Assembly of D1/D2 complexes of photosystem II: binding of pigments and a network of auxiliary proteins

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Running title: Earliest steps in PSII assembly

One sentence summary: Analysis of isolated assembly complexes provides new insights into the early stages of photosystem II biogenesis.

Author contributions

J.Kno and M.B. constructed strains and performed isolation of complexes and their electrophoretic analyses; R.S. analyzed data and calculated pigment ratios; J.P. performed HPLC pigment analyses; L.C. performed MS analyses of proteins; J.Y. and J.P.T. constructed strains expressing tagged D1 and D2 proteins; D.B. measured charge separation activity; J.K. made spectroscopic measurements and analysed electrophoretic and MS data; J.Kno., R.S., J.K., and P.J.N. wrote the article; P.J.N. and J.K. acquired the funding and supervised the project; all authors discussed the results and commented on the article.

ABSTRACT

Photosystem II (PSII) is the multi-subunit light-driven oxidoreductase that drives photosynthetic electron transport using electrons extracted from water. To investigate the initial steps of PSII assembly, we used strains of the cyanobacterium *Synechocystis* sp. PCC 6803 arrested at early stages of PSII biogenesis and expressing affinity-tagged PSII subunits to isolate PSII reaction center assembly (RCII) complexes and their precursor D1 and D2 modules ($D1_{mod}$ and $D2_{mod}$). RCII preparations isolated using either a His-tagged D2 or a FLAG-tagged PsbI subunit contained the previously described RCIIa and RCII* complexes that differ with respect to the presence of the Ycf39 assembly factor and high-light-inducible proteins (Hlips) and a larger complex consisting of RCIIa bound to monomeric PSI. All RCII complexes contained the PSII subunits D1, D2, PsbI, PsbE, and PsbF and the assembly factors rubredoxin A (RubA) and Ycf48, but we also detected PsbN, Slr1470, and the Slr0575 proteins, which all have plant homologs. The RCII preparations also contained prohibitins/stomatins (Phbs) of unknown function and FtsH protease subunits. RCII complexes were active in light-induced primary charge separation and bound chlorophylls, pheophytins, beta-carotenes, and heme. The isolated $D1_{mod}$ consisted of D1/PsbI/Ycf48 with some Ycf39 and Phb3, while $D2_{mod}$ contained D2/cytochrome b_{559} with co-purifying PsbY, Phb1, Phb3, FtsH2/FtsH3, CyanoP, and Slr1470. As stably bound chlorophyll was detected in $D1_{mod}$ but not $D2_{mod}$, formation of RCII appears to be important for stable binding of most of the chlorophylls and both pheophytins. We suggest that chlorophyll can be delivered to RCII from either monomeric PSI or Ycf39/Hlip complexes.

Keywords: photosystem II biogenesis, reaction center assembly complex, assembly factors, pigment binding

INTRODUCTION

Photosystem II (PSII) is the unique multi-subunit oxidoreductase embedded in the thylakoid membranes (TM) of cyanobacteria and chloroplasts that catalyzes the solar-powered oxidation of water. Electrons extracted from water are utilized for fixation of carbon into organic molecules while molecular oxygen, the by-product of PSII activity, is released into the atmosphere. In cyanobacteria, PSII consists of 17 intrinsic and three extrinsic protein subunits and numerous pigments and redox-active cofactors (Zouni et al., 2001; Umena et al., 2011). At the heart of PSII is a heterodimer of two homologous proteins, D1 and D2, both having five transmembrane helices, which are flanked by two large chlorophyll (Chl) - binding inner antennae, known as CP43 and CP47. There are also several low molecular mass (LMM) subunits which are rather featureless except for PsbE and PsbF which bind heme to form cytochrome b_{559} (Cyt b_{559}) (Barber, 2014). The extrinsic lumenal subunits PsbO, PsbU, PsbV, and CyanoQ serve to stabilize the oxygen-evolving Mn_4CaO_5 cluster. In chloroplasts of plants and green algae, the lumenal PsbU and PsbV subunits have been replaced by the structurally different PsbP and PsbQ proteins, while the intrinsic part of PSII is similar to that of cyanobacteria (Ifuku, 2015; Roose et al., 2016).

Biogenesis of this complicated molecular machine is an intricate and highly organized process. Analysis of PSII knock-out mutants of the cyanobacterium *Synechocystis* sp. PCC 6803 (hereafter *Synechocystis*) has revealed that the large pigment-binding proteins first associate with their neighboring LMM subunits to form small building blocks (or modules) which then sequentially assemble via a series of intermediates to form the functional PSII complex (Komenda et al., 2012b) .

In the earliest stage of PSII biogenesis, the D1 and D2 subunits form modules ($D1_{mod}$, $D2_{mod}$) with adjacent small PSII proteins and auxiliary factors. $D1_{mod}$ was shown to contain PsbI, Ycf48, Ycf39 and HliC/D (Dobáková et al., 2007; Knoppová et al., 2014) while $D2_{mod}$ contains both the PsbE and PsbF subunits of Cyt b_{559} (Komenda et al., 2008) and CyanoP (Knoppová et al., 2016). So far, no information about their pigment content has been reported. $D1_{mod}$ and $D2_{mod}$ then combine to form the PSII reaction center assembly complex, RCII (Komenda et al., 2008; Knoppová et al., 2014; Kiss et al., 2019). The subsequent attachment of $CP47_{mod}$ results in the formation of the RC47 intermediate (Boehm et al., 2011; Boehm et al., 2012) which then binds $CP43_{mod}$ (Boehm et al., 2011) to give rise to the monomeric PSII core complex, RCCII. The final

steps involve the light-driven formation of the Mn_4CaO_5 cluster, the attachment of the lumenal subunits, and dimerization of the PSII complex. The process of PSII biogenesis is assisted by several step-specific auxiliary or assembly factors, many of unclear function (Nixon et al., 2010; Komenda et al., 2012b; Heinz et al., 2016). PSII biogenesis in chloroplasts follows a similar sequence of events (Nickelsen and Rengstl, 2013) and is assisted by homologous as well as chloroplast-specific auxiliary factors (Lu, 2016).

The transient nature and low accumulation of assembly complexes have hindered their detailed analysis. Nevertheless, a combination of protein tagging (either His-tag or FLAG-tag) and the use of appropriate mutants that block assembly at a specific step has allowed the isolation and characterization of the CP43 and CP47 modules (Boehm et al., 2011; Bučinská et al., 2018) and the RC47 complex (Boehm et al., 2012).

Using a FLAG-tagged derivative of the Ycf39 assembly factor, we have also isolated and characterized the protein composition and spectral characteristics of one type of RCII complex (called RCII*) associated with Ycf39 and High-light inducible proteins (Hlips) (Knoppová et al., 2014). The Ycf39 and single-helix Chl- and β-carotene (β-car) binding Hlips (Komenda and Sobotka, 2016) form a stable sub-complex that is implicated in the photoprotection of RCII (Komenda et al., 2008; Knoppová et al., 2014).

Isolated thylakoid membranes also contain a smaller RCII complex called RCIIa (Komenda et al., 2008; Knoppová et al., 2014) which does not bind the Ycf39/Hlips complex. The Ycf39/Hlips complex can be detached from the isolated RCII* *in vitro* (Knoppová et al., 2014) but the relationship between RCIIa and RCII* during the biogenesis of PSII *in vivo* is not yet clear.

We show here that isolated RCII complexes perform light-driven primary charge separation and have a pigment composition that corresponds well to that found in the D1/D2 part of the PSII crystal structure (Umena et al., 2011). Several newly identified proteins that co-purify with RCII might play a role in PSII biogenesis. Moreover, our data suggest that RCII* and RCIIa are formed independently via distinct assembly pathways. We also isolated and characterized FLAG-tagged $D1_{mod}$ and $D2_{mod}$. Both modules lacked pheophytin *a* (Pheo) but contained neighboring small PSII subunits and several co-purifying proteins that might represent previously unidentified D1- and D2-specific assembly factors. Stably bound Chl was detected just in $D1_{mod}$.

RESULTS

Spectral properties and protein composition of RCII complexes isolated using a His-tagged D2 subunit

We have shown previously that RCII complexes accumulate to low but detectable levels when PSII assembly is blocked due to the lack of the PSII inner antenna CP47 (Komenda et al., 2004; Knoppová et al., 2014). To isolate RCII complexes, we first introduced a modified *psbDI* gene coding for D2 with an N-terminal 6xHis tag into the previously characterized Tol145 strain depleted in phycobilisomes and lacking both *psbD* genes (Tang et al., 1993). In a second step, we inactivated the *psbB* gene coding for the CP47 antenna to produce the His-D2/ΔCP47 strain. Two-dimensional gel electrophoresis consisting of clear native (CN) electrophoresis in the first dimension followed by SDS-PAGE in the second dimension confirmed that His-D2/ΔCP47 accumulated small amounts of RCII complexes, denoted RCII* and RCIIa, similar to the ΔCP47 strains characterized previously (Komenda et al., 2008; Knoppová et al., 2014). The slightly lower mobility of the D2 protein identified in the second dimension suggested successful incorporation of His-tagged D2 into RCII complexes (Fig. S1).

To purify RCII complexes, membranes isolated from the His-D2/ΔCP47 strain were solubilized using the mild detergent β-dodecyl-D-maltoside (DM) and subjected to nickel (Ni) affinity chromatography. After extensive washing the final eluate showed a similar absorption spectrum to that of plant or *Synechocystis* PSII reaction center complexes (PSII RC) prepared by detergent extraction of the CP47 and CP43 antennae from larger PSII core complexes (Fig. 1; compare with Fig. 2 in Oren-Shamir et al. (1995) and Tomo et al. (2008)). However, the Chl red absorption maximum of our preparation was blue-shifted by about 3 nm indicating the prevalence of more blue-absorbing Chl forms (with maxima at 670-671 nm) over the forms absorbing at longer wavelengths. The low temperature Chl fluorescence spectrum had a main maximum at 682 nm which is also similar to that identified by Tomo et al. (2008). A small fluorescence shoulder at 720 nm indicated the possible presence of PSI (see below).

To characterize the complexes present in the preparation, we used two-dimensional gel electrophoresis. As shown in Fig. 1C the preparation was heterogeneous and contained several types of complex in line with previously published data (Komenda et al., 2008; Knoppová et al., 2014). We detected the presence of the two complexes previously designated RCII* and RCIIa with masses of about 200 and 150 kDa, respectively. A combination of immunoblotting and mass

spectrometry confirmed that both RCII* and RCIIa contain the D1, D2, PsbE, PsbF and PsbI subunits of PSII, as well as the assembly factors Ycf48 (Komenda et al., 2008; Yu et al., 2018) and the more recently described RubA (Kiss et al., 2019). As already noted, RCII* differs from RCIIa by association with the Ycf39/Hlips complex (Knoppová et al. 2014), which was also confirmed in our new preparation (Fig. 1C).

Torabi et al. (2014) have suggested the involvement of PsbN in the formation of the RCII complex in tobacco (*Nicotiana tabacum*). Using specific antibodies raised against *Synechocystis* PsbN, we detected PsbN in both RCII* and RCIIa (Fig. 1C) with PsbN more abundant in RCII* than in RCIIa. Moreover, the preparation also contained another putative PSII-related protein, Slr0575, a homologue of the plant Acclimation of Photosynthesis to the Environment 1 protein, APE1 (Walters et al., 2003). After separation by CN-PAGE the Slr0575 protein was mostly found in the unassembled protein fraction, but some remained associated with RCII*. The presence of Slr0575 in RCII* was confirmed by re-analysis of the previously characterized FLAG-Ycf39 preparation (Fig. S2).

We also constructed a His-D2/ΔCP47 mutant that lacked Ycf39. As anticipated, the isolation of RCII from this strain confirmed the loss of the RCII* complex and the accumulation of the RCIIa complex (Fig. S3). RCIIa was separated into two closely spaced fluorescent bands with the larger one containing an additional protein identified as Slr1470 by mass spectrometry (MS, Table S1).

The amounts of RCII* and RCIIa detected in cells of CP47-less strains are usually very similar (see Fig. S1). To test whether there is a precursor-product relationship between them, with one converting into the other, we performed a classic radioactive pulse-chase experiment using cells of the CP47-less strain. Both D1 and D2 were labeled in both complexes to a similar extent during the pulse and the label in both proteins declined similarly in RCII* and RCIIa, indicating no clear inter-conversion between these two complexes (Fig. S4). Thus, we propose that each complex is formed independently via a distinct assembly pathway.

Identification of a RCIIa/PSI assembly complex

In addition to the RCII* and RCIIa complexes, the His-D2 preparations also contained larger Chl-containing complexes: a fluorescent one of mass about 350 kDa containing the same protein components as RCIIa and most probably representing its dimer, and a larger, much less

fluorescent one, of about 450 kDa. The latter complex also contained components of RCIIa, but, in addition, showed the presence of PSI subunits. Based on its size, the complex likely represents RCIIa bound to a PSI mononomer (RCIIa/PSI; Fig. 1C). The content of RCIIa/PSI in the preparations was highly variable as documented by its higher level in another preparation isolated under the same conditions (Fig. S5). To verify the composition of the complexes and judge whether the formation of the RCIIa/PSI complex was an isolation artifact of the Ni affinity purification procedure, we compared the His-D2 preparation with another preparation isolated using FLAG affinity chromatography from a CP47-less strain expressing a C-terminally FLAG-tagged PsbI subunit (PsbI-FLAG). Both preparations consisted of similar complexes including RCIIa/PSI (Fig. 1 and Fig. S6). In addition, the FLAG-based purification provided a preparation devoid of most of the non-specific components identified in the His-D2 preparation (compare the control His- and FLAG-tag pull-downs in Fig. S7 and Fig. S8, respectively).

Prohibitin 3 (Phb3, Slr1128; Boehm et al., 2009) and FtsH2/FtsH3 complexes were present in both the His-D2 and the control WT pull-down together with elongation factor EF-Tu, a typical contaminant of our His-tag preparations (Fig. S1 and S7). The PsbI-FLAG preparation (Fig. S6) also contained both Phb3 and FtsH2/FtsH3, and additionally prohibitin 1 (Phb1, Slr1106), but all these proteins were absent in the control FLAG-tag pull-down (Fig. S8). We therefore conclude that FtsH2/FtsH3, Phb1 and Phb3 are authentic components of RCII preparations. These components were also identified by MS analysis (Table S2).

Primary charge separation in RCII complexes

The D1-D2 heterodimer binds the Chl and Pheo molecules involved in the light-induced primary charge separation step within PSII. The primary electron donor, P680, is considered to be a collection of four special Chl molecules bound to D1 and D2 whereas the primary electron acceptor Pheo is bound specifically to D1 (Diner and Rappaport, 2002; Romero et al., 2012). Light-induced electron transfer from P680 to Pheo results in the formation of the $P680^{+}Pheo^{-}$ radical pair (Diner and Rappaport, 2002). Primary charge separation also occurs in plant and cyanobacterial PSII RC complexes prepared by detergent-induced removal of the CP43 and CP47 antenna complexes from larger PSII core complexes (Nanba and Satoh, 1987; Giorgi et al., 1996; Tomo et al., 2008).

To judge whether the RCII assembly complexes isolated here were capable of primary charge separation, we measured the reversible light-induced absorption difference spectra in the range 650-710 nm. This measurement was performed either in the presence of the electron acceptor, SiMo, to detect the accumulation of the oxidized primary donor, $P680^+$, or in the presence of sodium dithionite to detect reduction of the primary electron acceptor Pheo, according to Vácha et al. (2002). Both methods (Fig. 2) gave similar results to those obtained with the isolated plant PSII RC complex (Vácha et al., 2002). These data showed that at least some of the RCII assembly complexes can perform the primary photochemical reaction and therefore should contain the complete set of pigments required for these reactions. Because our RCII preparations contained variable amounts of the monomeric PSI complex associated with many more Chl molecules (Malavath et al., 2018) than the RCII complexes, precise quantification of the photoactive P680 or Pheo was not reliable. Instead, we focused on further purification of each RCII complex from the crude His-D2 preparation to assess their individual spectral properties and pigment composition.

Pigment composition of individual RCII complexes

To separate RCII* and RCIIa we used semi-preparative CN-PAGE as the similar size of RCII complexes did not allow their separation using gel filtration, and ionex chromatography was also unsuccessful (Fig. S9). Separated complexes from several CN gel lanes were eluted from the gel and characterized using absorption and 77 K fluorescence spectroscopies and pigment composition was determined using HPLC. The overlay of absorption spectra of the RCII* and RCIIa complexes normalized to the 415 nm absorption maximum showed that they have the same position of the red maximum of Chl at 672 nm and mainly differed in the content of carotenoids absorbing in the 430-520 nm region (Fig. 3A). The 77 K Chl fluorescence spectra were practically identical with a maximum at 685 nm, which is similar to the fluorescence maximum displayed by PSII RCs isolated by fragmentation (Tomo et al., 2008). The RCIIa complex did show a slightly higher satellite peak at 740 nm than RCII* (Fig. 3B).

We also measured both types of spectra for the RCIIa/PSI complex containing the PSI monomer and RCIIa complex. The red absorption peak was red-shifted by 4 nm due to the red-absorbing Chls of the PSI monomer (Fig. 3A). Interestingly, the 77 K Chl fluorescence spectrum showed a

typical peak of RCII at 680 nm and that of PSI at 720 nm but there was also a shoulder around 675 nm (Fig. 3B), which is typical for Chl molecules in detergent or lipid micelles (Trinugroho et al., 2020). No such 675-nm shoulder was observed in RCII* or in RCIIa.

We subsequently performed a detailed HPLC analysis of Chls, carotenoids and heme-*b* (Fig. 4A). Heme-*b* was used as a standard for normalization of the pigment content as exactly one heme-*b* (Cyt b_{559}) should be present in all types of RCII complexes (Table 1). The pigment composition of RCIIa was similar to that previously determined for the *Synechocystis* PSII RC complex prepared by stripping the CP47 and CP43 inner antennae from the complete PSII core complex (Tomo et al., 2008). Based on the crystal (Umena et al., 2011) and cryo-EM (Gisriel et al., 2020) structures of PSII, the 6 Chl are likely to represent the four central Chls belonging to the P680 oligomer plus $ChlZ_{D1}$ and $ChlZ_{D2}$ ligated by D1-His118 and D2-His117 on the periphery of the RCII complex, respectively. Our measurements also support the presence of 1-2 predicted carotenoids in the D1/D2 heterodimer. RCII* contained an additional three Chl and a single β-car molecule which can be attributed to the presence of the Ycf39/Hlip complex. We also analyzed the RCIIa/PSI complex and given the expected 1:1 ratio between PSI and RCII based on the estimated size of the complex, it is interesting that the PSI complex appears to contain on average only about 60 Chls, which is much less than the value of 95 Chls predicted from the known structure of monomeric PSI (Malavath et al., 2018).

Isolation of D1 and D2 assembly modules

To investigate the steps preceding RCII assembly, we isolated and characterized unassembled forms of both D1 and D2. Both these forms are associated with adjacent small PSII subunits and several auxiliary factors and are termed D1 and D2 modules ($D1_{mod}$ and $D2_{mod}$). For their isolation, we constructed new strains expressing N-terminal FLAG-tagged versions of D1 and D2 (FLAG-D1/ΔPSII and FLAG-D2/ΔPSII, respectively) in the previously constructed ΔPSII strain lacking the *psbA, psbB, psbC* and *psbD* genes encoding D1, CP47, CP43 and D2, respectively (Trinugroho et al., 2020). The corresponding FLAG-tagged D1 and D2 assembly modules were then isolated using immunoaffinity chromatography.

The Chl and carotenoid content of the isolated FLAG-D1 preparation was low and consequently the absorption spectra of the preparation (Fig. 5A) displayed an unusually large maximum around

620 nm indicating an increased level of contaminating phycocyanin which has been previously detected in FLAG-specific pulldowns (Knoppová et al., 2014). Its relatively high content is most probably related to the very low cellular level of unassembled D1 and the need for extensive concentration of the preparation, so that even small amounts of contaminating protein become strongly pronounced. The red peak of Chl absorption with a maximum of about 677 nm (Fig. 5A) suggested the presence of PSI and this was confirmed from the 77 K Chl fluorescence spectrum (Fig. 5B) which exhibited a lower maximum at 720 nm belonging to PSI-bound Chl. There was also a peak at 675 nm indicating weakly associated Chl bound in lipid/detergent envelope of the protein. On the other hand, the fluorescence maximum at 683 nm showed the presence of Chl specifically associated with FLAG-D1 since it was missing in the control preparation from the FLAG-free ΔPSII strain lacking all four large PSII Chl-proteins (Fig. S10B). The stable binding of Chl to the FLAG-D1 was also confirmed by the detection of Chl fluorescence (Fig. 6, 1D fluor and 2D Chl, red arrows) at the position of the main FLAG-D1 band (Fig. 6, 2D SYPRO stain and 2D blots, blue arrows) on the 1D and 2D gels. HPLC analysis of the preparation (Fig. 4B) confirmed the presence of Chl and β-car, which might, however, partly belong to contaminating PSI. Most importantly, no Pheo or heme was detected in the preparation.

A 2D immunoblot analysis confirmed the isolation of the FLAG-tagged D1 protein together with PsbI, Ycf39 and Ycf48. Based on its staining intensity, Ycf48 looked to be present at a ratio of close to 1:1 with the main D1 protein band (Fig. 6, F.D1 with a blue arrow). Additional protein bands were also present in the preparation. Apart from the PSI trimer and monomer there was also a large abundance of ribosomal subunits, which are all likely contaminants present in this highly concentrated preparation of a rare FLAG-tagged protein. This is confirmed by the high abundance of ribosomal subunits in the control preparation lacking a FLAG tag (Fig. S8, right panel). Immunoblotting also revealed the presence of the prohibitin homologue Phb3 and the FLAG-D1 specific co-elution of this protein together with the previously mentioned components Ycf39 and Ycf48 was confirmed by MS analysis (Tables 2 and S3).

The absorption spectrum of the isolated FLAG-D2 preparation was characterized by a small peak at 559 nm and a sharp main maximum at 429 nm suggesting the presence of Cyt b_{559} (Figs. 5A and S10A). The 77 K Chl fluorescence emission spectrum had a small peak at 721 nm again suggesting the presence of PSI while the dominating band peaked at 675 nm indicating the presence of Chl weakly bound in the lipid/detergent envelope of the protein (Figs. 5B and S10B).

This was confirmed by the absence of a distinct Chl fluorescence band in the 1D CN gel (Fig. 6, 1D fluor, red arrow) at the position of the main FLAG-D2 band (Fig. 6, 2D SYPRO stain and FLAG blot, blue arrows). Instead, we detected a smeared 2D Chl fluorescence underneath the broadened part of the Flag-D2 band (Fig. 6, 2D Chl, red rectangles). Protein analysis confirmed the presence of contaminating PSI complexes, phycocyanin and ribosomal subunits while the stained components were identified as FLAG-D2 and the PsbE and PsbF subunits of Cyt b_{559} (Fig.6). The other specific components detected in the preparation by immunoblotting were CyanoP, FtsH3 and the two prohibitin homologues Phb1 and Phb3. The small amount of Ycf48 can be considered unspecific, since similar trace amounts of the protein were found in the control (Fig. S8). MS analysis (Tables 2 and S4) confirmed the components detected by the immunoblotting and identified several other specific proteins including FtsH2, Slr1470 and PsbY, which is a PSII subunit localized in some structural models of PSII in the vicinity of PsbE (Guskov et al., 2009; Kato et al., 2021). Besides the presence of Chl and β-car, which are likely to be due to PSI contamination, we detected a high content of heme belonging to Cyt b_{559} but no Pheo (Fig. 4B). Interestingly, the absorption peak at 559 nm suggested that Cyt b_{559} was present in its reduced form which is unusual for isolated PSII complexes (Shinopoulos and Brudvig, 2012).

Discussion

RCII* and RCIIa are distinct assembly complexes

RCII assembly complexes are normally undetectable in WT and have only been observed in cells grown at low temperature (Komenda et al., 2008). Increased levels are, however, observed in either mutants lacking the CP47 subunit (Komenda et al., 2004) or mutants impaired in the de novo biosynthesis of Chl needed for efficient CP47 accumulation (Hollingshead et al., 2016). Although the level of RCII complexes in CP47 knock-out strains is less than 10 % of PSII in WT cells (Komenda et al., 2004), this was sufficient for isolation and characterization.

We show here that the main RCII complexes present in preparations isolated using either His-tagged D2 (Fig. 1) or FLAG-tagged PsbI (Fig. S6) in a CP47 deletion background are the previously identified RCII* and RCIIa complexes (Komenda et al., 2008). Both complexes contain D1, D2, the PsbE and PsbF subunits of Cyt b_{559}, PsbI and the lumenally exposed Ycf48

accessory factor but only RCII* contains the Ycf39/Hlip complex (see Knoppová et al., 2014). Ycf39 belongs to the family of short-chain dehydrogenase/reductases but it remains unclear whether Ycf39 has a catalytic activity or has evolved an alternative function such as binding mRNA (for review see Kavanagh et al. (2008)). Ycf39 binds to a pair of Hlips ligating 4-6 Chls and 2 β-cars, which can quench Chl excited states via energy transfer to an S_1 state of β-car (Staleva et al., 2015) thereby potentially protecting RCII* from photodamage (Knoppová et al., 2014).

Proteins that co-purify with RCII and the D1 and D2 modules

The only accessory protein factors present in close to stoichiometric amounts with respect to the main RCII components were Ycf48 in both complexes, Ycf39/Hlips in RCII* and RubA in RCIIa. We could not detect RubA in either $D1_{mod}$ or $D2_{mod}$ although we have previously co-isolated RubA and D1 with FLAG-Ycf39 from the D2-less strain (Knopová et al., 2014; Kiss et al., 2019). RubA is thought to facilitate the mutual binding of $D1_{mod}$ and $D2_{mod}$ so its greater abundance in RCII might reflect tighter binding due to interactions with both modules.

In addition, we have now identified several additional factors that are present at sub-stoichiometric levels and thus not easily observed in stained gels, but which are clearly detectable by immunoblotting and MS analyses. The presence of these factors causes microheterogeneity within the population of RCII complexes which is manifested in 1D native gels as double bands (Fig. S3) and on 2D gels by broader bands of the major components that extend horizontally towards higher molecular mass (for instance bands of D1 and D2 in RCII* in Figs. 1, S2, S5 and S6).

One of these factors is the PsbN protein, which has previously been detected in tobacco (*Nicotiana tabacum*) RC assembly complexes but not in any type of cyanobacterial PSII. The *Synechocystis* PsbN protein does not contain positively charged amino acids and is therefore less easy to detect by an ordinary MS bottom-up analysis after trypsin cleavage. Its higher level in RCII* appears to be complementary to lower levels of RubA in the same sub-complex (Fig. S6), possibly suggesting neighboring or overlapping binding sites for both components. The previous deletion of the *psbN* gene in *Synechocystis* (Mayes et al., 1993) did not exhibit any apparent effect on the photoautrotrophic growth of the mutant. However, PsbN in tobacco is required for

repair from photoinhibition and efficient assembly of RCII (Torabi et al., 2014), which is in agreement with our localization of the protein within this complex. Similarly, RubA is also needed for PSII repair and RCII formation in *Synechocystis* (Kiss et al., 2019) and *Chlamydomonas* (García-Cerdán et al., 2019).

Another component found in RCII is the Slr0575 protein which together with CyanoP appears to bind mostly to RCII* (Fig. S2). The *Arabidopsis* (*Arabidopsis thaliana*) homologue of Slr0575 is APE1 (corresponding gene is *At5g38660*), which has been detected in the *Arabidopsis* complex equivalent to RCII*, consisting of One Helix Protein 1 (OHP1) and One Helix Protein 2 (OHP2) (plant homologues of Hlips), D1, D2, High Chlorophyll Fluorescence 136 (HCF136, plant homologue of Ycf48) and High Chlorophyll Fluorescence 244 (HCF244, plant homologue of Ycf39) (Myouga et al., 2018). Our identification of Slr0575 in RCII* (Figs. 1 and S2) provides important support for a conserved role for Slr0575/APE1 in the early steps of PSII biogenesis. Such a role is consistent with the phenotype of the *ape1* mutant of *Arabidopsis* which is defective in increasing the levels of PSII upon a transition from low to high light (Walters et al., 2003). In contrast, the *Synechocystis* deletion mutant did not show any substantial phenotypic changes when compared with WT (Thompson, 2016).. However, when we deleted this gene in the ΔCP47 strain and compared protein synthesis in ΔCP47 and the ΔCP47/ΔSlr0575 double mutant, the latter showed a 50 % higher labelling of D2 within the RCIIa complex compared to CP43 and PSI large subunits PsaA/B (Fig. S11). Since protein staining of the 2D gel did not show any apparent increase in the accumulation of RCII in the double mutant, the result indicates that Slr0575 increases the stability of unassembled D2. In agreement with this, we found traces of Slr0575 in the Flag-D2 but not in the FLAG-D1 preparation.

The Slr1470 subunit was detected as a Coomassie Blue stained band in the RCIIa complex isolated from the Ycf39-less strain (Fig. S3) and was also detected in the Flag-D2 preparation but not in the FLAG-D1 or PsbI-FLAG preparations. Slr1470 is predicted in Uniprot to possess a single transmembrane helix and a bacterial pleckstrin homology domain (InterPro entry IPR012544) which is implicated in lipid binding (Xu et al., 2010). The gene for Slr1470 is conserved among oxygenic phototrophs (the homologous gene in *Arabidopsis* is *AT1G14345*) and our attempts to delete the gene were not successful indicating its crucial importance.

We also found that prohibitin subunits, Phb1 and Phb3, and FtsH2/FtsH3 protease complexes co-purified with PsbI-FLAG tagged RCII preparations (Fig. S6) but were absent from control non-FLAG preparations (Fig. S8). The presence of the FtsH2/FtsH3 complex might be related to the targeting of RCII for degradation (Krynická et al., 2015). Prohibitins are structurally related to HflC/K proteins that form large supercomplexes with FtsH protease complexes in *Escherichia coli* and regulate FtsH activity (Saikawa et al., 2004). Indeed, the *Synechocystis* prohibitin homologue, Phb1, has previously been detected in affinity-purified FtsH2/FtsH3 preparations but at a much lower level than that found here in the RCII preparations (Boehm et al., 2012). MS analysis of the PsbI-FLAG preparation (Table S2) also revealed the specific presence of another FtsH protease FtsH4 of unknown function, the thylakoid curvature homologue CurT (Slr0483) and CP43. Possibly these proteins are located in the vicinity of RCII complexes when CP47 is absent.

It is perhaps not surprising that the FtsH2/FtsH3 protease complex together with Phb1 were detected in $D2_{mod}$, since we have previously shown that the level of unassembled D2 is controlled by FtsH2/FtsH3 (Komenda et al., 2006). Therefore, $D2_{mod}$ is expected to be targeted by this protease complex for degradation. The absence of FtsH2/FtsH3 and Phb1 in $D1_{mod}$ is also in agreement with our previous data demonstrating a negligible effect of the FtsH2 deletion on the level of unassembled D1 (Komenda et al., 2010). Together these data suggest that either the site of $D1_{mod}$ formation in cells differs from that of $D2_{mod}$ as suggested by Schottkowski et al. (2009), or that $D1_{mod}$ is not recognized by FtsH2/FtsH3, for instance due to the binding of Ycf48.

More surprising is the association of $D1_{mod}$, $D2_{mod}$ and RCII with the Phb3 protein which has been previously detected in the plasma membrane fraction obtained using a two-phase partitioning technique (Boehm et al., 2009). Possibly Phb3 is located at the junction of the thylakoid and plasma membranes in a region called the thylapse where PSII biogenesis might be initiated (Rast et al., 2019). Phb3, which forms a large circular homocomplex, is more closely related to stomatins than the prohibitins (Boehm et al., 2009). The role of stomatins remains unclear but they have been implicated in regulating the structural organization of membranes which would be consistent with a location for Phb3 in the thylapse (Rast et al., 2019).

Although our previous characterization of Phb1 and Phb3 in *Synechocystis* did not reveal a crucial role for these proteins in PSII repair/biogenesis (Boehm et al., 2009), their association

418 with $D1_{mod}$, $D2_{mod}$ and RCII suggested a possible involvement in the stability of these PSII
419 assembly intermediates. To test the stabilizing role of Phb3, we deleted the *phb3* gene in the
420 ΔCP47 and ΔCYT strains lacking CP47 and Cyt b_{559}, respectively. In both cases the deletion did
421 not affect the levels of $D1_{mod}$, $D2_{mod}$ and RCII leaving the importance of Phb3 unclear.

422 As expected from previous analyses, the $D1_{mod}$ contains PsbI, which binds to the first and second
423 transmembrane helices of D1, plus Ycf48 which docks on to the lumenal surface of D1
424 (Komenda et al., 2008, Yu et al., 2018). $D2_{mod}$ contains Cyt b_{559} subunits at stoichiometric levels
425 (Komenda et al., 2008), and present at lower levels are PsbY, which binds to PsbE (Guskov et al.,
426 2009), and CyanoP, a lipoprotein that binds to lumenal surface of D2 (Cormann et al., 2014;
427 Knoppová et al., 2016) (Table 2).

428

429 **The complex of RCIIa with monomeric PSI and its physiological relevance**

430 Unlike RCII*, RCIIa was also detected in a binary complex with a PSI monomer (Fig. 1C, see
431 below) which added another layer of complexity to the RCII preparations. The RCIIa/PSI
432 complex was present in both the His-D2 and PsbI-FLAG preparations indicating that it is not an
433 artifact related to the unspecific binding of PSI to the isolation resin. When membranes of the
434 His-D2/ΔCP47 strain were solubilized with different concentrations of DM before His-D2
435 purification, the RCIIa/PSI complex was obtained preferentially at lower DM concentrations
436 while an increased amount of DM led to its decreased level while the content of both RCII
437 complexes in the preparation increased (Fig. S12). Surprisingly, we were unable to detect
438 RCIIa/PSI in solubilized membranes isolated from the CP47-less strain (see Fig. S1). Our
439 interpretation of these results is that RCIIa/PSI is unstable and can be stabilized by binding to the
440 isolation resins while at higher detergent concentration it becomes disrupted again. The marked
441 increase in yield of RCII* and RCIIa obtained by extracting at higher detergent levels suggests
442 that these complexes reside in membrane regions that are not as easily solubilized as standard
443 thylakoids which are normally solubilized completely at both 1.5 and 3 % (w/v) DM. These
444 regions might be related to so-called lipid rafts or detergent-resistant membranes (DRMs) which
445 were originally described in eukaryotic cells but are now recognised to be present in bacterial
446 membranes (for review see Lopez and Koch (2017)). A hallmark of DRMs is the presence of
447 members of the band 7 protein or SPFH family (consisting of the stomatin, prohibitin, flotillin,

and HflK/C proteins) (Rivera-Milla et al., 2006) and so it is notable that we could detect Phb1 and Phb3 in RCII preparations. Overall, our data support the concept that thylapses, the proposed site of PSII biogenesis (Rast et al., 2019), could represent a specialised detergent-resistant membrane.

Pigment composition of RCII complexes and D1/D2 assembly modules

The electrophoretically purified RCIIa complex showed absorption and low temperature fluorescence spectra that were in line with those previously reported for PSII RCs isolated after stripping CP43 and CP47 from plant PSII-enriched membrane particles (Nanba and Satoh, 1987; Gounaris et al., 1990) or from *Synechocystis* PSII complexes using high concentrations of Triton X-100 (Gounaris et al., 1989; Oren-Shamir et al., 1995; Tomo et al., 2008). The main difference was a blue-shifted maximum of the Chl red absorption peak (from 676 nm in plant to 672 nm in our *Synechocystis* complex; Fig. 3). The observed ratio of two Pheos, 1-2 carotenoids and six Chls per Cyt b_{559} suggests the binding of a full complement of pigments to the D1/D2 heterodimer in RCII. The low amounts of complex precluded the detection of plastoquinone by HPLC so the presence of bound quinone cannot be excluded. The RCII* complex had a higher content of Chl and β-car than RCIIa due to the presence of pigment-containing Ycf39/Hlips complex (Table 1; Knoppová et al., 2014; Staleva et al., 2015). However, the isolated Ycf39-Hlip complex alone binds 4 - 6 Chls and two β-cars (Staleva et al., 2015; Shukla et al., 2018), thus RCII* appears to have two or three Chls less than expected. One possible explanation is that Ycf39/Hlip has delivered some of its Chl to RCII. The presence of a sufficient set of pigments (Chls, pheophytins) needed for charge separation in both RCII* and RCIIa suggests that both complexes possess this basic photochemical activity. We did not directly measure the individual activities of RCII* and RCIIa due to their low yields; this measurement was feasible only for the crude His-D2 preparation containing both types of RCII complex (Fig. 2).

The analysis of $D1_{mod}$ and $D2_{mod}$ by CN gel electrophoresis and subsequent detection of Chl fluorescence indicated that the main band of $D1_{mod}$ is fluorescent and therefore stably binds Chl. In contrast, the D2 main band was not fluorescent and its broadened part showed just a weak fluorescence with a low temperature emission maximum at 675 nm (Fig. 5 and Fig. S10) corresponding to detergent/lipid-associated Chl or Chl bound to non-native binding sites in D2.

Thus, $D2_{mod}$ appears Chl-deficient or possibly has weakly bound Chl that is easily lost during isolation and/or analysis. This difference between $D1_{mod}$ and $D2_{mod}$ might be related to the binding of Ycf48 to the lumenal side of D1 and Ycf39 to the cytoplasmic side, thereby helping to stabilize the tertiary structure of D1 and binding of Chl (Knoppová et al., 2014; Yu et al., 2018). PsbI, which binds to the first and second transmembrane helices of D1 may also stabilize binding of Chl ligated by His118 of D1 (Dobáková et al., 2007; Umena et al., 2011). Importantly, we were unable to identify Pheo in either $D1_{mod}$ or $D2_{mod}$ suggesting that this pigment is inserted or formed within RCII or during the association of $D1_{mod}$ and $D2_{mod}$.

Two possible pathways for the assembly of RCII complexes

The existence of two main, seemingly independent populations of RCII complexes with specific protein components indicates that cyanobacteria might possess different pathways of RCII biogenesis. We have considered the possibility that the Ycf39/Hlips-containing RCII* with an incomplete complement of Chl (Table 1) could have its origin in a de novo process, while RCIIa could be formed in a repair-like process, in which pigments released from degraded D1 and D2 subunits would be immediately reintroduced into newly synthesized proteins, resulting in RCII with a complete set of pigments. To test this idea, we analyzed the RCII preparation from a His-D2/ΔCP47 strain additionally lacking the FtsH2 (Slr0228) subunit of the FtsH2/FtsH3 protease complex and so impaired in the degradation of damaged PSII (Komenda et al., 2006). This mutant accumulated similar amounts of both RCII* and RCIIa as in the original His-D2 strain and the complexes also had the same pigment composition as the corresponding complexes of the original strain (Fig. S13). The only apparent difference was the almost exclusive presence of the mature D1 protein in the FtsH2-lacking strain. This can be explained by an increase in the time available for D1 maturation due to inhibited turnover of D1 and D2 (Komenda et al., 2006). Thus, it is unlikely that RCIIa comes from the repair process.

The above results led us to the idea of two distinct pathways for the assembly of RCII complexes. One would lead to RCIIa via a combination of $D1_{mod}$ and $D2_{mod}$ without participation of the Ycf39/Hlips complex. Instead, PSI monomeric complexes located in the vicinity would provide photoprotection via energy spill-over and help deliver Chl to RCIIa to give rise to a Chl-depleted sub-population of PSI (Kopečná et al., 2012). Indeed, the gel-purified RCIIa-associated PSI(1)

complex was depleted by about 15 Chls in comparison to PS(1) complexes purified from the CP47-less strain. In agreement with this model, quantification of RCII complexes in membranes of a strain lacking PSI, CP43 and CP47 showed an approximate four-fold higher abundance of RCII* over RCIIa, with the latter possibly formed from release of the Ycf39/Hlip complex from RCII* during the analysis by CN PAGE (Fig. S14). A similar prevalence of RCII* over RCIIa has been observed in the PSI-deficient strains generated by deleting PsbH and both PsbH and Pam68 (Bučinská et al., 2018).

The second pathway requires the assistance of the Ycf39/Hlips complex both to provide photoprotection and deliver Chl to RCII. We hypothesize that this route is the only pathway for building PSII in chloroplasts as the Ycf39 homologue, HCF244, together with the Hlip homologues OHPs are strictly required for D1 synthesis, RCII formation and PSII accumulation (Link et al., 2012; Hey and Grimm, 2018) while in *Synechocystis* RCIIa is formed in the absence of Ycf39. Additional support for this hypothesis comes from the experiments of Muller and Eichacker (1999) who detected a D2 assembly complex in etioplasts that lacked Chl as is the case of the cyanobacterial $D2_{mod}$ described here.

MATERIALS AND METHODS

Construction and cultivation of the strains

The *Synechocystis* strain expressing exclusively the His-tagged version of D2 and lacking CP47 (His-D2/ΔCP47) was derived from the Tol145 mutant lacking both copies of D2 as described by Tang et al. (1993). Plasmid pDC074 carrying a kanamycin-resistance cassette was used as the parental vector for D2 mutagenesis (Suzuki et al., 2013). The coding sequence of 6xHis tag (CATCATCATCATCATCAT) was introduced after the start codon of *psbD1* gene by overlap extension PCR using the primer set *psbDC1F/psbD*-His-2R and *psbD*-His-3F/*psbDC*4R (Supplemental Table S5). The plasmid was then used to transform the Tol145 strain to yield the His-D2 mutant. Then the *psbB* gene was disrupted using the *pPsbB-GentA* vector where the *psbB (slr0906)* was replaced with gentamycin-resistance cassette oriented as the *slr0906* gene. First, an intermediate vector, namely pGEMT-*psbB*, was constructed via overlap extension PCR. Primer sets *slr0906*-1F/*slr0906*-2R and *slr0906*-3F/*slr0906*-4R (Supplemental Table S5) were used to amplify the upstream and downstream flanking sequences, respectively. Both upstream and downstream fragments were then mixed as the DNA template for overlap extension reaction

using the primers *slr0906*-1F and *slr0906*-4R. The resulting fragments, which carry an *EcoRV* restriction site instead of *slr0906,* were then cloned into the multiple cloning region of the pGEM-T Easy vector. Further modifications were then carried out by inserting a gentamycin-resistance cassette into the *EcoRV* site via restriction digestion and ligation to create the final transformation vector p*PsbB*-GentA. To disrupt the *ycf39* locus in the His-D2/ΔCP47 mutant we used the transformation vector p*Slr0399*-ErmA as in Knoppová et al. (2014). The genotypes of the mutants were confirmed by PCR and sequencing. The CP47-less strain ΔCP47 (Eaton-Rye and Vermaas, 1991) was used as a control strain for the evaluation of RCII level (Fig. S1) and for the purification of proteins using the Ni-affinity chromatography (Fig. S7).

The strain expressing the C-terminal FLAG-tagged variant of the PsbI subunit and lacking CP47 (PsbI-FLAG/ΔCP47) was obtained by transforming the CP47 deletion mutant (Eaton-Rye and Vermaas, 1991) using a synthetic DNA construct (Genscript, USA) in which the 3xFLAG coding sequence (Sigma) was inserted before the *psbI* STOP codon and an erythromycin-resistance cassette downstream the gene was used as a selection marker. The Ycf39-binding RCII* was isolated from the CP47-less strain expressing FLAG-Ycf39 as described in Knoppová et al. (2014). The PSII-lacking strains expressing either FLAG-D1 or FLAG-D2 were based on the PSII-less strain ΔD1/ΔD2/ΔCP43/ΔCP47 (Trinugroho et al., 2020) where the N-terminal FLAG-tagged versions of D1 or D2 proteins were introduced using the pPD-FLAG vector (Hollingshead et al., 2012). The *psaA/psaB* deletion strain ΔPSI, the *psbEFLJ* deletion strain ΔCYT, the *ftsH2* deletion strain ΔFtsH2 and *phb3* deletion strain ΔPhb3 were described in Shen et al. (1993), Pakrasi et al. (1988), Komenda et al. (2006) and Boehm et al. (2009), respectively. The corresponding multiple deletion mutants were obtained by transformation using the genomic DNA isolated from these strains.

For purification of the RCII, D1 and D2 complexes, 4-L cultures were grown in 10-L round bottomed flasks in BG11 medium supplemented with 5 mM glucose at 30°C at a surface irradiance of 100 µmol photons m^{-2} s^{-1}. The culture was agitated using a magnetic stirrer and bubbled with air. For thylakoid membrane protein analyses, the strains were grown in 100 mL of BG11 medium plus 5 mM glucose using 250 mL conical flasks on a rotary shaker under 40 µmol photons m^{-2} s^{-1} at 28°C.

Isolation of thylakoid membranes and tag-specific purification

Thylakoid membranes were isolated in buffer A (25 mM MES/NaOH, pH 6.5, 10 mM $CaCl_2$, 10 mM $MgCl_2$, 25% (v/v) glycerol) containing EDTA-free protease inhibitor cocktail (Sigma-Aldrich, USA) using a procedure described in Chidgey et al. (2014). His-tagged RCII was purified using Protino Ni-NTA agarose (MACHEREY-NAGEL, Germany) in a gravity-flow chromatography column at 10 °C after membrane solubilization with DM as described in Knoppová et al. (2021). The FLAG-affinity purification was performed as in Koskela et al. (2020).

Radioactive Labeling

Radioactive pulse and pulse-chase labeling of the cells was performed at 500 µmol photons m^{-2} s^{-1} and 30°C using a mixture of [^{35}S]Met and [^{35}S]Cys (Hartmann Analytic Gmbh, Braunschweig, Germany) as described in Dobáková et al. (2009).

Protein analyses

The composition of purified complexes was analyzed using two-dimensional system combining CN electrophoresis in a 4 % to 14 % (w/v) gradient polyacrylamide gel with SDS-PAGE in a denaturing 16 % to 20 % (w/v) gradient gel containing 7 M urea (Komenda et al., 2012a). The amount of the pull-down preparation loaded onto the gel corresponded to 0.5 µg of Chl. The first-dimensional native gels were photographed (1D color) and scanned for fluorescence (1D fluo). The proteins separated using the denaturing SDS gels were visualized by staining with either Coomassie Blue (CBB) or SYPRO Orange and detected by MS or immunoblotting. The primary antibodies against D1, D2, CP47, CP43, PsbE and PsbF used in this study were previously described by Komenda et al. (2004) and Dobáková et al. (2007). The antibody against Synechocystis Slr0575 and PsbN were raised in rabbit against peptides 161-172 and 32-43 of the *Synechocystis* proteins, respectively, conjugated to keyhole limpet hemocyanin (Moravian Biotechnologies, Czech Republic). We also used our own antibodies raised against *Escherichia coli*-expressed Ycf48 (Yu et al., 2018).

Enzymatic digestion and protein identification by mass spectrometry

The CBB-stained protein bands to be identified were cut from the gel and digested by trypsin. Resulting peptides were extracted, purified with ZipTip C18 pipette tips (Millipore, USA) and

analyzed using a NanoAcquity UPLC (Waters, USA) on-line coupled to an ESI Q-ToF Premier mass spectrometer (Waters, USA) as described in detail in Janouškovec et al. (2013).

The proteins in whole preparations were analyzed after their acetone precipitation. 50 µl of acetone cooled to -20°C was added to the whole protein fraction and after one hour of incubation at -20°C sample was spun down for 10 min at 20 000 g and 4°C. Supernatant was removed and the rest of the acetone was evaporated in a fume hood for approx. 30 minutes. The precipitate was dissolved in 10 µl of 40 mM ammonium bicarbonate in 9 % (v/v) acetonitrile containing 0.4 mg trypsin (proteomics grade; Sigma-Aldrich, USA) and incubated at 37°C overnight. Excessive liquid was removed by Speedvac, and 40 µl of solvent A (0.1 % (v/v) formic acid in water) was added to the 10 µl of tryptic digest. MS analysis was performed using a NanoElute UHPLC (Bruker) on-line coupled to the ESI Q-ToF a high-resolution mass spectrometer (Bruker Impact HD). Peptides were separated by UHPLC using Thermo Trap Cartridge as a trap column and Bruker Fifteen C18 analytical column (75 mm i.d. 150 mm length, particle size 1.9 mm, reverse phase; Bruker). The linear gradient elution ranged from 95 % solvent A (0.1 % (v/v) formic acid in water) to 95 % solvent B (0.1 % (v/v) formic acid in acetonitrile and water (90/10)) at a flow rate of 0.3 mL/min and time 60 min. Eluted peptides flowed directly into the ESI source. Raw data were acquired in the Dynamic MS/MS Spectra Acquisition with following settings: dry temperature 150 °C, drying gas flow 3 l/min, capillary voltage 1300 V, and endplate offset 500 V. The spectra were collected in the range 150–2000 m/z with spectra rate 2 Hz. The CID was set as a ramp from 20 to 60 eV on masses 200–1200, respectively. The acquired spectra were submitted for database search using the MaxQuant software against *Synechocystis* protein databases from the Uniprot Web site (https://www.uniprot.org/ proteomes/UP000001425). Acetyl N-terminal, deamidation N and Q, carbamidomethyl C, and oxidation M were set as variable modifications. Identification of three consecutive y-ions or b-ions was required for a positive peptide match.

Determination of pigment content

Chl content per cell was determined after methanol extraction of pigments according to Ritchie (2006). The ratio of Chl, Pheo and β-car for a single RCII (one heme-*b*) was determined essentially as described in Trinugroho et al. (2020).

Light-induced charge separation in RCII

Measurements of charge separation activity in the preparation of RCII were performed using a home-built kinetic photodiode array spectrophotometer with side illumination (maximum at 660 nm, 1000 μmol photons m^{-2} s^{-1}, provided by an LED source M660L3-C1; Thorlabs, USA) described in Bína et al. (2006). The measurement was performed as described in Vácha et al. (2002). Briefly, samples were diluted to a final Chl concentration of 5 μg ml^{-1} in a buffer A containing 0.04 % (w/v) DM and light-induced oxidation of primary donor was measured in the presence of silicomolybdate at a concentration of 200 μM. Light-induced Pheo reduction was measured in the presence of sodium dithionite and methylviologen at concentrations of 1 mg ml^{-1} and 10 μM, respectively.

Accession Numbers

The Uniprot database accession numbers of proteins identified in this article can be found in Table 2, Supplemental Figure S7 and Supplemental Tables S1, S2, S3, and S4.

Supplemental Data

Supplemental Figure S1. Two-dimensional protein analysis of membranes isolated from the CP47-less mutant expressing a native or a His-tagged variant of the D2 protein (ΔCP47 and His-D2/ΔCP47, respectively).

Supplemental Figure S2. Two-dimensional protein analysis of the FLAG-Ycf39 preparation isolated from the CP47-null mutant expressing FLAG-Ycf39.

Supplemental Figure S3. Two-dimensional protein analysis of preparations isolated from the CP47-less strains expressing His-tagged D2 protein and lacking Ycf39 (His-D2/ΔCP47/ΔYcf39).

Supplemental Figure S4. Two-dimensional protein analysis of the membranes isolated from the CP47-less strain (ΔCP47) after the radioactive pulse-labeling (p) followed by 30 and 60 min of the pulse-chase (pch).

Supplemental Figure S5. Two-dimensional protein analysis of preparation isolated from the CP47-less strains expressing His-tagged D2 protein.

Supplemental Figure S6. Two-dimensional protein analysis of preparations isolated from the CP47-less strains expressing FLAG-tagged PsbI protein using FLAG-specific affinity chromatography.

Supplemental Figure S7. Two-dimensional protein analysis of the control preparation isolated by Ni-affinity chromatography from the control CP47-less strains.

Supplemental Figure S8. Two-dimensional protein analysis of FLAG-D2 and the control preparation.

Supplemental Figure S9. Separation of RCII complexes by ionex chromatography and their absorption spectra.

Supplemental Figure S10. Room temperature absorption spectra and 77K chlorophyll fluorescence spectra of the FLAG-D2 and control FLAG-free preparations.

Supplemental Figure S11. Two-dimensional protein analysis of radioactively labelled membrane proteins of ΔCP47 and ΔCP47/ΔSlr0575 strains.

Supplemental Figure S12. Native gel analysis of His-D2 preparations isolated from membranes of the CP47-less strain expressing His-D2 solubilized with different concentrations of β-dodecyl-maltoside (DM).

Supplemental Figure S13. Two-dimensional protein analysis of preparation isolated from the CP47-less strain lacking FtsH2 and expressing His-tagged D2 protein and the stoichiometry of pigment cofactors in RCII* and RCIIa electrophoretically purified from this preparation.

Supplemental Figure S14. Two-dimensional protein analysis of membrane proteins of ΔPSI/ΔCP47/ΔCP43 strain.

Supplemental Table S1. List of PSII and PSI-related proteins identified by MS and Western blotting in the RCII complexes separated by 2D CN/SDS PAGE from the isolated His-D2/ΔCP47 preparation (see Fig. 1, arrows) and identification of Slr1470 in the His-D2/ΔCP47 preparation (Fig. S3).

Supplemental Table S2. List of the most abundant 40 proteins identified by MS in Flag-PsbI/ΔCP47 preparation.

Supplemental Table S3. List of the most abundant 40 proteins identified by MS in Flag-D1 preparation.

Supplemental Table S4. List of the most abundant 40 proteins identified by MS in Flag-D2 preparation.

Supplemental Table S5. List of primers.

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Tables

Table 1. The stoichiometry of pigment cofactors present in the crude His-D2 preparation and in the RCII*, RCIIa, and PSI/RCIIa complexes. The complexes were purified from the His-D2/ΔCP47 strain using a combination of Ni affinity chromatography and native gel electrophoresis. The ratio of all pigments is normalized to one heme. Data shown as mean of three independent replicates; see methods for details of pigment quantification.

Preparation	Stoichiometry of pigment cofactors			
	Chl-*a*	β-car	Pheo	Heme-*b*
Crude His-D2	9.7 ± 2.2	2.4 ± 0.4	1.2 ± 0.3	1.0
RCII*	9.8 ± 1.1	2.6 ± 0.6	1.8 ± 0.1	1.0
RCIIa	6.7 ± 0.25	1.4 ± 0.3	1.8 ± 0.05	1.0
RCIIa/ PSI(1)	65.2 ± 0.7	7.5 ± 1.2	1.9 ± 0.3	1.0

Table 2. List of proteins identified by MS specifically in the isolated FLAG-D1/ΔPSII and FLAG-D2/ΔPSII preparation after subtraction of proteins identified in the ΔPSII control.

Preparation	Protein UniProt KB No.	Mass spectrometric analysis			
		Size (Da) Length (AA)	Coverage (%)	Detected no. of peptides	Intensity
FLAG-D1	Ycf48 P73069	37267 342	70	15	54533000
	D1 P16033	39695 360	29	10	26748000
	Ycf39 P74429	36496 326	29	10	19328000
	Phb3 P72665	35727 321	61	18	17022000
FLAG-D2	D2 P09192	39466 352	21	5	33302000
	PsbE P09190	9442 81	26	2	18318000
	PsbY P73676	4202 39	21	1	2790100
	FtsH2 Q55700	68496 627	47	5	563000
	Phb3 P72665	35727 321	12	4	515600
	FtsH3 P72991	67250 616	8	4	353700
	PsbF P09191	4929 44	39	1	306100
	Slr1470 P74154	14902 131	7	1	182100
	CyanoP P73952	20747 188	5	1	51900

The analysis of proteins precipitated from the preparation was performed using NanoElute UHPLC (Bruker) on-line coupled to a high-resolution mass spectrometer (Bruker Impact HD).

Figure Legends

Fig. 1. Isolation and analysis of RCII complexes. Room temperature absorption spectra (A), 77 K chlorophyll fluorescence spectra (B) and 2D gel analysis of RCII preparation (C) isolated from cells of the CP47-less strain expressing His-tagged D2. The preparation was isolated using Ni affinity chromatography and analyzed using CN PAGE in the first dimension. The native gel was photographed (1D color) and scanned by LAS 4000 for fluorescence (1D fluor). After SDS-PAGE in the 2nd dimension, the gels were stained by SYPRO Orange (2D SYPRO stain). RCII* is the D1-D2 complex containing Ycf39/Hlips, RCIIa and RCIIa(2) are monomeric and dimeric forms of the D1/D2 complex lacking the Ycf39/Hlips complex, respectively, and RCIIa/PSI designates the PSI monomer bound to RCIIa. Designated proteins were identified by mass spectrometry (see Table S1); non-specifically interacting proteins are designated with an asterisk. PsbN and Slr0575 were identified using specific antibodies. 0.5 µg of Chl was loaded onto the gel.

Fig. 2. Photochemical activity of RCII complexes. The light-induced accumulation of $P680^+$ (A) and $Pheo^-$ (B) in two preparations isolated from the CP47-less strain expressing His-tagged D2 protein. The preparations differed in the content of PSI (green circles, sample analyzed in Fig. 1C; red squares, sample analyzed in Fig. S5), and in a pea 5-Chl reaction center complex isolated using Cu affinity chromatography (black diamonds). Accumulation of $P680^+$ (A) and $Pheo^-$ (B) was elicited by strong red actinic light (wavelength 660 nm, 1000 µmol photons m^{-2} s^{-1}) in the presence of silicomolybdate (electron acceptor) or sodium dithionite (electron donor), respectively. The curves were normalized to the same absorption value of the preparations at the red Qy absorption maximum of Chl.

Fig. 3. Spectroscopic analysis of RCII complexes. Room temperature absorption spectra (A) and 77 K chlorophyll fluorescence spectra (B) of RCIIa/PSI, RCII* and RCIIa complexes. CN gel electrophoresis was used to purify complexes from the preparation isolated using Ni affinity chromatography from cells of the CP47-less strain expressing the His-tagged D2 protein. The absorption spectra were normalized to the 415 nm maximum and the fluorescence spectra to the 680 nm maximum.

Fig. 4. HPLC analysis of pigments and heme in the RCII complexes. RCII* and RCIIa complexes (A) were purified by CN PAGE from the His-D2 preparation and FLAG-D1 and FLAG-D2 preparations (B) were isolated using FLAG affinity chromatography from strains lacking PSII Chl-binding subunits and expressing either FLAG-D1 or FLAG-D2. The analysis was performed as described in Materials and methods and the chromatograms were normalized to the highest peak in the preparation; in RCII*, RCIIa and FLAG-D1 to Chl; in FLAG-D2 to heme.

Fig. 5. Spectroscopic analysis of FLAG-D1 and FLAG-D2. Room temperature absorption spectra (A) and 77 K chlorophyll fluorescence spectra (B) of FLAG-D1 and FLAG-D2. The preparations were isolated using FLAG affinity chromatography from strains lacking PSII Chl-binding subunits and expressing either FLAG-D1 or FLAG-D2. The absorption spectra were normalized to blue maxima, while measurements of the fluorescence spectra were performed using identical volumes of the obtained preparations.

Fig. 6. Two-dimensional protein analysis of FLAG-D1 and FLAG-D2. The preparations were isolated using FLAG affinity chromatography from strains lacking PSII Chl-binding subunits and expressing either FLAG-D1 (FLAG-D1/ΔPSII) or FLAG-D2 (FLAG-D2/ΔPSII). The analysis was performed by CN PAGE in the first dimension and the native gel photographed (1D color) and scanned by LAS 4000 for fluorescence (1D fluor). Following SDS-PAGE in the 2nd dimension, the gel was scanned for Chl fluorescence (Chl, 2D fluor), then stained by SYPRO Orange (2D SYPRO stain) and probed with the designated antibodies (2D blots). Blue arrows mark the fastest native forms of FLAG-D1 (F.D1) and FLAG-D2 (F.D2) and red arrows their Chl fluorescence signals. Chl fluorescence of slower F.D1 and F.D2 forms is in a red dashed rectangle. FP designates free pigments. Both preparations were isolated from the same amount of cells and the analysis was performed on identical volumes of the obtained preparations.

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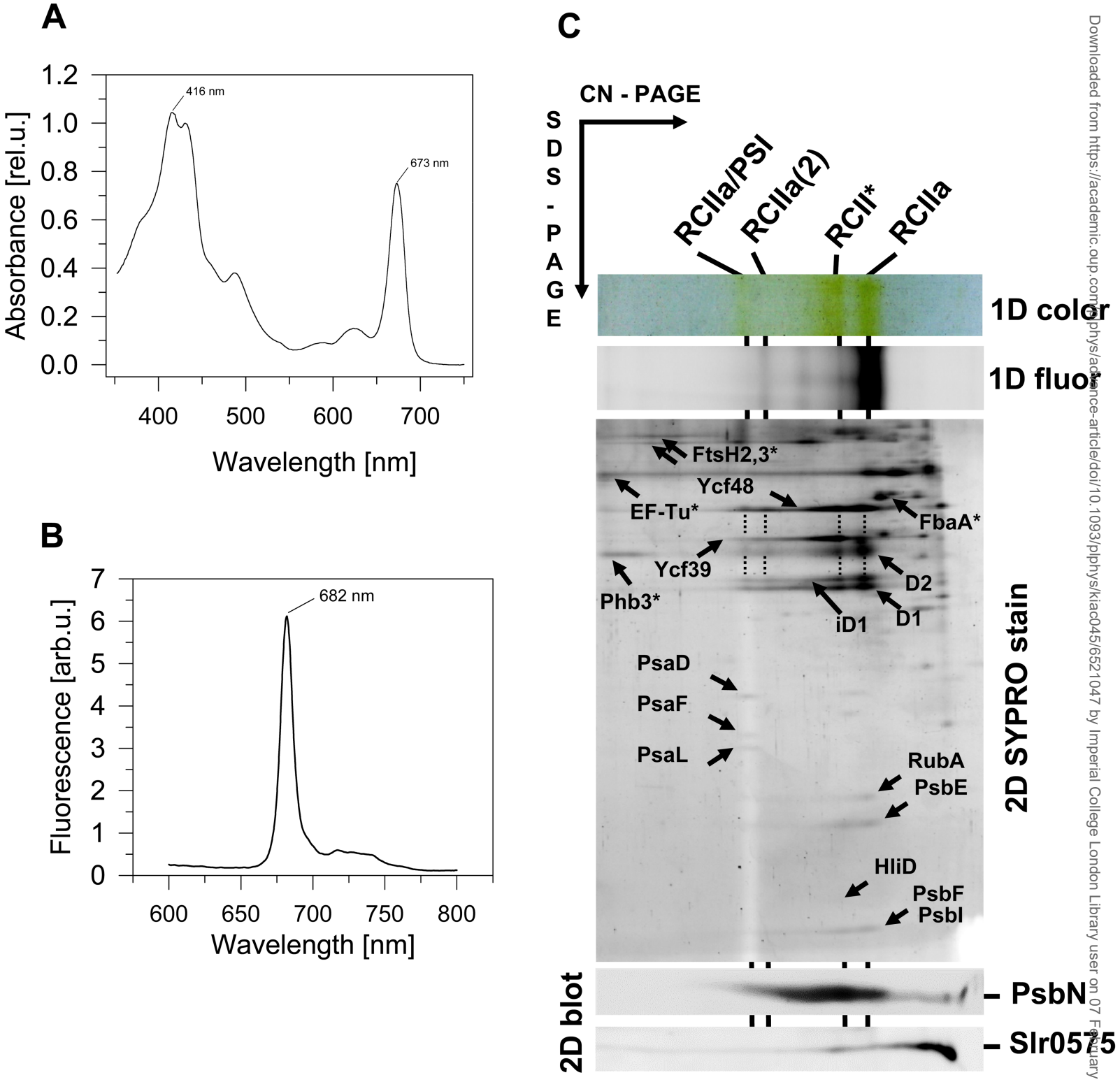

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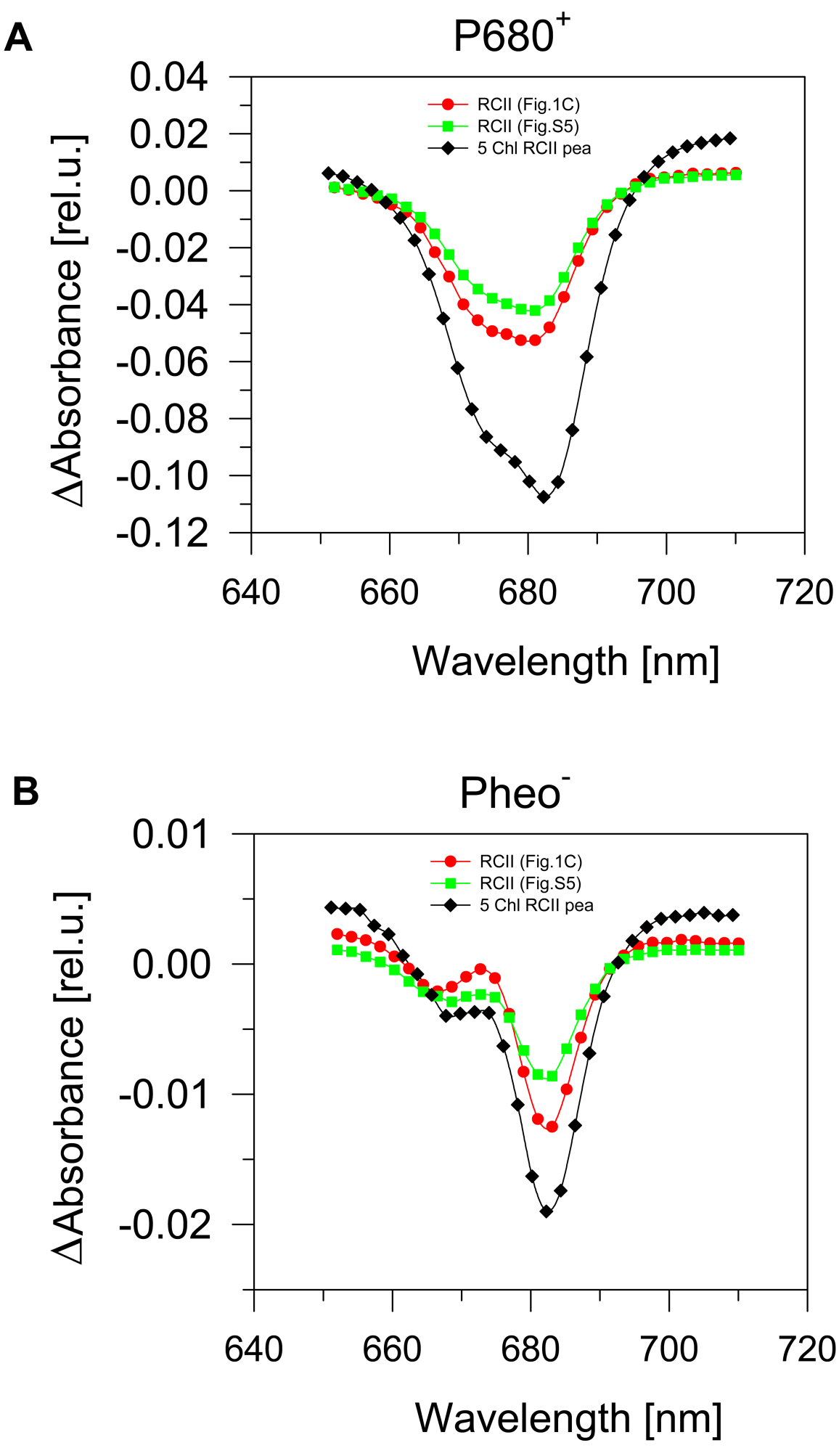

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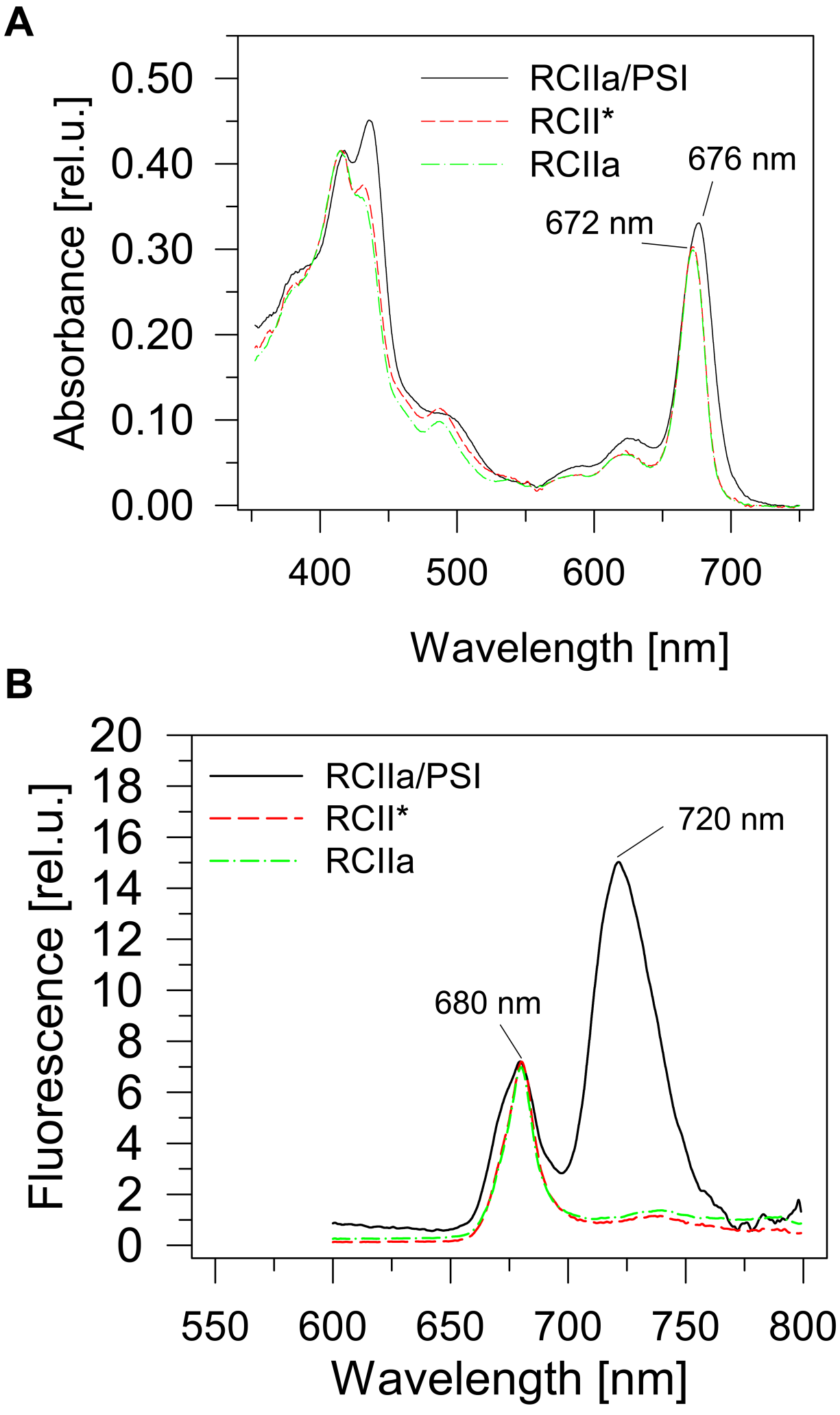

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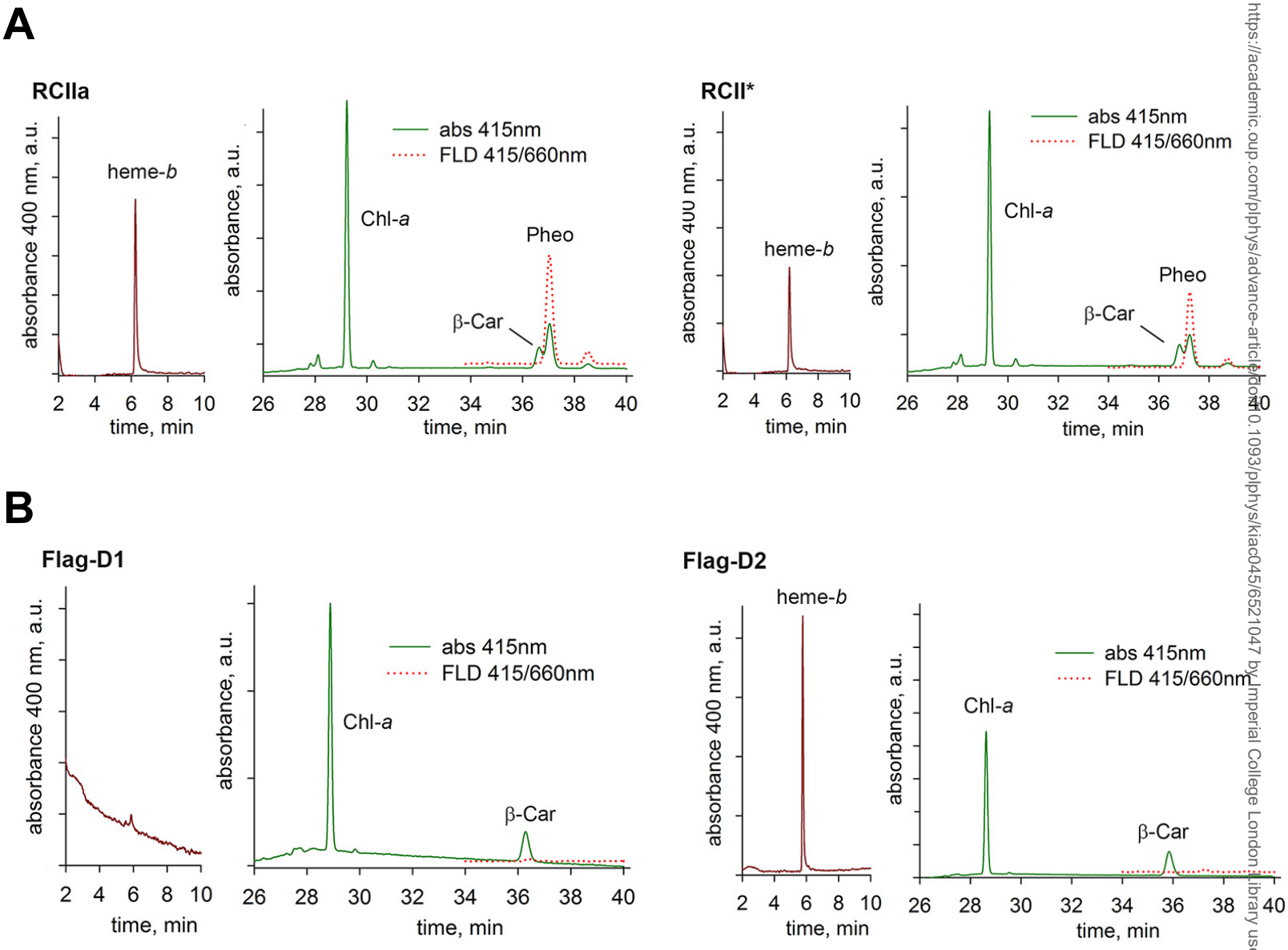

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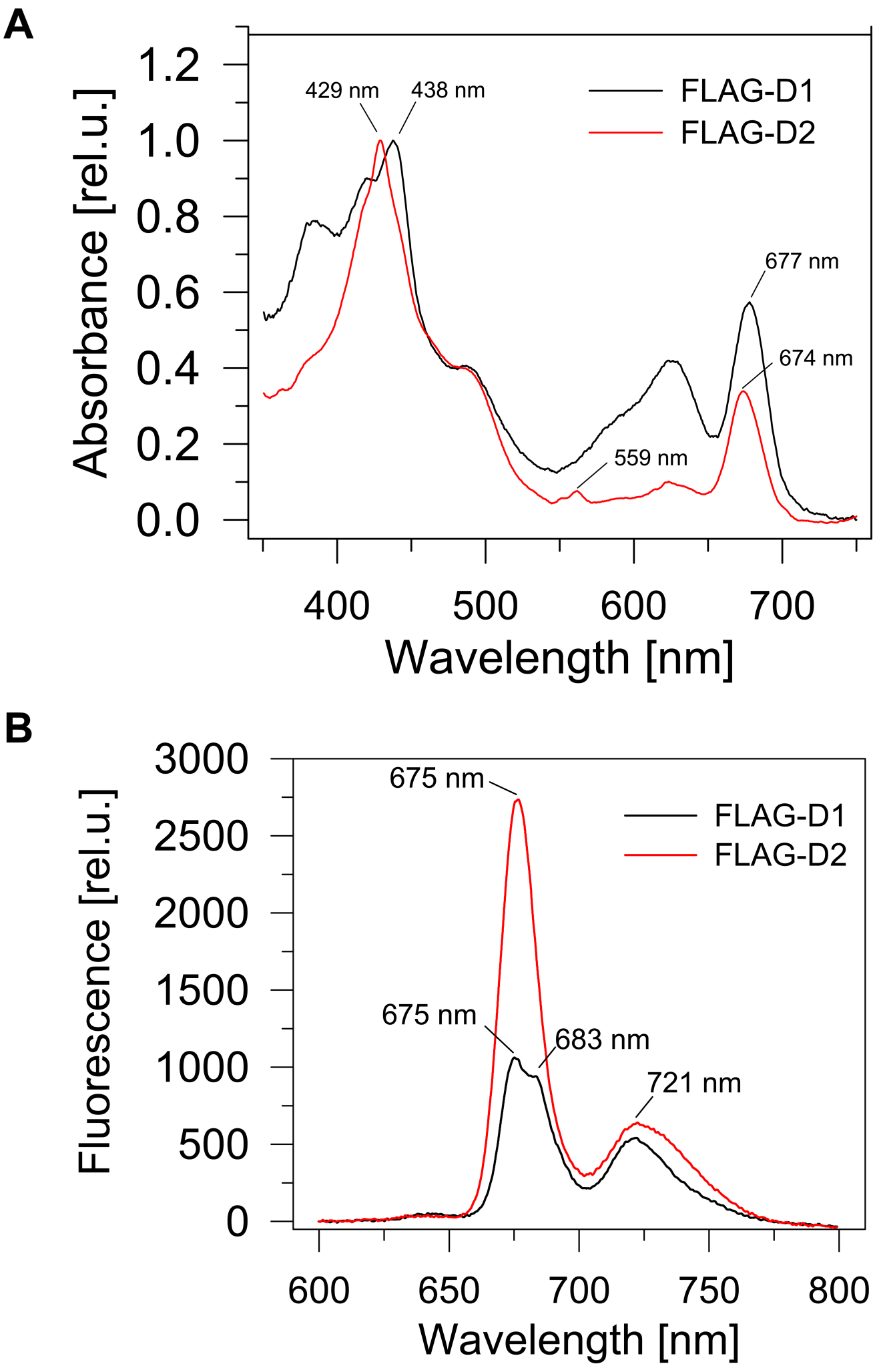

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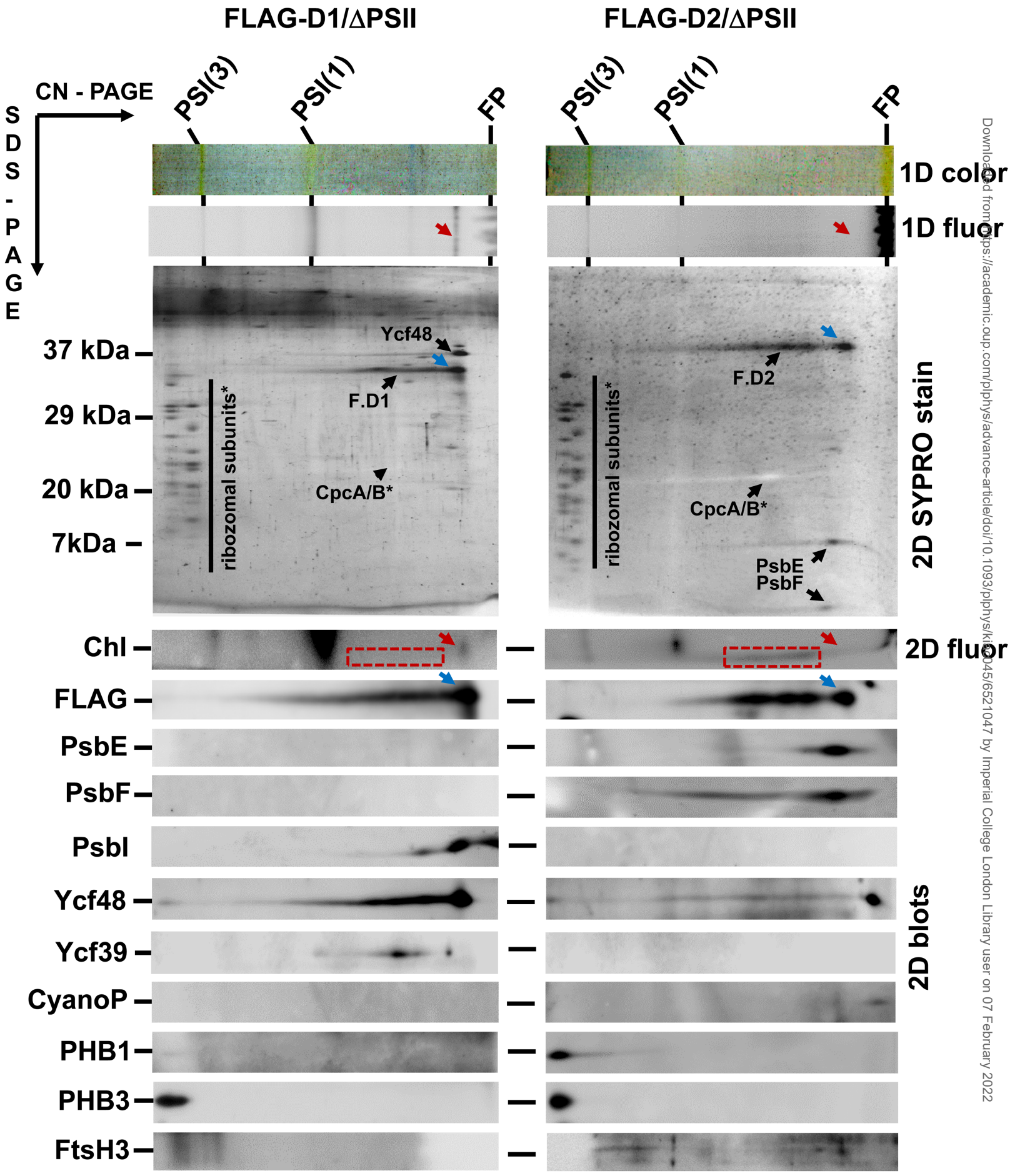

Figure 6.

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