Opinion Piece

Hypothesis: Entrapment of lipoprotein particles in the brain causes Alzheimer's disease

- Delphine Boche¹, James AR Nicoll^{1,2}
- ¹ Clinical Neurosciences, Clinical and Experimental Science, Faculty of Medicine, University of Southampton, Southampton, United Kingdom
- ² Department of Cellular Pathology, University Hospital Southampton NHS Foundation Trust, Southampton, United Kingdom

Corresponding author:

 $\label{eq:constraint} \begin{array}{l} \text{Delphine Boche} \cdot \text{Clinical Neurosciences, Clinical and Experimental Sciences Academic Unit } \text{Faculty of Medicine} \cdot \text{University of Southampton} \cdot \text{Southampton General Hospital} \cdot \text{Mailpoint 806} \cdot \text{Southampton SO16 6YD} \cdot \text{United Kingdom} \\ \hline \\ \underline{\text{d.boche}} \\ \underline{\text{soton.ac.uk}} \end{array}$

Submitted: 05 August 2021	•	Accepted: 21 October 2021	•	Copyedited by: Cinthya Agüero	 Published: 02 November 2021
---------------------------	---	---------------------------	---	-------------------------------	---

Abstract

We present for consideration a hypothesis that impaired movement of lipoprotein particles in the extracellular space in the brain in ageing is central to and causes all the key pathophysiological features of Alzheimer's disease (AD). The role of lipoprotein particles is to transport cholesterol from glial cells, where it is synthesised, to neurons, which require cholesterol for synaptic plasticity. The lipoprotein particles have a cholesterol-containing hydrophobic core, in which amyloid- β (A β) can be solubilised. The core is surrounded by a hydrophilic surface containing apolipoprotein E (APOE) which, as neurons bear receptors for APOE, determines the destination of the particles. The problem arises because the extracellular space is a narrow cleft, barely wider than the lipoprotein particles themselves, which they have to navigate in order to perform their crucial cholesterol-transporting function. We explain how lipoprotein particles could become trapped in the ageing extracellular matrix and that this primary abnormality results in reduced delivery of cholesterol to neurons leading to impaired synaptic plasticity, crucial for learning and memory. It can also explain extracellular Aβ accumulation, to which a microglial response generates a neurotoxic reaction, and intraneuronal tau aggregation, each of which exacerbate the problem. All these players have been known for many years to be important in Alzheimer's pathogenesis but a single unifying mechanism to explain how they are linked has been lacking. This proposed mechanism, with entrapment of lipoproteins particles as key to the development of AD, can explain the failure of so many clinical trials and points out new directions to be taken.

Keywords: Lipoprotein particles, Extracellular matrix, Cholesterol transport, Apolipoprotein E, Alzheimer's disease



Copyright: © 2021 The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International License (<u>https://creativecommons.org/licenses/by/4.0/</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited, a link to the Creative Commons license is provided, and any changes are indicated. The Creative Commons Public Domain Dedication waiver (<u>https://creativecommons.org/publicdomain/zero/r.o/</u>) applies to the data made available in this article, unless otherwise stated.

Genetics shows the overriding importance of apolipoprotein E

In a complex multifaceted disease process, such as Alzheimer's disease (AD), genetic factors are important in highlighting key elements relevant to the onset of the disease. By far the major genetic risk factor for AD, in terms of scale of effect, is the apolipoprotein E genotype (gene: APOE, protein: APOE). There are 3 common APOE gene alleles ($\epsilon 2$, ϵ 3 and ϵ 4); as each person inherits one allele from either parent there is a total of 6 APOE genotypes $(\epsilon 2/\epsilon 2, \epsilon 2/\epsilon 3, \epsilon 3/\epsilon 3, \epsilon 3/\epsilon 4 and \epsilon 4/\epsilon 4)$. The APOE $\epsilon 4$ allele carriage rate (i.e. the proportion of people in a population who possess one or two ε4 alleles) is typically about 25% of people of European ancestry [1], but varies considerably around the world among different populations [2]. A single copy of the APOE ε4 allele confers a two to fourfold increased risk of developing AD, with ε 4 homozygotes at fourteen times increased risk of developing AD, whereas the less common ɛ2 allele confers relative protection, approximately halving the risk [3]. The effect is so substantial that a person who is £4 homozygous and lives to 85 years of age has a lifetime risk of AD of more than 50%, comparable to the risk associated with BRCA1 mutation in breast cancer [4]. On the other hand, the protective effect of APOE ϵ 2 is so substantial that ϵ^2 homozygotes have a very low likelihood of developing AD; 2 homozygotes have a 87% lower odds ratio than ɛ3 homozygotes and a 99.6% lower odds ratio than ε 4 homozygotes [5]. The APOE gene polymorphism is lacking from nonhuman primates [6, 7] although, interestingly, Rhesus monkeys which have an ɛ4-like APOE sequence develop A β plaques as they age [8]. The fact that there are more than 200 animal models of AD highlights the difficulty in mimicking the complexity of the human disease in experimental animals. A complete understanding of the pathophysiology of AD ideally would explain all aspects of the human disease, and in particular incorporate the role of the APOE protein; how it causes and interacts with the accumulation of amyloid- β (A β) and tau proteins, the glial cell activity and most importantly, the neuronal and synaptic dysfunction and loss.

The function of APOE is to deliver cholesterol, packaged in lipoprotein particles, to neurons

APOE is the principal cholesterol carrier in the brain, acting as a detergent with hydrophobic and hydrophilic moieties. Its main role is to solubilise cholesterol and other lipids and lipid-soluble substances to enable them to be transported in the aqueous extracellular environment of the brain. The importance of cholesterol to the brain is highlighted by the fact that 25% of the body's cholesterol is contained within the brain, despite the brain representing only 2% of the body weight [9]. Cholesterol forms about 30% of the lipid bilayer of the membrane of all cells and is important in maintaining membrane fluidity. In the brain, the cell membrane is essential for conducting the action potential and for communication between neurons at synapses. However, despite its importance to neuronal function, neurons do not synthesise their own cholesterol, but rely on cholesterol which is synthesised by glial cells and then transported to neurons [10]. Delivery of cholesterol to neurons is in particular demand when the neuron changes, as in synaptic plasticity which underpins learning and memory which are particularly affected in AD [11]. Of note, genome-wide association studies have highlighted polymorphisms in genes in addition to APOE that are involved in cholesterol handling (CLU, PICALM, BIN1 and ABCA7) as risk factors for AD [12, 13], supporting an important role for cholesterol handling in the disease mechanism [14, 15].

APOE is located in the shell of lipoprotein particles, with hydrophobic substances including cholesterol, which is loaded onto the lipoprotein particle by the enzyme ABCA1, being transported in the core [16]. The destination of the lipoprotein particles is determined by the receptors for the proteins on the surface of the lipoprotein shell. APOE, on the outer surface of the lipoprotein particles, binds to receptors of the low-density lipoprotein (LDL) receptor family (principally LRP1/APOE receptor) present on neuronal cell membranes and, by this mechanism, the cholesterol is internalised within the neurons. Some CNS lipoprotein particles also bear APOJ (clusterin) on their surface; ependymal cells, but not neurons or other glia, bear APOJ receptors so it is unlikely to be relevant for cholesterol delivery to neurons [17]. However, it is notable that in genome-wide association studies, polymorphism of APOJ/clusterin gene has also been shown to influence risk for AD [18], raising the possible importance of clearance of lipoprotein particles and cholesterol to the CSF.

Outside the brain, in the rest of the body, there are several other lipoproteins that can fulfil the function of cholesterol transport in addition to APOE, including APOA which is synthesised in the liver [19]. APOA-containing lipoprotein particles are detectable in the CSF but appear to be excluded from the brain parenchyma by the blood-brain barrier. Consequently, the cholesterol required by neurons is synthesised within the brain and delivered to neurons by APOE-containing lipoprotein particles [20] [9, 21].

Lipoprotein particles become entrapped in the ageing extracellular matrix

The lipoprotein particles, which resemble the high-density lipoproteins (HDL) present in the bloodstream, are in the region of 11-20 nm in diameter [22]. In order to transport cholesterol to neurons, they have to travel in the extracellular space in the brain which itself measures only about 40 nm between adjacent cells [23], below the resolution of light microscopy. Small molecules can pass readily, by diffusion and potentially by active flow, through the extracellular space but diffusion even of conventional macromolecules much smaller that lipoprotein particles is hindered, particularly in pathological processes such as gliosis (activation of glial cells) [24], which occurs in association with ageing and the neurodegeneration in AD.

An additional complexity is that the extracellular space resembles a sponge, containing the extracellular matrix (ECM). In the brain, the ECM is a complex multimolecular three-dimensional structure consisting of proteoglycans/glycosaminoglycans, proteins, proteinases, and cytokines [25]. Expression of collagen IV, laminin and fibronectin is upregulated in the cerebral cortex in early AD. With ageing-related changes to the extracellular matrix, exacerbated by other risk factors for AD (hypertension, diabetes, obesity, inflammation and physical inactivity), the narrow extracellular space likely becomes compromised ('fibrosed'), impeding the movement of, and trapping, lipoprotein particles in between cells.

Interestingly, oxidative modification of plasma lipoproteins is one of the earliest steps in the development of atherosclerosis, involving accumulation of lipoprotein particles in the vessel wall provoking inflammatory reaction [26]. In AD patients, there is evidence of greater oxidation of plasma and CSF lipoproteins [26] and this could conceivably further impair the passage of lipoprotein particles through the extracellular space in the brain. Consequently, the APOE-mediated system for transporting cholesterol and lipids from glial cells to neurons in the brain is unique, crucial for neuronal function and plasticity and vulnerable to age-related failure (Figure 1). It is proposed that this results in a number of consequences leading to the key features of AD pathophysiology as follows.

Entrapment of lipoprotein particles can explain the key features of Alzheimer's disease

Aβ deposition

In the parenchyma as plaques:

A β peptide is notoriously insoluble in an aqueous environment, but it is lipid soluble and is transported in the hydrophobic core of lipoprotein particles [27-30]. Therefore, trapping lipoprotein particles would immobilise A β in the extracellular space. As the trapped particles degrade and rupture, we propose they release A β peptide into the surrounding aqueous environment where it aggregates, initiating the formation of extracellular A β plaques. Colocalisation of APOE with A β plaques is consistent with the origin of the A β as being from APOE-containing lipoprotein particles [31]. Immunohisto-



Figure 1. Entrapment of lipoprotein particles in the brain causes Alzheimer's disease.

Lipoprotein particles transport cholesterol from glial cells, where it is synthesised, to neurons which require cholesterol for synaptic plasticity. The lipoprotein particles have a cholesterol-containing hydrophobic core, in which AB is solubilised, and a hydrophilic surface containing apolipoprotein E. Receptors for APOE on the neuronal cell membrane determine the destination of the particles. The lipoprotein particles must navigate the extracellular space to reach the neurons, a narrow cleft barely wider than the particles themselves. As the extracellular matrix ages, lipoprotein particles become trapped between cells and this primary abnormality results in reduced delivery of cholesterol to neurons leading to impaired synaptic plasticity, crucial for learning and memory. Impaired movement of lipoprotein particles in the extracellular space in the brain in ageing is central to and causes all the key pathophysiological features of Alzheimer's disease: degeneration of entrapped lipoprotein particles releases AB which aggregates in the aqueous environment of the extracellular space; a microglial reaction to the AB results in secretion of neurotoxic substances; neuronal cholesterol deficiency causes intraneuronal tau accumulation. [Artwork by Dr Jennifer M Dewing]



chemistry for A β on semi-thin sections (1µm) shows that diffuse plaques, usually interpreted as the earliest stage of A β deposition, are formed from a cluster of dot-like structures [32], and electron microscopy shows small vesicles associated with amyloid fibrils in the extracellular space [32]. In the context of the current hypothesis, it is intriguing to speculate that these might be entrapped lipoprotein particles from which the amyloid has originated.

APP transgenic mice, over-expressing the V717F human amyloid precursor protein, develop A β plaques as they age. Interestingly, when crossed with *APOE* knock out mice so that they lack APOE, these mice do not accumulate plaques [33]. This finding indicates that APOE is essential for the deposition of A β plaques, supporting the mechanism we propose.

With ageing, the primary risk factor for AD, the human brain decreases in weight and size with an overall decline in cortical cholesterol that accelerates from the age of 80 [34, 35]. This is consistent with age-associated entrapment of the lipoprotein particles impairing delivery of cholesterol to neurons and resulting in neuronal cholesterol deficiency [36]. This has further effects on Aβ as cholesterol deficiency leads to thinning of the cell membrane, shifting the site for secretase cleavage of the amyloid protein precursor (APP) from producing predominantly the shorter form of A β (A β 40) to the longer form (Aβ42) which is relatively even more insoluble and prone to aggregation [37, 38]. This exacerbates the effect of trapped lipoprotein particles as the $A\beta 42$ then coalesces onto the initial plaque seeds facilitating their growth.

In physiological conditions, $A\beta$ is suggested to act as a regulator of cholesterol homeostasis as observed in experimental models [39]. In humans, the familial forms of Alzheimer's disease are due to an imbalance in $A\beta$ production and are associated with increased cholesterol levels [40]. The binding of APOE to lipoprotein receptor-related protein (LRP) [41], the major APOE receptor on neurons, is also involved in the cellular uptake of cholesterol and $A\beta$, consistent with APOE, cholesterol and $A\beta$ all being components of the lipoprotein particle. *In the blood vessel walls as cerebral amyloid angiopathy (CAA):*

A β also colocalises with APOE in the walls of cerebral blood vessels suggesting that lipoprotein particles become trapped in the extracellular space of the vessel wall basement membrane as a consequence of age-related changes. The blood vessel dysfunction caused by the presence of CAA further compromises brain function by haemorrhage, ischaemia and paralysing autoregulation of cerebral blood flow [42].

Impaired neuronal and synaptic function

Deficiency of neuronal cholesterol results in impaired communication between neurons at the synapse, and particularly it interferes with the alterations in neurons which underpin learning and memory (i.e. synaptic plasticity) [43-46]. Synaptic plasticity requires neuronal cell membrane to be synthesised to form and re-form synapses as they are remodelled. In neuronal cultures, the presence of glial cells enhances the formation and function of synapses and the essential factor mediating this effect has been identified as glia-derived cholesterol, delivered to the neurons by APOE-containing lipoprotein particles binding to the neuronal LDL receptors [11, 47, 48].

Interestingly, in a number of clinical studies of traumatic brain injury, the possession of APOE E4 is associated with a worse outcome and severe neurologic deficits [49-51]. This seems particularly so in young people in whom neuronal plasticity after injury might otherwise be more pronounced [50]. Experimental studies in APOE E4 transgenic mice confirmed impaired neuronal plasticity after brain injury [52-56]. Further evidence comes from experimental models of global cerebral ischaemia in which APOEdeficient mice have increased neuronal damage which is ameliorated by intraventricular infusion of lipoprotein particles [57]. In addition, agonists of Liver X receptors (LXR), which act as cholesterol sensors and promote lipidation of APOE by ATP-binding cassette transporter A1 (ABCA1), ameliorate neuronal injury in experimental models of trauma [58] and reverse deficits in mouse models of AD [59].

The effects of neuronal and synaptic dysfunction due to lack of cholesterol [60] would be expected to be most pronounced in neuroanatomical locations in which plasticity is greatest (i.e. hippocampus involved in memory, association cortex involved in interpretation of sensations) and less severe or absent where plasticity is least (i.e. primary motor and sensory cortex, cerebellum, spinal cord). This is consistent with the clinical observations that the hierarchical sequence of loss of functions over time in AD follows the distribution of neuroplasticity [61].

Tau protein accumulation

We propose that neuronal cholesterol deficiency, resulting from the entrapment of lipoprotein particles, leads to the intracellular aggregation of tau. Tau is involved in maintaining the cytoskeletal structure of neurons and, in particular, the transport of proteins from the cell body along axons to the synapses. Accumulation of hyperphosphorylated tau occurs early in regions where the cholesterol is most in demand, that is where the rate of plasticity and synaptic remodelling is greatest (i.e. hippocampus, association cortex) and later or not at all where plasticity is least (i.e. primary motor and sensory cortex, cerebellum, spinal cord). A difficulty in trying to explain a direct link between Aβ and tau pathology in AD has been that they appear to arise in different neuroanatomical locations; tau accumulation occurs earlier in the hippocampus and associated structures, only spreading later to the cerebral neocortex, whereas AB accumulation occurs early in the cerebral neocortex. The hypothesis proposed here that there is not a direct link between AB and tau pathology, but rather that they are each driven by cholesterol deficiency, resolves this conundrum.

Direct experimental support comes from cholesterol depletion in neuronal cultures which induces tau hyperphosphorylation that can be prevented by treatment with lipoproteins and cholesterol [62]. The link between neuronal cholesterol deficiency and tau accumulation is further highlighted by the occurrence of tangles at a young age in Niemann-Pick type disease type C, a rare progressive genetic disorder characterised by the inability of the body to transport cholesterol and lipids [63-64].

Additional circumstantial evidence supporting a link between cholesterol deficiency and tangle formation potentially comes from the study of chronic traumatic encephalopathy (CTE). CTE is a condition caused by repeated blows to the head, typically in boxing and other sports, and is associated with the development of dementia with the formation of tangles as the key pathological feature [65]. It has been found that after a head injury, levels of APOE and cholesterol-containing lipoproteins in the cerebrospinal fluid plummet, just at the time when there is an increased demand for cholesterol for neuronal repair [66-68]. This relative deficiency of cholesterol, when recurrent over time with repeated blows to the head, could explain the development of tangles in CTE [51-56] [65]

Studies using cerebral organoids derived from AD patients highlight an association of tau pathology with APOE ϵ 4 carriage [69].

Glial cell dysfunction

Astrocytes and microglia have a major role in supporting neurons and activation of glial cells is a hallmark of AD. In particular, they play an important role in the cycling of lipoprotein particles, scavenging lipid debris from degenerating neuron/synapses, lipidation the particles and releasing them for transport and uptake by neurons [70]. In APOE E4 carriers, the brain is relatively deficient in APOE and the lipoprotein particles are smaller [71], more prone to aggregate and carry less cholesterol, rendering them particularly vulnerable to the effects of age-related cholesterol deficiency. Human induced pluripotent cell (iPSC)-derived astrocytes from ɛ4 homozygotes produce lipoprotein particles that are smaller, carry less cholesterol and support neurons less well in terms of viability and expression of synaptic proteins compared with those from ɛ3 carriers [72].

Microglia are the immune cells of the brain and are markedly activated in AD [73, 74]. Many of the genes identified in genome-wide association studies are expressed by microglia, indicating that they play an important role in the development and/or progression of the disease [75]. The presence of extracellular A β expelled by trapped and degraded lipo-

protein particles is putatively recognised by the microglial pattern recognition receptors (PPRs) evolved to detect molecular patterns associated with the bacterial cell walls. The consequent pro-inflammatory state provoked by this response causes release of cytotoxic substance evolved to destroy invading micro-organisms, which inadvertently has a harmful effect on neurons, compounding the effects of cholesterol deficiency [73, 76].

Weaknesses/limitations of the hypothesis

The ideas presented here form a hypothesis to explain the development of AD which is coherent and explains many facets of the disease. It is important to emphasise that a hypothesis is also an extrapolation of what is currently known and so some of the statements above are not firmly established but are potentially controversial and remain to be explored in the testing of the hypothesis. Potential thorns in the side of this hypothesis include:

- Localisation of Aβ within APOE-containing lipoprotein particles has not been directly demonstrated in the human brain. This needs to be explored and represents an important gap in our knowledge. Available technology is a limitation because the lipoprotein particles are below the resolution of light/confocal microscopy but are potentially amenable to study by novel 3D electron microscopy methods combined with multilabel immunostaining.
- Direct evidence that lipoprotein particles become trapped in the extracellular space, disintegrate and release Aβ is currently lacking.
- Studies of immunohistochemistry for Aβ and APOE identify APOE in some, but not all plaques. APOE seems to be present particularly in cored plaques, which are interpreted as later stage plaques, rather than early diffuse plaques as might be predicted from the hypothesis. The presence of APOE in later stage plaques could possibly reflect involvement of APOE-containing lipoprotein particles in attempted removal of Aβ, in addition to a role in plaque formation.

Evidence that high peripheral cholesterol levels are associated with increased risk of AD and that statins, which reduce circulating cholesterol by inhibiting the cholesterol-synthesising enzyme HMG-CoA reductase, may reduce risk for AD [77] might seem to contradict the hypothesis presented here. It seems that statins do reduce cholesterol levels in the brain [78]. However, it may be that the relative amounts of cholesterol, A β and APOE are important i.e., that there is sufficient cholesterol as required by neurons, sufficient APOE to solubilise the cholesterol and a sufficient volume of lipophilic core within the lipoprotein particles to transport $A\beta$ and prevent its escape into the aqueous environment of the extracellular space where it is prone to aggregate.

Therapeutic consequences

Over the past decades, human clinical trials of new therapies for AD have been unrelentingly disappointing. This is despite there being no shortage of endeavour and expenditure, and no lack of successful therapeutic studies in animal models of specific aspects of AD. Why is this? We suggest it is because the wrong targets have been addressed. Whereas each of the major suspects listed above (AB, tau, glial cells) may exacerbate the neurodegeneration and set up self-perpetuating vicious cycles, we propose they are not the initiating factor in sporadic AD, either singly or in combination. The pathophysiological scheme outlined above indicates that, according to this hypothesis, $A\beta$ and tau are not the cause of AD but are a consequence of the primary abnormality, which is trapping of lipoprotein particles in the extracellular space resulting in disruption of the normal system of delivery of cholesterol to neurons. This implies that even if, for example, the AD brain can be completely cleared of AB plaques, this will not halt the neurodegenerative decline as observed first in the long-term follow up of AD patients immunised against AB using AN1792 [79] and also in subsequent trials. Instead, therapy for AD must be directed at the health of the extracellular space, cholesterol and APOE, lubricating the passage of cholesterol-containing lipoprotein particles.



References

- 1. Wolters FJ, Yang Q, Biggs ML, Jakobsdottir J, Li S, Evans DS, et al. The impact of APOE genotype on survival: Results of 38,537 participants from six population-based cohorts (E2-CHARGE). PLoS One. 2019;14(7):e0219668. https://doi.org/10.1371/journal.pone.0219668
- 2. Corbo RM, Scacchi R. Apolipoprotein E (APOE) allele distribution in the world. Is APOE*4 a 'thrifty' allele? Ann Hum Genet. 1999;63(Pt 4):301-10. https://doi.org/10.1046/j.1469-1809.1999.6340301.x
- 3. Saunders AM, Strittmatter WJ, Schmechel D, George-Hyslop PH, Pericak-Vance MA, Joo SH, et al. Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. Neurology. 1993;43(8):1467-72.
- 4. Genin E, Hannequin D, Wallon D, Sleegers K, Hiltunen M, Combarros O, et al. APOE and Alzheimer disease: a major gene with semi-dominant inheritance. Mol Psychiatry. 2011;16(9):903-7. https://doi.org/10.1038/mp.2011.52
- Reiman EM, Arboleda-Velasquez JF, Quiroz YT, Huentelman MJ, Beach TG, Caselli RJ, et al. Exceptionally low likelihood of Alzheimer's dementia in APOE2 homozygotes from a 5,000person neuropathological study. Nat Commun. 2020;11(1):667. https://doi.org/10.1038/s41467-019-14279-8
- 6. Zannis VI, Nicolosi RJ, Jensen E, Breslow JL, Hayes KC. Plasma and hepatic apoE isoproteins of nonhuman primates. Differences in apoE among humans, apes, and New and Old World monkeys. J Lipid Res. 1985;26(12):1421-30.
- 7. McIntosh AM, Bennett C, Dickson D, Anestis SF, Watts DP, Webster TH, et al. The apolipoprotein E (APOE) gene appears functionally monomorphic in chimpanzees (Pan troglodytes). PLoS One. 2012;7(10):e47760. https://doi.org/10.1371/journal.pone.0047760
- 8. Poduri A, Gearing M, Rebeck GW, Mirra SS, Tigges J, Hyman BT. Apolipoprotein E4 and beta amyloid in senile plaques and cerebral blood vessels of aged rhesus monkeys. Am J Pathol. 1994;144(6):1183-7.
- 9. Dietschy JM, Turley SD. Thematic review series: brain Lipids. Cholesterol metabolism in the central nervous system during early development and in the mature animal. J Lipid Res. 2004;45(8):1375-97. https://doi.org/10.1194/jlr.R400004-JLR200

- 10. Poirier J. Apolipoprotein E and cholesterol metabolism in the pathogenesis and treatment of Alzheimer's disease. Trends in Molecular Medicine. 2003;9(3):94-101. https://doi.org/10.1016/S1471-4914(03)00007-8
- 11. Pfrieger FW. Cholesterol homeostasis and function in neurons of the central nervous system. Cell Mol Life Sci. 2003;60(6):1158-71. https://doi.org/10.1007/s00018-003-3018-7
- 12. Jones L, Harold D, Williams J. Genetic evidence for the involvement of lipid metabolism in Alzheimer's disease. Biochim Biophys Acta. 2010;1801(8):754-61. https://doi.org/10.1016/j.bbalip.2010.04.005
- 13. Hollingworth P, Harold D, Sims R, Gerrish A, Lambert JC, Carrasquillo MM, et al. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. Nat Genet. 2011;43(5):429-35. https://doi.org/10.1038/ng.803

- 14. Lamartiniere Y, Boucau MC, Dehouck L, Krohn M, Pahnke J, Candela P, et al. ABCA7 Downregulation Modifies Cellular Cholesterol Homeostasis and Decreases Amyloid-beta Peptide Efflux in an in vitro Model of the Blood-Brain Barrier. J Alzheimers Dis. 2018;64(4):1195-211. https://doi.org/10.3233/JAD-170883
- 15. Dib S, Pahnke J, Gosselet F. Role of ABCA7 in Human Health and in Alzheimer's Disease. Int J Mol Sci. 2021;22(9):4603. https://doi.org/10.3390/ijms22094603
- 16. Wahrle SE, Jiang H, Parsadanian M, Legleiter J, Han X, Fryer JD, et al. ABCA1 is required for normal central nervous system ApoE levels and for lipidation of astrocyte-secreted apoE. J Biol Chem. 2004;279(39):40987-93. https://doi.org/10.1074/jbc.M407963200
- 17. LaDu MJ, Gilligan SM, Lukens JR, Cabana VG, Reardon CA, Van Eldik LJ, et al. Nascent astrocyte particles differ from lipoproteins in CSF. J Neurochem. 1998;70(5):2070-81. https://doi.org/10.1046/j.1471-4159.1998.70052070.x
- 18. Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, et al. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nat Genet. 2009;41(10):1094-9. https://doi.org/10.1038/ng.439
- 19. Koch S, Donarski N, Goetze K, Kreckel M, Stuerenburg H-J, Buhmann C, et al. Characterization of four lipoprotein classes in human cerebrospinal fluid. Journal of Lipid Research. 2001;42(7):1143-51. https://doi.org/10.1016/S0022-2275(20)31605-9
- 20. Mahley RW. Central Nervous System Lipoproteins: ApoE and Regulation of Cholesterol Metabolism. Arterioscler Thromb Vasc Biol. 2016;36(7):1305-15. https://doi.org/10.1161/ATVBAHA.116.307023
- 21. Linton MF, Gish R, Hubl ST, Bütler E, Esquivel C, Bry WI, et al. Phenotypes of apolipoprotein B and apolipoprotein E after liver transplantation. J Clin Invest. 1991;88(1):270-81. https://doi.org/10.1172/jci115288
- 22. Stukas S, Kulis I, Zareyan S, Wellington CL. Lipids and Lipoproteins in Alzheimer's Disease. In: Zerr I, editor. Alzheimer's Disease -Challenges for the Future: IntechOpen; 2015.
- 23. Nicholson C, Hrabetova S. Brain Extracellular Space: The Final Frontier of Neuroscience. Biophys J. 2017;113(10):2133-42. https://doi.org/10.1016/j.bpj.2017.06.052
- 24. Nicholson C, Sykova E. Extracellular space structure revealed by diffusion analysis. Trends Neurosci. 1998;21(5):207-15. https://doi.org/10.1016/s0166-2236(98)01261-2
- 25. Ma J, Ma C, Li J, Sun Y, Ye F, Liu K, et al. Extracellular Matrix Proteins Involved in Alzheimer's Disease. Chemistry. 2020;26(53):12101-10. https://doi.org/10.1002/chem.202000782
- 26. Martins IJ, Berger T, Sharman MJ, Verdile G, Fuller SJ, Martins RN. Cholesterol metabolism and transport in the pathogenesis of Alzheimer's disease. J Neurochem. 2009;111(6):1275-308. https://doi.org/10.1111/j.1471-4159.2009.06408.>
- 27. Beffert U, Poirier J. Apolipoprotein E, plaques, tangles and cholinergic dysfunction in Alzheimer's disease. Ann N Y Acad Sci. 1996;777:166-74. https://doi.org/10.1111/j.1749-6632.1996.tb34415.x

- Biere AL, Ostaszewski B, Stimson ER, Hyman BT, Maggio JE, Selkoe DJ. Amyloid beta-peptide is transported on lipoproteins and albumin in human plasma. J Biol Chem. 1996;271(51):32916-22. <u>https://doi.org/10.1074/jbc.271.51.32916</u>
- 29. Cole GM, Ard MD. Influence of lipoproteins on microglial degradation of Alzheimer's amyloid beta-protein. Microsc Res Tech. 2000;50(4):316-24. https://doi.org/10.1002/1097-0029(20000815)50:4%3C316::aid-jemt11%3E3.0.co;2-e
- Tamamizu-Kato S, Cohen JK, Drake CB, Kosaraju MG, Drury J, Narayanaswami V. Interaction with amyloid beta peptide compromises the lipid binding function of apolipoprotein E. Biochemistry. 2008;47(18):5225-34. <u>https://doi.org/10.1021/bi702097s</u>
- Arold S, Sullivan P, Bilousova T, Teng E, Miller CA, Poon WW, et al. Apolipoprotein E level and cholesterol are associated with reduced synaptic amyloid beta in Alzheimer's disease and apoE TR mouse cortex. Acta Neuropathol. 2012;123(1):39-52. https://doi.org/10.1007/s00401-011-0892-1
- Yamaguchi H, Nakazato Y, Hirai S, Shoji M, Harigaya Y. Electron micrograph of diffuse plaques. Initial stage of senile plaque formation in the Alzheimer brain. Am J Pathol. 1989;135(4):593-7.
- Bales KR, Verina T, Cummins DJ, Du Y, Dodel RC, Saura J, et al. Apolipoprotein E is essential for amyloid deposition in the APP(V717F) transgenic mouse model of Alzheimer's disease. Proc Natl Acad Sci U S A. 1999;96(26):15233-8. <u>https://doi.org/10.1073/pnas.96.26.15233</u>
- 34. Soderberg M, Edlund C, Kristensson K, Dallner G. Lipid compositions of different regions of the human brain during aging. J Neurochem. 1990;54(2):415-23. https://doi.org/10.1111/j.1471-4159.1990.tb01889.x
- Svennerholm L, Bostrom K, Jungbjer B, Olsson L. Membrane lipids of adult human brain: lipid composition of frontal and temporal lobe in subjects of age 20 to 100 years. J Neurochem. 1994;63(5):1802-11. https://doi.org/10.1046/j.1471-4159.1994.63051802.x
- de Chaves EP, Narayanaswami V. Apolipoprotein E and cholesterol in aging and disease in the brain. Future Lipidol. 2008;3(5):505-30. <u>https://doi.org/10.2217/17460875.3.5.505</u>
- Welander H, Frånberg J, Graff C, Sundström E, Winblad B, Tjernberg LO. Abeta43 is more frequent than Abeta40 in amyloid plaque cores from Alzheimer disease brains. J Neurochem. 2009;110(2):697-706. https://doi.org/10.1111/j.1471-4159.2009.06170.x
- Saito T, Suemoto T, Brouwers N, Sleegers K, Funamoto S, Mihira N, et al. Potent amyloidogenicity and pathogenicity of Abeta43. Nat Neurosci. 2011;14(8):1023-32. <u>https://doi.org/10.1038/nn.2858</u>
- 39. Grimm MO, Grimm HS, Hartmann T. Amyloid beta as a regulator of lipid homeostasis. Trends Mol Med. 2007;13(8):337-44. https://doi.org/10.1016/j.molmed.2007.06.004
- Grimm MO, Grimm HS, Patzold AJ, Zinser EG, Halonen R, Duering M, et al. Regulation of cholesterol and sphingomyelin metabolism by amyloid-beta and presenilin. Nat Cell Biol. 2005;7(11):1118-23. https://doi.org/10.1038/ncb1313
- Tachibana M, Holm ML, Liu CC, Shinohara M, Aikawa T, Oue H, et al. APOE4-mediated amyloid-beta pathology depends on its neuronal receptor LRP1. J Clin Invest. 2019;129(3):1272-7. https://doi.org/10.1172/JCl124853

- Dumas A, Dierksen GA, Gurol ME, Halpin A, Martinez-Ramirez S, Schwab K, et al. Functional magnetic resonance imaging detection of vascular reactivity in cerebral amyloid angiopathy. Ann Neurol. 2012;72(1):76-81. <u>https://doi.org/10.1002/ana.23566</u>
- Frank C, Rufini S, Tancredi V, Forcina R, Grossi D, D'Arcangelo G. Cholesterol depletion inhibits synaptic transmission and synaptic plasticity in rat hippocampus. Exp Neurol. 2008;212(2):407-14. https://doi.org/10.1016/j.expneurol.2008.04.019
- 44. Maggo S, Ashton JC. Effects of HMG-CoA reductase inhibitors on learning and memory in the guinea pig. European Journal of Pharmacology. 2014;723:294-304. https://doi.org/10.1016/j.ejphar.2013.11.018
- Korinek M, Gonzalez-Gonzalez IM, Smejkalova T, Hajdukovic D, Skrenkova K, Krusek J, et al. Cholesterol modulates presynaptic and postsynaptic properties of excitatory synaptic transmission. Scientific Reports. 2020;10(1):12651. <u>https://doi.org/10.1038/s41598-020-69454-5</u>
- 46. Yujun G, Guichang Z, Jin J, Lei Y, Keke Q, Yang P, et al. Simvastatin Impairs Hippocampal Synaptic Plasticity and Cognitive Function in Mice. Molecular Brain. 2020. <u>https://doi.org/10.21203/rs.3.rs-76680/v1</u>
- Mauch DH, Nagler K, Schumacher S, Goritz C, Muller EC, Otto A, et al. CNS synaptogenesis promoted by glia-derived cholesterol. Science. 2001;294(5545):1354-7. <u>https://doi.org/10.1126/science.294.5545.1354</u>
- Pfrieger FW. Role of cholesterol in synapse formation and function. Biochim Biophys Acta. 2003;1610(2):271-80. https://doi.org/10.1016/s0005-2736(03)00024-5
- Teasdale GM, Nicoll JA, Murray G, Fiddes M. Association of apolipoprotein E polymorphism with outcome after head injury. Lancet. 1997;350(9084):1069-71. https://doi.org/10.1016/S0140-6736(97)04318-3
- Teasdale GM, Murray GD, Nicoll JA. The association between APOE epsilon4, age and outcome after head injury: a prospective cohort study. Brain. 2005;128(Pt 11):2556-61. <u>https://doi.org/10.1093/brain/awh595</u>
- Deng H, Ordaz A, Upadhyayula PS, Gillis-Buck EM, Suen CG, Melhado CG, et al. Apolipoprotein E Epsilon 4 Genotype, Mild Traumatic Brain Injury, and the Development of Chronic Traumatic Encephalopathy. Med Sci (Basel). 2018;6(3):78. <u>https://doi.org/10.3390/medsci6030078</u>
- 52. Poirier J. Apolipoprotein E in animal models of CNS injury and in Alzheimer's disease. Trends Neurosci. 1994;17(12):525-30. https://doi.org/10.1016/0166-2236(94)90156-2
- 53. Sabo T, Lomnitski L, Nyska A, Beni S, Maronpot RR, Shohami E, et al. Susceptibility of transgenic mice expressing human apolipoprotein E to closed head injury: the allele E3 is neuroprotective whereas E4 increases fatalities. Neuroscience. 2000;101(4):879-84. https://doi.org/10.1016/s0306-4522(00)00438-3
- White F, Nicoll JA, Roses AD, Horsburgh K. Impaired neuronal plasticity in transgenic mice expressing human apolipoprotein E4 compared to E3 in a model of entorhinal cortex lesion. Neurobiol Dis. 2001;8(4):611-25. <u>https://doi.org/10.1006/nbdi.2001.0401</u>
- 55. Mannix RC, Zhang J, Park J, Zhang X, Bilal K, Walker K, et al. Agedependent effect of apolipoprotein E4 on functional outcome after controlled cortical impact in mice. J Cereb Blood Flow Metab. 2011;31(1):351-61. https://doi.org/10.1038/jcbfm.2010.99

- Tensaouti Y, Yu T-S, Kernie SG. Apolipoprotein E regulates the maturation of injury-induced adult-born hippocampal neurons following traumatic brain injury. PLOS ONE. 2020; 15(3):e0229240. <u>https://doi.org/10.1371/journal.pone.0229240</u>
- 57. Horsburgh K, McCulloch J, Nilsen M, McCracken E, Large C, Roses AD, et al. Intraventricular infusion of apolipoprotein E ameliorates acute neuronal damage after global cerebral ischemia in mice. J Cereb Blood Flow Metab. 2000;20(3):458-62. https://doi.org/10.1097/00004647-200003000-00003
- Báez-Becerra C, Filipello F, Sandoval-Hernández A, Arboleda H, Arboleda G. Liver X Receptor Agonist GW3965 Regulates Synaptic Function upon Amyloid Beta Exposure in Hippocampal Neurons. Neurotox Res. 2018;33(3):569-79. <u>https://doi.org/10.1007/s12640-017-9845-3</u>
- 59. Cramer PE, Cirrito JR, Wesson DW, Lee CYD, Karlo JC, Zinn AE, et al. ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models. Science (New York, NY). 2012;335(6075):1503-6. https://doi.org/10.1126/science.1217697
- Koudinov AR, Koudinova NV. Cholesterol homeostasis failure as a unifying cause of synaptic degeneration. J Neurol Sci. 2005;229-230:233-40. <u>https://doi.org/10.1016/j.jns.2004.11.036</u>
- 61. Mesulam MM. A plasticity-based theory of the pathogenesis of Alzheimer's disease. Ann N Y Acad Sci. 2000;924:42-52. https://doi.org/10.1111/j.1749-6632.2000.tb05559.x
- Fan QW, Yu W, Senda T, Yanagisawa K, Michikawa M. Cholesteroldependent modulation of tau phosphorylation in cultured neurons. J Neurochem. 2001;76(2):391-400. https://doi.org/10.1046/j.1471-4159.2001.00063.x
- 63. Love S, Bridges LR, Case CP. Neurofibrillary tangles in Niemann-Pick disease type C. Brain. 1995;118 (Pt 1):119-29. https://doi.org/10.1093/brain/118.1.119
- 64. Suzuki K, Parker CC, Pentchev PG, Katz D, Ghetti B, D'Agostino AN, et al. Neurofibrillary tangles in Niemann-Pick disease type C. Acta Neuropathol. 1995;89(3):227-38. <u>https://doi.org/10.1007/BF00309338</u>
- McKee AC, Cairns NJ, Dickson DW, Folkerth RD, Keene CD, Litvan I, et al. The first NINDS/NIBIB consensus meeting to define neuropathological criteria for the diagnosis of chronic traumatic encephalopathy. Acta Neuropathol. 2016;131(1):75-86. <u>https://doi.org/10.1007/s00401-015-1515-z</u>
- Kay AD, Petzold A, Kerr M, Keir G, Thompson EJ, Nicoll JA. Cerebrospinal fluid apolipoprotein E concentration decreases after traumatic brain injury. J Neurotrauma. 2003;20(3):243-50. https://doi.org/10.1089/089771503321532824
- Kay AD, Day SP, Kerr M, Nicoll JA, Packard CJ, Caslake MJ. Remodeling of cerebrospinal fluid lipoprotein particles after human traumatic brain injury. J Neurotrauma. 2003;20(8):717-23. <u>https://doi.org/10.1089/089771503767869953</u>
- Kay AD, Petzold A, Kerr M, Keir G, Thompson E, Nicoll JA. Alterations in cerebrospinal fluid apolipoprotein E and amyloid beta-protein after traumatic brain injury. J Neurotrauma. 2003;20(10):943-52. https://doi.org/10.1089/089771503770195795
- Zhao J, Fu Y, Yamazaki Y, Ren Y, Davis MD, Liu C-C, et al. APOE4 exacerbates synapse loss and neurodegeneration in Alzheimer's disease patient iPSC-derived cerebral organoids. Nature Communications. 2020;11(1):5540. https://doi.org/10.1038/s41467-020-19264-0

- Horsburgh K, McCarron MO, White F, Nicoll JA. The role of apolipoprotein E in Alzheimer's disease, acute brain injury and cerebrovascular disease: evidence of common mechanisms and utility of animal models. Neurobiol Aging. 2000;21(2):245-55. <u>https://doi.org/10.1016/s0197-4580(00)00097-x</u>
- Heinsinger NM, Gachechiladze MA, Rebeck GW. Apolipoprotein E Genotype Affects Size of ApoE Complexes in Cerebrospinal Fluid. Journal of Neuropathology & Experimental Neurology. 2016;75(10):918-24. <u>https://doi.org/10.1093/jnen/nlw067</u>
- 72. Zhao J, Davis MD, Martens YA, Shinohara M, Graff-Radford NR, Younkin SG, et al. APOE epsilon4/epsilon4 diminishes neurotrophic function of human iPSC-derived astrocytes. Hum Mol Genet. 2017;26(14):2690-700. <u>https://doi.org/10.1093/hmg/ddx155</u>
- Boche D, Nicoll JAR. Invited Review Understanding cause and effect in Alzheimer's pathophysiology: Implications for clinical trials. Neuropathol Appl Neurobiol. 2020;46(7):623-40. <u>https://doi.org/10.1111/nan.12642</u>
- Franco-Bocanegra DK, Gourari Y, McAuley C, Chatelet DS, Johnston DA, Nicoll JAR, et al. Microglial morphology in Alzheimer's disease and after Abeta immunotherapy. Sci Rep. 2021;11(1):15955. https://doi.org/10.1038/s41598-021-95535-0
- 75. Jones L, Holmans PA, Hamshere ML, Harold D, Moskvina V, Ivanov D, et al. Genetic evidence implicates the immune system and cholesterol metabolism in the aetiology of Alzheimer's disease. PLoS One. 2010;5(11):e13950. https://doi.org/10.1371/journal.pone.0013950
- 76. Boche D, Perry VH, Nicoll JAR. Review: activation patterns of microglia and their identification in the human brain. Neuropathology and applied neurobiology. 2013;39(1):3-18. https://doi.org/10.1111/nan.12011
- Torrandell-Haro G, Branigan GL, Vitali F, Geifman N, Zissimopoulos JM, Brinton RD. Statin therapy and risk of Alzheimer's and age-related neurodegenerative diseases. Alzheimers Dement (N Y). 2020;6(1):e12108. https://doi.org/10.1002/trc2.12108
- Cibičková L. Statins and their influence on brain cholesterol. J Clin Lipidol. 2011;5(5):373-9. https://doi.org/10.1016/j.jacl.2011.06.007
- 79. Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, et al. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebocontrolled phase I trial. Lancet. 2008;372(9634):216-23. https://doi.org/10.1016/S0140-6736(08)61075-2

