandaceae), myrtle leaves (*Myrtus communis* L., Myrtaceae), chamomilla flowers (*Matricaria chamomilla* L., Asteraceae) and rose flowers (*Rosa × damascena* Herrm., Rosaceae) (Kılıç et al., 2019). The two latter plants are also used in Indian dermatology (Table 5), as these herbal extracts demonstrated strong antibacterial and anti-inflammatory activity in preliminary trials, the topical application of these botanical extracts can be good candidates for local acne treatment (Kılıç et al., 2019).

Some Indian cosmetics listed in Table 5 (e.g., Antiacne cream, Active D pigmentation cream, or Anti-blemish gel) contain also jojoba oil (*Simmondsia chinensis* (Link) C.K. Schneid., Simmondsiaceae) that comprise a high content of wax esters makes it a good option for repairing dermatoses with an altered skin barrier, such as seborrhea, eczema or atopic dermatitis (AD) and acne (Fig. 9) (Lin et al., 2018).

Mediterranean plants can also be admixed with typical Indian plants as ingredients, e.g., Rosemary extract (*Rosmarinus officinalis* L., Lamiaceae) is also a component of the Face gold massage cream (Table 5). Namely, the addition of rosemary extract may have an anti-inflammatory effect in cosmeceutical or dermatological products, also confirmed by injection of rosemary extract which is not associated with skin irritation or inflammation in mice (Winkelman, 2018). Thanks to

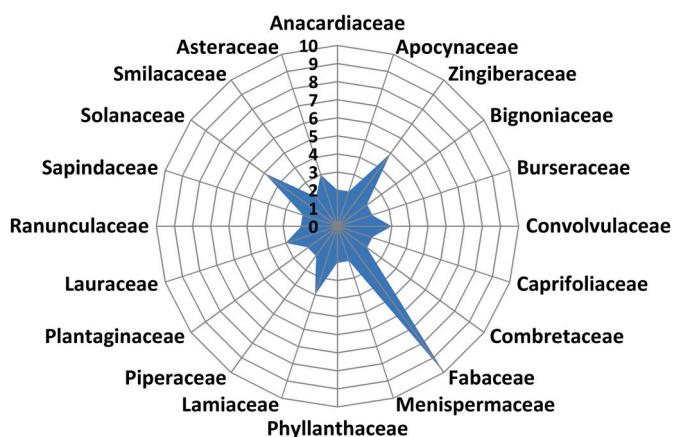


Fig. 4. Ayurvedic formulations in dermatology: how many species of the plant family are used (apart from single uses per family). The use of the Fabaceae family prevails significantly (ten species of this family are used in Ayurveda). It is followed by the following plant families: Solanaceae and Zingiberaceae (five species used in each family) and Lamiaceae (four species are used in Ayurveda).

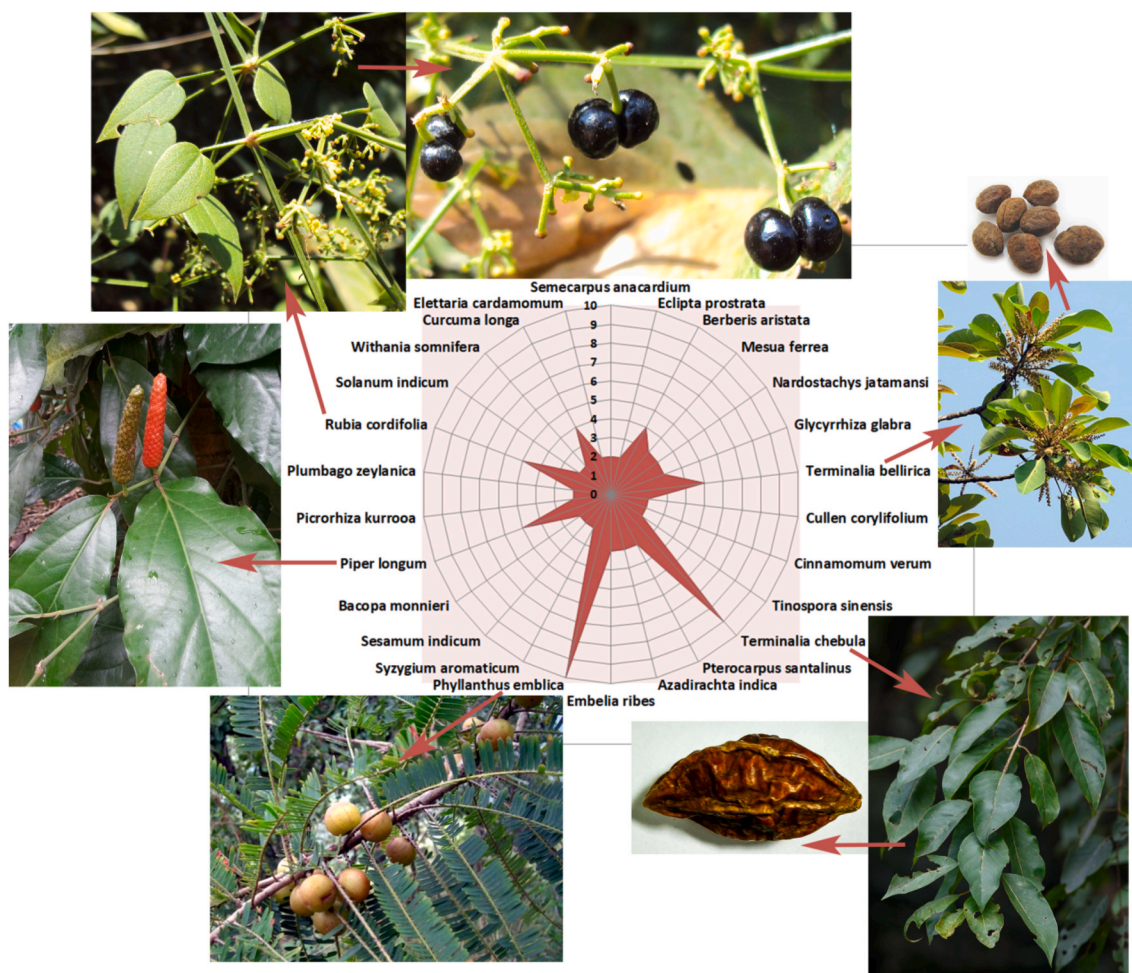


Fig. 5. Ayurvedic formulations in dermatology. Most often used plant species are: *Phyllanthus emblica* L. (Phyllanthaceae, bottom left), *Terminalia chebula* Retz. (Combretaceae, bottom and middle right), *Terminalia bellirica* (Gaertn.) Roxb. (Combretaceae, top and middle right), *Piper longum* L. (Piperaceae, middle left), and *Rubia cordifolia* L. (Rubiaceae, top left and middle). All photographs were reproduced on CC-BY-2.5, CC BY-SA 3.0, and CC BY-NC 4.0 licences.

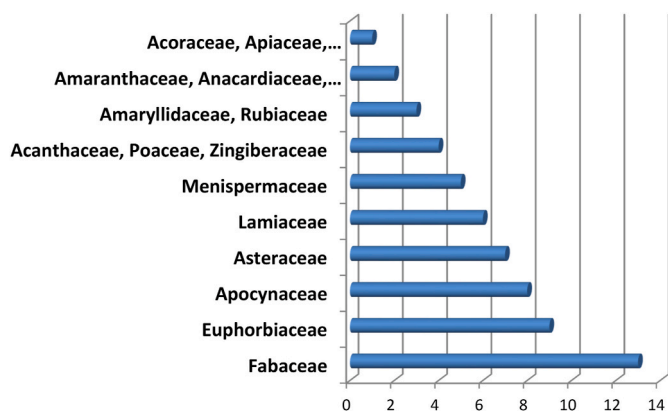


Fig. 6. Ethnodermatological drugs by plant families used in India. Plants of Fabaceae, Euphorbiaceae, Apocynaceae, Asteraceae, Lamiaceae prevail. X axis: number of plant species reported.

these properties, Rosemary is grown in many countries of the world.

Further comprehensive ethnobotanical and ethnopharmacological research could benefit the development of medicinal plants for skincare (Figs. 9 and 10) and treatment (A. Gupta et al., 2010). India offers a great deal of scope for ethnobotanical research on skin diseases, not only because of the richness of the flora but also because of the ancient culture of traditional medicine and hence the experience in applying these

treatments (Jeeva et al., 2007).

Natural plant oils are widely used worldwide for skin care (as topical therapy) because they are usually readily available and are relatively inexpensive. Many natural oils contain specific compounds with antibacterial, anti-inflammatory, antioxidant, and anti-itch properties, which make them an alternative and attractive complement to the treatment of xerotic and inflammatory dermatoses related to skin barrier disorders (Vaughn et al., 2018). Plant oils applied topically may have different effects on the skin depending on their composition and the pathophysiological properties of the skin (Lin et al., 2018). In addition to the olive oil (*Olea europaea* L., Oleaceae) listed above, in Indian medicine, some plant oils are also commonly used: sunflower seed oil (*Helianthus annuus* L., Asteraceae) in Antiacne cream and moisturizer cream (Table 5). Natural oils such as sunflower or sesame seed oil (*Sesamum indicum* L., Pedaliaceae) have been suggested to be a good option for promoting skin barrier homeostasis. In traditional Taiwanese medicine, sesame oil has been used to relieve inflammatory joint and wound pain and has shown chemopreventive effects in a two-step carcinogenesis mouse skin cancer model (Lin et al., 2018). In Ayurvedic Medicine, olive oil is an ingredient of Divya Kayakalpa Taila directed against skin disorders including ring worm, itching, sun burn, eczema, leucoderma, psoriasis, urticaria and allergy. Vipadikahara ghrita taila is used against skin lesions and contains sesame oil as well. However, in Ayurvedic Medicine, it was also found that excessive intake of sesame seed is the reason of obstinate skin state (Routh and Bhowmik, 1999). All of these ingredients are very helpful in skin treatment, but the most important

| Used against | Euphorbiaceae | Fabaceae | Apocynaceae | Asteraceae | Zingiberaceae |
|---|---------------|----------|-------------|------------|---------------|
| Cuts, wounds | 5 | 1 | 1 | 6 | 6 |
| Ringworm | 3 | 2 | 2 | 1 | 1 |
| Skin diseases | 3 | 1 | 1 | | 4 |
| Eczema | 2 | 2 | 1 | | 3 |
| Sores | 2 | 1 | | | 2 |
| Scabies | 1 | 1 | 2 | | 1 |
| Leucoderma | 1 | 2 | | | |
| Burning area | 1 | 1 | | | 6 |
| Wart | 1 | | 1 | | 5 |
| Herpes | 1 | | | 1 | 4 |
| Infective hepatitis | 1 | | | 1 | 3 |
| Affected area of skin | 1 | | | | 2 |
| Carbuncle | 1 | | | | 1 |
| Itching | 1 | | | | |
| Pimple | 1 | | | | 6 |
| Sole on legs | 1 | | | | 5 |
| Leprosy | | 2 | 1 | 1 | 4 |
| Psoriasis | | 2 | 1 | | 1 |
| Skin infection | | 2 | 1 | | 2 |
| Inflammatory diseases of skin | | 1 | | | 1 |
| Astringent | | 1 | | | |
| Boils | | 1 | | | 6 |
| Dandruff | | 1 | | | 5 |
| Dermatitis | | 1 | | | 4 |
| Skin allergies | | 1 | | | 3 |
| Swelling on hands and legs | | 1 | | | 2 |
| Ear ache | | | 2 | | 1 |
| Abdominal cramps | | | 1 | | |
| Rash | | | 1 | | 6 |
| Snake bites | | | 1 | | 5 |
| Syphilis | | | 1 | | 4 |
| Acne | | | | 1 | 3 |
| Bites | | | | 1 | 2 |
| White spot of skin | | | | 1 | 1 |
| Wormy skin sores | | | | 1 | |
| Hyperpigmentation | | | | | 2 |
| Leukoderma | | | | | 1 |
| Premature skin wrinkle | | | | | 1 |
| Skin allergy caused by insect bites or microbes | | | | | 1 |

Fig. 7. Quantification of use reports by dermatological problems and five most used plant families in Indian ethnodermatology. Euphorbiaceae and Asteraceae plants are mostly used on cuts and wounds. Euphorbiaceae plant species are also reported against ringworm and skin diseases (generally).

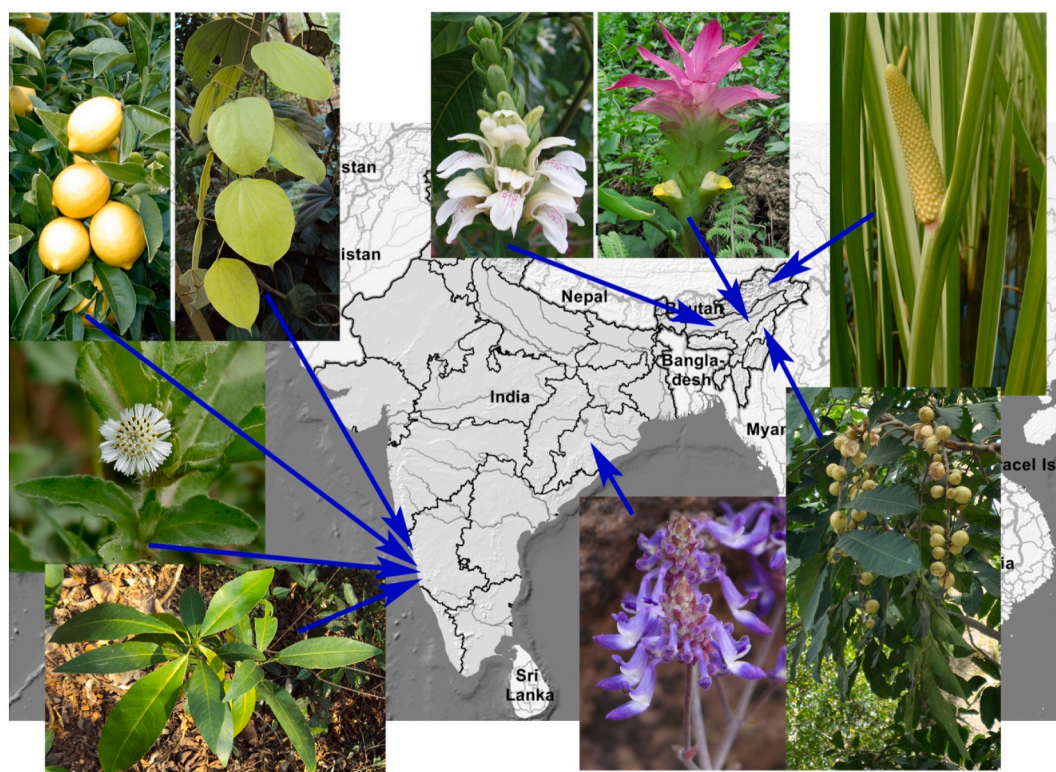


Fig. 8. Less commonly used plant species in Indian dermatology, only reported from specific regions of India. *Citrus limon* (L.) Osbeck, Rutaceae (top left), *Coscinium fenestratum* Colebr., Menispermaceae (second left), *Eclipta prostrata* (L.) L., Asteraceae (middle left), and *Croton persimilis* Müll. Arg., Euphorbiaceae (bottom left) are reported only in ethnodermatology of Coastal Karnataka. *Justicia adhatoda* L., Acanthaceae (middle top) and *Curcuma aromatica* Salisb., Zingiberaceae (second right) are only reported in ethnodermatology of Assam. *Acorus calamus* L., Acoraceae (top right) is only reported in ethnodermatology of Arunachal Pradesh. *Aphanamixis polystachya* (Wall.) R. Parker, Meliaceae (bottom right) is only reported in ethnodermatology of Nagaland. *Anisochilus carnosus* (L.f.) Wall., Lamiaceae (middle bottom) is only reported in ethnodermatology of Odisha. All photographs were reproduced with CC BY 2.0, CC BY-SA 3.0, or CC BY-SA 4.0 licences.

Ayurvedic issue to be considered is to tailor to the doshic imbalances and then learn how to correct the potential imbalances. Plant based carrier oils recommended for the skin according to the aromatherapeutic literature (Orchard and van Vuuren, 2019) are also used in Indian ethnodermatology (Tables 1–5); they are obtained from the following plant species: *Aloe vera* (L.) Burm.f. (Asphodelaceae), *Calendula officinalis* L. (Asteraceae), *Cocos nucifera* L. (Arecaceae), *Helianthus annuus* L. (Asteraceae), *Macadamia ternifolia* F. Muell. (Proteaceae), *Prunus dulcis* (Mill.) D.A.Webb (= *Prunus amygdalus* var. *dulcis* (Borkh. ex DC.) Koehne) (Rosaceae), *Simmondsia chinensis* (Link) C.K. Schneid. (Simmondsiaceae), *Triticum vulgare* Vill. (Poaceae), *Vitis vinifera* L. (Vitaceae). *V. vinifera* L. is also used in Ayurvedic formulations such as Chyawanprash, that accelerates wound closure (Lin et al., 2018). Thus, it is also included in Face Gold massage cream, anti-acne cream, anti-blemish gel, and moisturizer cream (Table 5). Direct topical application of grape seed oil to human skin has not yet been well researched, however, resveratrol has a direct antibacterial effect against pathogens, such as *Enterococcus faecalis*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* (Lin et al., 2018).

The ethanol plant extract of *Cocos nucifera* L. (Arecaceae) was not confirmed for antimicrobial activity against *Propionibacterium acnes* and *Staphylococcus epidermidis* (Kumar et al., 2007), but topical application of virgin coconut oil is effective in the promotion of wound healing through much faster epithelization also important in the treatment of acne (Lin et al., 2018). Additionally, in veterinary dermatology, it was found that the use of a topical spray that contains essential oils of plant origin and fatty acids and compounds with antimicrobial properties (e. g., Manuka oil) may accelerate the resolution of pyoderma and shorten the duration of antibacterial treatment (Bensignor et al., 2016).

Traditional medicine is a promising alternative therapeutic strategy

that can be used in oral and/or topical modes against various skin conditions. It highlights the significance of monoherbal as well as polyherbal formulations in the treatment of skin ailments. A plethora of investigations have been carried out to depict the multifaceted use of traditional medicines to treat different skin conditions. This information may be considered as valuable for dermatologists from the West to use such therapies as alternative or adjunct treatment strategies. In this review, an attempt has been made to present an ethnodermatological evidences across India with their efficacy, methods of preparation and administration. However, almost no literature was obtained describing the safety issues regarding possible contamination of the applied formulations. Moreover, more elaborate clinical placebo-controlled studies are needed for better understanding, systematic analysis and ramifications of the traditional dermatological applications in human.

However, it should be remembered that in Ayurvedic medicine there is no need to use preset formulations. There is no tailoring to each individual or looking at different individual nuances for each disease. Moreover, some evidence in Ayurvedic medicine is still only supported by *in vitro* testing, and *in vitro* evidence does not always successfully extend into the clinical realm. Therefore, extensive clinical studies are still required for most of these candidate plant extracts.

4. Concluding remarks

This review shows that the knowledge of herbal medicine is being used to treat different kinds of human skin disorders by local people. In the Indian subcontinent, complementary therapy for dermatological problems and treatment remains the main option for millions of people. This review of ethnobotanical dermatology in India confirms the belief that their analysis will accelerate the discovery of new, effective

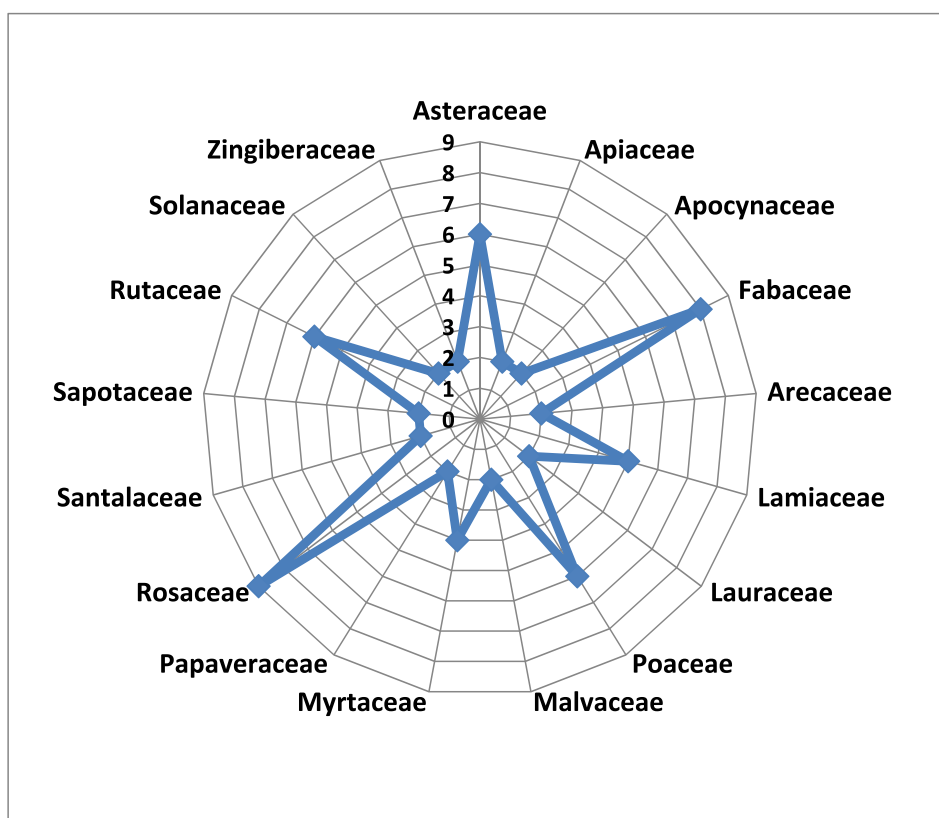


Fig. 9. Herbal drugs and cosmetics products in dermatology in India: the number of species of the plant family used. The utility of the Rosaceae family prevails (nine species of this family are used in cosmetics produced in India), followed by the following plant families: Fabaceae (eight species are used in cosmetics products in India) and Asteraceae, Poaceae, and Rutaceae (six species utilised from each family).

therapeutic agents for skin diseases. We conclude that the administration of extracts from most of these plant species is topical and few only are administered orally. We also compiled the pharmacological activity of the extracts of the most cited plants against mice, bacterial and fungal pathogens, and human cells. As confirmed by [Ariffin and Hasham \(2016\)](#), more studies and clinical evidence are still required to determine if the identified species may contribute to skin condition treatment, particularly in atopic eczema. Today, ethnodermatology is a well-accepted international discipline and many new practices have been initiated in numerous countries. The topic is of supreme significance since many skin conditions are reportedly treated by the traditional medicines alongside the Western medications as alternative or in conjunction. Substantial evidence supports the efficacy of traditional formulations, but the safety issues are also needed to be remembered that may accompany the application of such medicines. We hope this article will further accelerate the development of this area to identify a new generation of natural human skin treatments that will help meet the growing consumer demand for safe, sustainable, and natural treatments. In this context, research on plants utilised in ethnodermatology in India and elsewhere should be intensified with further systematic and rigorous research needed for analyses and testing of traditional dermatological preparations that may eventually guide to formulate novel therapeutics against skin disorders.

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Availability of data and materials

The datasets generated in the current study are available in the Tables attached to this paper.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

CRediT authorship contribution statement

Uttpal Anand: Writing – original draft, Figures and Tables Preparation, Project administration, Attempt Reviewers and Editor Comments. **Champa Keeya Tudu:** Writing – original draft, Figures and Tables Preparation. **Samapika Nandy:** Writing – original draft, Figures and Tables Preparation. **Kumari Sunita:** Writing – original draft, Writing – review & editing, Figures and Tables Preparation, Attempt Reviewers and Editor Comments. **Vijay Tripathi:** Conceptualization, Formal analysis, Data curation, Writing – review & editing, Figures and Tables Preparation, Resources, Supervision. **Gary J. Loake:** Overall Proofreading. **Abhijit Dey:** Conceptualization, Formal analysis, Data curation, Visualization, Attempt Reviewers and Editor Comments, Resources, Supervision, Project administration. **Jarosl w Pro k w:** Conceptualization, Software, Formal analysis, Data curation, Writing – review & editing, Figures and Tables Preparation, Visualization, Attempt Reviewers and Editor Comments, Resources, Supervision, Funding acquisition.

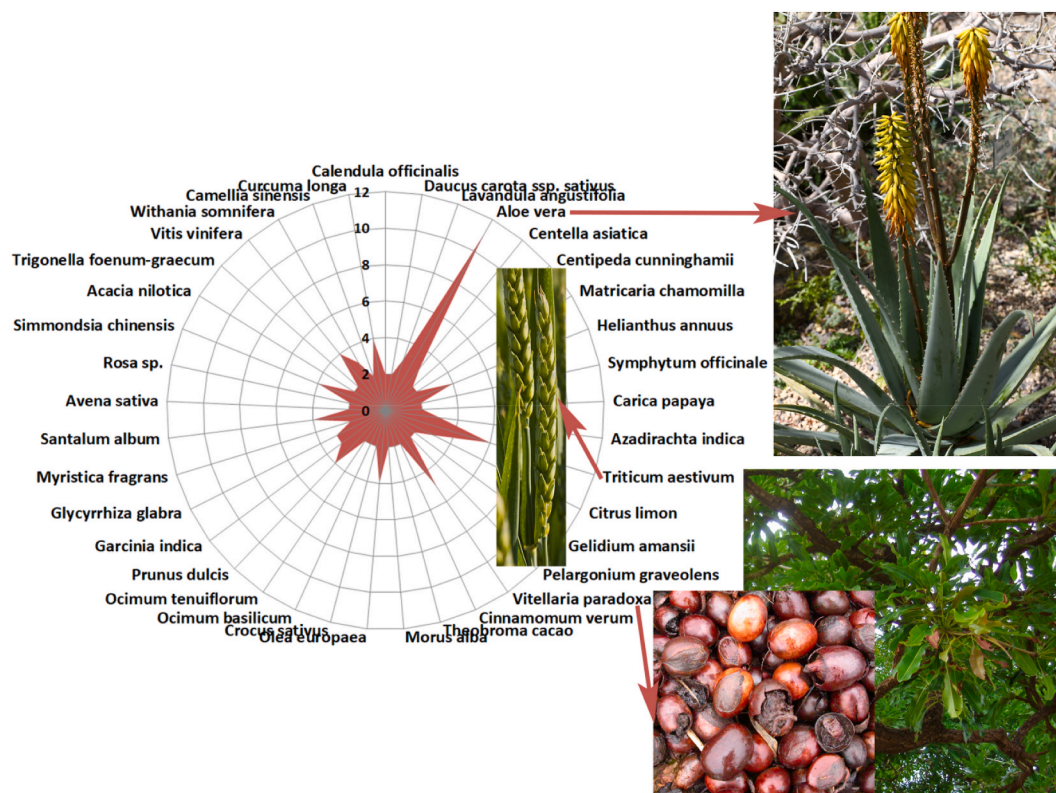


Fig. 10. Herbal drugs and cosmetics products in dermatology in India. Most frequently used plant species in cosmetics products are: *Aloe vera* (Asphodelaceae, top right), *Triticum aestivum* (Poaceae, centrally on the chart), and *Vitellaria paradoxa* (= *Butyrospermum parkii*) (Sapotaceae, both bottom). All photographs reproduced on CC BY 2.0, CC-BY-SA 2.5, and CC BY-SA 3.0 licences.

Declaration of competing interest

The other authors declared no potential conflicts of interests.

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Further reading

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