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Targeting eosinophils in respiratory diseases: biological axis, emerging therapeutics and treatment modalities

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Abstract

Eosinophils are bi-lobed, multi-functional innate immune cells with diverse cell surface receptors that regulate local immune and inflammatory responses. Several inflammatory and infectious diseases are triggered with their build up in the blood and tissues. The mobilization of eosinophils into the lungs is regulated by a cascade of processes guided by Th2 cytokine generating T-cells. Recruitment of eosinophils essentially leads to a characteristic immune response followed by airway hyperresponsiveness and remodelling, which are hallmarks of chronic respiratory diseases. By analysing the dynamic interactions of eosinophils with their extracellular environment, which also involve signalling molecules and tissues, various therapies have been invented and developed to target respiratory diseases. Having entered clinical testing, several eosinophil targeting therapeutic agents have shown much promise and have further bridged the gap between theory and practice. Moreover, researchers now have a clearer understanding of the roles and mechanisms of eosinophils. These factors have successfully assisted molecular biologists to block sreenic pathways in the growth, migration and activation of eosinophils. The primary purpose of this review is to provide an overview of the eosinophil biology with a special moharis on potential pharmacotherapeutic targets. The review also summarizes promisin, eosinophil-targeting agents, along with their mechanisms and rationale for use, including those in developmental pipeline, in clinical trials, or approved for other respirator / cisorders.

Keywords: Respiratory diseases: Pulmonary; Eosinophils; Targeted therapies; Immunity; Lung

1.0 Introduction

1.1 Role of eosinophils in host defense and immunity

In 1879, Paul Ehrlich was the first person to recognize the unique ability of eosinophils to stain using acidophilic dyes (1). For many years, they were recognized as cells with end-stage effector functions in helminth infections and tissue damage (1). However, the plethora of clinical studies carried out in previous years have helped establish the crucial role of eosinophils in host defense, allergic inflammation, innate and adaptive immunity (1,2). Importantly, the interaction of eosinophils with B-cells allo vs them to process antigens, stimulate T-cells and induce humoral responses (1). Inflammator, and adaptive responses can also be initiated by eosinophils through their bidirection at a treactions with dendritic cells (DCs) and T-cells (3). Activated eosinophils are able to release a large assortment of newly synthesized as well as pre-formed mediators, such vs cytotoxic granule proteins, cytokines, chemokines, and lipid mediators which contributes to the various activities of eosinophils in flammatory and infectious responses (4).

Eosinophils are now recognize a regulatory cells with the proven ability to influence and enhance local inflammation, estead of just simply effector granulocytes with cytotoxic activities (5). The immunor on latory role played by eosinophils include mediating aluminum hydroxide-induced B-cell, riming, acting as an antigen-presenting cell (APC) for T-cells, influencing T-cell differentiation (i.e. Th1 or Th2), and recruiting T-cells, DCs and macrophages to inflammatory .^{ites} in the host (6,7). Eosinophils were shown to regulate DCs and Th2 pulmonary impuner response following an allergen challenge in mouse models (8). Moreover, a unique incuon of eosinophil is the suppression of DC-mediated Th17 production (8). Thus, eosinophils are important modulatory cells to maintain the equilibrium between DC-mediated Th2 and Th17 signaling pathways following an allergen exposure and subsequently, the allergic airway inflammation progression (8). Recently, it was discovered that eosinophils are necessary for the long-term preservation of plasma cells in the bone marrow and eosinophil depletion induces bone marrow plasma cells apoptosis (9). Wu et al., also reported that eosinophils regulate glucose homeostasis by preserving adipose tissue alternatively-activated macrophages (AAMs) via the secretion of IL-4 cytokine (10). As a regulator of innate immune response, eosinophils are also responsible for apoptotic cell clearance in the thymus (11).

Similar to other granulocytes, eosinophils undergo development and differentiation in the bone marrow (12). Upon maturation, eosinophils are distributed in various organs in the body, such as blood, lung, uterus, thymus, spleen, mammary gland, adipose tissue, and gastrointestinal tract (GI), to carry out their physiological functions under homeostasis (6,12). The recruitment of mature eosinophils from the systemic circulation to the inflammatory sites occurs following the overexpression of eosinophil-specific chemokines in response to stimuli (12). Interleukin-5 (IL-5) is the most important cytokine responsible for eosinophil differentiation, priming and survival and its main sources of production are type 2 T-helper (Th2) cells and type 2 innate helper lymphoid cells (ILC2) (6,12).

1.2 Cytokines, chemokines, receptors, and surface marker

A broad variety of cytokines, lipid mediators and other major molecules are expressed and secreted by eosinophils (Figure 1). These molecules are stored in eosinophilic granules and rapidly secreted in response of a specific atimuli, hence altering the external microenvironment and cellular functions. Locinophils are distinguished from other lymphocytes, such as T-cells and B-cells, due to their ability to store and rapidly release preformed cytokines within minutes in response to stimuli (13,14). Besides pre-formed cytokines, eosinophils can carry out *d'a novo* synthesis and secretion of other immunological factors (14). It was hypothesized into the binding of soluble N-ethylmaleimide sensitive factor attachment protein recepore (SNARES), which are part of the membrane fusion complexes, regulates the final steps of cytokine secretion from eosinophilic crystalloid granules and secretory vesicles (14).

Eosinophils are a major sou ce of IL-5 cytokine, which is important for its differentiation, survival and chemotax. (15). Other Th2 immunomodulatory cytokines secreted by eosinophils include IL-4 and IL-13 (13). Besides Th2 cytokines, other cytokines with Th1 and regulatory capacities released by eosinophils include IL-6, IL-10, IL-12, TNF- α , TGF- β , and IFN- γ (13). IL-6, IFN- γ , and TNF- α causes tissue damage through their proinflammatory actions whereas TGF- β contributes to airway remodeling via its role in epithelial changes, subepithelial fibrosis and microvascular changes (16,17). As for the lipid mediators, eosinophils releases a large quantity of prostaglandins, leukotrienes, and plateletactivating factors (PAF) (16). Eosinophils express a wide variety of receptors and molecular surface markers on their cell surfaces, including IL-5R α , prostaglandins (CRTH2), CCchemokine receptor (CCR)-3, sialic acid-binding immunoglobulin-like lectin 8 (SIGLEC-8),

leukotriene B4 receptors (IL-4R, IL-5R, IL-33R, IFN- γ R, TGF- β R, CCR1, CCR3, CCR4, and TSLPR), PAF-receptor, F α R, F α PR, and pattern-recognition receptors (PRRs) (18). The most prominent cytokine receptor present on human and mice eosinophils is IL-5R α where the main receptors which define the distinct biology of eosinophils are CCR3 and SIGLEC-8 (19). Human eosinophils uniquely express SIGLEC-8 and the binding of antibodies or glycan ligands with this structure leads to apoptosis of eosinophils (20). The PRRs families expressed by human eosinophils include Toll-like receptor (TLR) family (TLR1-5, TLR7, TLR9), C-type lectin receptor (CLR) (Dectin-1), nucleotide-binding oligomerization domain (NOD)-like receptors (NOD1, NOD2), and receptors for advanced glycation end-products (RAGE) (19,21,22). Interaction with eosinophilic PRRs m.⁴uces eosinophil survival, oxidative burst, adhesion system activation, and mediator release (21).

Integrins are cell surface proteins commonly foun ¹ on immune cells which functions to merge the intracellular and extracellular domains of the immune system. Eosinophils express several integrins, such as $\alpha 4\beta 1$ (CD49d/CD29), $\alpha 6\beta 1$ (CD49f/ CD29), $\alpha L\beta 2$ (CD11a/CD18), $\alpha M\beta 2$ (CD11b/CD18), $\alpha X\beta 2$ (CD11c/CD18), $\alpha D\beta 2$, and $\alpha 4\beta 7$ (23). These integrins interact with not only their respective ligands, but also vascular adhesion molecule 1 (VCAM-1), intracellular adhesion molecule 1 (ICAM-1), periostin, laminin, and fibronectin located in the extracellular matrix or other cells (23).

Furthermore, blood eosinophils constitutively expresses selectins, including P-selectin glycoprotein ligand-1 (PSGL-1, CD162) and L-selectin (CD62L) (24). A high level of surface P-selectin results in excitation β_1 integrin activation *in vivo* and thus, stimulates eosinophilic adhesion to VCAM-1 and migration to the airway (24).

Notch receptors and Notch ligands are also expressed on human blood eosinophils, which serve as important components for eosinophil autoregulation (25). A study showed that granulocyte-macrophage colony-stimulating factor (GM-CSF) influences the expression and activation of Notch molecules and thus, affects the activity and survival of eosinophils (25).

1.3 Eosinophilic granule proteins

Eosinophils store pre-formed enzymatic and non-enzymatic cationic proteins in large secondary granules which are selectively secreted in response to specific stimuli (26). The release of these granule proteins have typically be considered as the primary effector mechanism of eosinophils against specific parasites and in allergic inflammation (27). The antibacterial effect of eosinophils *in vivo* is specifically mediated thorough the release of cationic secondary granule proteins (28). This dominant population of cytoplasmic crystalloid

granules containing cationic proteins distinguishes eosinophils from other leukocytes (29). The four notable granule proteins present in human eosinophils include eosinophil peroxidase (EPO), major basic protein (MBP), and eosinophil-associated RNases (EARs) which are eosinophil cationic protein (ECP) and eosinophil-derived neurotoxin (EDN) (19,29).

EPO is the cationic protein present most abundantly in the matrix of the crystalloid granules (16). EPO contributes to eosinophil function by generating reactive oxidizing species (ROS) as well as acting as a direct toxin against mammalian cells and parasites (16). The promotion of oxidative stress by EPO results in cell apoptosis and necrosis (19,30). A study carried out by Panagopoulos *et al.*, reported that EPO and other peroxidases play a part in angiogenesis in addition to cellular proliferation, migration, and invasion (31). Ochkur *et al.*, also demonstrated the use of a sensitive and specific E⁺JS/4 for the detection of EPO (32).

MBP exerts direct toxicity which contributes to its function in altering membrane permeability and enzyme functions in mammalian cella inicrobes, and helminths (16,19). As for its role in the pathophysiology of asthma, Mbr induces bronchoconstriction and epithelial tissue damage (16). MBP was also reported to have pro-angiogenic effects *in vitro* and *in vivo* (33).

ECP and EDN are part of the ibonuclease A superfamily (16). EDN demonstrates cytotoxicity, neurotoxicity, and ant vi. ¹ activity (single-stranded RNA virus), but it exerts low toxicity towards mammalian cells and parasites (16). Tsuda et al., recently reported that EDN stimulates matrix metallo proteinase 9 (MMP-9) production from nasal epithelium, thus indicating its potential role in the pathogenesis of eosinophilic chronic rhinosinusitis (ECRS) (34). Conversely, ECP snow distinct toxicity towards a large variety of bacteria, helminths, single-stranded RNA vn uses, and host tissues (16). ECP exerts its toxic effects through formation of pores in target membranes (19). Besides its cytotoxic effects, ECP also stimulates airway mucus hypersecretion and mast cell degranulation, as well as suppresses Bcell immunoglobulin synthesis and T-cell proliferative responses (19). EDN is known to possess multifunctional properties and may contribute to innate immunity by either killing or inactivating viral invaders (35). In 1998, the potential of EDN as an antiviral agent against respiratory syncytial virus was demonstrated under in-vitro settings (36). Domachowske and colleagues demonstrated a dose-dependent decline in the infectivity of respiratory syncytial virus B after the introduction of eosinophils into the viral suspension (36). Interestingly, the inclusion of ribonuclease inhibitor was found to reverse the antiviral effect observed by eosinophils, suggesting a role of EDN (eosinophil secretory ribonuclease). The suggestive

antiviral role of EDN was confirmed when a 40-fold decrease in infectivity, after the introduction of EDN into the suspension of respiratory syncytial virus B was obtained. Importantly, the inactivated form of EDN was reported to have no antiviral effects. Furthermore, the observed antiviral activity of EDN was found to be the direct ribonucleolytic destruction of extracellular virions. These findings collectively suggest that EDN may also be used as a therapeutic agent for the management of respiratory syncytial virus (36). Furthermore, Rugeles *et al.*, reported that EDN was responsible for the majority of anti-HIV-1 activity shown by alloantigen-stimulated factors in the supernatant of mixed lymphocyte reactions (37).

Additionally, another important protein found in primary granules of eosinophils is galectin-10, or Charcot-Leyden crystal protein. Galect n-1) was shown to be strongly associated with sputum eosinophilia and hence, can function as a potential alternative biomarker of eosinophil-associated airway inflammation (38).

1.4 Degranulation

The extracellular release of eosi. or all granule proteins is termed as degranulation. Piecemeal degranulation (PMD) is generally acknowledged as the most prevalent manner of eosinophilic degranulation (16,19) 'During PMD, the contents of the granule are selectively packaged into vesicles which are unnsported across the cytoplasm and fused with the cell membrane in order to extrace unapported across the granule proteins at the cell surface (29). Cytokines and chemokines which induces PMD are IFN- γ , CCL-11 (eotaxin-1), and TNF- α (16,39). Following the release of its granule proteins via PMD, the eosinophil remains fully functional and responsive to other stimuli. An example of PMD is the release of IL-4 from eosinophils stimulated with eotaxins, whereby secretory vesicles deliver IL-4/IL-4R α complex, which is initially formed inside the granule membrane, to the cell surface and subsequently released into the extracellular space (19).

Another mechanism of degranulation observed in eosinophils is associated with cytolysis, in which the eosinophil undergoes cell lysis in a manner that morphologically differs from either apoptosis or necrosis, thus releasing intact, cell-free, membrane-bound granule proteins which are fully competent (29). The manner of cell death in eosinophilic cytolysis is termed as extracellular trap cell death (ETosis), whereby the nuclear membrane disintegrates and DNA de-condenses into the surrounding cytoplasm (16). Cytolysis is

acknowledged as a common method for the release and deposition of cell-free eosinophilic granules in eosinophil-associated disorders (29).

Lastly, eosinophil degranulation can also occur by classical exocytosis, in which the fusion of intracellular granules with the plasma membrane precedes the extracellular release of the total granule contents (40). This mechanism of degranulation does not occur commonly in eosinophilic diseases, except in the presence of parasitic helminths or specific fungi (40).

Although degranulation is the primary mechanism in which eosinophils exert their function, this process does not occur during transit in blood circulation. Instead, granules are commonly released when the eosinophils arrive at the site of intermation (41).

1.5 Hypereosinophilic syndromes

Hypereosinophilic syndromes (HES) comprise of a heterogenous collection of disorders with common features of elevated blood and tissue eosinophils and tissue damage (42,43). The clinical manifestations are variable at J may include any organ system, but most commonly the skin (42). Since 1975, the definition of HES includes 3 criteria: (i) blood eosinophil count $\geq 1500/\text{mm}^3$ for >6 menths (or mortality within 6 months associated with signs and symptoms of hypereosinophilic disease), (ii) insufficient evidence of other causes of eosinophilia (e.g. parasite, allergel). and (iii) presumptive signs of organ involvement (e.g. GI dysfunction, heart failure, cer aral nervous system impairment, weight loss, or fever) (44). However, there are several sho.*comings with these definitions of HES. Marked eosinophilia is typically caused by kelm ath infections, but other non-infectious causes include malignancies, drug react ons. immunologic, allergic and inflammatory diseases (45).

2.0. Eosinophils and ast. ma

2.1 Eosinophils in the Pathophysiology of Asthma

2.1.1 Eosinophilic asthma

Asthma is inflammatory respiratory airway an disorder outlined by hyperresponsiveness obstruction, (AHR), airway mucus hypersecretion, airway inflammation, tissue damage and airway remodeling (16,46-48). According to a study conducted by the Global Burden of Disease (GBD), it was estimated that around 339.4 million people globally were affected by asthma in 2016 (49). Eosinophils have been closely related to the pathophysiology of asthma, as reported from the elevation of eosinophil numbers in peripheral blood and bronchoalveolar lavage fluid (BALF) of asthmatic patients

(16,46). In a certain percentage of patients, eosinophilic airway inflammation represents the occurrence of bronchial asthma (50). Eosinophils are pro-inflammatory granulocytes with major contributions to the inflammatory responses of asthmatic patients, especially in T2high asthma phenotypes such as severe eosinophilic asthma, by releasing inflammatory mediators to trigger an inflammatory cascade as well as exerting toxic effects directly on host tissues (51). Severe asthma is a heterogenous disorder as different phenotypes exists, such as eosinophilic asthma. Eosinophilic asthma can be clinically characterized by its severity and frequent exacerbations (50). Additionally, other features of eosinophilic asthma includes sputum and airway eosinophilia, elevated blood eosinophil count, adult-onset, and involvement of nasal polyps-associated chronic rhinosinusitis in 50% of the patients (51,52). Conversely, non-eosinophilic asthma can be characterized by low eosinophil counts, with the dominant inflammatory cell type being neutrophils and mixed granulocytes, or very few inflammatory cells (51). Generally, the higher the degree of eosinophilia, the greater the disease severity and exacerbation frequency in eostheranilic asthma (46). Eosinophilic and paucigranulocytic asthma (asthma with very fev 1) Pammatory cells) are among the dominant inflammatory asthma phenotypes.

2.1.2 Eosinophil recruitment, migration, development, and survival

In eosinophilic asthma, which is also known as Th2 asthma, eosinophils present as the hallmark as seen in their elevate.⁴ numbers (Figure 2) (53). The exposure of airway epithelium to allergens or a tigens triggers an immunological cascade which attracts eosinophils to the airway. by Th2 cytokines and chemoattractants (16). Th2 cytokines secretion contributes to the in crease in eosinophil counts in the BALF as they are responsible for eosinophil recruitment, migration and survival (16). The Th2 cytokines which induces eosinophil migration to the airways include IL-4, IL-5 and IL-13, which are produced by activated Th2 lymphocytes and ILC2s (16,53). Among them, IL-5 and the chemokine eotaxin-1 (CCL11) are the primary mediators for the release of bone marrow eosinophils and trafficking of eosinophils to the lungs (16,51). Eotaxin is produced by allergen-challenged endothelial and epithelial cells and enhances eosinophil migration by binding to CCR3 expressed on its surface (16). Besides stimulating eosinophil migration, IL-5 is also an important mediator of eosinophils differentiation, development, activation and survival (52,54). Other major sources of IL-5, other than Th2 cells and ILC2s, are CD34+ progenitor cells, mast cells, invariant natural killer (NK) T-cells, and eosinophils themselves (55). In addition to the regulation of eosinophil numbers, IL-5 produced by ILC2s also regulates their

circadian cycling (55). IL-4 and IL-13 do not directly mediate eosinophil trafficking; IL-4 sustains eosinophil migration by inducing B-cell isotype switching to immunoglobulin (Ig)E for the development of Th2 lymphocytes whereas IL-13 stimulates eotaxin production (53). IL-4 also induces the expression of VCAM-1 and eotaxin by epithelial cells which enhances eosinophil migration to the site of allergic inflammation (16). study by Beckert et al., provides evidence on the effects of the Th2 cytokines, either alone or in combination, on eosinophils in a mouse model (56). C57BL/6 mice were intranasally administered equimolar amounts of single agent or combination IL-4, IL-5 and IL-13. The results demonstrated that IL-4 and IL-13 were correlated with airway eosinophilia, progression of airway hyperresponsiveness, and goblet cell metaplasia (56). However, IL-4 demonstrated weaker effects than IL-13 and no synergism was observed when hese two cytokines were administered in combination (56). After the administration of IL-5, it was observed that the eosinophil count in bone marrow and lung tissues were increased but no structural changes of the eosinophils were observed (56). The combine¹ administration of IL-5 and IL-13 significantly increased the number of lung, a way, blood and bone marrow eosinophils, whereas IL-5 and IL-4 combined only increas 'd eosinophils count in lungs and bone marrow (56).

Besides Th2 cells, the danloged epithelial cells also contributes to eosinophils recruitment through the release of 'L 25, IL-33, and thymic stromal lymphopoietin (TSLP) (16). These cytokines activate IL 22 from the innate immune system to secrete IL-4, IL-5 and IL-13 (16). Additionally, IL-32 and granulocyte-macrophage colony-stimulating factor (GM-CSF) contributes to eosinophils differentiation and migration as well as mediate their survival in the airways (16). I -35 directly stimulates eosinophil differentiation from CD117⁺ hematopoietic progenite cells and contributes to the exacerbation of eosinophilic inflammation by elevating eosinophil, macrophage, lymphocyte, IL-13, TGF- β , CCL3, CCL17, and CCL24 levels in the airways (57). IL-18 was also recently identified as an important cytokine for the production, differentiation, and maturation of (CD101⁺CD274⁺) pathogenic eosinophils (54).

Fanat *et al.*, researched the effects of cells sourced from human airway smooth muscle (HASM) on eosinophils (58). In this study, peripheral blood progenitor cells collected from atopic asthmatics and control subjects with no atopy were cultured together with supernatant collected from HASM cells culture. At the end of the study, it was observed that HASM cell-derived cytokines stimulated eosinophil differentiation *via* the p38 mitogen-activated protein kinase (MAPK) pathway but not the src kinase (srcK) pathway (58). The cytokines released

by HASM cells were identified to be IL-5 and GM-CSF as eosinophil differentiation was inhibited by anti-IL-5 and anti-GM-CSF blocking antibodies (58). Thus, the researchers concluded that HASM cells have the ability to modulate differentiation and maturation of eosinophils from precursor cells including immature eosinophils, that may lead to eosinophilic inflammation and airway remodeling in severe asthmatics (58). However, haemopoietic progenitor cell migration and adhesive response *in vitro* are not influenced by HASM cells. (58).

Lung dendritic cell (DC) subsets also play a role in initial eosinophil recruitment to the site of allergen challenge. A specific sub-type of DCs in the lung, i.e., CD24CD11b DC2s, produce NO synthase which induces CCL17 and CCL2? on lung cDC1s to attract eosinophils for initial eosinophil infiltration. After the allergen challenge, eosinophil recruitment is inhibited by lung CD24 cDC2s through 'GF β 1 secretion (59). Hence, the impaired expression of CCL17 and CCL22 lead to the response seen during the late phase reaction. It was suggested that different lung APCs secrete specific soluble factors during the memory stage of chronic asthma after an allergen challenge, thus regulating lung cDC1-mediated eosinophil recruitment differently (52).

Several lipids, such as prostagion d_{ns} and leukotrienes, are key mediators of the inflammatory response by acting cs chemoattractant(s) of eosinophils. For example, leukotriene E4 and prostaglandin D2 (PGD2) induce eosinophil migration to the airways (16). PGD2 acts by binding to CN H2 receptor (DP2) in eosinophils which induces its recruitment and activation (16).

Nitric oxide (NO) synthesized from epithelial cells and vascular endothelial cells by inducible nitric oxide synthese enzyme is also responsible for the recruitment of eosinophils in addition to its function as a mediator of inflammation, and this correlates with the level of exhaled NO, which is a biomarker of asthma (16).

Following the recruitment phase comes the effector phase. Autocrine secretion of IL-5 by activated eosinophils enables them to survive within tissues in their own capacity as the secreted IL-5 inhibits apoptosis (16,60). Moreover, the adherence of eosinophils to fibronectin stimulates the secretion of other pro-survival mediators, such as GM-CSF and IL-3, by activated eosinophils and hence, promotes their own survival (16,60). The additional effects of these cytokines enable eosinophils to persist in the airways for long periods and prolong their effects. Eosinophils in inflamed tissues are also activated by antigen-specific IgGs and IgAs, but not IgE, as reported by a study conducted on the effects of antigens and antigen-specific immunoglobulins on eosinophil function (61).

2.1.3 Airway hyperresponsiveness (AHR) and Mucus Hypersecretion

AHR occurs when there is a dysregulation of airway homeostasis. Allergen exposure causes events such as airway epithelium damage, mucus hypersecretion and ASM proliferation which leads to homeostasis imbalance (62). Eosinophils are highly involved in the development of these events through the release of its granule cytotoxic proteins via cytolysis, such as MBP, EPO, ECP, and EDN (51,53). The release of these granule proteins was shown to be triggered through chemokines, including ota in-1 and interferon-gamma (IFN- γ) (53). MBP presents as two type of homologs, which are MBP-1 and MBP-2, with MBP-1 having a significantly alkaline pH and exerting direct cytotoxic effects on the host cells (51). MBP-1 also has a greater potency in hutanine induction and leukotriene C4 release from basophils as compared to MBP-2 (15). MBP and EPO were reported to cause AHR, although ECP and EDN did not (46) L was also reported that MBP induces bronchial hyperactivity by stimulating the release of istamine from mast cells and basophils (46,51). MBP can also cause bronchoconstriction by blocking acetylcholine receptor, M2, which is a negative feedback regulator for $a' et_{f}$ holine (53). Moreover, the combination of EPO, halides and hydrogen peroxide was shown to stimulate the release of mast cell mediators (51).

In addition to cytotexic sationic proteins, eosinophils secrete several cytokines which can contribute to the h.llm.rks of asthma. For example, IL-13 is a cytokine released by eosinophils which can schance the differentiation of goblet cells to promote mucus hypersecretion, as well as cause AHR. Moreover, Th2 cells and ILC2 also produces IL-13 (46).

Additionally, eosinophils also produce lipid mediators in eosinophil lipid bodies, such as leukotrienes, which can also cause AHR and mucus hypersecretion (46). Eosinophilic leukotrienes stimulate bronchoconstriction and activation of basophils and mast cells, which further sustains the ongoing inflammation via the secretion of histamine, prostaglandins, and additional leukotrienes (53).

The role of eosinophils in airway remodeling is further reinforced when mouse studies showed that mice with congenital eosinophil deficiency did not experience collagen and smooth muscle deposition in the airways (46).

2.1.4 Tissue damage and Airway remodeling

Eosinophils cause persistent inflammation in the airways which causes continuous damage to the airway walls (53). During the reconstruction of the damaged airway, airway remodeling occurs as the basement membrane thickens and fibrosis occurs due to airway smooth muscle proliferation, goblet cell hyperplasia, extracellular matrix (ECM) proteins deposition, and new blood vessels formation (angiogenesis) (16,46,53). Airway remodeling is commonly associated with severe asthma phenotypes (63).

Tissue damage occurs as a result of the cytotoxic effects of the eosinophilic granule proteins. The granule proteins exhibit varying strengths of torricity when tested *in vitro*. MBP, EPO and ECP exert toxicity to several tissues, including bronchial epithelium, skin, heart and brain, whereas EDN only exhibits marginal toxicity (51). MBP, EPO and ECP were found to cause damage to epithelial cells *in vitro* when equivalent concentrations as found in asthmatic patients were administered (16,53). ECP is able to bind to and alter the permeability of cell membranes in the airway (4 σ)

The process of airway remodeling is viso contributed by TNF- α , as well as IL-1- β according to recent studies, which activalies cosinophils and stimulates the secretion of matrix metalloproteinase-9 (16,53). TNF- γ and IL-1- β are also associated with persistent eosinophils recruitment (53). TGF- β is a growth factor secreted by eosinophils which can promote airway remodeling. Bronchial biopsies from asthmatic patients have shown that eosinophils are the primary source TGF- β (46). Studies have shown that TGF- β mRNA expression was upregulated in source as a chemoattractant for fibroblast and stimulates fibroblast proliferation and differentiation into myofibroblasts and smooth muscle cells (46,53). Additionally, TGF- β upregulates the synthesis of collagen and glycosaminoglycans leading to ECM production and resulting in tissue remodeling of the airways (46,53).

Besides granule proteins and cytokines, eosinophils also secrete reactive oxidant species (ROS), such as hydrogen peroxide, superoxide anion and hydroxyl radicals, which contributes to airway inflammation and airway remodeling by damaging the cells and tissues of the airways and inducing the fibroblast hyperplasia (16,51,53). EPO stimulates the production of cytotoxic ROS by catalyzing the oxidation process of halides and thiocyanate (46).

Zagai *et al.*, stated that conditioned media (CM) collected from cultured human peripheral eosinophils and eosinophil cationic protein (ECP) extracted from human peripheral eosinophils significantly stimulated the migration of human lung fibroblasts *in vitro* in a concentration- and time-dependent manner (64). Thus, it was suggested that eosinophils affect the fibrotic response in asthmatics. As airway remodeling is associated with fibroblast recruitment, it was proposed that the stimulation of fibroblast migration by ECP may be an important mechanism in the remodeling process of extracellular matrix leading to airway fibrosis in asthmatics.

2.1.5 ASM proliferation

The co-culture of eosinophils isolated from ast ma patients and ASM cells demonstrated an enhancement of ASM proliferation, which is inhibited by the presence of leukotriene antagonist, montelukast (46). A mutual relationship exists between eosinophils and ASM cells as ASM cells are able to produce eosinophilic cytokines (46).

Based on the study done by Halwani *et at*, cysteinyl leukotrienes (CysLTs) released from eosinophils when they are in direct co. tact with airway smooth muscle (ASM) cells enhanced ASM proliferation (65). The ichuir rement of direct cell contact between eosinophils and ASM cells to trigger CysLTs release were further confirmed when the administration of anti-adhesion molecule antibodies resulted in the inhibition of ASM proliferation (66). Additionally, it was proven that the increase in ASM proliferation was dependent on CysLTs released from eosinophils instead on ECM proteins as ASM proliferation was inhibited when leukotriene receptor antagenist (Montelukast) was co-cultured with eosinophils and ASM cells. The researchers concluded that airway remodeling in asthmatic patients can be attributed to eosinophil chrived CysLTs which function to increase ASM mass through the enhancement of ASM proliferation.

2.1.6 Eosinophilic pro-inflammatory cytokines and chemokines

Besides the direct cytotoxic effects of eosinophilic granule proteins on tissues, eosinophils produce and secrete a variety of pro-inflammatory mediators which contributes to the pathophysiology of asthma. For example, eosinophils produce a variety of Th2 cytokines in the airways of asthmatics, including IL-4, IL-5, and IL-13 (51). The recruitment of leukocytes to the airways is also regulated *via* the secretion of eosinophilic chemokines, such as CCL3, RANTES (CCL5), and eotaxin (CCL11) (51). Alternatively, neutrophil recruitment is induced by IL-8 and GM-CSF, which are also expressed by eosinophils following an

allergen challenge (51). Eosinophil activation is also mediated *via* Th1 cytokines released by eosinophils themselves, which is IFN- γ (51). Thus, it is evident that eosinophils exhibit pleiotropic effects, which includes destruction of host tissues, regulation and potentiation of inflammatory pathways, as well as host defense. The severity of asthma symptoms in patients is worsened by these eosinophilic functions. Therefore, therapies which target eosinophils can be a potential intervention for asthma and other eosinophilic inflammation-associated diseases.

2.2 Eosinophils as Biomarkers of Asthma

Heterogeneity exists in asthma with regards to its underlying pathophysiology, clinical signs and symptoms, and treatment response. The use of merely clinical and physiological assessment has become inadequate in the accurate prediction of the underlying mechanism of the disease or the treatment response. Thus, additional testing for biomarkers in conjunction with examination of clinical signs road symptoms of the patient will be able to help identify asthma phenotypes and endotypes, anticipate the disease progression and its prognosis, as well as enhance the precision derapy for asthma (67). Eosinophil counts have become a useful biomarker in the assessment of asthma.

Szefler *et al.*, reported that the measurement of total eosinophil counts and sputum eosinophils as biomarkers for as hnew are recommended as supplemental outcome measures (68). Supplemental outcome measures are defined as outcomes of asthma for which standardized definitions and specific methods of measurement exists, and their validity has been proven but they remain as an optional inclusion in funded clinical asthma research (68). The eosinophil measurements are obtained through complete blood counts. In addition to sputum eosinophils, nasal discharge eosinophils with asthma symptoms may also be a predictive factor for persistent asthma (69). As such, testing for nasal discharge eosinophils with asthma symptoms increases with age (69).

The severity of sputum eosinophilia in asthma can be assessed using various biomarkers. However, the biomarkers' diagnostic accuracy might vary between different asthma phenotypes. Westerhof *et al.*, carried out an investigation to determine the accuracy of several biomarkers for the detection of sputum eosinophilia in different phenotypes of adult asthma, such as total IgE, area under curve (AUC), and exhaled nitric oxide fraction (FeNO) (70). Among the different asthma phenotypes of adult patients used for comparison were severe and mild, obese and non-obese, atopic and non-atopic, and smoking/ex-smoker

and non-smoking asthma patients. The findings showed similarities in AUCs for blood eosinophils and FeNO among the various asthma phenotypes whereas total IgE showed higher accuracy in the detection of sputum eosinophilia in non-obese and non-atopic asthmatics as compared to obese and atopic asthmatics (70). The researchers concluded that the measurement of blood eosinophils and FeNO as sputum eosinophilia biomarkers, irrespective of asthma phenotype, showed superior diagnostic accuracy as compared to total IgE (70). A separate study also showed that blood eosinophils can most accurately identify sputum eosinophilia in patients with varying severities of asthma, such as mild, moderate or severe asthma (71). Thus, eosinophils found in blood circulation can be used to facilitate individualized asthma treatments.

Nevertheless, some studies have shown that cospile statistically significant associations, blood eosinophil and neutrophil counts, FeN D ard IgE levels, FEV1 percentage and age are poor surrogate biomarkers, both singularly and in a combination, for predicting sputum eosinophil and neutrophil percentages (72).

Phenotyping based on airway inflammation severity is often applied to asthmatic patients. According to a study done by W 1sh *et al.*, the relationship between rates of exacerbations and the phenotype of severe asthma was investigated based on longitudinal measures of sputum eosinophils and noutrophils (73). The results demonstrated that asthmatic patients with persistent eosinophilic parotype experienced shorter intervals before the first exacerbation and an increased risk of exacerbation over a 1-year period as compared to those with non-eosinophilic phenotype. However, no observable changes were noted in time to first exacerbation or exacerbation, risk among neutrophilic phenotypes.

A cohort study (arri d out in UK also reported that asthmatic patients with blood eosinophil numbers >40.) cells/ μ L experienced a worsening in asthma control and more severe exacerbations, as defined by the increased use of asthma relievers, asthma-related hospitalization, oral corticosteroids use, or antibiotics prescriptions (74).

Petsky *et al.*, reported, in a systemic review, that carrying out therapy adjustment according to airway eosinophilic markers, such as sputum eosinophil counts and exhaled nitric oxide, significantly decreased the risk of asthma exacerbations (75). However, no significant impact was observed with regards to dose of daily inhaled corticosteroid, asthma control or lung function.

Periostin is a proposed novel biomarker of eosinophilic airway inflammation. A specific asthma subset as characterized by the expression of Th2 cytokine-inducible genes in bronchial epithelial cells was recently discovered. Periostin is included in this gene signature

which is present in about 50% of asthma patients and is associated with eosinophil-mediated airway inflammation. In a study done by Jia et al., an assay for peripheral blood periostin protein was developed to investigate its potential as a biomarker in order to designate selective therapies for specific groups of asthma patients (76). The patients used for sample collection were asthmatics with persistent symptoms in spite of inhaled corticosteroid administration at maximum doses. The results showed that asthma patients with evident eosinophilic airway inflammation demonstrated a significant increase in levels of serum periostin as compared to those with minimal eosinophilic airway inflammation (76). Serum periostin levels also proved to be a more useful predictor of eosinophilia in airways than other indices, such as peripheral blood eosinophil, IgE and FeN. levels (76). The researchers concluded that the occurrence of eosinophilic airway inflar may lon can be predicted using periostin as a systemic biomarker in asthma patients and thus, this can potentially be a utility for selecting patients for emerging asthmatic therapeutics, such as ICS and biologics, targeting Th2 inflammation (76). Simpson et al., 20 reported that serum and sputum periostin levels have high correlation with statum eosinophil counts (77). Periostin concentrations were also found to be higher i. serum than in sputum. However, the ability of periostin levels to predict the presence of a sinophilic asthma was modest.

Besides periostin, eosinophil peroxidase (EPO) level are also reported in several studies to have correlation with sprare eosinophil levels. Rank *et al.*, carried out a study to compare nasal, pharyngeal, and putum EPO levels with induced sputum eosinophil percentage and found that there was a strong association between nasal and pharyngeal EPO levels and the percentage of ecsinophils of induced sputum. Another study carried out by Nair *et al.*, concluded that EPO levels assayed by ELISA was an appropriate surrogate marker of eosinophils and for eosinophil degranulation in sputum of respiratory patients (78).

ILC2s are recognized as another novel biomarker for eosinophilic airway inflammation. In an investigation carried out by Liu *et al.*, the inherent diagnostic ability of ILC2 was compared to standard biomarkers of eosinophilic asthma, including age, sex, BMI, number of blood eosinophils, IgE, and FeNO (79). The results concluded that ILC2 levels were significantly elevated in patients with eosinophilic asthma and the ILC2 percentage was the most significant component of eosinophilic airway inflammation as compared to the other biomarkers (79). Moreover, ILC2 percentage showed high sensitivity and specificity in differentiating patients with eosinophilic asthma from asthma patients without eosinophilic inflammation (79). Thus, ILC2 is a prospective substitute biomarker for eosinophilic airway inflammation in mild-moderate asthma patients and subsequently, can be used to select

patients for beneficial asthmatic therapies targeting Th2 inflammation. There is a six-gene signature paper that predicts exacerbations in the severe asthma patients, this six-gene sig has been proposed as a marker for severe asthma exacerbations, and more recently in COPD (80,81).

Biomarkers of eosinophilic inflammation appears to differ between severe asthmatic patients with concomitant obesity and normal asthmatic patients. In a study done by Desai *et al.*, obese severe asthmatic patients demonstrated significant elevations in submucosal eosinophils and sputum IL-5, however no changes were observed in sputum eosinophils (82). Hence, supplementary investigations should be performed to determine if eosinophil-targeted therapies or diet and lifestyle modifications are more effective in providing anti-inflammatory effects in obese patients with asthma.

2.3. Bacteria and eosinophils

A greater diversity of microorganisms wh altered compositions are known to be present in asthmatic patients, for example, or Proteobacteria and less Bacteroidetes as compared to healthy subjects (83). Hence, it was proposed that the composition of airway microbiome has correlations with the type of airway inflammation (84,85). A study carried out by Sverrild et al., revealed t'at 'he level of eosinophilic airway inflammation has correlations with the varying airv ay microbiome constituents in asthma patients (83). A more abundant bacterial profile was obtained from patients with the lowest eosinophil counts, for example, a greater proportion of Neisseria, Bacteroides, and Rothia species and a lower proportion of Sphingom may Halomonas and Aeribaccilus species as compared to asthma patients with greater eo inophil counts and healthy subjects (83). The type of bacterial species also affects eosinophil functionality differently. A study conducted by Hosoki et al., showed that Staphylococcus aureus (SA) stimulated the release of EDN in a dose-dependent manner, but Haemophilus influenzae (HI) and Prevotella sp. (PS) did not (86). SA, HI and PS all significantly enhanced superoxide generation, but SA had a greater effect which significantly induced a greater eosinophilic TNF-a production as compared to either HI or PS (86). Conversely, HI and PS induced IL-10 production more strongly than SA (86). Thus, it was concluded that SA may be associated with exacerbation of eosinophilic inflammation in asthma whereas HI and PS may be involved in its inhibition.

2.4 Drugs targeting eosinophils in the treatment of asthma

Asthma is conventionally treated with beta-agonists, anti-cholinergics and inhaled corticosteroids (ICS). In addition, biologics are commonly used as an alternative to treat severe and refractory asthma in which conventional treatments show limited efficacy or intolerable side effects. Monoclonal antibodies, such as those targeting IL-5, IL-5R α , IgE and IL-4R α , have shown favourable outcomes in clinical trials for the treatment of severe, uncontrolled asthma (87). Patients with severe eosinophilic asthma were also successfully managed with biologics which target IL-5, IL-4, IL-13, and IgE. This is because local eosinophilopoiesis is hypothesized to be the predominant mechanism that results in on-going airway eosinophilia and steroid requirements in patients with severe asthma, and these agents either directly or indirectly targets eosinophils which leads to improved asthma outcomes, such as reduced exacerbations and steroid-sparing effects. I mong these biologic therapies, anti-IL-5 drugs, including reslizumab, mepolizumab, and be iralizumab are most used and supported as adjuncts for severe eosinophilic asthma berapy and poor asthma control. Studies have shown that treatments that target IL-5/ T_{2} -5R α in severe eosinophilic asthma patients resulted in significant decrease of b'ocd and sputum eosinophilia, reduction of exacerbation episodes, along with major implyvement in clinical symptoms (15). Patients on anti-IL-5 therapies experienced approxime ely half of the asthma exacerbation rates (88). However, improvements in health-reacted quality of life (HRQoL) scores and lung function could not be confirmed due to ins if a evidence (88). With regards to safety, patients taking reslizumab, mepolizumal on benralizuamb did not experience any serious adverse effects (88). Moreover, novel uprapies targeting other eosinophil-associated factors and their receptors have also been tud ed, including those that target CC-chemokine receptor 3 (CCR3), prostagland in 1¹2 () GD2), thymic stromal lymphopoietin (TSLP), as well as novel oligonucleotide therapies.

2.4.1 Reslizumab

Reslizumab is a humanized, neutralizing anti-IL-5 monoclonal antibody which acts by binding to IL-5 to block its interaction with its receptor (89,90). IL-5 receptors (IL-5R) can be found on the surfaces of eosinophils. IL-5 functions to increase the activation and maintenance of eosinophils, which drives eosinophilic inflammation (89). As such, the blocking of IL-5 binding with its receptor on eosinophils leads to the disruption of eosinophil maturation and promotes apoptosis (91). Many studies have shown the beneficial outcomes from reslizumab treatment in patients with severe eosinophilic asthma. For example, in an

experiment comparing reslizumab and placebo use in asthma patients, the reslizumab group experienced significantly less asthma exacerbations as compared to the placebo group (91). The results of the study concluded that reslizumab can be effectively used for treating asthma patients with persistent eosinophilia who are inadequately controlled with corticosteroids (91). Another study reported that patients that were administered 3.0 mg/kg of IV reslizumab every month experienced better asthma control as well as an improved lung function (90). The study also highlighted that patients with an increased eosinophil load that is inadequately managed with moderate-high doses of ICS benefited the most from reslizumab therapy (90). When compared to benralizumab, which targets IL-5R α , it was shown that reslizumab confers higher efficacy than benralizumab in patients with eosinc philic asthma (89).

2.4.2 Mepolizumab

Another anti-IL-5 monoclonal antibody with the same mechanism of action as reslizumab is mepolizumab, which has shown to be useful for treating eosinophilic asthma in several studies. In a research carried out by Hal a *et al.*, the efficacy of mepolizumab in the treatment of patients with refractory eo. nophilic asthma and history of recurrent exacerbations was investigated, who ware administered mepolizumab therapy for 1 year experienced reductions in exacerbation rates as well as improvements in the Asthma Quality of Life Questionnaire (AQLQ) scor/2 (92). At the end of the study, the patients who undergone 1 year of therapy reported less exacerbations, scored higher in the Asthma Quality of Life Questionnaire (AQLQ, and showed significant reductions in levels of blood and sputum eosinophils (92). However, mepolizumab showed no significant distinction in symptoms, post- broncl odil tor FEV₁, or airway hyperresponsiveness when compared to placebo and other anti-L-5 therapies (92). Another study demonstrated the effects of mepolizumab in prednisone-sparing. Nair et al., reported that mepolizumab administered to asthma patients with high sputum eosinophils despite continuous prednisone therapy effectively lowered blood and sputum eosinophil counts and thus, proved the prednisonesparing effect of mepolizumab (93). Furthermore, Sehmi et al., demonstrated that subcutaneous mepolizumab therapy significantly attenuated systemic differentiation of eosinophils in severe eosinophilic asthma patients, as observed from the decrease in blood eosinophils and elevated eosinophil-lineage-committed progenitors (EoP) counts in the mepolizumab group (94). However, mepolizumab had no significant effects on mature eosinophils, sputum EoP counts or maintenance dose of prednisone, indicating that mepolizumab did not inhibit differentiation of local airway eosinophils to mature cells (94).

Thus, it is evident that airway eosinophilia and severe eosinophilic asthma can be optimally controlled by targeting IL-5 pathway-associated local airway eosinophil differentiation. In a comparison study on the effectiveness of anti-IL-5 therapies, mepolizumab showed superiority in reducing the frequency of asthma exacerbations and improving asthma control over reslizumab or benralizumab in severe eosinophilic asthma patients with identical baseline levels of blood eosinophils (95).

Despite these studies that showed a positive outcome from mepolizumab therapy, there exists some concerns regarding the potential risk of 'rebound' eosinophilia following treatment cessation (55). Another study has also showed that mepolizumab treatment cessation resulted in rapid blood eosinophilia and subsequent worsening of asthma symptoms and exacerbations (55). Thus, further studies should be carried out to ensure the safety of eosinophil-targeted therapies. Nonetheless, mepolizumab is approved by both FDA and EMA to be used in patients >6 years of age with severe eosinophilic asthma. The primary benefits of mepolizumab include improvements in the lung turction (forced expiratory volume) and marked reductions in asthma exacerbations (96)

2.4.3 Benralizumab

Benralizumab is a humaniz.¹, afucosylated, IgG1k monoclonal antibody which prevents the receptor interaction o D_{L} 5 by binding to IL-5R α via its Fab domain. As a result, eosinophilia is reduced in blood, tissue and bone marrow due to the inhibition of eosinophil differentiation and in aturation. In addition, benralizumab also depletes eosinophils in tissues and systemic circulation via antibody-dependent, cell-mediated cytotoxicity as it can bind to the RIIIa region of the $Fc\gamma$ receptor on NK cells, neutrophils and macrophages through its afucosylated 1^c domain (97,98). The dual-function of benralizumab allows for a more rapid and sustained reduction of eosinophilia than other IL-5-targeted monoclonal antibodies (97). Benralizumab therapy has shown positive outcomes in both moderate-severe asthma patients as well as eosinophilic chronic obstructive pulmonary disorder (COPD) patients (99). Sridhar et al. conducted a research in which eosinophilic asthma and eosinophilic COPD patients were given benralizumab subcutaneously and the results demonstrated that benralizumab could selectively modulate eosinophil- and basophilassociated blood proteins and genes (99) Following benralizumab administration, eosinophil chemokines (eotaxin 1 and 2) were upregulated significantly in both the asthmatic and COPD patients (99). Furthermore, the expression of eosinophil- and basophil-associated genes were significantly reduced following benralizumab therapy (99). Thus, benralizumab is shown to

selectively modulate blood proteins and eosinophil- or basophil-associated genes, with alterations occurring prominently in eosinophil-high patients rather than eosinophil-low patients (99). Randomized clinical trials have also provided evidence on the optimal safety profile of benralizumab, as well as its efficacy in reducing asthma exacerbations, steroid-sparing and improvement of lung function (98). When compared to reslizumab, benralizumab was associated with a significantly improved lung function in asthma patients with severe eosinophilia (\geq 400 cells/µL) (95). Benralizumab is approved by FDA as an add-on maintenance therapy for use in patients with severe eosinophilic asthma aged over 12 years. The primary benefits include significant reductions in the asthma exacerbations, improvements in lung function parameters (forced expiratory volume in one second) and marked reduction in prescription of inhaled corticosteroids (1)0).

2.4.4 Omalizumab

Bronchial asthma is not only associated while elevations in pro-inflammatory Th2 cytokines, but also high levels of IgE. Besides the roles of Th2 cytokines in eosinophil differentiation, maturation, migration, and su, vival, they also contribute to the production of IgE. Omalizumab is an anti-IgE monoc.on.1 antibody that can effectively reduce peripheral blood eosinophil (PBE) counts as well as airway eosinophil counts in asthmatic patients (101). Massanari et al., reported that or alizumab had an inhibitory effect on eosinophils as observed from the lowered PBE courts in moderate-severe persistent allergic asthma patients who were on moderate-high uses of ICS (102). It was observed that patients who were administered omalizumab therapy demonstrated a significant decrease in PBE counts from baseline, with patients that have post-treatment free IgE levels < 50 ng/mL showing a larger decrease (102). In both steroid-reduction and steroid-stable phases of the studies, larger decreases in PBE counts were observed as compared to placebo (102). Thus, omalizumab treatment groups consistently experienced a pattern of lowered PBE counts and favourable clinical outcomes. Omalizumab is approved by both FDA and EMA to be used in patients over 12 years with moderate allergic asthma demonstrating raised levels of IgE. Omalizumab reduces the asthma exacerbation rates and, in some cases, reduces the corticosteroid prescriptions (103).

2.4.5 Dupilumab

Dupilumab is a novel anti-cytokine biologic drug used in the treatment of asthma. Therapies targeted at IL-4 and IL-13 are also beneficial in asthma treatments as they are Th2

cytokines that can stimulate eosinophil recruitment and migration to local airways. IL-4 and IL-13 contributes to the pathophysiology of the typical characteristics of asthma, such as chronic airway inflammation, tissue remodeling and airway hyperresponsiveness (104). Dupilumab is a monoclonal antibody that can inhibit the interaction between IL-4 and IL-13 with IL-4R α . In a recent study, dupilumab significantly attenuated Th2 cell-associated inflammatory biomarkers, which is correlated with an improvement in lung function and airway symptoms as well as reduction of asthma exacerbations in patients with difficult-to-control asthma (104). Dupilumab is approved to be used as an add-on therapy in patients over 12 years of age and who demonstrate a poorly controlled moderate-to-severe asthma phenotype. Dupilumab has been shown to prevent asthma exacerbations and improvements in lung function (105).

2.4.6 MEDI-563

In addition to the commonly used anti-IL-5 a...,s, MEDI-563 is a novel humanized anti-IL-5R α monoclonal antibody that has keel developed with an enhanced effector function in the management of asthma (106). Kolbeck *et al.*, reported that MEDI-563 caused both eosinophils and basophils to undergo antibody-dependent cellular cytotoxicity (ADCC) *in vitro* (107). MEDI-563 administered to non-human primates also decreased peripheral blood and bone marrow eosinophil courts (107). Thus, this study suggests that the focus of anti-asthma biologic therapies should be shifted from the passive removal of IL-5 to the reduction of eosinophil and basophil courts via ADCC mechanism.

2.4.7 Novel CCR3 receptor a stagonist

Agents which tar et eosinophil cell surface-structures, for example CC-chemokine receptor 3 (CCR3), have also showed efficacies in decreasing blood and tissue eosinophil levels (108). Ki19003 (4-[[5-(2,4-dichlorobenzylureido) pentyl][1- (chlorophenyl) ethyl] amino] butanoic acid) is a novel CCR3 antagonist. CC-chemokine ligands (CCL), such as CCL11, CCL24 and CCL26, are associated with eosinophil chemotaxis (109). A study done by Komai *et al.*, administered Ki19003 to ovalbumin-induced BALB/c mice to study its effects on airway remodeling in a mouse model of allergic asthma (109). It was observed that Ki19003 inhibited antigen-induced elevations in the number of eosinophils found in bronchoalveolar lavage fluid (BALF), but other cells were not affected (109). Furthermore, a dose-dependent increase in TGF- β 1 production in BALF and hydroxyproline amount in lungs were observed following Ki19003 administration (109). Ki19003 treatment also

resulted in the attenuation of allergen-induced subepithelial and peribronchial fibrosis (109). Hence, it can be concluded that CCR3 antagonism can prevent eosinophil airway infiltration as well as the progression of subepithelial and peribronchial fibrosis following an allergen challenge. Therefore, the process of airway remodeling, which is a prominent feature of allergic asthma, can potentially be prevented using CCR3-targeted treatments.

2.4.8 PGD2 antagonist

Chemoattractant receptor-homologous molecules expressed on T-helper type 2 cells (CRTH2) that mediates chemotactic response to PGD2 are also expressed on eosinophils (110). PGD2 inhibitors, such as timapiprant and fevipiprant, have been studied for their effects on eosinophilia. Timapiprant was shown to improve conditions of mild-moderate eosinophil-mediated allergic asthma patients whereas the administration of fevipiprant therapy to patients with moderate-severe asthma who were not adequately managed with ICS resulted in reduced sputum eosinophil counts and subscipately, attenuation of eosinophilic airway inflammation (111). However, the phase 11 trial of fevipiprant did not show marked clinical improvements in patients with poorly pontrolled asthma (112).

2.4.9 Anti-TSLP

TSLP plays a part in eosine phase inflammation by recruiting eosinophils following epithelial damage. AMG 157 is a human anti-TSLP monoclonal IgG2 λ which inhibits the interaction and binding of TCLP to its receptor. A Phase I clinical trial conducted by Gauvreau *et al.*, reported that A MG 157 treatment administered to allergic asthma patients of mild severity significantly reduced FeNO in addition to blood and sputum eosinophil levels following and prior to all rgen (113). Moreover, the AMG 157 treatment group experienced a greater reduction in maximum percentage decrease in FEV₁ during late response than the placebo group, with a greater reduction occurring as the treatment progressed (113). Thus, the researchers concluded that AMG 157 attenuates bronchoconstriction and other indications of respiratory inflammation pre- and post-allergen challenge.

2.4.10 Novel oligonucleotide therapy

Oligonucleotides, such as siRNA and miRNA, are promising novel therapeutic strategies for the treatment of various respiratory disorders via their gene silencing or RNA interference abilities (114–117). An oligonucleotide-based therapy, TPI ASM8, was studied for its potential beneficial effects in lung function and sputum eosinophilia in patients with

mild allergic asthma. TPI ASM8 consists of two phosphorothioate antisense oligonucleotides (AON) which are modified, with one targeting CCR3 receptor and another targeting the beta chain (β c) which is common among IL-3, IL-5 and GM-CSF receptors. According to a study carried out by Imaoka *et al.*, patients with mild allergic asthma who were administered nebulized TPI ASM8 following an allergen challenge showed reductions in sputum eosinophils, early and late asthmatic responses, as well as airway eosinophil progenitor cells (118). Thus, it was concluded that the accumulation of eosinophils and their progenitor cells in airways of asthmatics can be inhibited successfully via TPI ASM8-mediated CCR3 and β c expression blockade. Therefore, further efforts of developing novel therapies that inhibit accumulation of airway progenitor cells should be taken.

2.4.11 Corticosteroids

Oral corticosteroids (OCS) are one of the manistay therapies for the long-term management of severe asthma, which is characterized by persistent asthma symptoms and airway inflammation despite maximum efforts using anti-asthma treatments. Corticosteroids effectively acts on various components of $t_1 > \ln 2$ inflammatory pathway and its use can result in rapid attenuation of eosinop, ils associated inflammation as well as long-term reduction in airway hyperresponsioness (119). As such, corticosteroids confer better responses in asthma patients with ecsipophilic inflammation as compared to neutrophilic inflammation (120). However, a major drawback to corticosteroid use is the development of corticosteroid resistance or a sensitivity, which frequently occurs in long-term use of corticosteroids. This feature is especially prominent in severe asthmatics with persistent Th2 inflammation despite regular OCS use. Thus, biological therapies are introduced as alternative therapies althe igh a large proportion of asthmatic patients will still require the use of OCS to control their asthma (119). The therapeutic action of OCS on inflammatory cytokines and eosinophils are proven in several studies. In a study conducted by Dente et al., short-term courses of OCS reduced both sputum eosinophilia and sputum pro-inflammatory cytokine concentrations, in addition to improving lung function in patients with severe refractory asthma (121). The improvement of pulmonary function was concluded from the significant increase in FEV₁ following OCS treatment/ The prednisone group also exhibited a significant decrease in sputum eosinophil percentages and concentrations of IL-5 and IL-8 (121). It was also noted that prednisone treatment only showed positive effects in patients with baseline sputum eosinophilia, whereas only a significant decrease of sputum IL-8 was observed in non-eosinophilic patients (121). Several studies also reported on the inverse

relationship between CS dose and PBE counts displayed in severe eosinophilic asthma patients. Prazma *et al.*, predicted a model in which reducing the daily dose of OCS by 5 mg/day resulted in a 41% increase in PBE counts (122). Another study reported that ICS dose increment from medium to high dose in patients with uncontrolled asthma resulted in a significant reduction of blood eosinophil concentrations (123).

2.5 Alternative therapies affecting eosinophils in the treatment of asthma

2.5.1 Quercetin

Known to be present as flavonoids in fruits and vegetable: quercetin was shown to be an anti-inflammatory agent with the potential use in the treatment of asthma (124). In this case, the clinical features of asthma were relieved by suparessing the activation of the signaling pathway of NF- κ B which results in the reduced production of proinflammatory mediators associated with airway hyperresponsiveness and airway inflammation (124). Two other studies have also insinuated quercetin's rore in the prevention of eosinophilic airway inflammation via the attenuation of eosino_F hil activation, particularly the production of chemokines (125,126). In the study by the κ Yong *et al.*, quercetin was encapsulated into a lipid crystalline nanoparticle (LCN) to overcome its limitation in terms of bioavailability as well as aqueous solubility which results of the successful exhibition of an improved antiinflammatory effect of quercetin when formulated as such (124).

2.5.2 Curcumin

Curcumin is a constituent originating from the spice, turmeric, that is thought to be a potential remedy for the management of asthma (127,128). Similarly, to quercetin, curcumin possesses anti-inflammatory properties which relieve airway inflammation of allergic nature as well as airway hyperresponsiveness through the suppression of the activation of the signaling pathway associated with NF- κ B and the attenuation of eosinophil production (128). Its clinical application is also limited by the same reasons seen in quercetin (128,129). Hence, Ng *et al.*, sought to load curcumin into liposomes as liposomes were noted to be able to modify a system's pharmacokinetic and biodistribution profiles (128–130). The study managed to overcome the limitation of curcumin's unfavourable solubility as well as demonstrate the sustained release properties attained through the encapsulation of curcumin in a liposome, indicating that curcumin loaded liposomes could be an appealing option for the treatment of asthma (128).

3.0 Eosinophils and COPD

Chronic obstructive pulmonary disease is described by the Global Initiative for Chronic Obstructive Lung Disease (GOLD) as a disease that can be commonly prevented and treated (131-134). However, COPD is noted to be among the most substantial causes of morbidity and mortality globally and this has caused a significant burden which is ever growing on the economy and society (131). This disease is characterized by the poorly reversible limitation of airflow which is commonly progressive and the elevated response to chronic inflammation that occurs in the peripheral airways and lung parenchyma due to the inhalation of noxious particles or gasses which is primarily (but not exclusively) caused by smoking tobacco (131,135–137). Emphysema and small, irw; y fibrosis may result from the chronic inflammation because of the destruction of painchymal tissue and disruption of mechanisms of repair and defense which causes the clise of the characteristic symptoms associated with COPD (131,135). As the dist ase progresses, so does the degree of inflammation which results in the elevation of neurophil, macrophage and lymphocyte levels within the airway lumen (137). The increase in numbers of neutrophil and macrophages occur due to the activation of pattern recognition receptors which precipitates an innate immune response that also gives rise in the activation of airway epithelial cells as well as mucus secretion (137,138). An i ici ase in T lymphocyte and B lymphocyte numbers in the lungs is seen when adaptive in munity is activated in the latter course of the disease which might result in the amplification of neutrophilic inflammation due to the increment in CD4+ Thelper 17 cells present in the lungs (137). Although various inflammatory cells are implicated in COPD, net trophils are by far the most abundant and has been extensively associated with the pathogenesis of COPD (139-143). However, it has been reported that some COPD patients without asthma have elevated numbers of eosinophils in their airways instead during exacerbations and clinical stability (Figure 3) (137,141,144,145). This is commonly seen in asthmatic patients where eosinophilic inflammation is known to be a distinguishing feature of the disease but the presence of a subdivision of COPD patients with eosinophilic airway inflammation has been reported (141,143,144,146–148). COPD patients with this phenotype are known to show the most significant response to corticosteroid treatment as well as an increased risk of exacerbation relapse and hospital readmissions (140,142,143,146,149,150).

3.0 Eosinophils and COPD

3.1 Role of eosinophils as biomarkers in COPD

Blood eosinophils have been recognised by several studies to be potential a biomarker of disease severity and clinical outcome in exacerbations of COPD (151–153). The measurement of eosinophils present in the blood, as a proxy for eosinophils present in tissues, has become an essential biomarker for the prediction of risk of exacerbations in COPD patients (154). Elevated counts of blood eosinophil in COPD has been well associated to the clinical features of the disease. Patients with an increment in blood eosinophil levels of \geq 450 cells/µl during stable disease reportedly reflected a higher rate of exacerbation by 13% in the following year compared to patients with lower numbers (152). Another studied showed, amid COPD patients in the general population, increment of blood eosinophil counts > 0.34×10^9 cells/L were affiliated with a heightened risk of ceve oping severe exacerbations by 1.76-fold (155). However, blood eosinophils singularly were not a dependable biomarker to predict the severity of the disease, the associated risk of exacerbations, or sputum eosinophilia (156). Instead, elevated sputum eosinophils for the identification of a subdivision of patients with more disease selecting and more persistent exacerbations (156).

Eosinophil concentrations may a o be a useful biomarker for the prediction of outcomes in COPD as increased number of blood eosinophil in COPD were related with a longer survival period and a higher in a dence of hospital readmissions (157–159). A study also suggested that the deterioration of pulmonary function tests could be predicted according to number of blood eosinophil (158). Low blood eosinophil counts on the other hand, is predictive of a poorer clinication come and longer durations of hospital admission (158–160). However, a few studies report that blood eosinophil counts do not appear to be related to mortality (158,161). It is also noted that blood eosinophil concentrations can be utilized as a biomarker for the prediction of readmissions of patients suffering from severe COPD exacerbations (162). Moreover, Bélanger *et al.*, suggests a relationship between greater blood eosinophil count upon admission of a COPD exacerbation and increased rate of readmissions (161–163).

3.2 Eosinophils as a guide for COPD treatment

Although airway eosinophilic inflammation in COPD may worsen the stabilization of symptoms, it can be used as to predict the benefit of treatment involving inhaled and oral corticosteroid (157,163,164). A review by Hillas *et al.*, states the benefit of corticosteroid use in patients experiencing acute exacerbation of COPD (AECOPD) who have increased blood

eosinophils counts (165). The review also covered a post hoc analysis of three randomized controlled trials (RCTs) which demonstrated that COPD patients with eosinophil counts > 100 cells had a reduced risk of AECOPD of 25% when a combination of budesonide and formoterol was used compared to formoterol alone (165,166). Two other studies also showed results of greater decrease of exacerbation frequency in patients as the blood eosinophil count increased when inhaled corticosteroids (ICS) was added to the therapy alongside a long-acting β -agonist (LABA) whereas patients who were on a single therapy regimen of LABA exhibited progressively increasing rate of exacerbations as the eosinophil counts increased (165,167–169). In addition, Vestbo *et al.*, has also demonstrated in his study that patients reflecting blood eosinophil counts $\geq 2\%$ experienced reduced concerbations of 30% with the use of either a regimen consisting of single inhaler extra fine triple therapy or long-acting muscarinic antagonist (LAMA) (170). Likewise, the 2019 GC LD COPD Strategy document states that blood eosinophil levels are to be used to grade ICS therapy in the treatment of stable COPD patients and frequent exacerbators as weil (171).

It was also suggested that ICS withdraw(1) 1 COPD patients with eosinophil counts \geq 4% and a history of exacerbations may increase the possibility of developing AECOPD (172,173). A trial reported that the continuous use of ICS reduced the exacerbations of moderate and severe grades in patients with either $\geq 4\%$ relative eosinophil count or ≥ 300 cells/mL absolute eosinophil count compared to patients who were tapered off ICS (173). Similarly, the SUNSET trial ider and that as ICS was tapered off, it led to a heightened risk of exacerbations among patient, with an eosinophil count of \geq 300 cells/mL (174). However, several studies including the Connical Practice Research Datalink (CPRD) reported that ICS withdrawal among COP D p; tient did not affect the exacerbation risk of moderate-to-severe severity significantly (172, 175-177). The difference in findings might be due to the use of different patient groups where the WISDOM trial had made use of patients with COPD of high severity, a past medical history of exacerbations and prior treatment with ICS before the trial, the SUNSET trial studied COPD patients treated with ICS regimens long-term and did not have persistent exacerbations whereas other studies such as CPRD utilised a patient population not at high exacerbation risk where the usage of bronchodilators and ICS are lesser than clinical trials (172-174). Nevertheless, both the WISDOM trial and CPRD reported a similar finding whereby patients with eosinophil counts more than 6.0% do not have an increased risk of exacerbations on withdrawal of ICS (172,173).

The study by Oshagbemi *et al.*, did not report findings of increased all-cause mortality risk among patients with elevated absolute or relative blood eosinophil counts who withdrew

from ICS regimen but there are several studies that have evidence which conflict this finding (172,178,179).

Even though, blood eosinophil counts can be regarded as a guide to treating COPD, it is critical to remember that COPD patients, blood eosinophils are not influenced by ICS use (165,180). This is because there is pronounced variation in blood eosinophil levels (165,181–183). Hence, one measurement of blood eosinophil counts may not be sufficient for the accurate choice of utilizing ICS in a COPD patient's regimen (165).

3.3 Therapeutic targets for eosinophilic inflammation in COPD

COPD treatment regimens are largely associated with use of bronchodilators which treat the symptoms but not the underlying inflamination nor disease progression (135). Biologics targeting immune mediators such as IL-5 are showing promising results in COPD patients with the eosinophilic phenotype (135-18-1). IL-5, a key eosinophil cytokine mediates its differentiation, proliferation, survival, and activation via the IL-5 receptor (184-187). Humanized monoclonal antibodies, mepo'ız mab and benralizumab reduces blood and tissue eosinophil counts via the inhibition of 1 -5 pinding to eosinophil surface receptors and binding to interleukin-5 receptor α respectively (185,186,188–190). These two drugs have been known to successfully reduce essinophilic inflammation and lower exacerbations rates in asthma (135). However, mepolizar, b was not able to obtain the approval from the US Food and Drug Administration ((D_{L})) to be used as an adjuvant to the maintenance regimen for COPD patients with eosil. philic inflammation because one of two phase III studies carried out showed no evidence of efficacy (135,184,186,187). Studies on benralizumab have been inconsistent in which it demonstrated a decrease in rate of exacerbations in a Phase IIa trial consisting of COPL patients with eosinophilic inflammation but failed to reach the primary end point in two phase III trials consisting of COPD patients with exacerbation histories of moderate to very severe grades (135,185,189,190).

Another target of interest in eosinophilic COPD is the transcription factor, GATA3 which plays a part in the activation of Th2 cells and action on the type 2 innate lymphoid cells (ILC2 cells) (191). These interactions result in increased cytokine IL-4, IL-5 and IL-13 production which are thought to moderate airway eosinophilia in non-allergic asthma and COPD (191). SB010 is a drug that consists of the active constituent, DNAzyme hgd40 which specifically binds to and cleaves the mRNA of GATA3 and has been evaluated in human and animal models (191). Turowska *et al.*, reports significantly reduced GATA3 mRNA along with reduction of Th2-specific cytokines production in murine models of allergic airway

inflammation when treated with SB010 (192). Greulich *et al.*, were able to prove the involvement of the GATA3 pathway in eosinophilic COPD patients through their phase IIa clinical trial which sought to demonstrate the feasibility to reduce sputum eosinophilia in COPD patients with elevated sputum eosinophil counts through treatment involving the inhalation of GATA3-specific DNAzyme SB010 for 4 weeks and were able to conclude that a decrease in airway eosinophilia is possible to attain via the GATA3-specific DNAzyme as such seen previously in asthmatic patients (191). Furthermore, the trial was able to identify the safety aspects associated with the use of SB010 in the studied COPD population whereby serious adverse events were not reported (191). However, the trial only included a small patient group so more studies including a larger patient groups and longer treatment period will be required for the identification of the long-term clinica' effectiveness and safety in this subgroup of COPD patients (191).

4.0 Eosinophils and Cystic Fibrosis

Cystic fibrosis is an autosomal recessive disorder that is an illness which is chronically progressive and life-limiting (193-196). The disease occurs due to a gene mutation of CF transmembrane conductance regulator (CFTR) causing a significant functional deficiency in the CFTR provin which are highly expressed in the tissues of the airways (193-195). In normal circumstances, the CFTR protein functions to regulate the movement of chloride and soa. I'm ions across the epithelial cell membranes but when either one or both copies of the gene is mutated, transport of the ions is deficient, resulting in the accumulation of thick mucus throughout the body which can lead to the development of respiratory insufficiency, consequential deficiencies in host anti-bacterial defences and various other systemic obstructions and abnormalities (193-195). Including being a chronically progressive lung illness, cystic fibrosis is also characterised by extensive inflammation and respiratory failure (195). This could be further exacerbated by the formation of biofilms on the exterior of medical appliances utilized for treatment causing lifethreatening infections in these patients (197). Cystic fibrosis is known to affect multiple organs and the common symptoms include breathlessness or wheeze, persistent cough along with frequent respiratory infections, increased appetite and bulky greasy stools (198). Deterioration in pulmonary symptoms and the loss of lung function are the key factors that determine the severity of cystic fibrosis. More than half of the patients may require a lung transplantation during severe form of the disease (199).

Cystic fibrosis associated inflammation is known to be dominated by neutrophils which releases oxidants and proteases such as elastase which can be found in airway secretions preceding the development of bronchiectasis in these patients (194,195,200,201). A few studies have also reported of the correlation between neutrophil elastase with the deterioration of lung function as well as respiratory exacerbations (194,200). However, Zhang et al., notes that a type II inflammation associated with an asthma phenotype can often be recognised in cystic fibrosis patients (195). The presence of this type of inflammation is indicated by the increased levels of total immunoglobulin E (IgE), specific IgE sensitization or absolute eosinophil count in the circulation independent of infection by the pathogen, Pseudomonas aeruginosa (195,202). This enables the differe viat on of the similar symptoms of asthma and cystic fibrosis in terms of variability of lun; function and bronchodilator response which in turn aids the recovery of lung function, airway remodeling, and rate of exacerbations in patients with eosinophilic inflammation related exacerbations through the focus of treatment on to the type II inflammatory response (195). Moreover, the study by Zhang et al., showed positive effects via a d. crease in eosinophil and IgE levels as well as dose of corticosteroids when mepolizun, by vas used to treat type II inflammation in patients with eosinophilic phenotype cystic fil rosis (195).

Apart from neutrophils taat are known to play an important inflammatory role in airway disease associated with cysic fibrosis, eosinophils are also found to be one of the components that contribute to it jury during cystic fibrosis. In two separate studies involving 42 and 20 patients with cysic fibrosis respectively, Koller *et al.*, showed significantly higher levels of ECP in sputh 1 of patients with cystic fibrosis compared to control subjects (203,204). Collectively, these findings indicated the destructive role played by eosinophils and an interrelationship between the clinical variables and secretory activity of eosinophils in cystic fibrosis. An increase in the propensity of eosinophils to release their granule proteins and a firm correlation between ECP levels and variables of pulmonary function like forced vital capacity has also been identified (204,205). Furthermore, cytokine profiles were found to be responsible for eosinophil activation and degranulation in patients with cystic fibrosis, especially IL-8 and IL-3 were found to be significantly correlated with the levels of ECP in sputa from 32 patients and therefore, these appeared to be responsible for elevated degranulation of eosinophils (206).

However, the mainstay drugs used to treat cystic fibrosis have been noted to cause various side effects (207). Thus, in order to diminish these drug associated side effects experienced by patients, tissue targeting strategies should be utilized to overcome the inadequate drug penetration due to the barrier formed by the accumulation of thick and viscous mucus (208,209).

5.0 Eosinophils and Pneumonia

Pneumonia is a disease which is relatively previlent and has caused a significant burden towards the global population (210). It is defined by the acute infection of the lung parenchyma and is used as a hypernym to describe a chuster of syndromes caused by not one disease but a group of specific infections, v much result in different manifestations and sequelae (210,211). Acute and chronic eosing phine pneumonia are two common pulmonary eosinophilic disorders which occur due to lung tissue damage by activated eosinophils (212,213). These disorders are characterized by the build-up of eosinophilic infiltrates in the pulmonary parenchyma, often accommon ranged by peripheral blood eosinophilia, which can be caused by either infectious or not -m.^cectious factors (212,214).

De Giacomi *et al.*, reports significantly elevated levels of IL-33 in acute eosinophilic pneumonia (AEP) patients and hypothesized that IL-33 plays an essential role in AEP (213). This is because attraction and activation of eosinophils is amplified when Th2-polarizing cytokines, IL-5 and IL-1, are rapidly produced due to the robust production of IL-33 as well as the enlistment and activation of ILC2 cells in the mucosa of the airway which takes place due to either epithelial or endothelial cell (213). However, the study by Katoh et. al. reports that IL-33 levels were not considerably elevated in the bronchial alveolar lavage fluid (BALF) of eosinophilic pneumonia patients whereas IL-25 and IL-5 levels were significantly heightened in the BALF of chronic eosinophilic pneumonia (CEP) patients but not AEP patients (215). Hence, it was postulated that CEP might be perpetuated by IL-25 which were thought to be produced by eosinophils via IL-5 stimulation (215,216). Despite these reports, the pathophysiology of both AEP and CEP are poorly understood and will require further studies to completely delineate it (213,214).

Treatment of AEP and CEP both utilize corticosteroids as the mainstays of treatment in which AEP is treated with systemic corticosteroids whereas CEP utilizes oral corticosteroid therapy with an aim to diminish the disease progression as well as lessen the risk of relapse (212–214,217–219). As an alternative therapy for CEP, biologic agents have been utilized for the regulation of eosinophilic inflammation (218,219). Studies have shown the effectiveness of omalizumab, an anti-IgE antibody and anti-IL-5 antibody's, mepolizumab and reslizumab in the treatment of CEP through the reduction or discontinuation of corticosteroid use in patients with relapsed CEP (218,219). IL-5 is a common aim in the treatment of CEP because its elevated levels are associated with the release of cytotoxic granular proteins from eosinophils which is $_{\rm P}$ ostulated to be an important underlying mechanism of CEP (219). However, there is instific ent information supporting the utilization of biologics for CEP treatment as well a the concerns associated with the adaptation and treatment duration of it (218,219). Hence, further studies regarding the use of steroid-sparing therapeutic regimens as an alternative dec.apy for CEP are required.

6.0 Eosinophils and Lung Cancer

Lung cancer is a common malignancy among both sexes and represents over 10% of all malignancies (220,221). The occurrence of eosinophilia is often attributed to hematological malignancies, with come cases of solid tumour-associated eosinophilia. Approximately 1% of malignant tun ours are associated eosinophilia (220). Bone marrow stimulation by IL-5 is theorized to be the primary causative factor of solid malignancy-associated eosinophilia. Para eoplastic eosinophilia cannot be treated with a specific treatment, besides treating the underlying malignancy using typical therapies, including surgery, chemotherapy, reacotherapy, or even novel treatments such as metformin (222).

6.1 Function of eosinophils in the pathophysiology of lung cancer

A distinct feature of cancer is sustained low-grade inflammation. Eosinophils have long been associated with cancer as they are one of the regulatory components of the tumour microenvironment (TME) responsible for tumour initiation and development (223). Eosinophils are commonly associated with negative connotations in allergic diseases, however they are able to provide immune protection against helminths, bacterial and viral pathogens (223).

There are evidences showing that the infiltration of eosinophils into tumour cells results in an improved prognosis of cancer (224). Tumour-associated eosinophilia can be

observed in many studies of patients with cancer as well as mouse models of cancer. Studies have reported that activated eosinophils play a vital role in tumour rejection. Activated tumour-honing eosinophils release chemoattractants which induces the migration of tumour-specific CD8(+) T-cells into the tumour, resulting in tumour eradication and thus, an increased chance of survival (225). Tumour rejection is also promoted by the significant alterations in the tumour microenvironment as initiated by the activated eosinophils, such as macrophage polarization and normalization of tumour vasculature (225).

Eosinophil peroxidase (EPO), which is an eosinophilic cationic granule protein, drives cell cycle progression and proliferation at non-cytotoxic levels, thus suggesting the role of eosinophils in tumorigenesis (226). According to a study conducted by Walsh *et al.*, EPO increased the expression and phosphorylation of epiderival growth factor-2 (HER2) in a sustained manner, which consequently induced extracelly lar- egulated kinase 1/2 activation (226). Subsequently, cyclin-dependent kinase inhibitor $p_2 r$ (kip) entered the cytoplasm from the nucleus in a focal adhesion kinase-dependent man.²⁶. Thus, the findings of the study led to the conclusion that EPO can induce the $p_1 p_2$ gulation of cell proliferation via HER2 mediation.

However, the actions of eosinop. its and eosinophil mediators varies according to the cancer type as eosinophils have been tinked to improved prognosis in certain neoplasias but poor prognosis in others. This is tecanse eosinophils can produce either anti-tumorigenic (e.g. TNF- α , IL-18, granzyme, and entronic proteins) or pro-tumorigenic molecules (e.g. pro-angiogenetic factors) subject to the internal environment (223). Melanoma, oral, gastric, colorectal, and prostate cancers are neoplasias in which eosinophils secrete anti-tumorigenic factors whereas a poor prognosis is expected from the actions of eosinophils in cervical carcinoma and Hodgkin's lymphoma (223).

Although there exist evidences for a positive correlation between eosinophils and better responses in some patients with melanoma, investigations to study the relationship between eosinophils and NSCLC are not frequently conducted. However, a case report showed that a patient with metastatic lung adenocarcinoma (AD), a type of non-small cell lung cancer (NSCLC), presented with shortness of breath, chest pain and associated hypereosinophilia, as defined by an absolute eosinophil count (AEC) of >1500 cells// μ L, in the absence of primary bone marrow disorder (220). Additionally, a study done by Lou *et al.*, reported that a patient with metastatic lung AD presented with asymptomatic hypereosinophilia following initiation of nivolumab therapy, which is an anti-programmed cell death 1 (PD-1) drug (227). Following a transient discontinuation of therapy, her

eosinophil counts transiently decreased, but increased again following re-initiation. The patient showed a favourable response throughout the therapy. Thus, eosinophils can act as potential peripheral biomarkers of favourable response to immunotherapy in patients with lung carcinoma, which warrants further studies of the role of eosinophils in lung carcinoma for the development of novel treatment strategies. As such, efforts should be focused on the delivery of eosinophil-targeting drugs for lung cancer in the form of nanocarriers in order to improve specificity and reduce adverse effects (228,229).

6.2 Eosinophils as biomarkers of lung cancer

Eosinophils can be used as a peripheral blood marker for detecting tumour-associated protein expression in cancer patients. Tumours cells in primary lung AD express surface immune factors such as indoleamine-2,3-dioxygenase-1 (DO) and programmed cell death-ligand-2 (PD-L2) (230). A study showed that AEC along with absolute monocyte count (AMC), could potentially act as a biomarker to predict IDO1 expression in patients with resected stage 1 to 3 primary lung AD (230). *Ford* is a tryptophan (Trp) catabolic enzyme that catalyzes the transformation of Trp to ky, urenine for immunosuppressive functions such as activation of myeloid-derived suppressor cells and T-regulatory cells, inhibition of effector T and NK cell functions, and promotion of neovascularization of solid tumours (231).

Eosinophils can also act as c_1 -ical outcome predictors in patients with lung AD. Tanizaki *et al.*, reported that NSLC₁-atients administered nivolumab therapy exhibited higher AECs which were significantly linked to a better progression-free and overall survival (232). Additionally, a cohort study reported that elevated eosinophil counts correlated with reduced mortality risk in specific sub} roups of cancer patients, such as colorectal cancer (233).

However, a study showed that only a small percentage of eosinophils (0.3%) were present in the immune infiltrate composition of NSCLC tumours (234).

Besides AEC, another eosinophil-related biomarker of lung AD is EPO. Ye *et al.*, demonstrated that EPO overexpression in lung AD patients can potentially act as a biomarker for poor prognosis (235). During the study, a significantly higher expression of EPO mRNA and protein in lung AD tissues as compared to adjacent normal tissues were observed (235). Moreover, patients with EPO overexpression were significantly associated with pathologic-tumour nodes metastases stage and lymph node metastasis, as well as decreased survival time as compared to patients with low levels of EPO (235).

7.0 Eosinophils and Acute Lung Injury

Acute lung injury (ALI) describes clinical syndromes of acute respiratory failure with prominent mortality and morbidity rates (236). The management of ALI after exclusion of infection consists of primarily supportive measures. Other diagnoses can be ruled out by performing surgical lung biopsy, however this measure has proven to be less useful for predicting therapy methods and outcomes in ALI (237). Peripheral blood and tissue eosinophil counts are often referred to as hallmarks of steroid-responsive acute eosinophilic pneumonia, but are not typically associated with ALI (237). Thus, a study conducted by Willetts *et al.* demonstrated that eosinophil peroxidase-rec ogl. zing monoclonal antibody (EPX-mAb) immunohistochemistry can be used as a nethod to assess eosinophil accumulation or degranulation in the lungs of ALI patie. 's as well as predict the prognosis of survival (237). Therefore, it is suggested that EFX-n Ab immunohistochemistry can be utilized as a diagnostic biomarker to identify a proportion of ALI patients with favourable clinical outcomes.

Various clinical trials conducted targeting cosinophils in respiratory diseases are shown inTables 1-3 and various important drug, targeting eosinophils in respiratory diseases areshowninFigure4andTable4

Conclusion

Respiratory disorders are chronic conditions requiring long-term therapy to prevent clinical symptoms. Morbidity and mortality are high for many respiratory disorders and existing available treatments are restricted due to lack of effectiveness, severe toxicity or both. Recent efforts to understand eosinophil biology has opened the door to many innovative biological treatments. Various drugs that target eosinophils are currently being tested in clinical trials and needs to be validated prior to medical use. No health issues are raised till date, despite theoretical concerns about the possible toxicity of rapidly declining eosinophil counts and the long-term effects of eosinophil depletion on the immune system and tumor monitoring. We expect to gain a better understanding with more research to recognize new biological factors for these respiratory disorders and eventually the implementation of this knowledge to patients.

References

- 1. Ravin KA, Loy M. The Eosinophil in Infection. Clin Rev Allergy Immunol. 2016 Apr;50(2):214–27.
- 2. Stone KD, Prussin C, Metcalfe DI · Ic, E, Mast Cells, Basophils, and Eosinophils. J Allergy Clin Immunol. 2010 Feb;125(? Suppl 2):S73-80.
- 3. Blanchard C, Rothenberg ME L'ology of the Eosinophil. Adv Immunol. 2009;101:81– 121.
- 4. Fulkerson PC, Rothenbe g N.E. Targeting eosinophils in allergy, inflammation and beyond. Nat Rev Drug Discov. 2013 Feb;12(2):117–29.
- 5. Mathur SK, Fichtinger PS, Kelly JT, Lee W-M, Gern JE, Jarjour NN. Interaction between allerg, and ir nate immunity: Model for eosinophil regulation of epithelial cell interferon expression. Ann Allergy Asthma Immunol Off Publ Am Coll Allergy Asthma Immunol. 2013 Jul;111(1):25-31.e1.
- 6. Wong TW, Doyle AD, Lee JJ, Jelinek DF. Eosinophils Regulate Peripheral B Cell Numbers in Both Mice and Humans. J Immunol. 2014 Apr 15;192(8):3548–58.
- 7. Akuthota P, Wang H, Weller PF. Eosinophils as Antigen-Presenting Cells in Allergic Upper Airway Disease. Curr Opin Allergy Clin Immunol. 2010 Feb;10(1):14–9.
- Jacobsen EA, Zellner KR, Colbert D, Lee NA, Lee JJ. Eosinophils Regulate Dendritic Cells and Th2 Pulmonary Immune Responses following Allergen Provocation. J Immunol. 2011 Dec 1;187(11):6059–68.
- 9. Chu VT, Fröhlich A, Steinhauser G, Scheel T, Roch T, Fillatreau S, et al. Eosinophils are required for the maintenance of plasma cells in the bone marrow. Nat Immunol. 2011 Feb;12(2):151–9.

- 10. Wu D, Molofsky AB, Liang H-E, Ricardo-Gonzalez RR, Jouihan HA, Bando JK, et al. Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. Science. 2011 Apr 8;332(6026):243–7.
- 11. Kim H-J, Alonzo ES, Dorothee G, Pollard JW, Sant'Angelo DB. Selective Depletion of Eosinophils or Neutrophils in Mice Impacts the Efficiency of Apoptotic Cell Clearance in the Thymus. PLoS ONE [Internet]. 2010 Jul 6 [cited 2020 Jan 19];5(7). Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2897847/
- Wen T, Rothenberg ME. The Regulatory Function of Eosinophils. In: Myeloid Cells in Health and Disease [Internet]. John Wiley & Sons, Ltd; 2017 [cited 2020 Jan 12]. p. 257–69. Available from: https://onlinelibrary.wiley.com/doi/abs/10.1128/9781555819194.ch14
- Spencer LA, Szela CT, Perez SAC, Kirchhoffer CL, Neves 'S, Radke AL, et al. Human eosinophils constitutively express multiple Th1, Th?, and immunoregulatory cytokines that are secreted rapidly and differentially 'Luckoc Biol. 2009 Jan;85(1):117–23.
- 14. Davoine F, Lacy P. Eosinophil Cytokines, Che notines, and Growth Factors: Emerging Roles in Immunity. Front Immunol parts (net]. 2014 [cited 2020 Jan 12];5. Available from: https://www.frontiersin.org/a.ticles/10.3389/fimmu.2014.00570/full
- 15. Varricchi G, Bagnasco D, Borriello F, L'effier E, Canonica GW. Interleukin-5 pathway inhibition in the treatment of eosin ph lic respiratory disorders: evidence and unmet needs. Curr Opin Allergy Clin Imn. 1 ol. 2016 Apr;16(2):186–200.
- Sastre B, Rodrigo-Muñoz JM, Garria-Sanchez DA, Cañas JA, Del Pozo V. Eosinophils: Old Players in a New Game. J Investig Allergol Clin Immunol. 2018;28(5):289–304.
- Halwani R, Al-Muhsen L, Al-Jahdali H, Hamid Q. Role of transforming growth factorβ in airway remodeling h. asthma. Am J Respir Cell Mol Biol. 2011 Feb;44(2):127–33.
- 18. Rosenberg HF, D' er k D, Foster PS. Eosinophils: changing perspectives in health and disease. Nat Rev Jmmunol. 2013 Jan;13(1):9–22.
- 19. Liao W, Long H, Chang CC-C, Lu Q. The Eosinophil in Health and Disease: from Bench to Bedside and Back. Clin Rev Allergy Immunol. 2016 Apr 1;50(2):125–39.
- 20. Kiwamoto T, Kawasaki N, Paulson JC, Bochner BS. Siglec-8 as a drugable target to treat eosinophil and mast cell-associated conditions. Pharmacol Ther. 2012 Sep;135(3):327–36.
- 21. Kvarnhammar AM, Cardell LO. Pattern-recognition receptors in human eosinophils. Immunology. 2012;136(1):11–20.
- Månsson A, Cardell L-O. Role of atopic status in Toll-like receptor (TLR)7- and TLR9-mediated activation of human eosinophils. J Leukoc Biol. 2009 Apr;85(4):719– 27.

- 23. Johansson MW. Activation states of blood eosinophils in asthma. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 2014 Apr;44(4):482–98.
- 24. Johansson MW, Mosher DF. Activation of beta1 integrins on blood eosinophils by P-selectin. Am J Respir Cell Mol Biol. 2011 Oct;45(4):889–97.
- 25. Radke AL, Reynolds LE, Melo RCN, Dvorak AM, Weller PF, Spencer LA. Mature human eosinophils express functional Notch ligands mediating eosinophil autocrine regulation. Blood. 2009 Mar 26;113(13):3092–101.
- 26. Acharya KR, Ackerman SJ. Eosinophil Granule Proteins: Form and Function. J Biol Chem. 2014 Jun 20;289(25):17406–15.
- 27. Shamri R, Xenakis JJ, Spencer LA. Eosinophils in innate immunity: an evolving story. Cell Tissue Res. 2011 Jan 1;343(1):57–83.
- Linch SN, Kelly AM, Danielson ET, Pero R, Lee JJ, Gc¹d 'A. Mouse Eosinophils Possess Potent Antibacterial Properties In Vivo. Inf ct 1 nmun. 2009 Nov;77(11):4976–82.
- 29. Muniz VS, Weller PF, Neves JS. Eosinophil ci_ysta loid granules: structure, function, and beyond. J Leukoc Biol. 2012 Aug;92(2):2 31-6.
- 30. Dua K, Malyla V, Singhvi G, Wadhwa R, Zrishna RV, Shukla SD, et al. Increasing complexity and interactions of oxidate 'estress in chronic respiratory diseases: An emerging need for novel drug deliger, systems. Chem Biol Interact. 2019 Feb 1;299:168–78.
- Panagopoulos V, Zinonos I, I each DA, Hay SJ, Liapis V, Zysk A, et al. Uncovering a new role for peroxidase enzymes as drivers of angiogenesis. Int J Biochem Cell Biol. 2015 Nov;68:128–38.
- 32. Ochkur SI, Kim JD, Procheroe CA, Colbert D, Moqbel R, Lacy P, et al. The Development of a Sencitive and Specific ELISA for Mouse Eosinophil Peroxidase: Assessment of Eo inophil Degranulation Ex Vivo and in Models of Human Disease. J Immunol Method's. 2012 Jan 31;375(1-2):138-47.
- Puxeddu I, Berkman N, Nissim Ben Efraim AH, Davies DE, Ribatti D, Gleich GJ, et al. The role of eosinophil major basic protein in angiogenesis. Allergy. 2009 Mar;64(3):368–74.
- 34. Tsuda T, Maeda Y, Nishide M, Koyama S, Hayama Y, Nojima S, et al. Eosinophilderived neurotoxin enhances airway remodeling in eosinophilic chronic rhinosinusitis and correlates with disease severity. Int Immunol. 2019 Feb 6;31(1):33–40.
- 35. Yang D, Chen Q, Rosenberg HF, Rybak SM, Newton DL, Wang ZY, et al. Human Ribonuclease A Superfamily Members, Eosinophil-Derived Neurotoxin and Pancreatic Ribonuclease, Induce Dendritic Cell Maturation and Activation. J Immunol. 2004 Nov 15;173(10):6134–42.

- 36. Domachowske JB, Dyer KD, Bonville CA, Rosenberg HF. Recombinant human eosinophil-derived neurotoxin/RNase 2 functions as an effective antiviral agent against respiratory syncytial virus. J Infect Dis. 1998 Jun;177(6):1458–64.
- 37. Rugeles MT, Trubey CM, Bedoya VI, Pinto LA, Oppenheim JJ, Rybak SM, et al. Ribonuclease is partly responsible for the HIV-1 inhibitory effect activated by HLA alloantigen recognition. AIDS Lond Engl. 2003 Mar 7;17(4):481–6.
- 38. Chua JC, Douglass JA, Gillman A, O'Hehir RE, Meeusen EN. Galectin-10, a potential biomarker of eosinophilic airway inflammation. PloS One. 2012;7(8):e42549.
- 39. Carmo LAS, Bonjour K, Ueki S, Neves JS, Liu L, Spencer LA, et al. CD63 is tightly associated with intracellular, secretory events chaperoning piecemeal degranulation and compound exocytosis in human eosinophils. J Leuk c Biol. 2016;100(2):391–401.
- 40. Spencer LA, Bonjour K, Melo RCN, Weller PF. Eosinoph' Secretion of Granule-Derived Cytokines. Front Immunol [Internet]. 2014 Oct 27 [cited 2020 Apr 9];5. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4209865/
- 41. Wright BL, Ochkur SI, Olson NS, Shim KP, Jacobsen EA, Rank MA, et al. Normalized serum eosinophil peroxidase levels are inversely correlated with esophageal eosinophilia in eosinophilic esophagitis. Dis Esophagus [Internet]. 2018 Feb 1 [cited 2020 Apr 9];31(2). Available from 1: https://academic.oup.com/dote/article/2 1/2/dox139/4714780
- 42. Gleich GJ, Leiferman KM. The hy_F ~ eosinophilic syndromes: current concepts and treatments. Br J Haematol. 2009 May; 45(3):271–85.
- 43. Mejia R, Nutman TB. Evalua 101 and differential diagnosis of marked, persistent eosinophilia. Semin Hematol 2012 Apr;49(2):149–59.
- 44. Simon H-U, Rothenberg ME, Bochner BS, Weller PF, Wardlaw AJ, Wechsler ME, et al. Refining the definitio. of hypereosinophilic syndrome. J Allergy Clin Immunol. 2010 Jul;126(1):45–9.
- 45. Mejia R, Nutma. L. Evaluation and differential diagnosis of marked, persistent eosinophilia. Semir Hematol. 2012 Apr;49(2):149–59.
- 46. McBrien CN, Menzies-Gow A. The Biology of Eosinophils and Their Role in Asthma. Front Med [Internet]. 2017 Jun 30 [cited 2020 Jan 19];4. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5491677/
- 47. Mehta M, Dhanjal DS, Paudel KR, Singh B, Gupta G, Rajeshkumar S, et al. Cellular signalling pathways mediating the pathogenesis of chronic inflammatory respiratory diseases: an update. Inflammopharmacology. 2020 Aug 1;28(4):795–817.
- Chan Y, Ng SW, Chellappan DK, Madheswaran T, Zeeshan F, Kumar P, et al. Celastrol-loaded liquid crystalline nanoparticles as an anti-inflammatory intervention for the treatment of asthma. Int J Polym Mater Polym Biomater. 2020 May 31;0(0):1– 10.

- 49. Dharmage SC, Perret JL, Custovic A. Epidemiology of Asthma in Children and Adults. Front Pediatr. 2019 Jun 18;7.
- 50. Matucci A, Maggi E, Vultaggio A. Eosinophils, the IL-5/IL-5Rα axis, and the biologic effects of benralizumab in severe asthma. Respir Med. 2019 Dec;160:105819.
- 51. Carr TF, Zeki AA, Kraft M. Eosinophilic and Noneosinophilic Asthma. Am J Respir Crit Care Med. 2018 Jan 1;197(1):22–37.
- 52. Brussino L, Heffler E, Bucca C, Nicola S, Rolla G. Eosinophils Target Therapy for Severe Asthma: Critical Points. BioMed Res Int. 2018;2018:7582057.
- 53. Bakakos A, Loukides S, Bakakos P. Severe Eosinophilic Asthma. J Clin Med. 2019 Sep 2;8(9).
- 54. Kandikattu HK, Upparahalli Venkateshaiah S, Mishra A. Dvnergy of Interleukin (IL)-5 and IL-18 in eosinophil mediated pathogenesis of allers in classes. Cytokine Growth Factor Rev. 2019 Jun;47:83–98.
- 55. Varricchi G, Bagnasco D, Ferrando M, Puggior F, Passalacqua G, Canonica GW. Mepolizumab in the management of severe eosinov hilic asthma in adults: current evidence and practical experience. Ther Adv F.esp.: Dis. 2017;11(1):40–5.
- 56. Beckert H, Meyer-Martin H, Buhl R, Teube C, Reuter S. Single and Synergistic Effects of Type 2 Cytokines on Eocine phils and Asthma Hallmarks. J Immunol Baltim Md 1950. 2019 Dec 20;
- 57. Stolarski B, Kurowska-Stolarska M, Kewin P, Xu D, Liew FY. IL-33 Exacerbates Eosinophil-Mediated Airway in Commation. J Immunol. 2010 Sep 15;185(6):3472–80.
- 58. Fanat AI, Thomson JV, Rodford K, Nair P, Sehmi R. Human airway smooth muscle promotes eosinophil differentiation. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 2009 Jul;39(7):1009–17.
- 59. Yi S, Zhai J, Niu K, Zhu G, Wang M, Liu J, et al. Eosinophil recruitment is dynamically regulated by interplay among lung dendritic cell subsets after allergen challenge. Nat Con mun. 2018 Sep 24;9(1):1–14.
- 60. Shen Z-J, Malter JS. Determinants of eosinophil survival and apoptotic cell death. Apoptosis. 2015 Feb 1;20(2):224–34.
- 61. Muraki M, Gleich GJ, Kita H. Antigen-specific IgG and IgA, but not IgE, activate the effector functions of eosinophils in the presence of antigen. Int Arch Allergy Immunol. 2011;154(2):119–27.
- Brannan JDP, Lougheed MDM. Airway Hyperresponsiveness in Asthma: Mechanisms, Clinical Significance, and Treatment. Front Physiol [Internet]. 2012 [cited 2020 Sep 17];3. Available from: https://www.frontiersin.org/articles/10.3389/fphys.2012.00460/full
- 63. Mehta M, Satija S, Paudel KR, Liu G, Chellappan DK, Hansbro PM, et al. Incipient need of targeting airway remodeling using advanced drug delivery in chronic

respiratory diseases. Future Med Chem [Internet]. [cited 2020 Sep 17]; Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7319495/

- 64. Zagai U, Lundahl J, Klominek J, Venge P, Sköld CM. Eosinophil cationic protein stimulates migration of human lung fibroblasts in vitro. Scand J Immunol. 2009 Apr;69(4):381–6.
- 65. Halwani R, Vazquez-Tello A, Sumi Y, Pureza MA, Bahammam A, Al-Jahdali H, et al. Eosinophils induce airway smooth muscle cell proliferation. J Clin Immunol. 2013 Apr;33(3):595–604.
- 66. Md S, Wc C, K D, Gm P, Rp P, Mq M, et al. Emerging concepts and directed therapeutics for the management of asthma: regulating the regulators. Inflammopharmacology. 2020 Nov 5;
- 67. Narendra D, Blixt J, Hanania NA. Immunological bion ark rs in severe asthma. Semin Immunol. 2019 Dec;46:101332.
- 68. Szefler SJ, Wenzel S, Brown R, Erzurum SC, Fal. Jv, Hamilton RG, et al. Asthma outcomes: biomarkers. J Allergy Clin Immuno¹. 2¹2 Mar;129(3 Suppl):S9-23.
- 69. Murayama N, Murayama K. Nasal Discharge Cosh ophils in Childhood Asthma Patients as a Predictive Factor for Persistent A sthma. Mediators Inflamm. 2018;2018:2563978.
- 70. Westerhof GA, Korevaar DA, An, ^{lir} K M, Nijs SB de, Groot JC de, Wang J, et al. Biomarkers to identify sputum eosino₁ hilia in different adult asthma phenotypes. Eur Respir J. 2015 Sep 1;46(3):688->5
- 71. Wagener AH, de Nijs SB, Jutter 'k, Sousa AR, Weersink EJM, Bel EH, et al. External validation of blood eosine; hils, FE(NO) and serum periostin as surrogates for sputum eosinophils in asthma. T. orax. 2015 Feb;70(2):115–20.
- 72. Hastie AT, Moore WC J. H, Rector BM, Ortega VE, Pascual RM, et al. Biomarker surrogates do not ccu ately predict sputum eosinophil and neutrophil percentages in asthmatic subject J Allergy Clin Immunol. 2013 Jul;132(1):72–80.
- Walsh CJ, Zaihra T, Benedetti A, Fugère C, Olivenstein R, Lemière C, et al. Exacerbation risk in severe asthma is stratified by inflammatory phenotype using longitudinal measures of sputum eosinophils. Clin Exp Allergy. 2016;46(10):1291– 302.
- 74. Price DB, Rigazio A, Campbell JD, Bleecker ER, Corrigan CJ, Thomas M, et al. Blood eosinophil count and prospective annual asthma disease burden: a UK cohort study. Lancet Respir Med. 2015 Nov;3(11):849–58.
- 75. Petsky HL, Cates CJ, Kew KM, Chang AB. Tailoring asthma treatment on eosinophilic markers (exhaled nitric oxide or sputum eosinophils): a systematic review and meta-analysis. Thorax. 2018;73(12):1110–9.

- 76. Jia G, Erickson RW, Choy DF, Mosesova S, Wu LC, Solberg OD, et al. Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients. J Allergy Clin Immunol. 2012 Sep;130(3):647-654.e10.
- Simpson JL, Yang IA, Upham JW, Reynolds PN, Hodge S, James AL, et al. Periostin levels and eosinophilic inflammation in poorly-controlled asthma. BMC Pulm Med. 2016 Apr 30;16(1):67.
- Nair P, Ochkur SI, Protheroe C, Radford K, Efthimiadis A, Lee NA, et al. Eosinophil peroxidase in sputum represents a unique biomarker of airway eosinophilia. Allergy. 2013 Sep;68(9):1177–84.
- 79. Liu T, Wu J, Zhao J, Wang J, Zhang Y, Liu L, et al. Type 2 innate lymphoid cells: A novel biomarker of eosinophilic airway inflammation in patients with mild to moderate asthma. Respir Med. 2015 Nov;109(11):1391–6.
- 80. Fricker M, Gibson PG, Powell H, Simpson JL, Yang A, Tpham JW, et al. A sputum 6-gene signature predicts future exacerbations of pc orly controlled asthma. J Allergy Clin Immunol. 2019 Jul 1;144(1):51-60.e11.
- 81. Baines KJ, Negewo NA, Gibson PG, Fu J-J, Shappon JL, Wark PA, et al. A Sputum 6 Gene Expression Signature Predicts Inflammetory Thenotypes and Future Exacerbations of COPD. Int J Chron Obs rust Pulmon Dis. 2020 Jul 2;15:1577–90.
- 82. Desai D, Newby C, Symon FA, He da P, Shah S, Gupta S, et al. Elevated Sputum Interleukin-5 and Submucosal Eosling philia in Obese Individuals with Severe Asthma. Am J Respir Crit Care Med. 2013 Sep 15;188(6):657–63.
- 83. Sverrild A, Kiilerich P, Brejn oc. A, Pedersen R, Porsbjerg C, Bergqvist A, et al. Eosinophilic airway inflammation in asthmatic patients is associated with an altered airway microbiome. J Allergy Clin Immunol. 2017 Aug 1;140(2):407-417.e11.
- Dua K, Gupta G, Chellar van DK, Shukla S, Hansbro PM. Targeting bacterial biofilms in pulmonary diseases in pediatric population. Minerva Pediatr. 2019 Jun;71(3):309– 10.
- 85. Chellappan DK, Sz Ning QL, Su Min SK, Bin SY, Chern PJ, Shi TP, et al. Interactions between microbiome and lungs: Paving new paths for microbiome based bio-engineered drug delivery systems in chronic respiratory diseases. Chem Biol Interact. 2019 Sep 1;310:108732.
- 86. Hosoki K, Nakamura A, Kainuma K, Sugimoto M, Nagao M, Hiraguchi Y, et al. Differential Activation of Eosinophils by Bacteria Associated with Asthma. Int Arch Allergy Immunol. 2013;161(Suppl. 2):16–22.
- 87. Patel SS, Casale TB, Cardet JC. Biological therapies for eosinophilic asthma. Expert Opin Biol Ther. 2018 Jul;18(7):747–54.
- 88. Farne HA, Wilson A, Powell C, Bax L, Milan SJ. Anti- IL5 therapies for asthma. Cochrane Database Syst Rev [Internet]. 2017 Sep 21 [cited 2020 Jan 30];2017(9). Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6483800/

- 89. Casale TB, Pacou M, Mesana L, Farge G, Sun SX, Castro M. Reslizumab Compared with Benralizumab in Patients with Eosinophilic Asthma: A Systematic Literature Review and Network Meta-Analysis. J Allergy Clin Immunol Pract. 2019;7(1):122-130.e1.
- 90. Lim HF, Nair P. Efficacy and safety of reslizumab in patients with moderate to severe eosinophilic asthma. Expert Rev Respir Med. 2015 Apr;9(2):135–42.
- 91. Castro M, Zangrilli J, Wechsler ME, Bateman ED, Brusselle GG, Bardin P, et al. Reslizumab for inadequately controlled asthma with elevated blood eosinophil counts: results from two multicentre, parallel, double-blind, randomised, placebo-controlled, phase 3 trials. Lancet Respir Med. 2015 May;3(5):355–66.
- 92. Haldar P, Brightling CE, Hargadon B, Gupta S, Monteir W, Sousa A, et al. Mepolizumab and Exacerbations of Refractory Eosinophin. Asthma. N Engl J Med. 2009 Mar 5;360(10):973–84.
- 93. Nair P, Pizzichini MMM, Kjarsgaard M, Inman ML Ef himiadis A, Pizzichini E, et al. Mepolizumab for prednisone-dependent asthma w⁻th sputum eosinophilia. N Engl J Med. 2009 Mar 5;360(10):985–93.
- 94. Sehmi R, Smith SG, Kjarsgaard M, Radford F., Boulet L-P, Lemiere C, et al. Role of local eosinophilopoietic processes in the (evel opment of airway eosinophilia in prednisone-dependent severe asthma. C in Exp Allergy J Br Soc Allergy Clin Immunol. 2016;46(6):793–802.
- 95. Busse W, Chupp G, Nagase H, Albers FC, Doyle S, Shen Q, et al. Anti-IL-5 treatments in patients with severe asthma by 0100d eosinophil thresholds: Indirect treatment comparison. J Allergy Clin Ir ur a. ol. 2019;143(1):190-200.e20.
- 96. Poulakos MN, Cargill SM, Waineo MF, Wolford AL. Mepolizumab for the treatment of severe eosinophilic as. ma. Am J Health-Syst Pharm AJHP Off J Am Soc Health-Syst Pharm. 2017 Jul 1; ⁴(13):963–9.
- Dávila González I Mc reno Benítez F, Quirce S. Benralizumab: A New Approach for the Treatment of Cevere Eosinophilic Asthma. J Investig Allergol Clin Immunol. 2019 Apr;29(2):84–93
- 98. Caminati M, Bagnasco D, Vaia R, Senna G. New horizons for the treatment of severe, eosinophilic asthma: benralizumab, a novel precision biologic. Biol Targets Ther. 2019 May 22;13:89–95.
- 99. Sridhar S, Liu H, Pham T-H, Damera G, Newbold P. Modulation of blood inflammatory markers by benralizumab in patients with eosinophilic airway diseases. Respir Res. 2019 Jan 18;20(1):14.
- 100. Benralizumab for asthma. Aust Prescr. 2018;41:164-5.
- Kupryś-Lipińska I, Molińska K, Kuna P. The effect of omalizumab on eosinophilic inflammation of the respiratory tract in patients with allergic asthma. Pneumonol Alergol Pol. 2016;84(4):232–43.

- Massanari M, Holgate ST, Busse WW, Jimenez P, Kianifard F, Zeldin R. Effect of omalizumab on peripheral blood eosinophilia in allergic asthma. Respir Med. 2010 Feb 1;104(2):188–96.
- 103. Busse W, Corren J, Lanier BQ, McAlary M, Fowler-Taylor A, Cioppa GD, et al. Omalizumab, anti-IgE recombinant humanized monoclonal antibody, for the treatment of severe allergic asthma. J Allergy Clin Immunol. 2001 Aug;108(2):184–90.
- 104. Vatrella A, Fabozzi I, Calabrese C, Maselli R, Pelaia G. Dupilumab: a novel treatment for asthma. J Asthma Allergy. 2014 Sep 4;7:123–30.
- Deeks ED. Dupilumab: A Review in Moderate to Severe Asthma. Drugs. 2019 Nov;79(17):1885–95.
- 106. Samitas K, Rådinger M, Bossios A. Current update on eos. pophilic lung diseases and anti-IL-5 treatment. Recent Patents Anti-Infect Drug D sc. 2011 Sep 1;6(3):189–205.
- 107. Kolbeck R, Kozhich A, Koike M, Peng L, Anderssen C, C, Damschroder MM, et al. MEDI-563, a humanized anti-IL-5 receptor alpha nAo with enhanced antibodydependent cell-mediated cytotoxicity function. *J Albergy Clin Immunol.* 2010 Jun;125(6):1344-1353.e2.
- 108. Bochner BS, Gleich GJ. What targeting e is a ophils has taught us about their role in diseases. J Allergy Clin Immunol. 2010 Jun 1,126(1):16–25.
- 109. Komai M, Tanaka H, Nagao K, Ishiz Ki M, Kajiwara D, Miura T, et al. A novel CCchemokine receptor 3 antagonist, Ki1> 003, inhibits airway eosinophilia and subepithelial/peribronchial fibrosic induced by repeated antigen challenge in mice. J Pharmacol Sci. 2010;112(2):? 0? - 13.
- 110. Bel EH, ten Brinke A. Net: And-Eosinophil Drugs for Asthma and COPD: Targeting the Trait! Chest. 2017 Dec 1; 152(6):1276-82.
- 111. Gonem S, Berair R, Shor puri A, Hartley R, Laurencin MFM, Bacher G, et al. Fevipiprant, a pro tagi indin D2 receptor 2 antagonist, in patients with persistent eosinophilic astinna. a single-centre, randomised, double-blind, parallel-group, placebo-controlled rial. Lancet Respir Med. 2016;4(9):699–707.
- 112. Watkins N. Novartis' Fevipiprant Fails in Phase 3 Trials for Treatment of Patients with Uncontrolled Asthma [Internet]. Specialty Pharma Journal. 2019 [cited 2020 Dec 17]. Available from: https://www.spjnews.com/2019/12/16/novartis-fevipiprant-fails-in-phase-3-trials-for-treatment-of-patients-with-uncontrolled-asthma/
- 113. Gauvreau GM, O'Byrne PM, Boulet L-P, Wang Y, Cockcroft D, Bigler J, et al. Effects of an Anti-TSLP Antibody on Allergen-Induced Asthmatic Responses [Internet]. http://dx.doi.org/10.1056/NEJMoa1402895. 2014 [cited 2020 Jan 31]. Available from: https://www.nejm.org/doi/10.1056/NEJMoa1402895
- 114. Mehta M, Deeksha null, Tewari D, Gupta G, Awasthi R, Singh H, et al. Oligonucleotide therapy: An emerging focus area for drug delivery in chronic inflammatory respiratory diseases. Chem Biol Interact. 2019 Aug 1;308:206–15.

- 115. Dua K, Wadhwa R, Singhvi G, Rapalli V, Shukla SD, Shastri MD, et al. The potential of siRNA based drug delivery in respiratory disorders: Recent advances and progress. Drug Dev Res. 2019;80(6):714–30.
- 116. Dua K, Chellappan DK, Singhvi G, de Jesus Andreoli Pinto T, Gupta G, Hansbro PM. Targeting microRNAs using nanotechnology in pulmonary diseases. Panminerva Med. 2018 Dec;60(4):230–1.
- 117. Mehta M, Chellappan DK, Wich PR, Hansbro NG, Hansbro PM, Dua K. miRNA nanotherapeutics: potential and challenges in respiratory disorders. Future Med Chem. 2020 Apr 9;12(11):987–90.
- 118. Imaoka H, Campbell H, Babirad I, Watson RM, Mistry M, Sehmi R, et al. TPI ASM8 reduces eosinophil progenitors in sputum after allergen challenge. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 2011 Dec;41(12):1740–6.
- 119. Ramsahai JM, Wark PA. Appropriate use of oral corticol toroids for severe asthma. Med J Aust. 2018 16;209(S2):S18–21.
- 120. Durham AL, Caramori G, Chung KF, Adcock Tvi. Largeted anti-inflammatory therapeutics in asthma and chronic obstructive rung disease. Transl Res J Lab Clin Med. 2016 Jan;167(1):192–203.
- 121. Dente FL, Bacci E, Bartoli ML, Cianchetti C, Costa F, Di Franco A, et al. Effects of oral prednisone on sputum eosinop'and and cytokines in patients with severe refractory asthma. Ann Allergy Asthma Immer of Off Publ Am Coll Allergy Asthma Immunol. 2010 Jun;104(6):464–70.
- 122. Prazma CM, Bel EH, Price R J, L radford ES, Albers FC, Yancey SW. Oral corticosteroid dose changes and impact on peripheral blood eosinophil counts in patients with severe eosinophilic asthma: a post hoc analysis. Respir Res. 2019 May 3;20(1):83.
- 123. Lommatzsch M, Klein M, Stoll P, Virchow JC. Impact of an increase in the inhaled corticosteroid dos on blood eosinophils in asthma. Thorax. 2019;74(4):417–8.
- 124. Cherk Yong DO, S ker SR, Wadhwa R, Chellappan DK, Madheswaran T, Panneerselvam J, et al. Preparation, characterization and in-vitro efficacy of quercetin loaded liquid crystalline nanoparticles for the treatment of asthma. J Drug Deliv Sci Technol. 2019 Dec 1;54:101297.
- 125. Sakai-Kashiwabara M, Asano K. Inhibitory Action of Quercetin on Eosinophil Activation In Vitro. Evid-Based Complement Altern Med ECAM [Internet]. 2013 [cited 2020 Apr 18];2013. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3690238/
- 126. Sakai-Kashiwabara M, Abe S, Asano K. Suppressive activity of quercetin on the production of eosinophil chemoattractants from eosinophils in vitro. Vivo Athens Greece. 2014 Aug;28(4):515–22.

- 127. Hewlings SJ, Kalman DS. Curcumin: A Review of Its' Effects on Human Health. Foods [Internet]. 2017 Oct 22 [cited 2020 Apr 18];6(10). Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5664031/
- 128. Ng ZY, Wong J-Y, Panneerselvam J, Madheswaran T, Kumar P, Pillay V, et al. Assessing the potential of liposomes loaded with curcumin as a therapeutic intervention in asthma. Colloids Surf B Biointerfaces. 2018 Dec 1;172:51–9.
- 129. Chellappan DK, Ng ZY, Wong J-Y, Hsu A, Wark P, Hansbro N, et al. Immunological axis of curcumin-loaded vesicular drug delivery systems. Future Med Chem. 2018 Apr 1;10(8):839–44.
- Chellappan DK, Hansbro PM, Dua K, Hsu A, Gupta G, Ng ZY, et al. Vesicular Systems Containing Curcumin and Their Applications ir Respiratory Disorders - A Mini Review. Pharm Nanotechnol. 2017;5(4):250–4.
- 131. Vestbo J, Hurd SS, Agustí AG, Jones PW, Vogelmeier C, Anzueto A, et al. Global Strategy for the Diagnosis, Management, and Prevention of Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med. 2013 Feb 15;187(4):347–65.
- 132. Wadhwa R, Aggarwal T, Malyla V, Kumar N, Sur ta G, Chellappan DK, et al. Identification of biomarkers and genetic appreaches toward chronic obstructive pulmonary disease. J Cell Physiol. 2019;2 34(10):16703–23.
- 133. Mehta M, Deeksha, Tewari D, Gur a G, A wasthi R, Singh H, et al. Oligonucleotide therapy: An emerging focus area to rug delivery in chronic inflammatory respiratory diseases. Chem Biol Interact. 2019 Au₅ 1;308:206–15.
- 134. Chellappan DK, Yee LW, Xu in TY, Kunalan K, Rou LC, Jean LS, et al. Targeting neutrophils using novel drug delivery systems in chronic respiratory diseases. Drug Dev Res. 2020;81(4):419-.36.
- 135. van Haarst A, McGarvey L, Paglialunga S. Review of Drug Development Guidance to Treat Chronic Obstructive Pulmonary Disease: US and EU Perspectives. Clin Pharmacol Ther. 2019 Dec;106(6):1222–35.
- 136. Hogg JC. A brief r^e view of chronic obstructive pulmonary disease. Can Respir J J Can Thorac Soc. 2012;19(6):381–4.
- 137. Barnes PJ, Burney PGJ, Silverman EK, Celli BR, Vestbo J, Wedzicha JA, et al. Chronic obstructive pulmonary disease. Nat Rev Dis Primer. 2015 03;1:15076.
- 138. Mehta M, Deeksha null, Sharma N, Vyas M, Khurana N, Maurya PK, et al. Interactions with the macrophages: An emerging targeted approach using novel drug delivery systems in respiratory diseases. Chem Biol Interact. 2019 May 1;304:10–9.
- Butler A, Walton GM, Sapey E. Neutrophilic Inflammation in the Pathogenesis of Chronic Obstructive Pulmonary Disease. COPD J Chronic Obstr Pulm Dis. 2018 Jul 4;15(4):392–404.
- 140. Tashkin DP, Wechsler ME. Role of eosinophils in airway inflammation of chronic obstructive pulmonary disease. Int J Chron Obstruct Pulmon Dis. 2018;13:335–49.

- 141. Kim VL, Coombs NA, Staples KJ, Ostridge KK, Williams NP, Wootton SA, et al. Impact and associations of eosinophilic inflammation in COPD: analysis of the AERIS cohort. Eur Respir J. 2017;50(4).
- 142. Tworek D, Antczak A. Eosinophilic COPD a distinct phenotype of the disease. Adv Respir Med. 2017;85(5):271–6.
- 143. George L, Brightling CE. Eosinophilic airway inflammation: role in asthma and chronic obstructive pulmonary disease. Ther Adv Chronic Dis. 2016 Jan;7(1):34–51.
- 144. Eltboli O, Bafadhel M, Hollins F, Wright A, Hargadon B, Kulkarni N, et al. COPD exacerbation severity and frequency is associated with impaired macrophage efferocytosis of eosinophils. BMC Pulm Med. 2014 Jul 9;14:112.
- 145. Kume H, Hojo M, Hashimoto N. Eosinophil Inflammation and Hyperresponsiveness in the Airways as Phenotypes of COPD, and Usefulness criminated Glucocorticosteroids. Front Pharmacol. 2019;10:765.
- 146. Singh D, Kolsum U, Brightling CE, Locantore N, Agusti A, Tal-Singer R. Eosinophilic inflammation in COPD: prevalence and clinical claracteristics. Eur Respir J. 2014 Dec 1;44(6):1697–700.
- 147. Cukic V, Lovre V, Dragisic D, Ustamujic A. Asthma and Chronic Obstructive Pulmonary Disease (COPD) – Differen⁴ es and Similarities. Mater Socio-Medica. 2012;24(2):100–5.
- 148. Pignatti P, Visca D, Cherubino F, Zan, Jogna E, Lucini E, Saderi L, et al. Do blood eosinophils strictly reflect airway inflammation in COPD? Comparison with asthmatic patients. Respir Res. 2019 Jul 10, 79(1):145.
- 149. Couillard S, Larivée P, Cc rteau J, Vanasse A. Eosinophils in COPD Exacerbations Are Associated With Inc. eased Readmissions. Chest. 2017 Feb;151(2):366–73.
- 150. Hyun D-G, Lee JH, G: Y-M, Lee SW, Lee SD, Lee JS. Association of plasma fibrinogen concentrations and blood eosinophil counts with clinical phenotypes of COPD. Int J Tubere Lung Dis Off J Int Union Tubere Lung Dis. 2019 01;23(9):1035– 41.
- 151. Thong L, O'Driscoll M, Casey C, Kennedy M, Plant BJ, Henry MT, et al. Eosinophils and COPD Readmission. Chest. 2017;151(3):724–5.
- 152. Fuschillo S, Molino A, Stellato C, Motta A, Maniscalco M. Blood eosinophils as biomarkers of therapeutic response to chronic obstructive pulmonary disease: Still work in progress. Eur J Intern Med. 2019 Oct;68:1–5.
- 153. Wadhwa R, Aggarwal T, Malyla V, Kumar N, Gupta G, Chellappan DK, et al. Identification of biomarkers and genetic approaches toward chronic obstructive pulmonary disease. J Cell Physiol. 2019 Aug;234(10):16703–23.
- 154. Tinè M, Biondini D, Semenzato U, Bazzan E, Cosio MG, Saetta M, et al. Reassessing the Role of Eosinophils as a Biomarker in Chronic Obstructive Pulmonary Disease. J Clin Med. 2019 Jul 2;8(7).

- 155. Vedel-Krogh S, Nielsen SF, Lange P, Vestbo J, Nordestgaard BG. Blood Eosinophils and Exacerbations in Chronic Obstructive Pulmonary Disease. The Copenhagen General Population Study. Am J Respir Crit Care Med. 2016 01;193(9):965–74.
- 156. Hastie AT, Martinez FJ, Curtis JL, Doerschuk CM, Hansel NN, Christenson S, et al. Association of sputum and blood eosinophil concentrations with clinical measures of COPD severity: an analysis of the SPIROMICS cohort. Lancet Respir Med. 2017;5(12):956–67.
- 157. Oh Y-M, Lee KS, Hong Y, Hwang SC, Kim JY, Kim DK, et al. Blood eosinophil count as a prognostic biomarker in COPD. Int J Chron Obstruct Pulmon Dis. 2018 Oct 31;13:3589–96.
- 158. DiSantostefano RL, Hinds D, Le HV, Barnes NC. Relationship between blood eosinophils and clinical characteristics in a cross-sectional tudy of a US population-based COPD cohort. Respir Med. 2016 Mar;112:88–9 ϵ .
- 159. Ko FWS, Chan KP, Ngai J, Ng S-S, Yip WH, Ip A, et a) Blood eosinophil count as a predictor of hospital length of stay in COPD exac. battons. Respirol Carlton Vic. 2019 Aug 6;
- 160. Mendy A, Forno E, Niyonsenga T, Gasana J. 'Jooc' biomarkers as predictors of long-term mortality in COPD. Clin Respir J. 2(15/14ay;12(5):1891–9.
- 161. Gonzalez-Barcala F-J, San-Jose M , Nie D-Fontarigo J-J, Calvo-Alvarez U, Carreira J-M, Garcia-Sanz M-T, et al. Blood e Jsinophils could be useful as a biomarker in chronic obstructive pulmonary disease exacerbations. Int J Clin Pract. 2019 Oct 1;e13423.
- 162. Couillard S, Larivée P, Courteal J, Vanasse A. Eosinophils in COPD Exacerbations Are Associated With Increased Readmissions. Chest. 2017 Feb;151(2):366–73.
- 163. Bélanger M, Couillard S, Courteau J, Larivée P, Poder TG, Carrier N, et al. Eosinophil counts in first COPD in repitalizations: a comparison of health service utilization. Int J Chron Obstruct Pi Imon Dis. 2018;13:3045–54.
- 164. Cheng S-L, Lin C-) I. Effectiveness using higher inhaled corticosteroid dosage in patients with COPD by different blood eosinophilic counts. Int J Chron Obstruct Pulmon Dis. 2016;11:2341–8.
- 165. Hillas G, Papaporfyriou A, Dimakou K, Papaioannou AI. Pharmacological treatment of stable COPD: need for a simplified approach. Postgrad Med. 2019 Dec 18;
- 166. Bafadhel M, Peterson S, Blas MAD, Calverley PM, Rennard SI, Richter K, et al. Predictors of exacerbation risk and response to budesonide in patients with chronic obstructive pulmonary disease: a post-hoc analysis of three randomised trials. Lancet Respir Med. 2018 Feb 1;6(2):117–26.
- 167. Pascoe S, Locantore N, Dransfield MT, Barnes NC, Pavord ID. Blood eosinophil counts, exacerbations, and response to the addition of inhaled fluticasone furoate to vilanterol in patients with chronic obstructive pulmonary disease: a secondary analysis

of data from two parallel randomised controlled trials. Lancet Respir Med. 2015 Jun;3(6):435–42.

- 168. Siddiqui SH, Guasconi A, Vestbo J, Jones P, Agusti A, Paggiaro P, et al. Blood Eosinophils: A Biomarker of Response to Extrafine Beclomethasone/Formoterol in Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med. 2015 Aug 15;192(4):523–5.
- 169. Pavord ID, Lettis S, Locantore N, Pascoe S, Jones PW, Wedzicha JA, et al. Blood eosinophils and inhaled corticosteroid/long-acting β -2 agonist efficacy in COPD. Thorax. 2016 Feb;71(2):118–25.
- 170. Vestbo J, Papi A, Corradi M, Blazhko V, Montagna I, Francisco C, et al. Single inhaler extrafine triple therapy versus long-acting muscarinic an agonist therapy for chronic obstructive pulmonary disease (TRINITY): a double-blina, parallel group, randomised controlled trial. The Lancet. 2017 May 13;389(10082): (91)–29.
- 171. McDonald CF. Eosinophils in chronic obstructive p umc nary disease: are they just another biomarker? Curr Opin Pulm Med. 2020 J. n 3;
- 172. Oshagbemi OA, Franssen FME, van Kraaij S, Lracken DCW, Wouters EFM, Maitland-van der Zee AH, et al. Blood Eosinc phil Counts, Withdrawal of Inhaled Corticosteroids and Risk of COPD Exace bations and Mortality in the Clinical Practice Research Datalink (CPRD). COPD. 2019;10(2):152–9.
- 173. Watz H, Tetzlaff K, Wouters EFM, Virsten A, Magnussen H, Rodriguez-Roisin R, et al. Blood eosinophil count and exacerbations in severe chronic obstructive pulmonary disease after withdrawal of inhale.¹ corticosteroids: a post-hoc analysis of the WISDOM trial. Lancet Respi Mic.¹. 2016 May 1;4(5):390–8.
- 174. Chapman KR, Hurst JR, Front S-M, Larbig M, Fogel R, Guerin T, et al. Long-Term Triple Therapy De-escalation to Indacaterol/Glycopyrronium in Patients with Chronic Obstructive Pulmonary Disease (SUNSET): A Randomized, Double-Blind, Triple-Dummy Clinical Tripl. Am J Respir Crit Care Med. 2018 May 20;198(3):329–39.
- 175. Magnussen H, D. se B, Rodriguez-Roisin R, Kirsten A, Watz H, Tetzlaff K, et al. Withdrawal of Inbraed Glucocorticoids and Exacerbations of COPD [Internet]. http://dx.doi.org/10.1056/NEJMoa1407154. Massachusetts Medical Society; 2014 [cited 2020 Mar 6]. Available from: https://www.nejm.org/doi/10.1056/NEJMoa1407154
- 176. Calzetta L, Matera MG, Braido F, Contoli M, Corsico A, Di Marco F, et al. Withdrawal of inhaled corticosteroids in COPD: A meta-analysis. Pulm Pharmacol Ther. 2017 Aug 1;45:148–58.
- 177. Vestbo J, Anderson JA, Brook RD, Calverley PMA, Celli B, Crim CC, et al. Effect of Treatment Withdrawal on Outcomes in the SUMMIT Study. In: C41 LONG ACTING BRONCHODILATOR THERAPY IN COPD II [Internet]. American Thoracic Society; 2017 [cited 2020 Mar 6]. p. A5483–A5483. (American Thoracic Society International Conference Abstracts). Available from:

https://www.atsjournals.org/doi/abs/10.1164/ajrccm-conference.2017.195.1_MeetingAbstracts.A5483

- 178. Vestbo J, Fabbri L, Papi A, Petruzzelli S, Scuri M, Guasconi A, et al. Inhaled corticosteroid containing combinations and mortality in COPD. Eur Respir J [Internet]. 2018 Dec 1 [cited 2020 Apr 11];52(6). Available from: https://erj.ersjournals.com/content/52/6/1801230
- 179. Vestbo J, Anderson JA, Brook RD, Calverley PMA, Celli BR, Crim C, et al. Fluticasone furoate and vilanterol and survival in chronic obstructive pulmonary disease with heightened cardiovascular risk (SUMMIT): a double-blind randomised controlled trial. Lancet Lond Engl. 2016 Apr 30;387(10030):1817–26.
- 180. Kreindler JL, Watkins ML, Lettis S, Tal-Singer R, Loca^{*} tore N. Effect of inhaled corticosteroids on blood eosinophil count in steroid-naïve patients with COPD. BMJ Open Respir Res. 2016;3(1):e000151.
- 181. Schumann DM, Tamm M, Kostikas K, Stolz D. Stavility of the Blood Eosinophilic Phenotype in Stable and Exacerbated COPD. Che. t. 2019 Sep;156(3):456–65.
- 182. Negewo NA, McDonald VM, Baines KJ, Wark PA, Simpson JL, Jones PW, et al. Peripheral blood eosinophils: a surrogate mar'.er for airway eosinophilia in stable COPD. Int J Chron Obstruct Pulmon Dis. 27/15;11:1495–504.
- 183. Oshagbemi OA, Burden AM, Brae'en DCW, Henskens Y, Wouters EFM, Driessen JHM, et al. Stability of Blood Eosthephils in Patients with Chronic Obstructive Pulmonary Disease and in Control Subjects, and the Impact of Sex, Age, Smoking, and Baseline Counts. Am J Respir Crit. Care Med. 2017 15;195(10):1402–4.
- 184. Lakshmi SP, Reddy AT, Reddy C. Emerging pharmaceutical therapies for COPD. Int J Chron Obstruct Pulmon Dis. 2017 Jul 21;12:2141–56.
- 185. Brightling CE, Bleecker Fk, Panettieri RA, Bafadhel M, She D, Ward CK, et al. Benralizumab for chro.ic obstructive pulmonary disease and sputum eosinophilia: a randomised, doub e-bi nd, placebo-controlled, phase 2a study. Lancet Respir Med. 2014 Nov;2(11):C91–901.
- 186. Pavord ID, Chanez P, Criner GJ, Kerstjens HAM, Korn S, Lugogo N, et al. Mepolizumab for Eosinophilic Chronic Obstructive Pulmonary Disease. N Engl J Med. 2017 26;377(17):1613–29.
- Assaf S, Hanania NA. Novel therapeutic targets and drug development for the precision treatment of COPD. Expert Rev Precis Med Drug Dev. 2019 May 4;4(3):121–8.
- 188. Gross NJ, Barnes PJ. New Therapies for Asthma and Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med. 2016 Dec 6;195(2):159–66.
- 189. Criner GJ, Celli BR, Brightling CE, Agusti A, Papi A, Singh D, et al. Benralizumab for the Prevention of COPD Exacerbations. N Engl J Med. 2019 12;381(11):1023–34.

- 190. Sridhar S, Liu H, Pham T-H, Damera G, Newbold P. Modulation of blood inflammatory markers by benralizumab in patients with eosinophilic airway diseases. Respir Res. 2019 Jan 18;20(1):14.
- 191. Greulich T, Hohlfeld JM, Neuser P, Lueer K, Klemmer A, Schade-Brittinger C, et al. A GATA3-specific DNAzyme attenuates sputum eosinophilia in eosinophilic COPD patients: a feasibility randomized clinical trial. Respir Res. 2018 04;19(1):55.
- 192. Turowska A, Librizzi D, Baumgartl N, Kuhlmann J, Dicke T, Merkel O, et al. Biodistribution of the GATA-3-specific DNAzyme hgd40 after inhalative exposure in mice, rats and dogs. Toxicol Appl Pharmacol. 2013 Oct 15;272(2):365–72.
- 193. Brown SD, White R, Tobin P. Keep them breathing: Cystic fibrosis pathophysiology, diagnosis, and treatment. J Am Acad PAs. 2017 May;30'5):23–27.
- 194. Bergeron C, Cantin AM. Cystic Fibrosis: Pathophysiology of Lung Disease. Semin Respir Crit Care Med. 2019 Dec;40(6):715–26.
- 195. Zhang L, Borish L, Smith A, Somerville L, Albor D. Ose of mepolizumab in adult patients with cystic fibrosis and an eosinophilic plenotype: case series. Allergy Asthma Clin Immunol Off J Can Soc Allergy Clin Immunol. 2020;16:3.
- 196. Haack A, Aragão GG, Novaes MRCG. Path of hysiology of cystic fibrosis and drugs used in associated digestive tract diseases. World J Gastroenterol WJG. 2013 Dec 14;19(46):8552–61.
- 197. Dua K, De Jesus Andreoli Pinto T, Chellappan DK, Gupta G, Bebawy M, Hansbro PM. Advancements in nano drug delivery systems: A challenge for biofilms in respiratory diseases. 2018 Mar 1 [cited 2/12/1 Apr 18]; Available from: https://opus.lib.uts.edu.au/hand.c.10453/128257
- 198. Davies JC, Alton EWFV, Bush A. Cystic fibrosis. BMJ. 2007 Dec 13;335(7632):1255–9.
- 199. Rey MM, Bonk MP, Eadjiliadis D. Cystic Fibrosis: Emerging Understanding and Therapies. Annu Rev Med. 2019 27;70:197–210.
- Cohen-Cymberknoh M, Kerem E, Ferkol T, Elizur A. Airway inflammation in cystic fibrosis: molecular mechanisms and clinical implications. Thorax. 2013 Dec 1;68(12):1157–62.
- 201. Roesch EA, Nichols DP, Chmiel JF. Inflammation in cystic fibrosis: An update. Pediatr Pulmonol. 2018;53(S3):S30–50.
- 202. Tiringer K, Treis A, Fucik P, Gona M, Gruber S, Renner S, et al. A Th17- and Th2skewed Cytokine Profile in Cystic Fibrosis Lungs Represents a Potential Risk Factor for Pseudomonas aeruginosa Infection. Am J Respir Crit Care Med. 2013 Mar 15;187(6):621–9.
- 203. Koller DY, Götz M, Eichler I, Urbanek R. Eosinophilic activation in cystic fibrosis. Thorax. 1994 May;49(5):496–9.

- 204. Dy K, R U, M G. Increased degranulation of eosinophil and neutrophil granulocytes in cystic fibrosis. Am J Respir Crit Care Med. 1995 Aug 1;152(2):629–33.
- 205. Koller DY, Nilsson M, Enander I, Venge P, Eichler I. Serum eosinophil cationic protein, eosinophil protein X and eosinophil peroxidase in relation to pulmonary function in cystic fibrosis. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 1998 Feb;28(2):241–8.
- 206. Dy K, I N, J O, R U, I E. Cytokine concentrations in sputum from patients with cystic fibrosis and their relation to eosinophil activity. Am J Respir Crit Care Med. 1997 Mar 1;155(3):1050–4.
- 207. Condren ME, Bradshaw MD. Ivacaftor: A Novel Gene-Based Therapeutic Approach for Cystic Fibrosis. J Pediatr Pharmacol Ther JPPT. 2017;18(1):8–13.
- 208. Dua K, Awasthi R, Madan JR, Chellappan DK, Nalluri BN Gupta G, et al. Novel drug delivery approaches in treating pulmonary fibrosis. Panatterva Med. 2018 Dec;60(4):238–40.
- 209. Porsio B, Craparo EF, Mauro N, Giammona G C, vallaro G. Mucus and Cell-Penetrating Nanoparticles Embedded in Nano-Arto Micro Formulations for Pulmonary Delivery of Ivacaftor in Patients with Cystic Fabrosis. ACS Appl Mater Interfaces. 2018 Jan 10;10(1):165–81.
- 210. Jain V, Bhardwaj A. Pneumonia Princlogy. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2020 [Sited 2020 Apr 6]. Available from: http://www.ncbi.nlm.nih.gov/books/NDK526116/
- 211. Mackenzie G. The definition in plassification of pneumonia. Pneumonia [Internet].
 2016 Aug 22 [cited 2020 A⁻⁻₂ 6];5. Available from: https://www.ncbi.nlm.nih.₂ ov/pmc/articles/PMC5471962/
- 212. Pahal P, Sharma S. Eosh opnilic Pneumonia. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishir.g; 2020 [cited 2020 Apr 7]. Available from: http://www.ncbi.n m.n h.gov/books/NBK537169/
- 213. De Giacomi F, Vas allo R, Yi ES, Ryu JH. Acute Eosinophilic Pneumonia. Causes, Diagnosis, and Management. Am J Respir Crit Care Med. 2017 Dec 5;197(6):728–36.
- 214. Akuthota P, Weller PF. Eosinophilic Pneumonias. Clin Microbiol Rev. 2012 Oct 1;25(4):649–60.
- 215. Katoh S, Ikeda M, Matsumoto N, Shimizu H, Abe M, Ohue Y, et al. Possible Role of IL-25 in Eosinophilic Lung Inflammation in Patients with Chronic Eosinophilic Pneumonia. Lung. 2017 Dec 1;195(6):707–12.
- 216. Endo Y, Nakayama T. Pathogenic Th2 (Tpath2) cells in airway inflammation. Oncotarget. 2015 Oct 8;6(32):32303–4.
- 217. Cottin V. Eosinophilic Lung Diseases. Clin Chest Med. 2016 Sep 1;37(3):535–56.

- 218. Suzuki Y, Suda T. Eosinophilic pneumonia: A review of the previous literature, causes, diagnosis, and management. Allergol Int. 2019 Oct 1;68(4):413–9.
- 219. Crowe M, Robinson D, Sagar M, Chen L, Ghamande S. Chronic eosinophilic pneumonia: clinical perspectives. Ther Clin Risk Manag. 2019 Mar 13;15:397–403.
- 220. Abughanimeh O, Tahboub M, Abu Ghanimeh M. Metastatic Lung Adenocarcinoma Presenting with Hypereosinophilia. Cureus. 2018 Jun 22;10(6):e2866.
- 221. Sharma P, Mehta M, Dhanjal DS, Kaur S, Gupta G, Singh H, et al. Emerging trends in the novel drug delivery approaches for the treatment of lung cancer. Chem Biol Interact. 2019 Aug 25;309:108720.
- 222. Gupta G, de Jesus Andreoli Pinto T, Chellappan DK, Mishra A, Malipeddi H, Dua K. A clinical update on metformin and lung cancer in diabet. patients. Panminerva Med. 2018 Jun;60(2):70–5.
- 223. Varricchi G, Galdiero MR, Loffredo S, Lucarini V, Mar me G, Mattei F, et al. Eosinophils: The unsung heroes in cancer? Oncoi muuology. 2017 Nov 13;7(2).
- 224. Davis BP, Rothenberg ME. Eosinophils and ca. cer. Cancer Immunol Res. 2014 Jan;2(1):1–8.
- 225. Carretero R, Sektioglu IM, Garbi N, Salgato JC, Beckhove P, Hämmerling GJ. Eosinophils orchestrate cancer rejection by normalizing tumor vessels and enhancing infiltration of CD8(+) T cells. Nat Immunol. 2015 Jun;16(6):609–17.
- 226. Walsh M-T, Connell K, Sheahan AM, Gleich GJ, Costello RW. Eosinophil peroxidase signals via epidermal growth factor-2 to induce cell proliferation. Am J Respir Cell Mol Biol. 2011 Nov;45(5):946-5'2.
- 227. Lou Y, Marin-Acevedo A, Vishnu P, Manochakian R, Dholaria B, Soyano A, et al. Hypereosinophilia in a Patient with metastatic non-small-cell lung cancer treated with antiprogrammed cell Coat. 1 (anti-PD-1) therapy. Immunotherapy. 2019;11(7):577–84.
- 228. Sharma P, Mehu, M, Dhanjal DS, Kaur S, Gupta G, Singh H, et al. Emerging trends in the novel drug delivery approaches for the treatment of lung cancer. Chem Biol Interact. 2019 Aug 25;309:108720.
- 229. Dua K, Madan JR, Chellappan DK, Gupta G. Nanotechnology in drug delivery gaining new perspectives in respiratory diseases. Panminerva Med. 2018;60(3):135–6.
- 230. Takada K, Shimokawa M, Tanaka K, Kohashi K, Haro A, Osoegawa A, et al. Association between peripheral blood markers and immune-related factors on tumor cells in patients with resected primary lung adenocarcinoma. PloS One. 2019;14(6):e0217991.
- 231. Liu M, Wang X, Wang L, Ma X, Gong Z, Zhang S, et al. Targeting the IDO1 pathway in cancer: from bench to bedside. J Hematol OncolJ Hematol Oncol [Internet]. 2018 Aug 2 [cited 2020 Apr 7];11. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6090955/

- 232. Tanizaki J, Haratani K, Hayashi H, Chiba Y, Nakamura Y, Yonesaka K, et al. Peripheral Blood Biomarkers Associated with Clinical Outcome in Non-Small Cell Lung Cancer Patients Treated with Nivolumab. J Thorac Oncol Off Publ Int Assoc Study Lung Cancer. 2018;13(1):97–105.
- 233. Taghizadeh N, Vonk JM, Hospers JJ, Postma DS, de Vries EGE, Schouten JP, et al. Objective allergy markers and risk of cancer mortality and hospitalization in a large population-based cohort. Cancer Causes Control CCC. 2015 Jan;26(1):99–109.
- 234. Stankovic B, Bjørhovde HAK, Skarshaug R, Aamodt H, Frafjord A, Müller E, et al. Immune Cell Composition in Human Non-small Cell Lung Cancer. Front Immunol. 2018;9:3101.
- 235. Ye L, Wang H, Li H, Liu H, Lv T, Song Y, et al. Eosino bil peroxidase overexpression predicts the clinical outcome of patients with ρ. mary lung adenocarcinoma. J Cancer. 2019;10(4):1032–8.
- 236. Johnson ER, Matthay MA. Acute Lung Injury: Epicemielogy, Pathogenesis, and Treatment. J Aerosol Med Pulm Drug Deliv. 2010 Aug;23(4):243–52.
- 237. Willetts L, Parker K, Wesselius LJ, Protheroe CA Jaben E, Graziano P, et al. Immunodetection of occult eosinophils in lung tissue biopsies may help predict survival in acute lung injury. Respir Res. 2011;12(1):116.
- 238. Zilaee M, Hosseini SA, Jafarirad S, A oh. Zhadian F, Cheraghian B, Namjoyan F, et al. An evaluation of the effects of sof con supplementation on the asthma clinical symptoms and asthma severity in patients with mild and moderate persistent allergic asthma: a double-blind, randomized placebo-controlled trial. Respir Res. 2019;20(1):39–39.
- 239. Kurosawa M, Sutoh E. Propective Open-Label Study of 48-Week Subcutaneous Administration of Mepol. 7 umab in Japanese Patients With Severe Eosinophilic Asthma. J Investig Allerz ol Clin Immunol. 2019;29(1):40–5.
- 240. Ramos-Martínez I, Lc pez-Vancell MR, Fernández de Córdova-Aguirre JC, Rojas-Serrano J, Chava, ría A, Velasco-Medina A, et al. Reduction of respiratory infections in asthma patients sur plemented with vitamin D is related to increased serum IL-10 and IFNγ levels and cathelicidin expression. Cytokine. 2018;108:239–46.
- 241. Bleecker ER, Wechsler ME, FitzGerald JM, Menzies-Gow A, Wu Y, Hirsch I, et al. Baseline patient factors impact on the clinical efficacy of benralizumab for severe asthma. Eur Respir J. 2018;52(4).
- 242. Chipps BE, Newbold P, Hirsch I, Trudo F, Goldman M. Benralizumab efficacy by atopy status and serum immunoglobulin E for patients with severe, uncontrolled asthma. Ann Allergy Asthma Immunol Off Publ Am Coll Allergy Asthma Immunol. 2018;120(5):504-511.e4.
- 243. Lugogo N, Domingo C, Chanez P, Leigh R, Gilson MJ, Price RG, et al. Long-term Efficacy and Safety of Mepolizumab in Patients With Severe Eosinophilic Asthma: A Multi-center, Open-label, Phase IIIb Study. Clin Ther. 2016;38(9):2058-2070.e1.

- 244. Rossios C, Pavlidis S, Hoda U, Kuo C-H, Wiegman C, Russell K, et al. Sputum transcriptomics reveal upregulation of IL-1 receptor family members in patients with severe asthma. J Allergy Clin Immunol. 2018;141(2):560–70.
- 245. Casale TB, Chipps BE, Rosén K, Trzaskoma B, Haselkorn T, Omachi TA, et al. Response to omalizumab using patient enrichment criteria from trials of novel biologics in asthma. Allergy. 2018;73(2):490–7.
- 246. Ortega H, Yancey SW, Keene ON, Gunsoy NB, Albers FC, Howarth PH. Asthma Exacerbations Associated with Lung Function Decline in Patients with Severe Eosinophilic Asthma. J Allergy Clin Immunol Pract. 2018;6(3):980-986.e1.
- 247. Casale TB, Bateman ED, Vandewalker M, Virchow JC, Schmidt H, Engel M, et al. Tiotropium Respimat Add-on Is Efficacious in Sympton atic Asthma, Independent of T2 Phenotype. J Allergy Clin Immunol Pract. 2018;6(3):923-935.e9.
- 248. Chupp GL, Bradford ES, Albers FC, Bratton DJ, Wang-Jeiraj J, Nelsen LM, et al. Efficacy of mepolizumab add-on therapy on health-related quality of life and markers of asthma control in severe eosinophilic asthma (LUSCA): a randomised, doubleblind, placebo-controlled, parallel-group, multicentre, phase 3b trial. Lancet Respir Med. 2017;5(5):390–400.
- 249. Denlinger LC, Phillips BR, Ramratnam S F.o. s K, Bhakta NR, Cardet JC, et al. Inflammatory and Comorbid Features c Patients with Severe Asthma and Frequent Exacerbations. Am J Respir Crit C are Med. 2017;195(3):302–13.
- 250. Ledford D, Busse W, Trzaskorna B, Omachi TA, Rosén K, Chipps BE, et al. A randomized multicenter study evaluating Xolair persistence of response after long-term therapy. J Allergy Clin Immu 10., 2017;140(1):162-169.e2.
- 251. Ferguson GT, FitzGerald JM, Bleecker ER, Laviolette M, Bernstein D, LaForce C, et al. Benralizumab for patients with mild to moderate, persistent asthma (BISE): a randomised, double-blind placebo-controlled, phase 3 trial. Lancet Respir Med. 2017;5(7):568–76.
- 252. Shimoda T, Odajima H, Okamasa A, Kawase M, Komatsubara M, Mayer B, et al. Efficacy and safety of mepolizumab in Japanese patients with severe eosinophilic asthma. Allergol Int Off J Jpn Soc Allergol. 2017;66(3):445–51.
- 253. Koshak A, Wei L, Koshak E, Wali S, Alamoudi O, Demerdash A, et al. Nigella sativa Supplementation Improves Asthma Control and Biomarkers: A Randomized, Double-Blind, Placebo-Controlled Trial. Phytother Res PTR. 2017;31(3):403–9.
- 254. Oishi K, Hirano T, Suetake R, Ohata S, Yamaji Y, Ito K, et al. A trial of oral corticosteroids for persistent systemic and airway inflammation in severe asthma. Immun Inflamm Dis. 2017;5(3):261–4.
- 255. Bjerregaard A, Laing IA, Backer V, Sverrild A, Khoo S-K, Chidlow G, et al. High fractional exhaled nitric oxide and sputum eosinophils are associated with an increased risk of future virus-induced exacerbations: A prospective cohort study. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 2017;47(8):1007–13.

- 256. Ortega HG, Yancey SW, Mayer B, Gunsoy NB, Keene ON, Bleecker ER, et al. Severe eosinophilic asthma treated with mepolizumab stratified by baseline eosinophil thresholds: a secondary analysis of the DREAM and MENSA studies. Lancet Respir Med. 2016;4(7):549–56.
- 257. Wenzel S, Castro M, Corren J, Maspero J, Wang L, Zhang B, et al. Dupilumab efficacy and safety in adults with uncontrolled persistent asthma despite use of medium-to-high-dose inhaled corticosteroids plus a long-acting β2 agonist: a randomised double-blind placebo-controlled pivotal phase 2b dose-ranging trial. Lancet Lond Engl. 2016;388(10039):31–44.
- 258. Fitzpatrick AM, Jackson DJ, Mauger DT, Boehmer SJ, Phipatanakul W, Sheehan WJ, et al. Individualized Therapy for Persistent Asthma in Young Children. J Allergy Clin Immunol. 2016;138(6):1608-1618.e12.
- 259. Hanania NA, Korenblat P, Chapman KR, Bateman ED Kopecky P, Paggiaro P, et al. Efficacy and safety of lebrikizumab in patients with <u>meconscolled</u> asthma (LAVOLTA I and LAVOLTA II): replicate, phase 3, randomisec do ible-blind, placebo-controlled trials. Lancet Respir Med. 2016;4(10):781–96.
- 260. Buhl R, Korn S, Menzies-Gow A, Aubier M, Cherman KR, Canonica GW, et al. Assessing biomarkers in a real-world severe asthma study (ARIETTA). Respir Med. 2016;115:7–12.
- 261. Tuomisto LE, Ilmarinen P, Niemel'a O Haanpää J, Kankaanranta T, Kankaanranta H. A 12-year prognosis of adult-onset C thma: Seinäjoki Adult Asthma Study. Respir Med. 2016;117:223–9.
- 262. Westerhof GA, de Groot JC, / melink M, de Nijs SB, Ten Brinke A, Weersink EJ, et al. Predictors of frequent exactivations in (ex)smoking and never smoking adults with severe asthma. Respir Mou. 2016;118:122–7.
- 263. Park H-S, Kim M-K. Im.[:] N, Nakanishi T, Adachi M, Ohta K, et al. A Phase 2a Study of Benralizumab for Pallents with Eosinophilic Asthma in South Korea and Japan. Int Arch Allergy Jmn uno . 2016;169(3):135–45.
- 264. Busse WW, Holgate ST, Wenzel SW, Klekotka P, Chon Y, Feng J, et al. Biomarker Profiles in Asthma With High vs Low Airway Reversibility and Poor Disease Control. Chest. 2015;148(6):1489–96.
- 265. Tagaya E, Kondo M, Kirishi S, Kawagoe M, Kubota N, Tamaoki J. Effects of regular treatment with combination of salmeterol/fluticasone propionate and salmeterol alone in cough variant asthma. J Asthma Off J Assoc Care Asthma. 2015;52(5):512–8.
- 266. Ortega HG, Liu MC, Pavord ID, Brusselle GG, FitzGerald JM, Chetta A, et al. Mepolizumab Treatment in Patients with Severe Eosinophilic Asthma. N Engl J Med. 2014;371(13):1198–207.
- 267. Liu W, Chu J, Sun L, Shen Z, Liu Y, Peng Q, et al. Effect of age and eosinophil number on fractional exhaled nitric oxide level in non-asthmatic children in shanghai. Iran J Allergy Asthma Immunol. 2014;13(5):343–7.

- 268. Castro M, Wenzel SE, Bleecker ER, Pizzichini E, Kuna P, Busse WW, et al. Benralizumab, an anti-interleukin 5 receptor α monoclonal antibody, versus placebo for uncontrolled eosinophilic asthma: a phase 2b randomised dose-ranging study. Lancet Respir Med. 2014;2(11):879–90.
- 269. Pettipher R, Hunter MG, Perkins CM, Collins LP, Lewis T, Baillet M, et al. Heightened response of eosinophilic asthmatic patients to the CRTH2 antagonist OC000459. Allergy. 2014;69(9):1223–32.
- 270. Ortega H, Li H, Suruki R, Albers F, Gordon D, Yancey S. Cluster analysis and characterization of response to mepolizumab. A step closer to personalized medicine for patients with severe asthma. Ann Am Thorac Soc. 2014;11(7):1011–7.
- 271. Steinke JW, Negri J, Payne SC, Borish L. Biological Effects of Leukotriene E4 on Eosinophils. Prostaglandins Leukot Essent Fatty Acids. 20:4;91(3):105–10.
- 272. Kupczyk M, Dahlén B, Sterk PJ, Nizankowska-Mogiluc E, Papi A, Bel EH, et al. Stability of phenotypes defined by physiological valiables and biomarkers in adults with asthma. Allergy. 2014;69(9):1198–204.
- 273. Nair P, Denis S, Cancelliere L, Radford K, Efthimiadis A, Rosano M, et al. The effects of an epithelial barrier protective cationic aerc sol on allergen-induced airway inflammation in asthma: a randomized, placet p-controlled clinical trial. Clin Exp Allergy J Br Soc Allergy Clin Immuno! 2014;44(9):1200–3.
- 274. Hanania NA, Wenzel S, Rosén K, ¹⁴c.eh H-J, Mosesova S, Choy DF, et al. Exploring the effects of omalizumab in a¹¹ergic asthma: an analysis of biomarkers in the EXTRA study. Am J Respir Crit Care Med. 2013;187(8):804–11.
- 275. Brusselle GG, Vanderstichele C, Jordens P, Deman R, Slabbynck H, Ringoet V, et al. Azithromycin for prevention of exacerbations in severe asthma (AZISAST): a multicentre randomised couble-blind placebo-controlled trial. Thorax. 2013;68(4):322–9.
- 276. Samary C dos S, Antu les MA, Silva JD, Silva da AL, Araújo de CC, Bakker-Abreu I, et al. Impact of Econus Calmette-Guérin Moreau vaccine on lung remodeling in experimental asthma. Respir Physiol Neurobiol. 2013;189(3):614–23.
- 277. Kupczyk M, Haque S, Middelveld RJM, Dahlén B, Dahlén S-E, Investigators B. Phenotypic predictors of response to oral glucocorticosteroids in severe asthma. Respir Med. 2013;107(10):1521–30.
- 278. McGrath KW, Icitovic N, Boushey HA, Lazarus SC, Sutherland ER, Chinchilli VM, et al. A large subgroup of mild-to-moderate asthma is persistently noneosinophilic. Am J Respir Crit Care Med. 2012;185(6):612–9.
- 279. Ricciardolo FLM, Di Stefano A, Silvestri M, Van Schadewijk AM, Malerba M, Hiemstra PS, et al. Exhaled nitric oxide is related to bronchial eosinophilia and airway hyperresponsiveness to bradykinin in allergen-induced asthma exacerbation. Int J Immunopathol Pharmacol. 2012;25(1):175–82.

- 280. Mendes FAR, Almeida FM, Cukier A, Stelmach R, Jacob-Filho W, Martins MA, et al. Effects of aerobic training on airway inflammation in asthmatic patients. Med Sci Sports Exerc. 2011;43(2):197–203.
- 281. Vaickus LJ, Bouchard J, Kim J, Natarajan S, Remick DG. Oral tolerance inhibits pulmonary eosinophilia in a cockroach allergen induced model of asthma: a randomized laboratory study. Respir Res. 2010;11:160–160.
- 282. Maneechotesuwan K, Ekjiratrakul W, Kasetsinsombat K, Wongkajornsilp A, Barnes PJ. Statins enhance the anti-inflammatory effects of inhaled corticosteroids in asthmatic patients through increased induction of indoleamine 2, 3-dioxygenase. J Allergy Clin Immunol. 2010;126(4):754-762.e1.
- 283. Lee SH, Lee J-H, Yoon HI, Park HY, Kim T-H, Yoo KF, et al. Change in inhaled corticosteroid treatment and COPD exacerbations: an analysis of real-world data from the KOLD/KOCOSS cohorts. Respir Res. 2019;20(1):(2-t.?.
- 284. George L, Wright A, Mistry V, Sutcliffe A, Chachi (, Hıldar K, et al. Sputum Streptococcus pneumoniae is reduced in COPD fc 'lowing treatment with benralizumab. Int J Chron Obstruct Pulmon Dis. 2019;14:1177–25.
- 285. Bafadhel M, Peterson S, De Blas MA, Calverley PiA, Rennard SI, Richter K, et al. Predictors of exacerbation risk and response to budesonide in patients with chronic obstructive pulmonary disease: a post-hoc analysis of three randomised trials. Lancet Respir Med. 2018;6(2):117–26.
- 286. Suissa S, Dell'Aniello S, Ernst P. Comparative effectiveness of LABA-ICS versus LAMA as initial treatment in COPD targeted by blood eosinophils: a population-based cohort study. Lancet Respir Net. 2018;6(11):855–62.
- 287. Shute JK, Calzetta L, Cardici V, di Toro S, Page CP, Cazzola M. Inhaled nebulised unfractionated heparin in proves lung function in moderate to very severe COPD: A pilot study. Pulm Pharma of Ther. 2018;48:88–96.
- 288. Rabe KF, Watz H Bai ildo S, Pedersen F, Biondini D, Bagul N, et al. Antiinflammatory etc. cts of roflumilast in chronic obstructive pulmonary disease (ROBERT): a 16-v eek, randomised, placebo-controlled trial. Lancet Respir Med. 2018;6(11):827–36.
- 289. Roche N, Chapman KR, Vogelmeier CF, Herth FJF, Thach C, Fogel R, et al. Blood Eosinophils and Response to Maintenance Chronic Obstructive Pulmonary Disease Treatment. Data from the FLAME Trial. Am J Respir Crit Care Med. 2017;195(9):1189–97.
- 290. Contoli M, Pauletti A, Rossi MR, Spanevello A, Casolari P, Marcellini A, et al. Longterm effects of inhaled corticosteroids on sputum bacterial and viral loads in COPD. Eur Respir J. 2017;50(4).
- 291. Papi A, Dokic D, Tzimas W, Mészáros I, Olech-Cudzik A, Koroknai Z, et al. Fluticasone propionate/formoterol for COPD management: a randomized controlled trial. Int J Chron Obstruct Pulmon Dis. 2017;12:1961–71.

- 292. Dasgupta A, Kjarsgaard M, Capaldi D, Radford K, Aleman F, Boylan C, et al. A pilot randomised clinical trial of mepolizumab in COPD with eosinophilic bronchitis. Eur Respir J [Internet]. 2017 Mar;49(3). Available from: https://erj.ersjournals.com/content/49/3/1602486
- 293. Sivapalan P, Moberg M, Eklöf J, Janner J, Vestbo J, Laub RR, et al. A multi-center randomized, controlled, open-label trial evaluating the effects of eosinophil-guided corticosteroid-sparing therapy in hospitalised patients with COPD exacerbations The CORTICO steroid reduction in COPD (CORTICO-COP) study protocol. BMC Pulm Med. 2017;17(1):114–114.
- 294. Pascoe S, Costa M, Marks-Konczalik J, McKie E, Yang S, Scherbovsky PS. Biological effects of p38 MAPK inhibitor losmapimod does not translate to clinical benefits in COPD. Respir Med. 2017;130:20–6.
- 295. Wedzicha JA, Banerji D, Chapman KR, Vestbo J, Roche N Ayers RT, et al. Indacaterol–Glycopyrronium versus Salmeterol–Fluticascute for COPD. N Engl J Med. 2016;374(23):2222–34.
- 296. Barnes NC, Sharma R, Lettis S, Calverley PM.'s. Plood eosinophils as a marker of response to inhaled corticosteroids in COPD. Eur Lespir J. 2016;47(5):1374–82.
- 297. Hinds DR, DiSantostefano RL, Le HV, P scov S. Identification of responders to inhaled corticosteroids in a chronic obstructive pulmonary disease population using cluster analysis. BMJ Open. 2016: / (6) e010099–e010099.
- 298. Marks-Konczalik J, Costa M, Pobertson J, McKie E, Yang S, Pascoe S. A post-hoc subgroup analysis of data from a six month clinical trial comparing the efficacy and safety of losmapimod in mod rrate severe COPD patients with ≤2% and >2% blood eosinophils. Respir Med. 2015;109(7):860–9.
- 299. Bafadhel M, McKenna S, Terry S, Mistry V, Pancholi M, Venge P, et al. Blood eosinophils to direct cord costeroid treatment of exacerbations of chronic obstructive pulmonary disease: a randomized placebo-controlled trial. Am J Respir Crit Care Med. 2012;186(1):48–55.
- 300. de Nijs SB, Fens N, Lutter R, Dijkers E, Krouwels FH, Smids-Dierdorp BS, et al. Airway inflammation and mannitol challenge test in COPD. Respir Res. 2011;12(1):11–11.
- 301. Hirano I, Collins MH, Assouline-Dayan Y, Evans L, Gupta S, Schoepfer AM, et al. RPC4046, a Monoclonal Antibody Against IL13, Reduces Histologic and Endoscopic Activity in Patients With Eosinophilic Esophagitis. Gastroenterology. 2019;156(3):592-603.e10.
- 302. de Ruiter K, Tahapary DL, Sartono E, Nutman TB, Smit JWA, Koenderman L, et al. The Effect of Helminths on Granulocyte Activation: A Cluster-Randomized Placebo-Controlled Trial in Indonesia. J Infect Dis. 2019;219(9):1474–82.
- 303. Johnson K, Iyer V, Katzka D, Ravi K, Lennon R, Pendegraft R, et al. Poor Relationship Between Fractionated Exhaled Nitric Oxide and Disease Activity in Eosinophilic Esophagitis. Dysphagia. 2019;34(1):138–44.

- 304. Kanagalingam S, Shehab SS, Kaminsky DA, Wise RA, Lang JE, Dixon AE. Effect of obesity on sinonasal disease in asthma. J Asthma Off J Assoc Care Asthma. 2018;55(5):525–31.
- 305. Wechsler ME, Akuthota P, Jayne D, Khoury P, Klion A, Langford CA, et al. Mepolizumab or Placebo for Eosinophilic Granulomatosis with Polyangiitis. N Engl J Med. 2017;376(20):1921–32.
- 306. Bachert C, Sousa AR, Lund VJ, Scadding GK, Gevaert P, Nasser S, et al. Reduced need for surgery in severe nasal polyposis with mepolizumab: Randomized trial. J Allergy Clin Immunol. 2017;140(4):1024-1031.e14.
- 307. Jerschow E, Edin ML, Pelletier T, Abuzeid WM, Akbar NA, Gibber M, et al. Plasma 15-Hydroxyeicosatetraenoic Acid Predicts Treatment Ot tcomes in Aspirin-Exacerbated Respiratory Disease. J Allergy Clin Immunot Pract. 2017;5(4):998-1007.e2.
- 308. Eastman JJ, Cavagnero KJ, Deconde AS, Kim AS. Kart: MR, Broide DH, et al. Group 2 innate lymphoid cells are recruited to the nasal success in patients with aspirin-exacerbated respiratory disease. J Allergy Clin instrunol. 2017;140(1):101-108.e3.
- 309. Liu T, Kanaoka Y, Barrett NA, Feng C, Garofalo L, Lai J, et al. ,Aspirin-exacerbated respiratory disease involves a cysteinyl le 1k of iene-driven IL-33-mediated mast cell activation pathway. J Immunol Baltim Md 1750. 2015;195(8):3537–45.
- 310. Oyama Y, Fujisawa T, Hashimoto N Enomoto N, Nakamura Y, Inui N, et al. Efficacy of short-term prednisolone treatment n. patients with chronic eosinophilic pneumonia. Eur Respir J. 2015;45(6):1624–31.
- 311. Steinke JW, Negri J, Liu L Payne SC, Borish L. Aspirin Activation of Eosinophils and Mast Cells: Implications in the Pathogenesis of Aspirin-Exacerbated Respiratory Disease. J Immunol Bah. n Md 1950. 2014;193(1):41-7.
- 312. Choi G-S, Kim J-H Shin Y-S, Ye Y-M, Kim S-H, Park H-S. Eosinophil activation and novel mediators in the aspirin-induced nasal response in AERD. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 2013;43(7):730–40.
- 313. Spergel JM, Rothenberg ME, Collins MH, Furuta GT, Markowitz JE, Fuchs G, et al. Reslizumab in children and adolescents with eosinophilic esophagitis: results of a double-blind, randomized, placebo-controlled trial. J Allergy Clin Immunol. 2012;129(2):456–63, 463.e1-3.
- Cai C, He M, Zhong S, Tang Y, Sun B, Chen Q, et al. Add-on montelukast vs doubledose budesonide in nonasthmatic eosinophilic bronchitis: a pilot study. Respir Med. 2012;106(10):1369–75.
- 315. Uller L, Ahlström Emanuelsson C, Andersson M, Erjefält JS, Greiff L, Persson CG. Early phase resolution of mucosal eosinophilic inflammation in allergic rhinitis. Respir Res. 2010;11(1):54–54.

- 316. Kim C-K, Choi J, Kim HB, Callaway Z, Shin BM, Kim J-T, et al. A randomized intervention of montelukast for post-bronchiolitis: effect on eosinophil degranulation. J Pediatr. 2010;156(5):749–54.
- 317. Sahota J, Robinson DS. Update on new biologics for intractable eosinophilic asthma: impact of reslizumab. Drug Des Devel Ther. 2018 May 8;12:1173–81.
- Pavord ID, Chanez P, Criner GJ, Kerstjens HAM, Korn S, Lugogo N, et al. Mepolizumab for Eosinophilic Chronic Obstructive Pulmonary Disease. N Engl J Med. 2017 26;377(17):1613–29.

Figure 1: Schematic representation of surface receptors and immunological moieties of eosinophils. Eosinophils are bi-lobed, multi-functional innate immune cells with diverse cell surface receptors, including those crucial for chemotaxis, cell adhesion, activation (via cytokine/growth factors), lipid mediation, and immune modulation (CD40/80/86/siglec-8/Fc/MHC class-II). Eosinophils also comprise of a variety of intracellular functional moieties, such as lipid bodies and granules that play a key role in immune regulation and eosinophil functionality.

Abbreviations: PR – Pattern recognition receptors; MBP – Major basic protein; EPX – Eosinophil peroxidase; MHC – Major histocompatibility complex; CD – Cluster of differentiation; CCR - CC-chemokine receptor; CXCR - CXC-chemokine receptor; FPR -Formyl peptide receptor; C5aR - Complement component 5a receptor; C3aR - Complement component 3a receptor; LFA-1 - Lymphocyte function-abociated antigen 1; CR – Complement receptor; LTC4 – Leukotriene C4; LTE4 - Leukotriene E4; Leukotriene D4; PAF - Platelet-activating factor; PAR – Protease activated activated eceptor; PAFR - Plateletactivating factor receptor; CRTH2 - Chemoattractat t receptor-homologous molecule expressed on T-helper type 2 cells; DP1 - Prostaglandin D2 receptor 1; EP4 - Prostaglandin E2 receptor 4; LTB₄ - Leukotriene B4

Figure 2: Pathophysiology of eosinophi's in a thma Abbreviations: IL (Interleukin); TSLP (thymic stromal lymphopoietin); PC (Antigen-presenting cell); Th0 (naïve T-cell); Th2 (T-helper type 2 cells); MBP (Major basic protein); EPO (Eosinophil peroxidase); ECP (Eosinophil cationic protein); EDN (Eosinophil-derived neurotoxin); NGF (Nerve growth factor); SCF (Stem cell factor); ASM (Airway smooth muscle); AHR (Airway hyperresponsiveness); TGF- β (Transforming growth factor beta); ECM (Extracellular matrix)

Figure 3: Pathophysiolog, of cosinophil on COPD Abbreviations: CPE - Cytopathogenic effect; CD4⁺TH2 - CD4⁺ 1 helper 2 cells; IL - Interleukin; PAF - Platelet activating factor; MBP - Major basic protein: \pm PO - Eosinophil peroxidase; ECP - Eosinophil cationic protein

Figure 4: Current drugs acting on eosinophils

Abbreviations: IL (Interleukin); TSLP (thymic stromal lymphopoietin); Th0 (naïve T-cell); Th2 (T-helper type 2 cells); ILC2 (Type 2 innate lymphoid cells); IL-5R α (Interleukin-5 receptor alpha); CRTH2 (chemoattractant receptor-homologous molecule expressed on Th2 cells); PGD2 (Prostaglandin D2); GM-CSF (Granulocyte-macrophage colony-stimulating factor); IL-4R α (Interleukin-4 receptor alpha); IgE (Immunoglobulin E); Fc ϵ RI (High-affinity immunoglobin E receptor); DC (Dendritic cell)

S.	Intervention	No. of	Area of trial	Sponsor	Ref
Ν		recruitments/Country/Pha			
0		se of study/Ongoing/concolled			
1.	Saffron	study/Ongoing/cancelled 86/Phase 1/Iran/Ongoing	Effect on	Ahvaz-	(238
1.	Samon	60/1 hase 1/ frail/ Ongoing	allergic asthma	Jundishapur	(250
			patient's	university of	/
			clinical	medical	
			symptoms,	sciences	
			blood pressure,		
			lipid panels,		
			basophils and		
			eosinophile		
			receiving		
			saffror		
			supplem.~atio		
2		22/1	$\frac{n}{r}$		(220
2.	Mepolizuma	32/Japan	E valuation of	Sutoh Hospital	(239
	b		r lepc ¹ izumab's)
			safety in severe		
			eosinophilic		
			asthma patients		
			receiving		
			treatment with		
			it long-term.		
3.	Vitamin D	86/Mexico	Evaluation of	General	(240
			vitamin D's	Hospital of)
			effect as a	Mexico	
			supplement on		
			the pathogenic		
			bacteria colonization in		
			the upper		
			respiratory tract		
			of patients with		
			allergic asthma.		
4	Benralizuma	2681/Phase III/Completed	Effects of	AstraZeneca	(241
	b	-	baseline factors)
			of patients with		
			severe asthma		
			on the efficacy		
			of		
~			benralizumab.		(0.10
5	Benralizuma	2508/Phase III/Completed	Efficacy of	AstraZeneca	(242
	b		benralizumab)
			based on atopic status and		
			status and serum		
			sciulli		

Table 1 : List of clinical trials on eosinophils for asthma
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			concentrations of IgE.		
6	Fevipiprant	61/Phase II/United Kingdom/ Completed	Potential reduction of eosinophilic airway inflammation in moderate-to- severe eosinophilic asthma receiving fevipiprant (QAW039).	Novartis Pharmaceuticals	(111)
7	Mepolizuma b	651/Phase III/Completed	Assessmeric of the effectiveness and long term safety a soc ated with the upatment of severe cosinophilic asthma patients with subcutaneous mepolizumab.	GlaxoSmithKli ne	(243)
8	Induced sputum	300/Multiple countries/Phase III/Complex 1	Comparison of different gene and protein expression in sputum samples of severe asthma and non-smoking mild/moderate asthma patients	Academisch Medisch Centrum - Universiteit van Amsterdam (AMC-UvA)	(244)
9	Omalizumab	1071/USA/Phase III/Completed	Evaluation of factors associated with the prediction of response to omalizumab for the purpose of identifying patients with highest potential to attain greatest clinical	-	(245)

			advantage from		
			treatment.		
12	Mepolizuma	621/Multiple	Association of	GlaxoSmithKli	(246
	b	countries/Phase	asthma	ne)
		II/Completed	exacerbations		
			with enhanced		
			decline in lung		
			function.		
13	Tiotropium	459/Multiple	Influence of T2	Boehringer	(247
	Respimat	countries/Phase	status on	Ingelheim)
		III/Completed	responses to		
			tiotropium		
			Respimat add-		
			on therapy.		
14	Mepolizuma	556/Multiple	Assessmera	GlaxoSmithKli	(248
	b	countries/Phase	mepolizu. ab's	ne)
		III/Completed	effect on sovere		
			eosi oplilic		
			asthma		
			potier t's		
			Feahl -related		
		0	cuality of life.		
15	-	120/USA/Phase I/Ongoi ¹ g	Description of	University of	(249
		(Suspensed)	exacerbation-	Wisconsin,)
			prone asthma's	Madison	
			clinical		
			features.		
16	Omalizumab	-	Benefit and	-	(250
			persistence of)
			response in		
			subjects		
			continuing or		
			withdrawing		
			from long-term		
			omalizumab		
		9	treatment.		
17	Benralizuma	211/Multiple	Assessment of	AstraZeneca	(251
	b	countries/Phase	benralizumab)
		III/Completed	as a treatment		
			for mild-to-		
			moderate		
			persistent		
			asthma patients		
			in terms of its		
			safety and		
			efficacy.		
18	Mepolizuma	580/Japan/Phase	Description of	GlaxoSmithKli	(252
	b	III/Completed	mepolizumab's	ne)
			safety and		
			efficacy in the		

			treatment of		
			Japanese		
			patients with		
			severe		
			eosinophilic		
			asthma from		
			the MENSA		
			trial.		
19	Nigella	80/Saudi Arabia/Phase	Evaluation of	University	(253
	sativa	II/Completed	nigella sativa	College,)
		-	oil benefits as a	London	
			supplement in		
			the treatment of		
			asthma bas		
			on clinica ¹ and		
			inflamma.ory		
			param ter.		
20	Oral	20/Japan/Phase	Iden ification	Japanese	(254
	corticosteroid	1/Completed	of the	Society for the)
	S		1, latic nship	Promotion of	/
	5		hetween	Science and	
		0	<i>s</i> thmatic	Wakayama	
			patients FeNO	Medical Award	
			levels and	for Young	
			blood	Researchers	
			eosinophils.		
21	-	112/Phase I	Assessment of	Bispebjerg	(255
			the relationship	University)
			between type 2	Hospital, the	·
			inflammation	University of	
			and risk of	Copenhagen	
			virus-induced	1 0	
			asthma		
			exacerbations		
22	Mepolizuma	580. Multiple	• Assessment	GlaxoSmithKli	(256
	b	comatries/Phase	of the	ne)
		III/Completed	relationship		Í
		1	of baseline		
			blood		
			eosinophil		
			counts and		
			mepolizum		
			ab's		
			efficacy.		
23	Dupilumab	776/Multile countries/Phase	Assessment	Sanofi-	(257
_	L	II/Completed	of	Genzyme and)
			dupilumab'	Regeneron	Í
			s efficacy	Pharmaceuticals	
			and safety		
			aspects as		
			aspects us		1

			an add-on		
			therapy in		
			uncontrolle		
			d and		
			persistent		
			asthmatics		
			on medium-		
			to-high		
			doses of		
			ICS/LABA		
			therapy,		
			disregardin		
			g the		
			baselin		
			eosinorhil		
			count		
24	Personalised	300/USA/Phase	Persor allz tion	Milton S.	(258
24	therapy	III/Completed	of as hu?	Hershey	(230
	ulerapy	m/completed	therap,	Medical Center)
25	Lebrikizuma	1068/Multiple	A sessment of	Hoffmann-La	(250
23		1068/Multiple			(259
	b	countries/Phase	1.brh. zumab's	Roche)
		II/Completed	efficacy and		
			safety aspects		
			when used for		
			the treatment of		
			uncontrolled		
			asthmatics		
			despite ICS and		
			at least a		
			second		
			controller		
			medication.		
26	-	483/Muliple	Addressment of	Hoffmann-La	(260
		ountrie 3/Phase	unanswered	Roche)
		IV/ Completed	fundamental		
			queries		
			associated with		
			biomarkers of		
			asthma.		
			Assessment of		
			the relationship		
			between		
			biomarkers of		
			asthma and		
			health		
			outcomes		
			associated with		
			disease.		
27	-	259/Finland/Phase	Evaluation of	Seinajoki	(261
- 1		257/1 manu/1 mase		Somujoki	(201

Journal Pre-proof

		I/Commissional	4h a 10 yraan	Control Hourital	
		I/Completed	the 12-year	Central Hospital)
			prognosis in		
			patients who		
			develop asthma		
			as adults and		
			the aspects		
			related to its		
			prognosis.		
28	-	571/Netherland	Investigation of	GlaxoSmithKli	(262
			factors related	ne)
			to frequent		
			exacerbations		
			in non-smoker		
			asthma path ts		
			and asthm ² .		
			patients with ?		
			history of		
			smolin		
29	Benralizuma	106/South Korea &	Evoluation of	MedImmune	(263
29	b	Japan/Phase I/Completed	l nra izumab's	LLC	(203
	U	Japan/Fliase I/Completed)
			elfect as a		
			treatment of		
			uncontrolled		
			eosinophilic		
			asthma who		
			experienced 2-6		
			exacerbations		
			in the previous		
			year and treated		
			with		
			medium/high		
			dosages of ICS		
			and LABA.		
30	-	315 Mu tiple	Comparison	Amgen	(264
		courtries/Phase	between asthma)
		II/C_mpleted	associated with		
			high and low		
			airway		
			reversibility in		
			terms of		
			function of		
			lungs,		
			biomarker		
			panel, and		
			control of		
			disease.		
21	SD010				(101
31	SB010	-	Evaluation of	-	(191
			SB010 in terms)
			of its safety and		
			efficacy in the		

		1			i
			treatment of		
			allergic asthma		
			patients with		
			sputum		
			eosinophilia.		
32	Salmeterol/	-	Evaluation of	-	(265
	fluticasone		efficacy of)
	propionate		combination		<i>,</i>
	Salmeterol		therapy in the		
	200000000		treatment of		
			cough variant		
			asthma.		
33	Mepolizuma	580/Multiple	Determination	GlaxoSmithKli	(266
55	b	countries/Phase	of	ne	(200
	U			ne)
		III/Completed	mepolizur'		
			treatm ent f		
			seve e		
			eosino _F hilic		
			a. thm a to		
			r ster fially		
			reduce frequent		
			corticosteroid		
			usage in these		
			patients.		
34	-	132/China	Identification	-	(267
			of the)
			association		
			between levels		
			of FeNO and		
			possible factors		
			in children		
			without asthma.		
35	Benralizuma	964 Mu tiple	Assessment of	MedImmune	(268
	b	countries/Phase	benralizumab's	LLC)
		II/Completed	in terms of its		Í
		1	effectiveness		
			and safety in		
			and safety in the treatment of		
			the treatment of		
			the treatment of adult patients		
			the treatment of adult patients with		
			the treatment of adult patients with uncontrolled		
			the treatment of adult patients with uncontrolled eosinophilic		
26	0000450	40/United Vinedom/Dhoos	the treatment of adult patients with uncontrolled eosinophilic asthma.	Chiosi	(260
36	OC000459	40/United Kingdom/Phase	the treatment of adult patients with uncontrolled eosinophilic asthma. Determination	Chiesi	(269
36	OC000459	40/United Kingdom/Phase II/Completed	the treatment of adult patients with uncontrolled eosinophilic asthma. Determination of effect	Farmaceutici	(269)
36	OC000459	-	the treatment of adult patients with uncontrolled eosinophilic asthma. Determination of effect associated with		(269)
36	OC000459	-	the treatment of adult patients with uncontrolled eosinophilic asthma. Determination of effect associated with lower doses of	Farmaceutici	(269)
36	OC000459	-	the treatment of adult patients with uncontrolled eosinophilic asthma. Determination of effect associated with	Farmaceutici	(269)

					ı
			Defined		
			phenotype of		
			patients with		
			highest		
			response to		
			OC00459		
			treatment.		
37	Mepolizuma	621/Multiple	Identification	GlaxoSmithKli	(270
	b	countries/Phase	of subgroups of	ne)
		II/Completed	severe asthma		
		-	patients at risk		
			for		
			exacerbations		
			with distinc ⁴		
			characteristics.		
			Determin each		
			patien'		
			subg our's		
			response to		
			t. atn ent.		
38		-	J rves 'igation of	Merck	(271
50			$P2Y_{12}$ and	Pharmaceuticals)
			gpr99	1 Indifinaced field is	'
			expression by		
			eosinophils and		
			their LTE ₄		
			response		
			-		
39	_	160/Europa	capacity. Assessment of	_	(272
39	-	169/Europe		-	(272)
			phenotype stability based)
			on biomarkers		
			0r physical scient		
			physiological variables.		
40	Inhaled		Evaluation of		(272
40	cationic	-	the role of	-	(273
)
	airway lining modulator		enhancing		
	mouulator		epithelial barrier in		
			reducing		
			inflammation		
			of the airway due to inhaled		
			particles.		
41	Omalizumab	850/Multipla	Assessment of	Conontach Inc	(274
41	Omanzumab	850/Multiple countries/Phase		Genentech, Inc	(274
			FeNO,)
		III/Completed	peripheral		
			blood		

		I	r	I	
			eosinophils		
			count, and		
			serum periostin		
			as potential		
			biomarkers of		
			Th2		
			inflammation		
			and potential		
			predictors of		
			omalizumab		
			treatment		
			outcomes.		
42	Azithromyci	109/Belgium/Phase	Benefit of	University	(275
	n	4/Completed	macrolides 1	Hospital, Ghent)
	11	4/Completed	neutrophil ³ .	mospital, Olient	,
			-		
43	Bacillus		airway di ease.		()76
43		-	Analy' is c ^c	-	(276
	Calmette-)
	Guérin		diffare. t BCG-		
	Moreau		L'ore lu strain		
	vaccine		? Jm. istration		
	(BCG-		routes and		
	Moreau)		application		
			periods on		
			inflammation		
			of the airways		
			and lungs as		
			well as		
			remodelling in		
			an allergic		
			asthma murine		
			model.		
44	Oral	233/Swein/NA/Active	Assessment of	Karolinska	(277
	prednisolone		the practicality	Institutet)
	P		to predict oral		/
			prednisolone		
			response based		
			on patient's		
			medical		
			history,		
			physiological		
			variables and		
1.5			biomarkers.		(070
45	-	0/USA/Phase 3/Withdarwn	Determination	Milton S.	(278
			of non-	Hershey)
			eosinophilic	Medical Center	
			asthma		
			phenotype's		
			prevalence and		
1			clinical	1	

			features.		
46	Allergen	_	Identification	-	(279
10	rineigen		that loss of)
			asthma control		,
			is associated		
			with increased		
			eosinophilic inflammation		
			of the airway		
			and airway		
			responsiveness		
			to bradykinin		
			due to exposure		
			to allergens		
			causing exhaust		
			NO levels to he		
			elevate d		
47	Aerobic	58/Brazil/Phase 3/Ongoing	Eval ration of		(280
	training		aerobic training	Instituto de)
			e. fect; on	Investigação em	
			ebsnophil	Imunologia	
		G	inflammation		
			and nitric oxide		
			of moderate or		
			severe		
			persistent		
			asthma		
			patients.		
48	Allergen	-	Examination of	-	(281
			potential to)
			ameliorate		
			asthma-like		
			pulmonary		
			inflammation		
			via induction of		
		2	oral tolerance		
			to cockroach		
			allergen.		
			Determination		
			of mechanisms		
			associated with		
			the		
			effectiveness of		
			oral tolerance.		
49	Simvastatin	-	Enhancement	-	(282
			of anti-)
			inflammatory		Í
			effects of		
			corticosteroids		
			by statins.		
		1	J	1	I

Table 2	2: Clinical trials o	on eosinophils act. g in COPD	5
S. No	Intervention	No. of	Area of trial
		recruitmu:s/Country/Phase of study/Ongoing/cancelled	
1	Inhaled corticosteroids (ICS)	1144, Korea	Evaluation of ICS prescription status accord the revision of the 2017 GOLD guidelines
2	Benralizumab	29/Finland/Phase II	Effect on bacterial load in the airways assoc with the decline in eosinophilic airway inflammation when treated with benralizum
3	Budesonide	1200/USA/Phase III/Completed	Establishment of characteristics that determinisk of exacerbation and clinical response to treatment with ICS in COPD patients using modelled continuous variable, eosinophil continuous varia
4	LABA-ICS/ LAMA	645/Germany/Completed	Comparison of efficacy and safety of treatment initiation guided by blood eosinophils with a LABA-ICS or LAMA in COPD patients.
5	Unfractionated heparin	-	Demonstration of improve lung function in pulmonary rehabilitation COPD patients rec unfractionated heparin. Demonstration of the novel, safe and effecti aspects of unfractionated heparin when used

			treatment of COPD.
6	Roflumilast	158/Multiple countries/Phase III/Completed	Assessment of roflumilast's anti-inflammate effects on bronchial mucosal inflammation moderate-to-severe COPD patients and chro bronchitis patients
7	ICS-LABA LABA-LAMA	3362/Multiple countries/Phase III/completed	Evaluation of blood eosinophil's value as a predictor of responsiveness in the treatment COPD exacerbations using ICS/LABA vers LABA/LAMA therapy.
8	Salmeterol/Flu ticasone propionate Salmeterol	60/Italy/NA/Completed	Determination of ICS effects on microbial l airways of COPD patients. Evaluation of the underlying inflammatory mechanisms associated with the colonisatio microbiome of airways.
9	Fluticasone propionate/ formoterol	0/NA/Phase III/Withdrawn	Enfect of fluticasone propionate/formoterol (.FP/FORM) in COPD
10	Mepolizumab	19/Canada/Phase III/Completing	Determination of mepolizumab's potential t decrease percentage of sputum eosinophil in cigarette smoke associated COPD patients v persistent sputum eosinophilia. Assessment of mepolizumab's effects on cli features of cigarette smoke associated COPD patients with persistent sputum eosinophilia
11	Eosinophil- guided corticosteroid- sparing therapy	318/Denmark/Phase IV/Completed	Determination of potential reduction of syst corticosteroid use in the treatment of AECC without affecting the outcome.
12	Losmapimod	72/United Kingdom/Phase	Evaluation of exacerbation reduction by losmapimod in moderate-to-severe COPD p
13	Indacaterol- Glycopyrroniu m Salmeterol- Fluticasone	3362/Multiple countries/Phase III/Completed	Identification of the role of LABA-LAMA in COPD patients with at least one exacerbathe past year.
14	ICS withdrawal	444/Denmark/Phase IV/Ongoing	Identification of relationship between baseli blood eosinophil count and rate of lung fund decline. Evaluation of risk/benefit ratio in COPD pa receiving ICS therapy through the prediction patient response to ICS using a biomarker
15	Fluticasone furoate/vilanter ol	1635/Multiple countries/Phase III/Completed	Identification of COPD patient clusters with potential to attain benefit from ICS/LABA treatment compared to LABA singularly.

	Vilanterol		Validation of COPD clusters identified in th
16	Losmapimod	604/Multiple countries/Phase	Analysis of losmapimod's effect as a treatm
		II/Completed	reduce moderate/severe exacerbation rates i
			subgroups of COPD patients with a baseline
			and $>2\%$ blood eosinophils.
17	Prednisolone	-	Investigation of the functionality of blood
			eosinophils in the direction of corticosteroid
			treatment during COPD exacerbations.
18	Hypertonic	-	Examination of the association between airv
	saline		hyperresponsiveness to mannitol and the
	Mannitol		inflammatory markers in the collected sputu
	challenge		induced by hypertonic saline, blood and exh

Table 3: Clinical trials on eosinophils acting in other respiratory diseases

S.	Intervention	Condition	No. of	Area of trial
No			recruiti ie at 5	
			/Cour try,Tn	
			6 JE 6.	
			s.יי גאי/Ongoi	
1		F 1 11	ng/cancelled	
1	RPC4046	Eosinophilic	1.00/Multiple	Evaluation of RPC4046 in eosinophilic esophagitis p
		esophagitis	Juntries/Pha	terms of efficacy and safety
			se U/Completed	
2	Albendazole	Helminth	II/Completed	Effect of infection by helminths on the status of activ
Ζ	Albenuazoie	infection	-	granulocytes.
3	Fractionated	Ecsicopullic	120/United	Utility of FeNO in predicting severity of eosinophilic
5	exhaled	es on gitis	States/NA/O	activity.
	nitric oxide		ngoing	activity.
	(FeNO))		
	testing			
	C			
4	Obesity	Sinonasal	-	Determination of the association between obesity and
		disease in		severity of sinonasal disease, and/or effects on treatm
		asthma		nasal corticosteroid response in patients with asthma
5	Mepolizuma	Eosinophilic	136/Multiple	Comparison between mepolizumab and a placebo as
	b	granulomatosi	countries/Pha	regimen in relapse or refractory eosinophilic granulo
		s with	se	patients with polyangiitis in terms of its efficacy and
		Polyangiitis	III/Complete	a duration of 52 weeks.
(M	N 1 1	d 100/United	
6	Mepolizuma	Nasal polyps	160/United	Assessment between mepolizumab and a placebo in the second office on for treatment of accurate hildered and
	b		states &	safety and efficacy for treatment of severe bilateral n
			Europe/Phase	polyposis.

			III/Ongoing	
7	Aspirin	Aspirin- exacerbated respiratory disease	-	Investigation of the relationship between aspirin ther associated clinical outcomes and levels of plasma eic aspirin-exacerbated respiratory disease.
8	Aspirin	Aspirin- exacerbated respiratory disease	-	Determination of change ILC2 levles in peripheral binasal mucosa in aspirin-exacerbated respiratory disea COX-1 inhibitor-induced reactions.
9	-	Aspirin- Exacerbated Respiratory Disease	-	Identification of the mechanistic basis for mast cell a
10	Prednisolone	Chronis eosinophilic pneumonia	50/Japan/Pha se IV/completed	Compa, son between short-term and long-term cortic treatment in chronic eosinophilic pneumonia patients its santy and effectiveness.
11	Aspirin	Aspirin- exacerbated respiratory disease	-	In /est. 3 ation of aspirin's capacity to trigger activation of aspirin's capacity to trigger activation of aspirines and mast cells in aspirine xacerbated response.
12	Aspirin	Aspirin- exacerbated respiratory disease	- O	h.vestigation of mechanism associated with the active eosinophils.Identification of novel inflammatory mediators through utilization of proteomics.
13	Reslizumab	Eosinophilic esophagitis	227 United States & Conada/Phas	Evaluate the effectiveness of reslizumab in children a adolescent eosinophilic esophagitis subjects.
14	Budesonide/ Montelukast Double-dose budesonide	Non- asthmatic eosiroph?lic branahus	63/China/Pha se IV/Unknown	Comparison of effectiveness and tolerance between r as an add-on therapy to budesonide and double dose budesonide for the suppression of airway eosinophili reduction of severity of cough in non-asthmatic eosin bronchitis.
15	Allergen	Al'ergic filinitis	-	Determination of eosinophilia in the mucosa, apopto eosinophils, general cell apoptosis, cell proliferation and CCL11 expression in allergic airway tissues of h resolution of symptomatic eosinophilic inflammation
16	Montelukast	Bronchiolitis	146/Iran/NA/ Completed	Investigation of montelukast's effect on degranulation eosinophils and recurrent episodes of wheezing in particular post-respiratory syncytial virus bronchiolitis.

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S.	Drug	Route of	Indicati	Mechanis	Biologica	Clinical	Referenc
N	2108	administr	on	m of	l effert3	outcomes	e(s)
							- (-)
o. 1.	Reslizum ab	IV	Adjuncti ve treatmen t in severe eosinoph ilic asthma	action Humanized anti-IL-5 monoclonal antibody	 Binds an 1 an 1 an 1 an 1 an 1 an 1 and binds an 1 and activit 	 50% reductio n in asthma exacerb ation rates Improve ment in lung function , asthma control and QOL 	(89– 91,317)
					y of eosino phils		
2.	Mepolizu mab	SC injection	Adjuncti ve treatmen t in refractor y eosinoph ilic asthma and	Humanized anti-IL-5 monoclonal antibody	 Binds and seques trates to IL- 5 which inhibit s recept 	 Reducti on in blood and sputum eosinop hil counts Reducti on in 	(55,92– 94,317,31 8)

 Table 4: Current drugs acting on eosinophils in respiratory diseases

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						depen		effect	
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4			A 11			ion		~	
4.	Omalizu	SC	Adjuncti	Humanied	•	Binds	٠	Signific	(87,102)
	mab	injection	ve	anti Ig.'		to free		ant	
			treatmen	In mostonal		IgE to		reductio	
			t in	ar.cibody		preve		n in	
			moderate			nt		PBE	
			-severe			intera		counts	
			allergic			ction		in	
			asthma			with		patients	
						FceRI		receivin	
						on		g	
						mast		concom	
						cells,		itant	
						dendri		ICS	
						tic	•	Improve	
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						hils		es	
					•	Decre	•	Improve	
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						pro-		asthma	
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						mator		and	
						y		QOL	
						y media		YOL	
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						releas			
						e from			
1						e nom			
						mast			

					•	cells and basop hils Inhibit s allerg en- induce d T- cell differe ntiatio n			
5.	Dupiluma b	SC injection	Adjuncti ve therapy in severe eosinoph ilic asthma	Humanized anti-IL- 4/IL-13 monoclonal antibody	•	Bind's tc TL- 4F α block IL-4 and IL-13 signali ng pathw ay Inhibit s B- cell IgE class switch ing Reduc es airwa y recruit ment of eosino phils	•	Reducti on in asthma exacerb ation rates Improve ment in AQLQ, ACQ and FEV ₁ Transie nt increase in peripher al eosinop hil counts (especia lly in patients with baseline blood eosinop hil counts of \geq 300 eosinop hils/µL)	(87,104,1 10)

6.	MEDI-	IV	Asthma	Humanized	•	Binds	•	Depleti	(107)
	563			anti-IL-		to		on of	
				5Rα		epitop		peripher	
				monoclonal		eon		al blood	
				antibody		IL-		and	
				5		5Rα		bone	
						near		marrow	
						IL-5		eosinop	
						site of		hils and	
						cataly		basophil	
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						ina, e		primate	
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						אר אר		5.	
						Cof			
						Jsino			
						phils			
						and			
						basop			
						hils <i>in</i>			
						vitro			
7.	Ki19003	Oral	Allergic	CITR3	•	Inhibit	•	Inhibiti	(109)
			asthma	a .tagonist		S		on of	
						CCL1		allergen	
						1-		-	
						induce		induced	
						d		airway	
						migrat		eosinop	
						ion of		hilia at	
						CCR3		an oral	
						-		dose of	
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						e pre-		on in	
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1					•	Inhibit		BALF	
1						s	_	Attenua	
1						s CCL1	•		
						1-		tion of	
1						induce		allergen	

					d migrat ion of eosino philic granul ocytes from bone marro W		induced subepith elial and peribron chial fibrosis	
8.	Fevipipra nt, Timapipr ant	Oral	Fevipipr ant: Mild- moderate eosinoph il- dominan t allergic asthma Timapipr ant: Persisten t eosinoph ilic asthm a	Small molecule PGD2 inhibitors	Antag onist of CRTH 2 expres cud on eosino phils which media tes chemo tactic respon se to PGD2	•	Reducti on in sputum eosinop hil counts Improve ment of asthmati c sympto ms	(110,111)
9.	AMG 157	IV	Mild Phyrgic arthma	Humanized anti-TSLP monoclonal antibody	Binds to TSLP, which is an alarmi n, to preve nt recept or intera ction and stimul ation of eosino philic inflam matio n via	•	Reducti on in maximu m percenta ge decreas e of FEV ₁ Reducti on of blood and sputum eosinop hil counts	(113)

						ILC2 and Th2 pathw			
10	TPI ASM8	Inhalation	Mild allergic asthma	Antisense oligonucleo tides	•	ay Block s expres sion of comm on βc of IL- $3/h \cdot 5/GM$ -'SF revept CSF revept CSF revent Slock s CCR3 expres sion	•	Reducti on in allergen - induced airway sputum eosinop hils Reducti on in airway eosinop hil progenit or cells	(118)
11	Prednison e	Oral	Severe asthma	A ti- iv.flammato ry OCS	•	Induc es eosino phil apopt osis via down- regula tion of IL-5 and GM- CSF which promo tes eosino phil surviv al Inhibit s IL-4 and IL-5 transc ription	•	Reducti on of eosinop hilic/typ e 2 inflamm ation Reducti on in sputum eosinop hil counts Reducti on in pro- inflamm atory cytokin e concent rations Improve ment in pulmon ary function	(119,121, 122)

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		geno	
		mic	
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Abbreviations: IV (Intravenous); QOL (Quality of L^{ifa}); SC (Subcutaneous); ACQ (Asthma Control Questionnaire); FccRI (High-affinity immunog obin E receptor); PBE (Peripheral blood eosinophil); ICS (Inhaled corticosteroids); AQLQ (Asthma Quality of Life Questionnaire); FEV₁ (Forced expiratory volume); ADCC (Antibody-dependent cellular cytotoxicity); CCR3 (CC-chemokine receptor $_{2}$); CCL (CC-chemokine ligand); TGF- β 1 (Transforming growth factor beta 1); ALF (Bronchoalveolar lavage fluid); PGD2 (Prostaglandin D2); CRTH2 (Chemoature that receptor-homologous molecule expressed on Th2 cells); TSLP (Thymic stromal lymphop.ietin); FeNO (Fraction of exhaled nitric oxide); ILC2 (Type 2 innate lymphoid cells); ρ^{-1} (Beta chain); OCS (Oral corticosteroid); GM-CSF (Granulocyte-macrophage colony-st multiplication).

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