

Mast cells: new therapeutic target in helminth immune modulation

K. V. VUKMAN,^{1,2} R. LALOR,² A. ALDRIDGE² & S. M. O'NEILL²

¹Department of Genetics, Cell- and Immunobiology, Semmelweis University, Nagyvarad ter 4., H-1089, Budapest, Hungry, ²Parasite Immune Modulation Group, School of Biotechnology, Faculty of Science and Health, Dublin City University, Glasnevin, Dublin, Ireland

SUMMARY

Helminth infection and their secreted antigens have a protective role in many immune-mediated inflammatory disorders such as inflammatory bowel disease, rheumatoid arthritis and multiple sclerosis. However, studies have focused primarily on identifying immune protective mechanisms of helminth infection and their secreted molecules on dendritic cells and macrophages. Given that mast cells have been shown to be implicated in the pathogenesis and progression of many inflammatory disorders, their role should also be examined and considered as cellular target for helminth-based therapies. As there is a dearth of studies examining the interaction of helminth-derived antigens and mast cells, this review will focus on the role of mast cells during helminth infection and examine our current understanding of the involvement of mast cells in $T_H 1/T_H 17$ -mediated immune disorders. In this context, potential mechanisms by which helminths could target the $T_H l/T_H l7$ promoting properties of mast cells can be identified to unveil novel therapeutic mast cell driven targets in combating these inflammatory disorders.

Keywords autoimmunity < disease, immune modulation < immunological terms, inflammation < disease, mast cell < cell, parasite

INTRODUCTION

Parasitic worms or helminths are highly successful metazoans that infect an estimated third of the world's population, causing chronic infections (1). While helminth infection is not associated with high mortality rates, high degrees of morbidity are associated with affected populations. The disability-adjusted life years (DALYs) of individuals is generally used as a measure of the burden of helminth infection (2) and a report in 2010 estimated that approximately 15 million DALYs were a direct result of helminth infection (3–5). Helminths have developed immunomodulatory strategies to evade host immune responses enabling them to persist within their host for prolonged periods of time. While studies have shown that the immunomodulatory effects of helminth infection can increase the susceptibility of a host to a number of secondary infections (6,7), they also have been demonstrated to have a protective role in many noncommunicable immune-mediated inflammatory disorders (8,9).

Helminth therapy is being explored as a viable treatment of $T_H 1/T_H 17$ -mediated inflammatory disorders such as multiple sclerosis (10) and inflammatory bowel diseases, including ulcerative colitis (11,12) and Crohn's disease (13). Human clinical trials to date have demonstrated that this therapy is both safe and effective (10,11,14–16). Furthermore, studies in mouse models such as experimental autoimmune encephalomyelitis (EAE) (17,18), type 1 diabetes (19), rheumatoid arthritis (20) and colitis (21) have suggested that helminth infection and the molecules it secretes are also protective in these disease models. Therefore, understanding the immune protective properties of helminths could offer potential therapeutic targets for a wide range of diseases.

Mastocytosis is a pathological infiltration of mast cells that is a common feature of helminth infection and a major component of protective immunity against helminth infection in the intestinal tract (22,23). Moreover, mast cells are associated with many $T_H 1/T_H 17$ immunemediated disorders including multiple sclerosis, inflammatory bowel disease and rheumatoid arthritis (24). While studies have established the impact of helminth infection on the ability of innate immune cells such as dendritic cell and macrophages to drive inflammatory responses (25),

Correspondence: Dr. Sandra O'Neill, Parasite Immune Modulation Group, Biotechnology, Faculty of Science and Health, Dublin City University, Glasnevin, Dublin 9, Ireland (e-mail: Sandra.ONeill@dcu.ie). Disclosure: None. Received: 7 August 2015 Accepted for publication: 2 November 2015

relatively few studies have examined the role of mast cells in this context. This review will examine the role of mast cells in helminth infection and inflammatory disorders with a view to highlight mast cells as an important cellular target in the development of helminth-derived therapies.

THE ROLE OF MAST CELLS IN HELMINTH INFECTION

Helminth infections are associated with increased mast cell numbers which are primarily redistributed to the site of infection (26). Activated mast cells secrete serine proteases, chymase and tryptase that have a direct cytotoxic effect on the helminth (27,28). In addition, mast cell-derived mouse mast cell protease 1 has been shown to loosen tight junction spaces in epithelial barrier, increases intestinal permeability resulting in increased luminal flow, leading to the expulsion of the parasite (29). However, the importance of mast cells in clearance is species specific (8). While the ablation of mast cells in murine models of Trichuris muris is not critical to its expulsion (30), there is strong evidence to support the involvement of mast cells in intestinal nematode infection (29). In vivo studies of Heligmosomoides polygyrus infection with mast cell-deficient Kit^{W} Kit^{W-v} mice show higher rates of nematode fecundity, compared to wild-type controls, highlighting their importance in intestinal helminth immunity (31). Enhanced mast cell number were associated with the clearance of Strongloides ratti, Trichinella spiralis, Nippostrongylus brasiliensis and Strongyloides ratti in rodent models, further implicating their importance (29, 32, 33).

The role of mast cells in tissue dwelling helminth infection, such as Schistosomais or Fasciolosis is not clearly understood, despite mast cells being implicated with both the acute and chronic stages of the infection (34). The early stages of Schistosoma infection are predominately skewed towards T_H1 immune responses (35). Given that mast cells are an important source of T_H1 inflammatory cytokines, their relative contributions to the development of T_H1 immune responses at this stage remains unclear. Similarly, mast cells are observed in the chronic stages of Schistosoma infection, when $T_H 2/T_{reg}$ immune responses are predominant (36), again implicating an important role for mast cells in shaping the adaptive immune response. Increased mast cell infiltration in the liver remains a key feature of Fasciola hepatica infection (37,38), and in rodent models, this is dominated by T_H2/T_{reg} responses within hours of infection.

Extensive migration is a common feature with tissue dwelling helminths, *F. hepatica* for example, migrates from the intestine to the peritoneal cavity, finally residing in the

bile ducts of the liver (39). This migration is correlated with increased mast cell infiltration in the gut mucosa, peritoneal cavity and liver (40–42). This increased mast cell population may be involved in promoting the balance between inflammation and wound healing as mast cells secrete mediators, such as histamine, serotonin, enzymes and cytokines that are important in inducing fibroblast proliferation; a marker for wound healing responses, while also increasing vascular permeability and recruiting neutrophils (43,44). In mast cell-deficient mice, wound closure is significantly impaired compared to normal or mast cell reconstructed mice (44).

During helminth infection, the activation of mast cells is mainly studied in the context of adaptive T_H2 immune responses. Protective immunity to helminths is thought to be mediated by the T_H2 subset of $CD4^+$ T cells; however, mast cells are observed in the early stages of infection suggesting that these cells may have an important role to play in shaping the T_H2 immune response. Cytokines, namely IL-4 and IL-13, secreted by T_H2 cells direct B-cells to produce helminth-specific IgE antibody (8,45). Mast cells express the high affinity FccRI receptor, which in the presence of helminth-specific antigens and IgE antibody induce mast cell degranulation, resulting in the release of inflammatory mediators such as histamine, cytokines and chemokines (46,47).

The release of these mediators by mast cells was shown to contribute to the development of T_{H2} immune responses towards gastrointestinal helminths by activating other cells involved in T_H2 immunity, (48). A recent study demonstrated the importance of mast cell crosstalk in the early stages of *H. polygyrus* infection. T_H2 immune responses and the clearance of the helminth was associated with mast cell released IL-25, IL-33 and thymic stromal lymphopoietins (TSLP) (49). Wild-type mice were characterized by high expression levels of T_H2 cytokines, namely IL-4, IL-5, IL-9, IL-10 and IL-13. However mast cell-deficient Kit^{W}/Kit^{W-v} mice showed a significantly impaired T_H2 response. Mast cell-derived IL-25, IL-33 and TSLP was shown to be crucial for driving T_{H2} cell priming through the activation of dendritic cells (48). Other studies have demonstrated that mast cells can indirectly modulate T-cell responses by crosstalking with dendritic cells, influencing their maturation (50). In contact hypersensitivity mouse models, activated dendritic cells bind to mast cells, promoting Ca²⁺ influx and the induction of tumour necrosis factor alpha (TNF- α) production. Activated mast cell-derived TNF-a was also shown to induce the in vivo migration of DCs (51).

Further work is required to define the phenotype of mast cells that promotes early $T_H 2$ immune responses in the absence of antigen-specific IgE. The definition of these

subsets could be based on analogy to dendritic cells and macrophages in helminth infection. Helminths alternatively activate innate immune cells, which display unique phenotypical and functional properties. Alternatively activated dendritic cells have been shown to display partial maturation (52,53) characterized by low expression of the co-stimulatory, major histocompatibility complex (MHC) molecules and a restricted cytokine and chemokine secretion profile compared to bacterial activated dendritic cells. Alternatively activated macrophages have also been shown to exhibit an M2-phenotype characterized by the expression of ARG1, YM1/2, RELM α genes and secretion of TGF- β , PGE2 and IL-10. The alternate activation of these cell populations contributes to the induction of T_H2 responses (54,55).

While there is strong evidence to support the influence mast cells can have in inducing $T_H l$, $T_H l7$ and $T_H 2$ immune responses (50), a subset of mast cells that drive T_{reg} immune responses has yet to be defined. However, considering mast cells secrete IL-10 and TGF- β , two cytokines important in the induction and maintenance of T_{reg} cells, further work is required to examine whether mast cells contribute to tolerogenic immune response (56,57).

POSITIVE BYSTANDER EFFECTS OF HELMINTHS UPON T_H1/T_H17 INFLAMMATORY DISORDERS

Numerous studies in helminth endemic regions reported a reduced risk of individuals developing T_H1/T_H17-mediated autoimmunity or inflammatory bowel disease (IBD) at the population level (58-60) and this led to the hypothesis that helminths conferred protection against inflammatory disorders. Leading on from these initial observations and overwhelming evidence from experimental models, helminth therapy is currently being used in phase I clinical trials as a novel approach for the treatment of a range of inflammatory disorders. Initial reports from these clinical trials have demonstrated that treatments are safe (10). While helminth therapy was shown as a good therapeutic candidate in individuals with multiple sclerosis; results were less promising for allergic rhinitis (61). There is also still debate to the effectiveness of helminth therapy on treating IBD. Despite positive results in initial studies using Trichuris suis ova to treat Ulcerative colitis or Crohn's disease (62), a more recent study in a large clinical trial of patients with Crohn's disease, showed no signs of improving disease activity index or remission rates and was subsequently stopped due to a lack of efficacy (63,64).

While human trials with worm therapy are a new development, there is overwhelming evidence in experi-

mental models that helminth infection and the products they release exert immune-suppressive effects that prevent the initiation and perpetuation of inflammatory disorders. Infection with *Schistosoma mansoni* reduces the incidence of autoimmune disease in mice by 50% (65) while gastrointestinal nematodes can suppress innate and adaptive pro-inflammatory immune responses, which are linked to the suppression of inflammation associated with IBD (66). *Hymenolepsis diminuta* was shown to have beneficial effect in a murine model of colitis while *T. spiralis* infection was observed to have a protective effect in the same model, reviewed elsewhere (60,67).

Similar to infection, helminth-derived excretory-secretory (ES) products and extracts have been shown to have a protective effect in the treatment of a range of inflammatory disorders such as murine arthritis, allergy and diabetes which have also been extensively reviewed elsewhere (68-71). The use of these parasitic antigens or synthetic analogues may allow for the development of specific and or more effective drugs to cure inflammatory disorders.

MAST CELLS, $T_H 1/T_H 17$ and inflammatory disorders

Similar to dendritic cells and macrophages, upon activation mast cells can elicit immune responses, by interacting with other cells through adhesion molecules, co-stimulatory/co-inhibitory molecules and the secretion of cytokines (72). MHC class I and II play central roles in antigen presentation. Mast cells express high levels of MHCI but very low levels of MHCII, although this can be up-regulated by LPS or IFN- γ stimulation, or during bacterial infection (73). Expression of ICAM-1, VCAM-1, OX40L, CD40L, LFA-1 and many other molecules by mast cells would suggest a broad ability to directly mediate T-cell activation, although the mechanism is yet unclear (74,75).

Mast cells indirectly activate the adaptive $T_{\rm H}1$ immune system by secreted cytokines, chemokines and also acting as antigen presenting cells. These mast cells are characterized by the lack of degranulation and the production of pro-inflammatory mediators, such as TNF- α . These cells have been shown to also contribute to pathology in inflammatory disorders (26,76). Mast cells are therefore thought to be a potential novel cellular target in the treatment of a range of $T_{\rm H}1/T_{\rm H}17$ immune-mediated diseases where mast cells contribute to pathology (26). Studies using mast cell knockout mice show a critical role for mast cells in many inflammatory disorders such as multiple sclerosis (MS), rheumatoid arthritis (RA) and inflammatory bowel disease (IBD) (77,78). In experimental autoimmune encephalitis (EAE), the murine model for MS, the release of pro-inflammatory mediators by mast cells was shown to contribute to the severity of disease (79), while others have implicated mast cells in the induction of the disease rather than as effector cells contributing to disease severity (80). Studies in mast cell-deficient mice showed mast cell-derived pro-inflammatory cytokines, such as TNF- α , are integral to the development of EAE (81,82).

In collagen-induced murine RA models, mast cells were shown to accumulate and de-granulate in the affected joints (83). Mast cell-deficient mice are resistant to anti-glucose-6-phosphate isomerize (GPI) antibody-induced RA, while wild type and mast cell reconstituted mice retain their sensitivity (84). The precise pathogenesis of RA is still unclear. $T_H 1/T_H 17$ cells are currently considered to be the key participants in the pathophysiology of this disease (85–87), and mast cells may have a critical role in skewing lymphocytes towards $T_H 1/T_H 17$ responses and in the development of these pathological processes (88–90).

Enhanced mast cell numbers have been observed in animal models of IBD at the site of inflammation, where mast cell associated inflammatory mediators are found in abundance and are positively linked with pathogenesis and disease progression. In these studies, mast cells were shown to undergo degranulation, release histamine and pro-inflammatory cytokines, such as IL-6 and TNF- α (91). Considering the longevity of these cells, mast cells could exert influence in IBD development and progression at multiple checkpoints. In contrast, studies based on IL-10-deficient mice that are highly susceptible to developing IBD, demonstrated that mast cells may have a protective role (92).

Studies have shown that mast cell inhibitors have therapeutic potential in the treatment of IBD (93). While there still remains conflicting data on the involvement and distribution of mast cells in the intestine of patients presenting with ulcerative colitis (94–96), it was demonstrated by Kurosawa and Nagai that ulcerative colitis patients were successfully treated with anti-allergic drugs which targeted mast cell activation and T_H2 -polarized immune responses (97). The role of mast cells is alluded to, by the elevation of UC severity in patients treated with drugs which directly target mast cell activation. Mast cells play a prominent role in inflammatory disorders, and yet there is a dearth of studies examining the potential of helminths



Figure 1 Potential mechanisms of action of Helminth-treated mast cells: evidence in the literature would suggest that helminths could have the potential modes of action on mast cells that would lead to the suppression of Th1/Th17 immune response.
I. Inhibiting mast cell proliferation (42).
Suppressing TLR-induced cytokine production (18,53).
Promoting regulatory cytokines (19,25,54).
Inhibiting TLR pathway (52,53).
Inducing Th2 promoting cytokines (18,19).
Inducing suppressive phenotypes (104).

to modulate these cells as targets for helminth-derived therapies. This highlights the need of further studies.

MAST CELLS AS THERAPEUTIC TARGETS FOR HELMINTH THERAPY

A strong rationale exists to analyse mast cells as cellular targets for helminth-based therapies. We have shown that helminth-derived molecules namely F. hepatica tegumental coat antigens (FhTeg) target mast cells inhibiting their ability to drive T_H1 immune responses. FhTeg suppress LPS-induced NF-KB and MAPK pathway (ERK) activation in mast cells (98). NF-KB and MAPKs are important signalling molecules leading to the expression of ICAM1 (99) and the secretion of pro-inflammatory cytokines. We demonstrated that the expression of ICAM1 was important in mast cell-T-cell communication, as inhibiting its expression in conjunction with the release of pro-inflammatory cytokines blocked the induction of T_H1 responses (98). This inhibition of $T_{\rm H}1$ immune responses is thought to be due in part by the expression of suppressor of cytokine signalling-3 (SOCS3) (a pathway not previously described in mast cells) (98), a negative regulator of $T_H 1/$ $T_{\rm H}17$ inflammatory processes (100). We also demonstrated that FhTeg does not induce mast cell proliferation while promoting migration of mast cells in vitro suggesting that the increase in mast cell numbers observed in the peritoneal cavity and liver of mice may be the result of mast cell migration and not proliferation (42).

Some other studies on *T. spiralis* have also demonstrated the immunomodulatory effects of helminths on mast cells. The *T. spiralis*-secreted molecule ES-62 was shown to block calcium mobilization and bind toll-like receptor 4, inhibiting downstream signalling of NF- κ B, antagonizing mast cell degranulation (47). *T. spiralis*-secreted molecules have also been shown to selectively modulate the secretion profiles of activated mast cells while *T. spiralis* muscle larval antigens were demonstrated to induce the release of histamine but inhibit β -hexosaminidase in mast cells (101,102). *T. spiralis*-secreted enzymes were also shown to inhibit mouse mast cell protease 1 (103).

REFERENCES

- McSorley HJ & Maizels RM. Helminth infections and host immune regulation. *Clin Microbiol Rev* 2012; 25: 585–608.
- 2 King CH. Health metrics for helminthic infections. Adv Parasitol 2010; 73: 51–69.
- 3 King CH. Health metrics for helminth infections. *Acta Trop* 2015; **141b**: 150–160.
- 4 Vos T, Flaxman AD, Naghavi M, et al. Years lived with disability (YLDs) for 1160

There is strong evidence in the literature demonstrating the mechanisms by which helminth infection and their secreted molecules target innate immune cells. These mechanisms are well described for macrophages and dendritic cells, many of which we have discussed are shared by mast cells. While there are relatively few studies, we can hypothesise the potential modulatory interaction (Figure 1). Given the prominent role mast cells play during inflammatory disorders and the interest in using parasitic helminths to treat these disorders, future studies should be focused on the mechanisms used by helminths to suppress mast cell responses for the potential discovery of novel therapies.

SUMMARY

In summary, mast cells are involved in multiple inflammatory and autoimmune disorders (26), where TNF- α secretion from these cells is critical to disease pathology (81,82). It is therefore possible that the protective effect of helminths against immune disorders may be the result of its molecules directly blocking the release of pro-inflammatory mediators from mast cells. These findings might lead to the development of a therapeutic inhibitors for pathogenic mast cell phenotypes.

ACKNOWLEDGEMENTS

All individuals listed as authors contributed equally to the manuscript. Research associated with this group is funded by Science Foundation Ireland (11/RFP.1/BIC/ 3109) and the Programme for Research in Third Level Institutions (PRTLI) Cycle 4. The PRTLI is co-funded through the European Regional Development Fund (ERDF), part of the European Union Structural Funds Programme 2007–2013. The first author is supported by OTKA PD 112085 and the Janos Bolyai Research Fellowship of the Hungarian Academy of Sciences. We also would like to acknowledge support by the COST Action BM1007 'Mast cells and basophils – targets for innovative therapies'.

- sequelae of 289 diseases and injuries 1990–2010: a systematic analysis for the global burden of disease study 2010. *Lancet* 2012; **380**: 2163–2196.
- 5 Murray CJL, Vos T, Lozano R, et al. Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990– 2010: a systematic analysis for the global burden of disease study 2010. Lancet 2012; 380: 2197–2223.
- 6 Resende Co T, Hirsch CS, Toossi Z, et al. Intestinal helminth co-infection has a negative impact on both anti-Mycobacterium tuberculosis immunity and clinical response to tuberculosis therapy. Clin Exp Immunol 2007; 147: 45– 52.
- 7 Edwards MJ, Buchatska O, Ashton M, et al. Reciprocal immunomodulation in a schistosome and hepatotropic virus coinfec-

tion model. J Immunol 2005; 175:6275-6285.

- 8 Anthony RM, Rutitzky LI, Urban JF, et al. Protective immune mechanisms in helminth infection. Nat Rev Immunol 2007; 7: 975–987.
- 9 Helmby H. Human helminth therapy to treat inflammatory disorders where do we stand? *BMC Immunol* 2015; **16**: 12.
- 10 Fleming JO, Isaak A, Lee JE, et al. Probiotic helminth administration in relapsingremitting multiple sclerosis: a phase 1 study. *Mult Scler* 2011; 17: 743–754.
- 11 Summers RW, Elliott DE, Urban JF Jr, et al. Trichuris suis therapy for active ulcerative colitis: a randomized controlled trial. *Gastroenterology* 2005; **128**: 825–832.
- 12 Broadhurst MJ, Leung JM, Kashyap V, et al. IL-22+ CD4+ T cells are associated with therapeutic trichuris trichiura infection in an ulcerative colitis patient. Sci Transl Med 2010; 2: 60ra88.
- 13 Summers RW, Elliott DE, Urban JF, et al. Trichuris suis therapy in Crohn's disease. Gut 2004; 54: 87–90.
- 14 Daveson AJ, Jones DM, Gaze S, et al. Effect of hookworm infection on wheat challenge in celiac disease – a randomised double-blinded placebo controlled trial. PLoS One 2011; 6: e17366.
- 15 Rosche B, Wernecke K, Ohlraun S, et al. Trichuris suis ova in relapsing-remitting multiple sclerosis and clinically isolated syndrome (TRIOMS): study protocol for a randomized controlled trial. Trials 2013; 14: 112.
- 16 Fleming J, Hartman L, Maksimovic M, et al. Clinical trial of helminth-induced immunomodulatory therapy (HINT 2) in relapsing-remitting multiple sclerosis. *Neurology* 2014; 82: P3.149.
- 17 La Flamme AC, Ruddenklau K & Bäckström BT. Schistosomiasis decreases central nervous system inflammation and alters the progression of experimental autoimmune encephalomyelitis. *Infect Immun* 2003; 71: 4996–5004.
- 18 Kuijk LM, Klaver EJ, Kooij G, et al. Soluble helminth products suppress clinical signs in murine experimental autoimmune encephalomyelitis and differentially modulate human dendritic cell activation. *Mol Immunol* 2012; **51**: 210–218.
- 19 Liu Q, Sundar K, Mishra PK, et al. Helminth infection can reduce insulitis and type 1 diabetes through CD25- and IL-10independent mechanisms. *Infect Immun* 2009; 77: 5347–5358.
- 20 Pineda MA, McGrath MA, Smith PC, et al. The parasitic helminth product ES-62 suppresses pathogenesis in collagen-induced arthritis by targeting the interleukin-17-producing cellular network at multiple sites. *Arthritis Rheum* 2012; 64: 3168–3178.
- 21 Smith P, Mangan NE, Walsh CM, et al. Infection with a helminth parasite prevents experimental colitis via a macrophagemediated mechanism. J Immunol 2007; 178: 4557–4566.

- 22 Behnke JM, Wahid FN, Grencis RK, et al. Immunological relationships during primary infection with *Heligmosomoides polygyrus* (*Nematospiroides dubius*): downregulation of specific cytokine secretion (IL-9 and IL-10) correlates with poor mastocytosis and chronic survival of adult worms. *Parasite Immunol* 1993; 15: 415–421.
- 23 Helmby H & Grencis RK. IL-18 regulates intestinal mastocytosis and Th2 cytokine production independently of IFN-gamma during *Trichinella spiralis* infection. J Immunol 2002; 169: 2553–2560.
- 24 Xu Y & Chen G. Mast cell and autoimmune diseases. *Mediators Inflamm* 2015; 2015: 246126.
- 25 Walsh KP, Brady MT, Finlay CM, et al. Infection with a helminth parasite attenuates autoimmunity through TGF-betamediated suppression of Th17 and Th1 responses. J Immunol 2009; 183: 1577–1586.
- 26 Weller CL, Collington SJ, Williams T & Lamb JR. Mast cells in health and disease. *Clin Sci (Lond)* 2011; **120**: 473–484.
- 27 Metz M & Maurer M. Mast cells-key effector cells in immune responses. *Trends Immunol* 2007; 28: 234–241.
- 28 McKean PG & Pritchard DI. The action of a mast cell protease on the cuticular collagens of *Necator americanus*. *Parasite Immunol* 1989; 11: 293–297.
- 29 McDermott JR, Bartram RE, Knight PA, et al. Mast cells disrupt epithelial barrier function during enteric nematode infection. *Proc Natl Acad Sci USA* 2003; 100: 7761–7766.
- 30 Betts CJ & Else KJ. Mast cells, eosinophils and antibody-mediated cellular cytotoxicity are not critical in resistance to *Trichuris muris*. *Parasite Immunol* 1999; 21: 45–52.
- 31 Hashimoto K, Uchikawa R, Tegoshi T, et al. Immunity-mediated regulation of fecundity in the nematode *Heligmosomoides* polygyrus-the potential role of mast cells. *Parasitology* 2009; **137**: 881–887.
- 32 Abe T & Nawa Y. Worm expulsion and mucosal mast cell response induced by repetitive IL-3 administration in *Strongyloides ratti-*infected nude mice. *Immunology* 1988; 63: 181–185.
- 33 Marshall JS. Mast-cell responses to pathogens. Nat Rev Immunol 2004; 4: 787– 799.
- 34 Gerken SE, Mota-Santos TA & Vaz NM. Evidence for the participation of mast cells in the innate resistance of mice to Schistosoma mansoni: effects on in vivo treatment with the ionophore 48-80. Braz J Med Biol Res 1990; 23: 559–565.
- 35 de Jesus AR, Silva A, Santana LB, et al. Clinical and immunologic evaluation of 31 patients with acute schistosomiasis mansoni. J Infect Dis 2002; 185: 98–105.
- 36 Caldas IR, Campi-Azevedo AC, Oliveira LFA, et al. Human schistosomiasis mansoni: immune responses during acute and chronic phases of the infection. Acta Trop 2008; 108: 109–117.

Parasite Immunology

- 37 Rahko T. The pathology of natural *Fasciola hepatica* infection in cattle. *Pathol Vet* 1969;
 6: 244–256.
- 38 Ferreras MC, Garcia-Iglesias MJ, Manga-Gonzalez MY, et al. Histopathological and immunohistochemical study of lambs experimentally infected with Fasciola hepatica and Schistosoma bovis. J Vet Med B Infect Dis Vet Public Health 2000; 47: 763–773.
- 39 Dawes B & Hughes DL. Fascioliasis: the invasive stages in mammals. *Adv Parasitol* 1970; 8: 259–274.
- 40 Burden DJ, Bland AP, Hammet NC & Hughes DL. *Fasciola hepatica*: migration of newly excysted juveniles in resistant rats. *Exp Parasitol* 1983; 56: 277–288.
- 41 van Milligen FJ, Cornelissen JB, Hendriks IM, et al. Protection of Fasciola hepatica in the gut mucosa of immune rats is associated with infiltrates of eosinophils, IgG1 and IgG2a antibodies around the parasites. Parasite Immunol 1998; 20: 285–292.
- 42 Vukman KV, Adams PN, Dowling D, *et al.* The effects of *Fasciola hepatica* tegumental antigens on mast cell function. *Int J Parasitol* 2013; **43**: 531–539.
- 43 Maurer M, Theoharides T, Granstein RD, et al. What is the physiological function of mast cells? Exp Dermatol 2003; 12: 886–910.
- 44 Weller K, Foitzik K, Paus R, *et al.* Mast cells are required for normal healing of skin wounds in mice. *FASEB J* 2006; **20**: 2366–2368.
- 45 Erb KJ. Helminths, allergic disorders and IgE-mediated immune responses: where do we stand? *Eur J Immunol* 2007; 37: 1170–1173.
- 46 Melendez AJ, Harnett MM, Pushparaj PN, et al. Inhibition of Fc epsilon RI-mediated mast cell responses by ES-62, a product of parasitic filarial nematodes. *Nat Med* 2007; 13: 1375–1381.
- 47 Pearce EJ. Worms tame mast cells. Nat Med 2007; 13: 1288–1289.
- 48 Hepworth MR, Maurer M & Hartmann S. Regulation of type 2 immunity to helminths by mast cells. *Gut Microbes* 2012b; 3: 476–481.
- 49 Hepworth MR, Danilowicz-Luebert E, Rausch S, et al. Mast cells orchestrate type 2 immunity to helminths through regulation of tissue-derived cytokines. *Proc Natl Acad Sci USA* 2012a; **109**: 6644–6649.
- 50 Dudeck A, Suender CA, Kostka SL, et al. Mast cells promote Th1 and Th17 responses by modulating dendritic cell maturation and function. Eur J Immunol 2011; 41: 1883–1893.
- 51 Otsuka A, Kubo M, Honda T, et al. Requirement of interaction between mast cells and skin dendritic cells to establish contact hypersensitivity. PLoS One 2011; 6: e25538.
- 52 Kane CM, Cervi L, Sun J, et al. Helminth antigens modulate TLR-initiated dendritic cell activation. J Immunol 2004; 173: 7454–7461.

- 53 Hamilton CM, Dowling DJ, Loscher CE, et al. The Fasciola hepatica tegumental antigen suppresses dendritic cell maturation and function. Infect Immun 2009; 77: 2488–2498.
- 54 Balic A, Harcus Y, Holland MJ & Maizels RM. Selective maturation of dendritic cells by *Nippostrongylus brasiliensis*-secreted proteins drives Th2 immune responses. *Eur J Immunol* 2004; 34: 3047–3059.
- 55 Cook PC, Jones LH, Jenkins SJ, et al. Alternatively activated dendritic cells regulate CD4⁺ T-cell polarization in vitro and in vivo. Proc Natl Acad Sci U S A 2012; 109: 9977–9982.
- 56 Levings MK, Bacchetta R, Schulz U & Roncarolo MG. The role of IL-10 and TGF-beta in the differentiation and effector function of T regulatory cells. *Int Arch Allergy Immunol* 2002; **129**: 263–276.
- 57 Kosiewicz M & Alard P. Tolerogenic antigen-presenting cells. *Immunol Res* 2004; 30: 155–170.
- 58 Carvalho EM, Bastos LS & Araujo MI. Worms and allergy. *Parasite Immunol* 2006; 28: 525–534.
- 59 MacDonald AS, Araujo MI & Pearce EJ. Immunology of parasitic helminth infections. *Infect Immun* 2002; 70: 427–433.
- 60 Reddy A & Fried B. An update on the use of helminths to treat Crohn's and other autoimmunune diseases. *Parasitol Res* 2009; 104: 217–221.
- 61 Croft AM, Bager P & Kumar S. Helminth therapy (worms) for allergic rhinitis. *Cochrane Database Syst Rev* 2012; 4: CD009238.
- 62 Summers RW, Elliott DE, Qadir K, et al. Trichuris suis seems to be safe and possibly effective in the treatment of inflammatory bowel disease. Am J Gastroenterol 2003; 98: 2034–2041.
- 63 Coronado Biosciences announces top-line results from its TRUST-1 phase 2 clinical trial of TSO for the treatment of Crohr's disease. [http://ir.coronadobiosciences.com/ Cache/1500053219.PDF?Y=&O=PDF&D= &FID=1500053219&T=&IID=4308955]
- 64 Coronado biosciences announces indpendent data monitoring committee recommendation to discontinue falk phase 2 trials of TSO in Chrohns disease. [http://ir.coronadobiosciences.com/Cache/1500053915.PDF? Y=&O=PDF&D=&FID=1500053915&-T=&IID=4308955]
- 65 Wilson MS & Maizels RM. Regulation of allergy and autoimmunity in helminth infection. *Clin Rev Allergy Immunol* 2004; 26: 35–50.
- 66 Whelan RA, Hartmann S & Rausch S. Nematode modulation of inflammatory bowel disease. *Protoplasma* 2011; 249: 871– 886.
- 67 Ruyssers NE, De Winter BY, De Man JG, et al. Worms and the treatment of inflammatory bowel disease: are molecules the answer? *Clin Dev Immunol* 2008; 2008: 567314.
- 68 McInnes IB, Leung BP, Harnett M, et al. A novel therapeutic approach targeting

articular inflammation using the filarial nematode-derived phosphorylcholine-containing glycoprotein ES-62. *J Immunol* 2003; **171**: 2127–2133.

- 69 Harnett W & Harnett MM. Therapeutic immunomodulators from nematode parasites. *Expert Rev Mol Med* 2008; 10: e18.
- 70 Imai S, Tezuka H & Fujita K. A factor of inducing IgE from a filarial parasite prevents insulin-dependent diabetes mellitus in nonobese diabetic mice. *Biochem Biophys Res Commun* 2001; 286: 1051–1058.
- 71 Lund ME, O'Brien BA, Hutchinson AT, et al. Secreted proteins from the helminth Fasciola hepatica inhibit the initiation of autoreactive T cell responses and prevent diabetes in the NOD mouse. PLoS One 2014; 9: e86289.
- 72 Reber LL, Sibilano R, Mukai K & Galli SJ. Potential effector and immunoregulatory functions of mast cells in mucosal immunity. *Mucosal Immunol* 2015; 8: 444–463.
- 73 Kambayashi T, Allenspach EJ, Chang JT, et al. Inducible MHC class II expression by mast cells supports effector and regulatory T cell activation. J Immunol 2009; 182: 4686–4695.
- 74 Kashiwakura J, Yokoi H, Saito H & Okayama Y. T cell proliferation by direct crosstalk between OX40 ligand on human mast cells and OX40 on human T cells: comparison of gene expression profiles between human tonsillar and lung-cultured mast cells. J Immunol 2004; 173: 5247–5257.
- 75 Sayed BA & Brown MA. Mast cells as modulators of T-cell responses. *Immunol Rev* 2007; 217: 53–64.
- 76 Sayed BA, Christy AL, Walker ME & Brown MA. Meningeal mast cells affect early T cell central nervous system infiltration and blood-brain barrier integrity through TNF: a role for neutrophil recruitment? J Immunol 2010; 184: 6891–6900.
- 77 Secor VH, Secor WE, Gutekunst CA & Brown MA. Mast cells are essential for early onset and severe disease in a murine model of multiple sclerosis. J Exp Med 2000; 191: 813–822.
- 78 Lee DM, Friend DS, Gurish MF, et al. Mast cells: a cellular link between autoantibodies and inflammatory arthritis. *Science* 2002; **297**: 1689–1692.
- 79 Brenner T, Soffer D, Shalit M & Levi-Schaffer F. Mast cells in experimental allergic encephalomyelitis: characterization, distribution in the CNS and in vitro activation by myelin basic protein and neuropeptides. *J Neurol Sci* 1994; **122**: 210–213.
- 80 Levi-Schaffer F, Riesel N, Soffer D, et al. Mast cell activity in experimental allergic encephalomyelitis. *Mol Chem Neuropathol* 1991; **15**: 173–184.
- 81 Christy AL, Walker ME, Hessner MJ & Brown MA. Mast cell activation and neutrophil recruitment promotes early and robust inflammation in the meninges in EAE. J Autoimmun 2012; 42: 50–61.
- 82 Costanza M, Colombo MP & Pedotti R. Mast cells in the pathogenesis of multiple

sclerosis and experimental autoimmune encephalomyelitis. *Int J Mol Sci* 2012; **13**: 15107–15125.

- 83 Malfait AM, Malik AS, Marinova-Mutafchieva L, et al. The beta2-adrenergic agonist salbutamol is a potent suppressor of established collagen-induced arthritis: mechanisms of action. J Immunol 1999; 162: 6278–6283.
- 84 Benoist C & Mathis D. Mast cells in autoimmune disease. *Nature* 2002; 420: 875–878.
- 85 Omoyinmi E, Hamaoui R, Pesenacker A, et al. Th1 and Th17 cell subpopulations are enriched in the peripheral blood of patients with systemic juvenile idiopathic arthritis. *Rheumatology (Oxford)* 2012; 51: 1881–1886.
- 86 Rodeghero R, Cao Y, Olalekan SA, et al. Location of CD4+ T cell priming regulates the differentiation of Th1 and Th17 cells and their contribution to arthritis. J Immunol 2013; 190: 5423–5435.
- 87 Simons DM, Oh S, Kropf E, et al. Autoreactive Th1 cells activate monocytes to support regional Th17 responses in inflammatoryarthritis. J Immunol 2013; 190: 3134–3141.
- 88 Nigrovic PA & Lee DM. Mast cells in inflammatory arthritis. *Arthritis Res Ther* 2005; 7: 1–11.
- 89 Anand P, Singh B, Jaggi AS & Singh N. Mast cells: an expanding pathophysiological role from allergy to other disorders. *Naunyn Schmiedebergs Arch Pharmacol* 2012; 385: 657–670.
- 90 Hueber AJ, Asquith DL, Miller AM, et al. Mast cells express IL-17A in rheumatoid arthritis synovium. J Immunol 2010; 184: 3336–3340.
- 91 de Winter BY, van den Wijngaard RM & de Jonge WJ. Intestinal mast cells in gut inflammation and motility disturbances. *Biochim Biophys Acta* 2011; 1822: 66–73.
- 92 Chichlowski M, Westwood GS, Abraham SN & Hale LP. Role of mast cells in inflammatory bowel disease and inflammation-associated colorectal neoplasia in IL-10-deficient mice. *PLoS One* 2010; 5: e12220.
- 93 He SH. Key role of mast cells and their major secretory products in inflammatory bowel disease. World J Gastroenterol 2004; 10: 309–318.
- 94 King T, Biddle W, Bhatia P, et al. Colonic mucosal mast cell distribution at line of demarcation of active ulcerative colitis. *Dig Dis Sci* 1992; 37: 490–495.
- 95 Bischoff SC, Wedemeyer J, Herrmann A, et al. Quantitative assessment of intestinal eosinophils and mast cells in inflammatory bowel disease. *Histopathology* 1996; 28: 1–13.
- 96 Lloyd G, Green FH, Fox H, et al. Mast cells and immunoglobulin E in inflammatory bowel disease. Gut 1975; 16: 861–865.
- 97 Kurosawa M & Nagai H. Accumulation of mast cells in the lesions and effects of antiallergic drugs on the patients with

© 2015 John Wiley & Sons Ltd, Parasite Immunology, 38, 45-52

inflammatory bowel disease. Ulcers 2013; 2013: e714807.

- 98 Vukman KV, Adams PN, Metz M, et al. Fasciola hepatica tegumental coat impairs mast cells' ability to drive Th1 immune responses. J Immunol 2013; 190: 2873–2879.
- 99 Tsang CM, Wong CK, Ip WK & Lam CW. Synergistic effect of SCF and TNF-alpha on the up-regulation of cell-surface expression of ICAM-1 on human leukemic mast cell line (HMC)-1 cells. *J Leukoc Biol* 2005; 78: 239–247.
- 100 Carow B & Rottenberg ME. SOCS3, a major regulator of infection and inflammation. *Front Immunol* 2014; 5: 58.
- 101 Yépez-Mulia L, Montaño-Escalona C, Fonseca-Liñán R, et al. Differential activation of mast cells by antigens from *Trichinella* spiralis muscle larvae, adults, and newborn larvae. Vet Parasitol 2009; 159: 253–257.
- 102 Arizmendi-Puga NG, Enciso JA, Ortega-Pierres G, et al. Trichinella spiralis: histamine secretion induced by TSL-1 antigens from unsensitized mast cells. Exp Parasitol 2006; 114: 67–76.
- 103 Afferson HC, Eleftheriou E, Selkirk ME & Gounaris K. *Trichinella spiralis* secreted enzymes regulate nucleotide-induced mast cell activation and release of mouse mast cell protease 1. *Infect Immun* 2012; 80: 3761–3767.
- 104 Dowling DJ, Hamilton CM, Donnelly S, et al. Major secretory antigens of the helminth fasciola hepatica activate a suppressive dendritic cell phenotype that attenuates Th17 cells but fails to activate Th2 immune responses. *Infect Immun* 2010; **78**: 793–801.