

Majoritary triterpenic compounds in some angiosperms of the central region of Rio Grande do Sul state

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ABSTRACT

Five species of angiosperms collected in the Central Region of Rio Grande do Sul was evaluated due to their pharmacological properties. The objective of this study was to extract and characterize the triterpenic content of angiosperm species. The extracts were obtained by ultrasonic extraction using chloroform as solvent. Identification and quantification were performed using high performance liquid chromatography with UV detection (HPLC-UV). The presence of arjunic acid, maslinic acid, oleanolic acid, erythrodiol, uvaol, lupeol, β -amirin, α -amirin, stigmasterol, and β -sitosterol was observed. All species presented β -sitosterol. *Polygala pulchella* presented the greatest diversity of triterpenic compounds, while *Ruellia angustiflora* and *Moquiniastrum mollissimum* the least. *Paspalum rawitscheri* and *Hesperozygis ringens* presented the same constituents, differing from each other only quantitatively. Therefore, an overview of the triterpene constitution in species of the central region of the state of RS is presented, along with a review on the pharmacological properties of the investigated compounds. The results obtained in this work are relevant since they

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establish an overview of the chemical constitution of the extracts whose biology is still little known. The pharmacological potential of the species can be attributed the identified triterpenic constituents thus supporting the use for medicinal purposes.

Keywords: Triterpenes; Moquiniastrum; Paspalum; Hesperozygis; Ruellia; Polygala

RESUMO

Cinco espécies de angiospermas coletadas na Região Central do Rio Grande do Sul foram avaliadas tendo em vista às suas propriedades farmacológicas. O objetivo deste estudo foi extrair e caracterizar o conteúdo triterpênico de espécies de angiospermas. Os extratos foram obtidos por extração ultrasônica usando clorofórmio como solvente. A identificação e quantificação foram realizadas por cromatografia líquida de alta eficiência com detecção UV (HPLC-UV). Foi verificado a presença de ácido arjunic, ácido maslínico, ácido oleanólico, eritrodiol, uvaol, lupeol, β -amirina, α -amirina, estigmasterol e β -sitosterol. Todas as espécies apresentaram β -sitosterol. *Polygala pulchella* apresentou a maior diversidade de compostos triterpênicos, enquanto *Ruellia angustiflora* e *Moquiniastrum mollissimum* o mínimo. *Paspalum rawitscheri* e *Hesperozygis ringens* apresentaram os mesmos constituintes, diferindo entre si apenas quantitativamente. Portanto, é apresentada uma visão geral da constituição dos triterpenos em espécies da região central do RS, juntamente com uma revisão das propriedades farmacológicas dos compostos investigados. Os resultados obtidos neste trabalho são relevantes, pois estabelecem uma visão geral da constituição química dos extratos cuja biologia ainda é pouco conhecida. O potencial farmacológico da espécie pode ser atribuído aos constituintes triterpênicos identificados, apoiando assim o uso para fins medicinais.

Palavras-chaves: Triterpenos; Moquiniastrum; Paspalum; Hesperozygis; Ruellia; Polygala

1 INTRODUCTION

Medicinal plants have been used for centuries in traditional medicine and are associated with promoting health and preventing/curing diseases (LASZCZYK, 2009; ROMERO et al., 2010).

In Brazil the use of medicinal plants is eased by the fact that the country presents the greatest biodiversity of the planet, resulting in a multitude of active principles (CHISTÉ et al., 2013). In contrast, it is known that many of the plants of this biodiversity are only little studied.

According to the Brazilian Institute of Geography and Statistics (IBGE, 2004), presents six distinct biomes. All with high biodiversity and phytophysiological diversity, but some of them are still poorly investigated and have been the focus of many studies in the last decade. One of these biomes is the Pampa, which covers only the state of Rio Grande do Sul in Brazil. In the last decades, about half of the area originally covered with fields in the state of Rio Grande do Sul has been transformed into other types of vegetation cover (PILLAR, 2009). This degradation is particularly worrying, since, despite recent advances, most of the Brazilian Campos Sulinos remains insufficiently known (GIULIETTI et al., 2005). Besides, out of the approximately 2,220 species of plants in the fields of Rio Grande do Sul, 213 are considered endangered (PILLAR, 2009), what makes more urgent the task of gathering knowledge and chemical characterization of this biodiversity.

The terpenes are natural hydrocarbons produced by a wide variety of plants and animals. They act in the defense and adaptation system of plants, show important pharmacological properties (Table 1), and therefore, are products of high interest for the chemical-pharmaceutical industry (RAMAWAT; MÉRILLIN, 2013). The triterpenes are terpenes which are composed by thirty

carbons (C_{30}) with promising pharmacological properties (MUFFLER et al., 2011). The triterpenic compounds involved in this study have their chemical structures represented in Figure 1 and their main pharmacological activities presented in Table 1.

Figure 1 - Chemical structures the compounds involved in this study

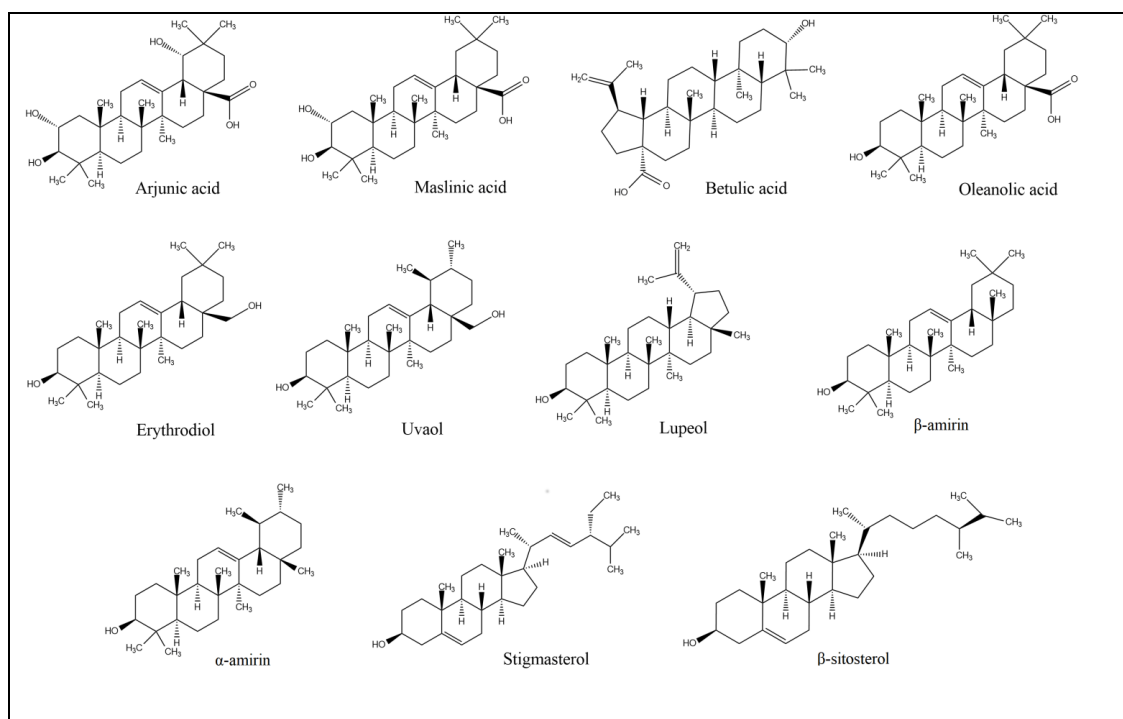


Table 1- Triterpenic compounds and their pharmacological activities:

Triterpenes	Pharmacological Activities
Arjunic acid	<i>Antioxidant, antimicrobial, antibacterial, neuroprotective, antidiabetic, scarring, inhibits the growth of insects, cardioprotective, antifungal, antioxidant, anti-inflammatory, cytotoxic and antitumoral (JOO, 2016).</i>
Maslinic acid	Inhibits the growth of insects (SÁNCHEZ-QUESADA et al., 2013), antifungal, antidiabetic, gastroprotector (SÁNCHEZ-ÁVILA et al., 2009), antitumoral (SIEWRT and CSUK, 2014).
Betulic acid	Antiviral, acts against the growth of parasites, antibacterial, anti-inflammatory, inhibits the growth of cancer cells (PAI et al., 2011, antimicrobial, antioxidant (MORE et al., 2012), hepatoprotective (DONFACK et al., 2010) and antifungal (GARCEZ et al., 2016).
Oleanolic acid	Anti-inflammatory, hepatoprotector, antitumoral, antimicrobial (MARTELANC; VOVK; SIMONOVSKA, 2009) antiviral, antifungal, antidiabetic, gastroprotector, reduces blood lipid levels (SÁNCHEZ-ÁVILA et al., 2009), antioxidant (DONFACK et al., 2010, antiprotozoal (XU; SU; ZANG, 2012) and antitumor (SHANMUGAM et al., 2014).
Erythrodiol	Antioxidant and antithrombotic (ALLOUCHE et al., 2010), antitumoral (SÁNCHEZ-QUESADA et al., 2013), antibacterial (DOUGLAS, 2016).
Uvaol	Antioxidant and antithrombotic (ALLOUCHE et al., 2010), antibacterial (DOUGLAS, 2016), anti-inflammatory and antioxidant (AGRA et al., 2016).
Lupeol	Antiprotozoa, anti-inflammatory, anticarcinogenic, cardioprotective, antimicrobial, hepatoprotector (UMARI; KAKKAR, 2012), antiarthritic, antimutagenic, antioxidant, antimalarial (SALEEM et al., 2008), antidiabetic, nephroprotector (SIDDIQUE; SALEEM, 2011) antioxidant (SANTIAGO; MAYOR, 2014).

β -amirin	Gastroprotector, antipruritic, hepatoprotector (MARTELANC; VOVK; SIMONOVSKA, 2007), anti-inflammatory and analgesic (DIAS; HAMERSKI; PINTO, 2011).
α -amirin	Gastroprotector, antipruritic, hepatoprotector, anti-inflammatory (MARTELANC; VOVK; SIMONOVSKA, 2007), antiarthritic (DIAS; HAMERSKI; PINTO, 2011), reduces sensitivity to pain stimuli (BACKHOUSE et al., 2008) and antimicrobial (MORE et al., 2012).
Stigmasterol	Antitumoral (LEE et al., 2012), cholesterol reducer and gastroprotector (QUÍLEZ et al., 2010), antifungal (MBAMBO; ODHAV; MOHANLALL, 2012) and larvicidal (GADE et al., 2017).
β -sitosterol	Gastroprotector (QUÍLEZ et al., 2010), analgesic (SAEIDNIA et al., 2014), anthelmintic, antimutagenic (SAEIDNIA et al., 2014; VILLASEÑOR et al., 2002), antimicrobial (SANTOS; MORENO, 2004) antitumoral (LEE et al., 2012, LOMENICK et al., 2015) and anti-inflammatory (SAEIDNIA et al., 2014; LOMENICK et al., 2015).

Different species of angiosperms were collected for the purpose of this study, considering the occurrence in the region and lack of scientific information about these compounds. The selected species belong to five different families, commonly found in grassland formations or edge of forests of the biomes Pampa and Atlantic Forest in the central region of Rio Grande do Sul. Three rare species, listed in the national list of endangered species (BRASIL, 2008) and poorly investigated biology were selected *Moquiniastrum mollissimum* (Malme) G. Sancho, *Paspalum rawitscheri* (Parodi) Chase ex G.H. Rua & Valls, and *Hesperozygis ringens* (Benth.) Epling). The other two species *Ruellia angustiflora* (Nees) Lindau ex Rambo and *Polygala pulchella* (A. St.-Hil. & Moq.), on the other hand, present broader distribution and belong to genera with citations in traditional medicine.

Moquiniastrum mollissimum (= *Gochnatia molissima* (Malme) Cabrera) is a species of the Pampa biome, which in Brazil is restricted to the state of Rio Grande do Sul, that belongs to the family Asteraceae (FLORA DO BRASIL, 2020).

The genus Moquiniastrum includes 21 species. In Brazil, 19 species are found distributed in the main phytogeographic domains that include Cerrado, Pampa and Atlantic Forest. Ten of these species occur in the South region (LOMENICK et al., 2015). Strapasson et al. (2014) assigns the following constituents to some species of the genus: sesquiterpenes, diterpenes, triterpenes, coumarins, flavonoids, caffeic acid derivatives, acetylene, and amino acid.

Paspalum rawitscheri (= *Thrasypopsis juergensii* (Hack.) Soderstr. & AG Burm.), known as “capim-dos-descampados”, is an endemic grass of southern Brazil. This

species is in the list of endangered species of Brazilian flora due to the progressive reduction of the number of individuals in known populations (RUA; VALLS, 2012; MARCHIORI, et al., 2014). Little is known about this species biology, so genetic, chemical and ecological studies are encouraged.

The genus *Hesperozygis* (Lamiaceae) arouses scientific interest for medicinal use, being already recognized by its characteristic odor (LAWRENCE, 1992). Among the most important species we find *Hesperozygis ringens*. It is an aromatic plant, endemic to southern Brazil and listed as a threatened species (BRASIL, 2008). Popularly, the plant is used as an insecticide, which has already been scientifically proven. Such activity is attributed to the pulegone, main component of its essential oil, which is found in concentrations above 80%. It is also reported in the literature that pulegone confers nematicidal, larvicidal, acaricidal and bactericidal activity to the plant (VON POSER, 1996; SILVA et al., 2013; SILVA et al., 2014). Chemical constituents such as α -pinene, sabinene, β -pinene, limonene, linalool and caryophyllene were reported by Silva et al. (2014). They also show antioxidant, anesthetic (SILVA et al., 2013), allelopathic, antiparasitic and antifungal (VON POSER, 1996).

Ruellia angustiflora is an herbaceous forest-edge species. Plants from the genus *Ruellia* present secondary metabolites such as flavonoids, steroids, triterpenes, coumarins and alkaloids (SAMMY et al., 2015; CHOTHANI et al., 2010; SALAH et al., 2002). These are responsible for cardiovascular, antihyperglycemic, antibacterial, antioxidant, antinociceptive, anti-inflammatory, and gastroprotective activities, among others that are usually associated to some species of *Ruellia* (SAMMY et al., 2015). *Ruellia angustiflora* occurs in Brazil, Uruguay, Paraguay and Argentina. It is popularly known as "flor-de-fogo", due to the red color of its flowers (ALICE, 1995; FUHRO; VARGAS; LAROCCA, 2005). Its leaves are referred to as scarring in traditional medicine (ALICE, 1995).

Polygala is the most diversified genus of the family Polygalaceae, with approximately 725 species (LUDTKE; CHIES; MIOTTO, 2008). Chemical research studies of the genus have revealed the occurrence of a variety of secondary metabolites, such as xanthonenes, saponins, flavonoids, and coumarins, as mentioned by Johann et al.

(2012). In addition to these compounds, the species of *Polygala* are also characterized by the presence of methyl salicylate found in its roots, which is used as pain killer, mainly for muscular pain (ROCHA et al., 2012). Martins-Nucci et al. (2016) reported the use of these species in the treatment of pain and inflammation. This species is used in popular medicine for the treatment of bronchitis, asthma, whooping cough, rheumatism as well as antiofidic (MARQUES; PEIXOTO, 2007).

Therefore, this article aims to identify and quantify triterpenic in *Moquiniastrum mollissimum*, *Paspalum rawitscheri*, *Ruellia angustiflora*, *Polygala pulchella* and *Hesperozygis ringens* collected in distinct locations of the central region of Rio Grande do Sul. Also, we aim to present a literature review of the compounds investigated.

2 MATERIALS AND METHODS

2.1 Chemicals

All the chemical standards used, α -amirin (98%), β -amirin (98,5%), β -sitosterol (85%), stigmasterol (95%), lupeol (90%), uvaol (95%), erythrodiol (97%), oleanolic acid (97%), betulic acid (97%), arjunic acid (88%) and maslinic acid (95%), were of analytical grade from Sigma-Aldrich (St. Louis, MO, USA). The solvents acetonitrile (ACN), methanol (MeOH) and tetrahydrofuran (THF) were of HPLC grade from Tedia (Fairfield, OH, EUA). The Chloroform (CHCl_3) used was of analytical grade, from Merck (Darmstadt, Germany). Stock solutions of α -amirin, β -amirin, uvaol, erythrodiol, oleanolic acid, arjunic acid and maslinic acid 1000 mg/l, stigmasterol 481 mg/l, lupeol 365 mg/l, betulic acid 196 mg/l and β -sitosterol 873 mg/l were prepared in methanol. The working analytical solutions for the analytical curve were obtained by diluting the analytical solutions in acetonitrile with the following concentrations: 0.7; 6.5; 12.2; 18.0; 23.6; 29.3 and 35.0 mg/l. All solutions were stored at -20°C until the moment of analysis.

2.2 Plant Material

The species were collected from different locations in the Central Region of the state of Rio Grande do Sul. Endangered species were collected without compromising their reproductive structures (leaves only). Table 2 shows the species involved in this study, as well as the part of the plant that was collected, month and year of collection and location.

Table 2 - Collection data of the studied species

Species	Collected part	Voucher
<i>Hesperozygis ringens</i> (Benth.) Epling	Leaf	Brazil, Rio Grande do Sul, São Pedro do Sul: Morro Itaquiatiá. <i>L. Essi</i> 612. 23. Oct 2013. (SMDB)
<i>Paspalum rawitscheri</i> (Parodi) Chase ex G.H. Rua & Valls	Leaf	Brazil, Rio Grande do Sul, Santa Maria: Em afloramento arenítico Morro Pedra do Lagarto. <i>L. Essi</i> 799 <i>et al.</i> 15 Oct 2014. (SMDB)
<i>Moquiniastrum mollissimum</i> (Malme) G. Sancho	Leaf	Brazil, Rio Grande do Sul, Santa Maria: Distrito de Santo Antônio, Pedra do Lagarto. <i>L. Essi</i> 904, <i>J. Schaefer</i> & <i>H.F. Menezes</i> . 25 Mar 2015. (SMDB)
<i>Ruellia angustiflora</i> (Nees) Lindau ex Rambo	Leaf	Brazil, Rio Grande do Sul, Santa Maria: Distrito de Santo Antônio, estrada entre a Pedra do Lagarto e a Capelo de Santo Antônio. <i>L. Essi</i> 760 & <i>J. Freitas</i> . 14 Oct 2016. (SMDB)
<i>Polygala pulchella</i> A. St.- Hil. & Moq.	root and aerial part	Brazil, Rio Grande do Sul, Santa Maria: em campo úmido modificado, atrás da FATEC. <i>L. Essi</i> 745. 16 Jul 2016. (SMDB)

The parts of interest were dried in oven at 40 °C until constant mass. Afterwards, they were ground in a knife mill, packed in waterproof packages and protected from light until the moment of analysis.

2.3 Extraction Procedure

The extractions were conducted according to the methodology described by Pires et al (2017) with some modifications regarding the extraction solvent. Plant extracts were prepared via ultrasound-assisted extraction (Bandelin, Sonorex Super

RK 510 H). Glass tubes containing approximately 0.2 g of the samples received 10 mL of the extraction solvent (chloroform) and were placed in an ultrasonic bath for 4 h at room temperature. The resulting liquid from the extraction was filtered with filter paper, placed in glass beaker and then oven-dried at 40 °C for evaporation of the solvent. Afterwards, they were resuspended in acetonitrile (HPLC) and filtered by a 0.22 µm membrane (Sorblin Tecnologie). The initial concentrations were maintained. The extracted samples remained stored at -30 °C until the analysis. Before injection into the chromatograph, all samples were diluted to 0.01 g/ml with acetonitrile (HPLC grade).

2.4 Quantitative Analysis of Triterpenic Compounds by HPLC-RP

The identification and quantification of apolar constituents in the species were performed according to the method described by Schmidt et al (2018) that uses reverse phase chromatography and ultraviolet detection. Chromatographic measurements were performed on a Dionex® model P680 (Sunnyvale, CA, USA) liquid chromatograph equipped with a UV-vis detector model UVD170U, Rheodyne® injection valve model 8125 (Cotati, CA, USA) with loop of 100 µL. The analyses were carried out with a Kinetex reversed-phase C₁₈ column (250 mm × 4.6 mm, I.D., 5 µm particle size; Phenomenex, Torrance, CA, USA) which was preceded by a Security Guard C₁₈ pre-column (Phenomenex, Torrance, CA, USA). The mobile phase consisted of acetonitrile: tetrahydrofuran (90:10, v/v) and the flow-rate was set at 0.5 ml/min. Spectrophotometry detection of the analytes was performed at the wavelength of 210 nm. Evaluation and quantification were made on a Chromeleon 6.7 Workstation. The general validation parameters were: linearity (0.74-40.0 mg/kg), LOD (0.06-1.04 mg/kg), LOQ (0.13-2.2 mg/kg), correlation coefficient r^2 (0.990-0.996) accuracy (80-120%), inter-day precision (1.84-4.40%) and intra-day precision (2.01-9.37%). All validation parameters are presented in detail in Schimdt et al (2018).

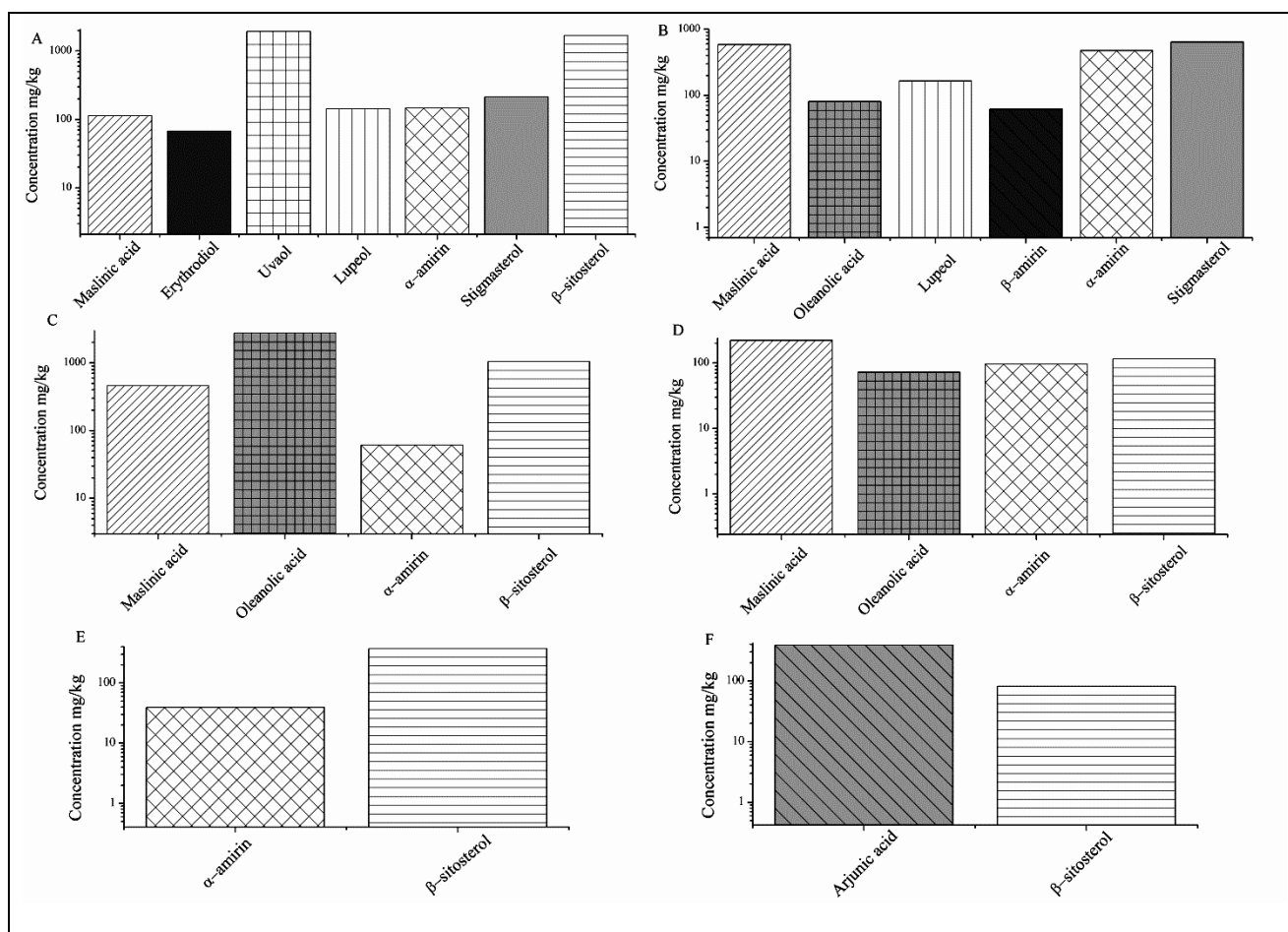
The quantification was performed using an external standardization method according to Ribani et al. (2004). The analytical curves were obtained by preparing the mixture of seven concentrations of the standards with injections in triplicates, where

the analytical curve was expressed by the equation of the line. Samples were analyzed based on injections in triplicate, where the average of the areas corresponding to each compound presented by the sample was obtained. By taking the average of the area under the analytical curves, we obtained the concentration of the constituent in the sample.

3 RESULTS

The distribution of the triterpenic compounds in the investigated species are shown in Figure 2.

Figure 2 - **A)** *Polygala pulchella* (root), **B)** *Polygala pulchella* (aerial part), **C)** *Hesperozygis ringens*, **D)** *Paspalum rawitsheri*, **E)** *Ruellia angustiflora*, **and F)** *Moquiniastrum mollissium*



4 DISCUSSION

Among the studied species, *Polygala pulchella* presented the greater diversity of compounds, as may be seen in graphs (a) and (b) in Figure 2. Although the literature mentions the roots much more often regarding medicinal purposes, it was verified that the aerial part is also rich in triterpenes (ROCHA et al., 2012).

The presence of β -sitosterol was verified in all species studied. This constituent was not found only in the aerial part of *Polygala pulchella*. β -sitosterol is described in the literature as presenting several pharmacological activities such as: hypocholesterolemic (CHAUHAN; RUBY; DWIVEDI, 2013), gastroprotective (QUÍLEZ et al., 2010), anti-inflammatory, analgesic (SAEIDNIA et al., 2014), anthelmintic, antimutagenic (VILLASEÑOR et al., 2002; SAEIDNIA et al., 2014) antimicrobial and antitumor (SANTOS and MORENO, 2004). The anthelmintic action of β -sitosterol may support the use of *Hesperozygis ringens*. Also, its anti-inflammatory and analgesic potential may endorse the use of *Polygala pulchella*.

In *Polygala pulchella*, in addition to β -sitosterol, β -amirin, α -amirin and lupeol may also justify the use of this species in the treatment of pain and inflammation (BACKHOUSE et al., 2008; DIAS; HAMERSKI; PINTO 2011; KUMARI; KAKKAR, 2012). Uvaol and erythrodiol were only identified in the roots of *Polygala pulchella*, and oleanolic acid only in its aerial part. Stigmasterol, maslinic acid, lupeol and α -amirin were found on both parts of the plant. Antifungal activity (SÁNCHEZ-ÁVILA., 2009) is attributed to the maslinic acid since it has already been proven in other species of the genus *Polygala* (JOHANN et al., 2012).

Polygala pulchella (aerial part), *Hesperozygis ringens* and *Paspalum rawitscheri* presented both oleanolic and maslinic acid. It should be highlighted that *Hesperozygis ringens* and *Paspalum rawitscheri* presented the same constituents, only differing quantitatively from each other, as shown in graphs (c) and (d) in Figure 2.

Moquiniastrum mollissimum and *Ruellia angustiflora* (Figure 2) presented lower diversity of compounds. However, the results are of great relevance due to the lack of

studies about these species. *Moquiniastrum mollissimum* was the only species that presented arjunic acid.

Paspalum rawitscheri is also an unexplored species, but presented important constituents, as shown in the graphic (d) from Figure 2. This species needs further investigation, especially regarding its biology.

According to the results obtained, at least two triterpenic compounds are present in the chemical composition of the extracts of the analyzed species. Among the compounds investigated, β -sitosterol was the only one found in all species. *Polygala pulchella* presented the greatest variety of compounds. *Hesperozygis ringens* and *Paspalum rawitscheri* presented the same chemical composition, but in differed quantities. *Moquiniastrum mollissimum* and *Ruellia angustiflora* showed the least diversity of compounds. The results obtained in this work are relevant since they stablish an overview of the chemical constitution of the extracts. This allows the evaluation of the medicinal potential that such species may present regarding the presence of important triterpenic constituents with pharmacological activities. Our results also build knowledge around rare species, whose biology is still little known, drawing attention to the need for their conservation.

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REFERENCES

AGRA, Lais Costa et al. Uvaol attenuates pleuritis and eosinophilic inflammation in ovalbumin-induced allergy in mice. *European journal of pharmacology*, v. 780, p. 232-242, 2016.

ALICE, Cecilia Ballvé. *Plantas medicinais de uso popular: atlas farmacognóstico*. Editora da ULBRA, 1995.

ALLOUCHE, Yosra et al. Antioxidant and antiatherogenic activities of pentacyclic triterpenic diols and acids. *Food and Chemical Toxicology*, v. 48, n. 10, p. 2885-2890, 2010.

BACKHOUSE, N. et al. Analgesic, anti-inflammatory and antioxidant properties of *Buddleja globosa*, Buddlejaceae. *Journal of ethnopharmacology*, v. 116, n. 2, p. 263-269, 2008.

Brasil. Ministério do Meio Ambiente. **Instrução Normativa nº 002, de 19 de setembro de 2008**. Institui a Nova Lista Oficial das Espécies da Flora Brasileira Ameaçada de Extinção. Availabe: http://www.mma.gov.br/estruturas/ascom_boletins/_arquivos/83_19092008034949.pdf.

CHAUHAN, RAJANI; RUBY, K.; DWIVEDI, JAYA. Secondary metabolites found in *Bergenia* species: a compendious review. *reactions*, v. 15, p. 17, 2013.

CHISTÉ, Renan Campos et al. The phenolic compounds and the antioxidant potential of infusion of herbs from the Brazilian Amazonian region. *Food Research International*, v. 53, n. 2, p. 875-881, 2013.

CHOTHANI, Daya L. et al. Review on *Ruellia tuberosa* (cracker plant). *Pharmacognosy Journal*, v. 2, n. 12, p. 506-512, 2010.

DIAS, Marluce Oliveira; HAMERSKI, Lidilhone; PINTO, Angelo C. Semi-preparative separation of α and β -amyrin by high performance liquid chromatographic. *Química Nova*, v. 34, n. 4, p. 704-706, 2011.

DONFACK, J. Hubert et al. In vitro hepatoprotective and antioxidant activities of the crude extract and isolated compounds from *Irvingia gabonensis*. *Asian Journal of Traditional Medicines*, v. 5, n. 3, p. 79-88, 2010.

DOUGLAS, Kemboi. Phytochemistry and Antimicrobial Activity of Extracts from Medicinal Plant *Olea africana* and *Olea europea*. *International Journal of Biochemistry Research & Review* 12(2): 1-7, 2016.

Flora do Brasil, 2017. *Moquiniastrum* In: Flora do Brasil 2020 em construção. Jardim Botânico do Rio de Janeiro. Availabe: <http://floradobrasil.jbrj.gov.br/reflora/floradobrasil/FB129778>_[accessed 2018 June 01].

FUHRO, Daniela; VARGAS, D.; LAROCCA, João. Levantamento florístico das espécies herbáceas, arbustivas e lianas da floresta de encosta da Ponta do Cego, Reserva Biológica do Lami (RBL), Porto Alegre, Rio Grande do Sul, Brasil. *Pesquisas, ser. Botânica*, v. 56, p. 239-256, 2005.

GADE, Sriramy et al. Acetylcholinesterase inhibitory activity of stigmasterol & hexacosanol is responsible for larvicidal and repellent properties of *Chromolaena odorata*. *Biochimica et Biophysica Acta (BBA)-General Subjects*, v. 1861, n. 3, p. 541-550, 2017.

GARCEZ, Fernanda R. et al. A diversidade dos constituintes químicos da flora de Mato Grosso do Sul e sua relevância como fonte de substâncias bioativas. *Revista Virtual de Química*, v. 8, n. 1, p. 97-129, 2016.

GIULIETTI, Ana Maria et al. Biodiversity and conservation of plants in Brazil. *Conservation Biology*, v. 19, n. 3, p. 632-639, 2005.

IBGE. Instituto Brasileiro de Geografia e Estatística. [internet]. Mapa da vegetação do Brasil e Mapa de Biomas do Brasil; 2004. [cited 2018 June 01]. Available: <https://ww2.ibge.gov.br/home/presidencia/noticias/21052004biomashtml.shtm>.

JOHANN, Susana et al. Antifungal activity of five species of *Polygala*. *Brazilian Journal of Microbiology*, v. 42, n. 3, p. 1065-1075, 2011.

JOO, HyeEun et al. c-Jun N-terminal Kinase-Dependent Endoplasmic Reticulum Stress Pathway is Critically Involved in Arjunic Acid Induced Apoptosis in Non-Small Cell Lung Cancer Cells. *Phytotherapy Research*, v. 30, n. 4, p. 596-603, 2016.

KUMARI, Archana; KAKKAR, Poonam. Lupeol protects against acetaminophen-induced oxidative stress and cell death in rat primary hepatocytes. *Food and chemical toxicology*, v. 50, n. 5, p. 1781-1789, 2012.

LASZCZYK, Melanie N. Pentacyclic triterpenes of the lupane, oleanane and ursane group as tools in cancer therapy. *Planta medica*, v. 75, n. 15, p. 1549-1560, 2009.

LAWRENCE, B. M. Chemical components of Labiatae oil sand their exploitation. *Advances in Lamiaceae Science*. Edits., RM Harley and T. Reynolds, p. 399-436, 1992.

LEE, Seung Rok et al. Determination of phytosterols in *Oryza sativa* L. cultivars by liquid chromatography atmospheric chemical ionization time-of-flight mass spectrometer (LC-APCI-TOF-MS) coupled with a modified QuEChERS method. *Journal of Agricultural Chemistry and Environment*, v. 1, n. 01, p. 15, 2012.

LOMENICK, Brett et al. Identification and characterization of β -sitosterol target proteins. *Bioorganic & medicinal chemistry letters*, v. 25, n. 21, p. 4976-4979, 2015.

LOMENICK, Brett et al. Identification and characterization of β -sitosterol target proteins. *Bioorganic & medicinal chemistry letters*, v. 25, n. 21, p. 4976-4979, 2015.

LUDTKE, Raquel; CHIES, Tatiana Teixeira de Souza; MIOTTO, Silvia Teresinha Sfoggia. Bredemeyera Willd. e Securidaca L. (Polygalaceae) na região sul do Brasil. Revista Brasileira de Biociências= Brazilian Journal of Biosciences. Vol. 6, n. 1 (jan./mar. 2008), p. 69-79, 2008.

MARCHIORI, José Newton Cardoso et al. Campos e florestas no curso médio do Rio Toropi, Rio Grande do Sul (Brasil). Retrato de um admirável patrimônio ameaçado. Balduinia, n. 45, p. 01-16, 2014.

MARQUES, Maria do Carmo Mendes; PEIXOTO, Ariane Luna. Taxonomic study of *Polygala subgenus Ligustrina* (Chodat) Paiva (Polygalaceae). Rodriguésia, v. 58, n. 1, p. 95-146, 2007.

MARTELANC, Mitja; VOVK, Irena; SIMONOVSKA, Breda. Determination of three major triterpenoids in epicuticular wax of cabbage (*Brassica oleracea* L.) by high-performance liquid chromatography with UV and mass spectrometric detection. Journal of Chromatography A, v. 1164, n. 1-2, p. 145-152, 2007.

MARTELANC, Mitja; VOVK, Irena; SIMONOVSKA, Breda. Separation and identification of some common isomeric plant triterpenoids by thin-layer chromatography and high-performance liquid chromatography. Journal of Chromatography A, v. 1216, n. 38, p. 6662-6670, 2009.

MBAMBO, B.; ODHAV, B.; MOHANLALL, V. Antifungal activity of stigmasterol, sitosterol and ergosterol from *Bulbine natalensis* Baker (Asphodelaceae). Journal of Medicinal Plants Research, v. 6, n. 38, p. 5135-5141, 2012.

MORE, Garland et al. Antimicrobial constituents of *Artemisia afra* Jacq. ex Willd. against periodontal pathogens. Evidence-Based Complementary and Alternative Medicine, v. 2012, 2012.

MUFFLER, Kai et al. Biotransformation of triterpenes. Process Biochemistry, v. 46, n. 1, p. 1-15, 2011.

NUCCI-MARTINS, Catharina et al. Antinociceptive effect of hydroalcoholic extract and isoflavone isolated from *Polygala molluginifolia* in mice: Evidence for the involvement of opioid receptors and TRPV1 and TRPA1 channels. Phytomedicine, v. 23, n. 5, p. 429-440, 2016.

PAI, Sandeep R. et al. Optimization of extraction techniques and quantification of Betulinic Acid (BA) by RP-HPLC method from *Ancistrocladus heyneanus* Wall. Ex Grah. Industrial Crops and Products, v. 34, n. 3, p. 1458-1464, 2011.

PILLAR, V. de P. Campos Sulinos: conservação e uso sustentável da biodiversidade. Ministério do Meio Ambiente, 2009.

PIRES, Fernanda B. et al. Qualitative and quantitative analysis of the phenolic content of *Connarus var. angustifolius*, *Cecropia obtusa*, *Cecropia palmata* and *Mansoa alliacea* based on HPLC-DAD and UHPLC-ESI-MS/MS. *Revista Brasileira de Farmacognosia*, v. 27, n. 4, p. 426-433, 2017.

QUÍLEZ, A. et al. Anti-secretory, anti-inflammatory and anti-*Helicobacter pylori* activities of several fractions isolated from *Piper carpunya* Ruiz & Pav. *Journal of ethnopharmacology*, v. 128, n. 3, p. 583-589, 2010.

RAMAWAT, Kishan Gopal; MÉRILLON, Jean-Michel (Ed.). *Natural products: phytochemistry, botany and metabolism of alkaloids, phenolics and terpenes*. Heidelberg, Germany:: Springer, 2013.

RIBANI, Marcelo et al. Validação em métodos cromatográficos e eletroforéticos. *Química nova*, 2004.

ROCHA, J. L. C. et al. Quantificação de salicilato de metila em quatro gêneros de Polygalaceae, por CLAE-DAD. *Quim. Nova*, v. 35, p. 2263-2266, 2012.

ROMERO, Concepción et al. Triterpenic acids in table olives. *Food Chemistry*, v. 118, n. 3, p. 670-674, 2010.

RUA, Gabriel H.; VALLS, Jose Francisco M. On the taxonomic status of the genus *Thrasypsis* (Poaceae, Panicoideae, Paspaleae): new combinations in *Paspalum*. *Phytotaxa*, v. 73, n. 1, p. 60-66, 2012.

SAEIDNIA, Soodabeh et al. The story of beta-sitosterol-a review. *European Journal of Medicinal Plants*, v. 4, n. 5, p. 590, 2014.

SALAH, A. M. et al. Estrogenic and cholinergic properties of the methanol extract of *Ruellia praetermissa* Scief. ex. Lindau (Acanthaceae) in female rats. *Phytomedicine*, v. 9, n. 1, p. 52-55, 2002.

SALEEM, Mohammad et al. Lupeol inhibits growth of highly aggressive human metastatic melanoma cells in vitro and in vivo by inducing apoptosis. *Clinical Cancer Research*, v. 14, n. 7, p. 2119-2127, 2008.

SAMY, Mamdouh Nabil et al. Chemical constituents and biological activities of genus *Ruellia*. *Int. J. Pharmacog*, v. 2, n. 6, p. 270-279, 2015.

SÁNCHEZ-ÁVILA, N. et al. Fast and selective determination of triterpenic compounds in olive leaves by liquid chromatography-tandem mass spectrometry with multiple reaction monitoring after microwave-assisted extraction. *Talanta*, v. 78, n. 1, p. 40-48, 2009.

SÁNCHEZ-QUESADA, Cristina et al. Bioactive properties of the main triterpenes found in olives, virgin olive oil, and leaves of *Olea europaea*. Journal of Agricultural and Food Chemistry, v. 61, n. 50, p. 12173-12182, 2013.

SANTIAGO, Librado A.; MAYOR, Anna Beatriz R. Lupeol: an antioxidant triterpene in *Ficus pseudopalma* Blanco (Moraceae). Asian Pacific journal of tropical biomedicine, v. 4, n. 2, p. 109-118, 2014.

SANTOS, Ana Paula; MORENO, Paulo Roberto Hrihorowitsch. Pilocarpus spp.: A survey of its chemical constituents and biological activities. Revista brasileira de ciências farmacêuticas, v. 40, n. 2, p. 116-137, 2004.

SCHMIDT, Marcella Emilia Pietra et al. Some triterpenic compounds in extracts of *Cecropia* and *Bauhinia* species for different sampling years. Revista Brasileira de Farmacognosia, v. 28, n. 1, p. 21-26, 2018.

SHANMUGAM, Muthu K. et al. Oleanolic acid and its synthetic derivatives for the prevention and therapy of cancer: preclinical and clinical evidence. Cancer letters, v. 346, n. 2, p. 206-216, 2014.

SIDDIQUE, Hifzur Rahman; SALEEM, Mohammad. Beneficial health effects of lupeol triterpene: a review of preclinical studies. Life sciences, v. 88, n. 7-8, p. 285-293, 2011.

SIEWERT, Bianka; CSUK, René. Membrane damaging activity of a maslinic acid analog. European journal of medicinal chemistry, v. 74, p. 1-6, 2014.

SILVA, D. T. et al. Larvicidal activity of Brazilian plant essential oils against *Coenagrionidae* larvae. Journal of economic entomology, v. 107, n. 4, p. 1713-1720, 2014.

SILVA, Lenise de Lima et al. Anesthetic activity of Brazilian native plants in silver catfish (*Rhamdia quelen*). Neotropical Ichthyology, v. 11, n. 2, p. 443-451, 2013.

STRAPASSON, Regiane LB et al. A new sesquiterpene lactone and other constituents of *Moquiniastrum polymorphum* subsp. *floccosum* (Asteraceae). Natural product communications, v. 9, n. 11, p. 1934578X1400901102, 2014.

VILLASEÑOR, Irene M. et al. Bioactivity studies on β -sitosterol and its glucoside. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, v. 16, n. 5, p. 417-421, 2002.

VON POSER, Gilsane L. et al. Essential oil composition and allelopathic effect of the Brazilian Lamiaceae *Hesperozygis ringens* (Benth.) Epling and *Hesperozygis rhododon* Epling. Journal of agricultural and food chemistry, v. 44, n. 7, p. 1829-1832, 1996.

XU, Xiao-Hong; SU, Qing; ZANG, Zhi-He. Simultaneous determination of oleanolic acid and ursolic acid by RP-HPLC in the leaves of *Eriobotrya japonica* Lindl. *Journal of pharmaceutical analysis*, v. 2, n. 3, p. 238-240, 2012.