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Original Article

DESIGN, 2³ FACTORIAL OPTIMIZATION AND *IN VITRO–IN VIVO* PHARMACOKINETIC EVALUATION OF ROSUVASTATIN CALCIUM LOADED POLYMERIC NANOPARTICLES

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ABSTRACT

Objective: The objective is to incorporate low bioavailable Rosuvastatin Calcium (20%) into polymeric nanoparticles (PNs) to improve its biopharmaceutical properties of Rosuvastatin Calcium.

Methods: The PNs were prepared by solvent evaporation method by applying 2³ factorial designs. The formulated PN are investigated for particle size (PS) and shape, zeta potential (ZP), polydispersity index (PI) and entrapment efficiency (EE), *in vivo* pharmacokinetic.

Results: Among 8 formulations, PN7 shows least PS of 159.9 ± 16.1 nm, which enhance the dissolution, surface area and permeability; ZP of 33.5 ± 1.54 mV, which shows good stability; PI of 0.587 ± 0.16 shows monodisperse distribution pattern; high EE of about 94.20 ± 2.46 %; percentage yield of 96.80 ± 2.08 %; maximum *in vitro* drug release of about 96.54 ± 2.02 % at 24 h with controlled and predetermined release pattern. PN7 drug release obeys zero-order release kinetics, non-fickian diffusion mechanism with r^2 value>0.96 and release exponent 'n' value falls between 0.5-0.8 for peppas kinetic model i.e., the mechanism of drug diffusion is based on polymer relaxation. *In vivo* pharmacokinetic studies illustrate enhance in AUC_{0- α} in mg/ml, which proves a significant enhancement of bioavailability of Rosuvastatin Calcium by PNs.

Conclusion: This PN shows a significant enhancement of bioavailability by minimizing the dose-dependent adverse side effects of rosuvastatin calcium.

Keywords: Rosuvastatin calcium, Polymeric nanoparticle, Bioavailability, Pharmacokinetic

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INTRODUCTION

The term Polymeric Nanoparticles (PNs) is defined as particulate dispersion in which the solid particle size range from 60-200 nm, where the drug particle can be encapsulated, dissolved, entrapped or can be attached to a nanoparticle matrix [1]. The main perspective of PNs is to reduce the drug volatility; also with the help of a multitude method it can be easily formulated in greater quantity. They also furnish a remarkable improvement over traditional method in terms of effectiveness and efficiency for an administration of a medicament. PNs show high specificity for the delivery of any pharmaceutical agent to their desired location. Due to certain ideal characteristic features of PNs, such as opting of polymer and its potential to revise drug release pattern makes PNs a potent candidate as for cancer, contraceptive, antibiotics, vaccines [2].

The limitations of PNs are high-cost formulation with low yield, productivity is more difficult, on the industrial aspect the technology transfer to commercial production is also very difficult. Stability of PNs is a big issue owing to its nano size. Highly sophisticated technology is required for manufacturing PNs [3].

Rosuvastatin Calcium is one of the most potent statins for reducing lowdensity lipoprotein cholesterol (LDL-C) level. Rosuvastatin calcium used as a lipid-lowering agent by acting as HMG CoA reductase inhibitor, and it has low extrahepatic tissue penetration, Rosuvastatin Calcium elimination half-life is 19 h with low bioavailability of about 20 %, with the maximum amount of drug excreted through faeces (90 %) which leads to a reduction in bioavailable dose in blood that reflects in less therapeutic effect [4]. This above biopharmaceutics parameter is a reason for the selection to fabricate Rosuvastatin Calcium into PNs to enhance the bioavailability of it. Hence the hypothesis of the research is as follows; Rosuvastatin Calcium bioavailability is very less because of sparingly solubility and low permeability (BCS Class II). It may lead to more toxicity due to its 19 h half-life without clearance.

So, polymeric nanoparticles were chosen to enhance the solubility, bioavailability and to decrease the toxicity of Rosuvastatin Calcium.

Another objective of this research is to optimize the polymeric nanoparticles by applying 2³ factorial designs and to perform *in vivo* pharmacokinetic evaluation of designed polymeric nanoparticles to evince the intensification of bioavailability.

MATERIALS AND METHODS

Materials

Rosuvastatin Calcium was procured from Microlabs Pvt. Ltd. India. Spans 80, Chitosan were obtained from LOBA Chemie Pvt. Ltd 107, Mumbai, India. Different instruments were used in the formulation and evaluation of polymeric nanoparticles like Magnetic Stirrer (REMI, India), Ultra Sonicator (Q Sonica, Germany), FT-IR Spectrophotometer (Bruker, India), Nanoparticles Size Analyzer (HORIBA, Japan), High-Performance Liquid Chromatography (HPLC, Shimadzu, Japan), Ultracentrifuge (REMI, India), Scanning Electron Microscopy (Zeiss Evo, USA).

Methods

Preparation of polymeric nanoparticles (PNs)

Polymeric nanoparticle (60-200 nm) was developed by using the solvent evaporation method. In this procedure, chitosan solution was done by using solvents like chloroform. To this organic phase required quantity of Rosuvastatin Calcium (20 mg) was dispersed. This organic phase is homogenized or ultrasonicated with an aqueous phase containing stabilizers like Span 80 to form 0/W type of emulsion. The emulsion is converted into polymeric nanoparticles by evaporating the solvent, in which diffusion of the solvent take place through the continuous phase of the emulsion. Continuous magnetic stirring under reduced pressure or at room temperature is carried out in order to evaporate the solvent from PNs and once the PNs is solidified they are collected with the help of ultracentrifugation followed by washing of PNs with distilled water in order to remove the additives like surfactants, At last the resultant product is lyophilized with 5% of manitol to get dried PNs and highspeed homogenization or ultrasonication is carried out in order to produce small particle size [5-8].

Experimental design for formulation of PNs

In the ongoing study, 2³ statistical designs are used with 3 levels, 2 factors and 8 runs were employed for the optimization study, which is carried out with the help of design expert software (State easy Inc, Minneapolis USA, design Expert 11). The independent variables are selected such as polymer concentration (A in mg), surfactant concentration (B in ml) and ultrasonication time (C in min) and they were set at high or low level based upon the result of the variable. According to this design, 8 PNs formulations are prepared and characterized for particle size (Y1), zeta potential (Y2), which are dependent variable i.e. which was chosen as a response parameter. These designs elucidate the main effect of independent variable over dependent variable. Its optimization design is shown in table 1 [5-8].

Evaluation parameters of PNs

Compatibility studies of drug and excipients

This test has been done by using Fourier transform infrared spectroscopy (FTIR). Pure drug and optimized formulation FTIR spectra analyzed separately, later the values are correlated for compatibility studies through reproducibility of functional groups in pure drug versus optimized formulation [9].

Percentage yield

The yield of any nanoparticles was decided by way of comparing the load of nanoparticles formed towards the total load of the drug and polymer used in PNs. The percentage yield is calculated as follows [9-11].

$$\% Yield = \frac{\text{Amount of Nanoparticles}}{\text{Amount of Drug and polymer}} X100$$

Entrapment efficiency

The PNs is subjected to centrifugation, after centrifugation, the supernatant was analyzed by UV spectrophotometer in order to verify the quantity of the drug in the supernatant liquid. The standard calibration curve method is used to determine the concentration of drug. The drug moiety quantity present in the supernatant liquid (W) was subtracted from the total quantity of drug used for the formulation of nanoparticle (w) was determined, as the result the % drug entrapment was determined by using following formula [12-14].

% Drug entrapment =
$$\frac{(W-w)}{W} X 100$$

Particulate characterization

The average particle size of PNs was found out by dynamic light scattering (DLS) at 90° angle and the sample holder's temperature is about 25 °C by using (Nanopartica SZ-100 HORIBA Scientific, Japan). For ensuring the scattering intensity of light in the range of instruments, the sample had been diluted with proportion of 1:10 v/v by double distilled water. Zeta potential is defined as the difference in potential that exist in-between the surface of the solid particle that is dispersed or immersed in a conducting liquid or in the bulk of the liquid. Zeta potential is one of the key factors that affect the stability of colloidal dispersion; zeta potential was determined by using a Zetasizer (Nanopartica SZ-100 HORIBA Scientific, Japan). For a stable nanoparticle, the zeta potential should be in the range of±30 to±60 mV and the polydispersity index should be<0.7, so that the nanoparticle will be dispersed in the continuous phase for a prolonged period of time [15, 16].

Scanning electron microscopy studies (SEM)

The SEM (Zeiss Evo, USA) technique is mainly used to determine the surface morphology, size and shape of PNs with direct visualization of the nanoparticle. The SEM posses various advantages in terms of sizing and morphological analysis. Initially, the PNs solution is converted into a dry powder and mounted on a sample holder later it was coated with a conducting metal for e.g. gold. Then the beam of electrons is focused on the sample surface results in the emission of the secondary electron from the surface of the sample, which determine the morphology of the sample. There may be the chance of damage to the polymer of the nanoparticles, which must be with

stand vaccum. The average mean size that is obtained by SEM is compared with results obtained by dynamic light scattering [17-19].

In vitro drug release study

It was executed by the dissolution USP type I (basket type) equipment. The prepared PNs was filled in a capsule and taken in the basket. The basket shaft with PNs capsule was immersed into a dissolution jar contains 900 ml of phosphate buffer (7.4 pH) solution and 37 ± 0.20 °C was maintained as bath temperature. Later 5 ml of sample is withdrawn from the dissolution jar at a specific time period i.e. 0,1,2,4,8,12,16,20 and 24 h, since in order to maintain a constant volume, the same 5 ml was restored in the dissolution jar. The sample withdrawn from the dissolution jar was examined by using UV spectrophotometer at 248 nm [20].

In vitro release kinetic study

The drug release study of PNs were fixed in different release kinetic parameters like, first-order (time vs. log % drug remaining); zero order (time vs. % cumulative release), Higuchi's model (square root of time vs. cumulative % drug release); Peppas model (Time Vs. log of drug concentration) and their regression (r^2) and k values are calculated in order to obtain a linear regression analysis to confirm the effect and mechanism of release over time [21, 22].

Stability studies

From the above-calculated results, only the optimized Rosuvastatin Calcium PNs was subjected to stability studies. PNs were filled in capsules and stored in 4 °C±2 °C. After 3 mo these capsules were analyzed at a specified period of time and measured to determine the polydispersity index (PI), particle size (nm), zeta potential (mV). Each formulation was checked for their reproducibility of results while manufacturing [23].

In vivo pharmacokinetic studies of rosuvastatin calcium loaded PNs

The *In vivo* Pharmacokinetic (PK) data of PNs was determined by orally administering Rosuvastatin Calcium loaded PNs to male albino wistar rats. The plasma drug concentration data is determined with the help of PK solver software. In this study male albino wistar rats are used of 180-250 g body weight. The animals were sorted into two groups with 6 animals each, on which the single-dose study was performed. The groups of the animals are dividing as follows:

Control	: Conventional Rosuvastatin Calcium Tablet (Arvast
group	20 mg/kg) p. o.
Test Group	: Rosuvastatin Calcium PNs Capsules (PN7) (20
	mg/kg) p. o.

Before administering the drug, the animals were fasted for 24h and they are free to access to water. Conventional Rosuvastatin Calcium (Arvast tablet) equivalent to animal dose was powdered and orally administered with the help of oral feeding needle. Rosuvastatin Calcium-loaded PNs; equivalent to animal dose was administered orally with the help of feeding needle. 0.5 ml of blood samples was collected at time intervals of about 0.5, 2, 4, 8, 12, 24 h by retro-orbital puncture by capillary tube from there into glass tubes which priorly heparinized by anticoagulant (ammonium oxalate 1% solution). After collection of the blood sample, the sample was centrifuged with the help of microcentrifugator at 4000 rpm once the centrifugation is completed, the supernatant plasma is collected and stored at-20 °C for further analysis. The plasma drug concentration was determined using HPLC by reversed-phase C18 column (250 mm X 4.6 mm i.d., 5 µm Particle size); 80:20 v/v 500 µl of 0.5% formic acid and 2 mmol ammonium acetate, which act as a mobile phase with rate of flow 1.0 ml/min with 5 µl injection volume [23-28].

Statistical analysis

The plasma drug concentration versus time data was obtained by HPLC as plotted in PK solver software. The discrete plasma drug concentration time profile gives data like C_{max} (peak plasma concentration), tmax (the time at which drugs attain peak plasma concentration), AUC_{0-∞} and AUC_{0-t}. Biological half-life (t_{1/2}), MRT are also determined by using PK solver software. The ANOVA studies were used to differentiate various pK parameters which are statistically evaluated [29, 30].

RESULTS

From the data obtained from FTIR spectra (fig. 1), it was inferred that the desired frequencies of fingerprint regions were reproducible in a physical mixture of Rosuvastatin Calcium and polymer, when compared to Rosuvastatin Calcium. It was confirmed that the drugs and excipients used in the formulations were found to be compatible with each other.

Particle size

The mean particle sizes for all the formulations are presented in table 1 and fig. 2. In general, the particle sizes for all the Rosuvastatin

Calcium PNs formulations were found to be in the range of 157.0±15.4 to 310.1±69.3 nm, based on the effect of the independent variable in the process of formulation. But the acceptance criteria, the PS of polymeric nanoparticle should be 60-200 nm. As per the acceptance criteria, the formulation PN1 (5 mg A factor; 0.5 ml of B factor and 10 min of C factor) shows 157.0±15.4 nm. The formulation PN2 (10 mg A factor; 1.0 ml of B factor and 10 min of C factor) shows 189.0±50.9 nm. The formulation PN5 (5 mg A factor; 0.5 ml of B factor and 20 min of C factor) shows 165.3±31.6 nm. The formulation PN7 (5 mg A factor; 1.0 ml of B factor and 20 min of C factor) shows 159.9±16.1 nm. Remaining formulation PN3, PN4, PN6 and PN8 formulations particle size was found to be more than desired range.



Fig. 1: Drug excipients compatibility studies-FTIR studies of (A) Rosuvastatin calcium pure drug and (B) Optimized rosuvastatin calcium PN

Zeta potential

The zeta potential for all the formulations is presented in table 1 and fig. 2. In general, the zeta potential for all Rosuvastatin Calcium PNs was found to be in the range of 4.7 ± 0.12 mV to 33.5 ± 1.54 mV, primarily based on the effect of surfactant in the

process of formulation. But the acceptance criteria of ZP of polymeric nanoparticles should be found between ± 30 to ± 60 mV. As per the acceptance criteria, the formulation PN7 (5 mg A factor; 1.0 ml of B factor and 20 min of C factor) shows a maximum ZP of- 33.5 ± 1.54 mV. The remaining formulation was found to be less than desired range.



Fig. 2: Particle characteristics; (A) PS, PI; (B) ZP of PN 7; (C) SEM images of optimized polymeric nanoparticle (PN7)

 Table 1: Optimization of the polymeric nanoparticle by 2³ factorial design and evaluation of the effect of independent variables on dependent variables by 2³ factorial design

Run	Independent variables			Dependant variables		Other variables		
	Factor A: polymer	Factor B:	Factor C:	PS*	ZP*	PI*	%EE*	%Yield*
	concentration	surfactant	ultra sonication	Y1 nm	Y2 mV			
	[Level code/mg]	concentration	time					
		[Level code/ml]	[Level code/min]					
PN1	-1/5	-1/0.5	-1/10	157.0±15.4	-4.7±0.12	1.341±0.12	90.40±2.42	93.54±2.80
PN2	1/10	-1/1.0	-1/10	189.0±50.9	-4.2±0.34	1.272±0.16	84.64±3.26	86.52±2.98
PN 3	-1/5	1/1.0	-1/10	210.3±53.8	-7.6±0.32	0.913±0.10	79.28±3.84	80.66±4.42
PN 4	1/10	1/1.0	-1/10	285.4±60.7	-5.9±0.26	0.557 ± 0.14	72.80±3.44	76.54±4.58
PN 5	-1/5	-1/0.5	1/20	165.3±31.6	-14.9±1.28	0.928±0.12	89.40±2.44	92.34±3.56
PN 6	1/10	-1/0.5	1/20	310.1±69.3	-14.3±1.36	0.547 ± 0.14	64.84±3.60	68.26±4.78
PN 7	-1/5	1/1.0	1/20	159.9±16.1	-33.5±1.54	0.587 ± 0.16	94.20±2.46	96.80±2.08
PN 8	1/10	1/1.0	1/20	230.5±60.2	-21.4±1.62	0.376±0.18	76.84±3.66	79.66±2.86

Acceptance criteria: Particle Size (PS) = 60-200 nm; Zeta Potential = ± 30 to ± 60 mV; Polydispersity Index (PI) =<0.7 for monodisperse particles; percentage entrapment efficiency (%EE) =>85 %; Percentage yield (% yield) =>85 %: *All data are measured as mean \pm SD, n=3.

Polydispersity index

The polydispersity index for all the formulations was shown in table 1 and fig. 2. The polydispersity index for Rosuvastatin Calcium PNs was found to be in the range of 0.376 ± 0.18 to 1.341 ± 0.12 , primarily based on the effect of homogenization speed or ultrasonication time in the process of formulation. But the acceptance criteria of PI should be<0.7 for monodisperse nanoparticles. As per the acceptance criteria, the formulation PN4, PN6, PN7 and PN8 shows good polydispersity index like 0.557 ± 0.14 , 0.547 ± 0.14 , 0.587 ± 0.16 and 0.376 ± 0.18 . Remaining formulations were found to be greater than 0.7.

Percentage entrapment efficiency and percentage yield

The desired Percentage entrapment efficiency and yield for polymeric nanoparticle should be more than 85 %. As per the results shown in table 1, the efficiency of entrapment was found to be 64.84 ± 3.60 % to 94.20 ± 2.46 % and % yield was found to be in the range of 68.26 ± 4.78 to 96.80 ± 2.08 %. By comparing all the

formulations, PN6 shows the very least amount of percentage entrapment efficiency and percentage yield.

In vitro drug release and in vitro release kinetics studies

The percentage amount of drug release studies were done for PN7 optimized formulation (table 2 and fig. 3). The percentage amount of drug release of PN7 was found to be 96.54 ± 2.02 % in 24 h. The regression value (r^2) for Zero order, First order, Higuchi model, Hixson crowell model and Korsmeyer Peppas model was determined to be 0.982 ± 0.02 , 0.702 ± 0.02 , 0.966 ± 0.04 , 0.842 ± 0.02 and 0.981 ± 0.04 .

In vivo pharmacokinetic studies

The comparative *in vivo* pharmacokinetic studies data between ARVAST (20 mg/kg) conventional Rosuvastatin Calcium tablet vs. polymeric nanoparticle suspension (PN7) (20 mg/kg) orally administered was shown in table 3. The T_{max} (h), C_{max} (µg/ml), AUC_{0- α} (µg/ml/h) and MRT_{0- α} (h) of ARVAST was found to be 02 h, 0.2634 µg/ml, 7.542 µg/ml/h, 08 h and PN7 was found to be 06 h, 0.1982 µg/ml, 44.2780 µg/ml/h, 12 h.



Fig. 3: Comparative In vitro drug release studies between PN7 vs. ARVAST tablet (Error bars are given mean±SD, n=3)

Table 2: In vitro drug release kinetic study data of optimized polymeric nanoparticle (PN

S. No.	Kinetic model and its regression value	
1	Zero order (r ²)*	0.982±0.02
2	First order (r ²)*	0.702±0.02
3	Higuchi model (r²)*	0.966±0.04
4	Hixson crowell model (r ²)*	0.842±0.02
5	Korsmeyer Peppas model (r ²)*	0.981±0.04
6	Korsmeyer Peppas release exponent (n)*	0.502±0.02

*All data are measured as mean±SD, n=3

Tab	ole	3:	Compara	tive <i>in</i>	<i>vivo</i> p	harr	nacol	kineti	c stud	lies (lata
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Parameter	ARVAST (20 mg/kg) conventional rosuvastatin	Polymeric nanoparticle suspension (PN7) (20 mg/kg)-
	calcium tablet-oral administration	oral administration
T _{max} (h)	02±0.0	06±0.0
C _{max} (µg/ml)	0.2634±0.002	0.1982±0.002
AUC $_{0-\alpha}$ (µg/ml/h)	7.542±0.12	44.2780±0.24
MRT $_{0-\alpha}$ (h)	08±1.0	12±1.0
F rel= (AUC) test. (Dose)std, (AUC)std. (Dose)test		Bioavailability enhanced by 5.80%

Note: Increase in AUC_{0- ∞}; MRT; T_{max}; Decrease in C_{max} in PNs shows better bioavailability than conventional dosage form (All data are measured as mean±SD, n=3)

Table 4: Comparative stability study data for PN7 b	before and after conducting stability studies
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Parameter (PN7)	Initial values during preparation*	Final values after conducting stability studies for 3 mo (4°±2° C)*
PS nm	159.9±36.1	161.1±16.1
ZP mV	-33.5±1.54	-30.5±1.40
PI	0.587±0.16	0.580±0.12

*All data are measured as mean±SD, n=3



Fig. 4: (A) PS, PI and (B) ZP report for optimized polymeric nanoparticle (PN7) at 4°±2° C after 3 mo

Stability studies

The comparative stability study data for PN7 before and after conducting stability studies was shown in table 4. The PS nm, ZP mV and PI of PN7 during preparation was found to be 159.9 ± 36.1 nm,- 33.5 ± 1.54 mV, 0.587 ± 0.16 and PN7 after performing stability studies i.e. after 3 mo on storing in $4^{\circ}\pm2^{\circ}$ C was found to be 161.1 ± 16.1 nm,- 30.5 ± 1.40 mV, 0.580 ± 0.12 .

RESULTS AND DISCUSSION

Optimization of polymeric nanoparticle

The 2³ optimization design table 1 and fig. 1 revealed the outcome of independent variables on dependent variables on Rosuvastatin Calcium PNs. From the above data, it was concluded that there was a powerful tie-up between particle size and polymer concentration, i.e. the particle size of PNs was bigger by raising the concentration of polymer. Among all the formulations (PN1-PN8), PN7 formulation showed desired particle size of about 159.9±36.1 nm at low-1 level polymer (5 mg). The minimization of particle size was due to low level of polymer concentration with high level of surfactant (table 1). And also the particle size reduction was resulted due to increase in ultrasonication time, which separated the big particle aggregates into small separated particles. Increase in surfactant concentration in the preparation of PNs showed a simultaneous increase in the zeta potential with the decrease in particle size, which confirmed the good stability of PNs in phase, so that the utmost conductivity of the particle was achieved. On enhancing the surfactant concentration, the charge distribution will be distributed uniformly on split small particle, which may lead to enhancement of zeta potential or surface charge potential, leads to good stability of nanoparticles and also keep the particle in motion without sedimentation. Among all formulations (PN1-PN8), PN7 formulation showed the required zeta potential of about-33.5±1.54 mV at high+1 level (1 ml of surfactant concentration). Increase in concentration of the surfactant and homogenization time showed a parallel increase in the ZP in mV and % EE of about 94.20±2.46. The Optimized Rosuvastatin Calcium PNs (PN7) were analyzed for surface morphology studies by SEM (fig. 2), in which the PNs were visualized as smooth spherical surfaced particles. By this it was observed that it will lead to improvement of drug loading efficiency and easy diffusion of the drug into physiological barriers due to its spherical, smooth nanometric surface. The Rosuvastatin Calcium PNs (PN7) formulation showed the maximum %yield and %entrapment efficiency of about 96.80±2.08 and 94.20±2.46%, respectively. From the abovementioned data, it may also conclude that the drug concentration was distributed uniformly in the PNs [1-23].

In vitro drug release studies

In vitro drug release studies for Rosuvastatin Calcium PNs (PN7) formulation showed a better-controlled drug release of 96.54 ± 2.02

% in 24 h compared to the conventional ARVAST tablet 20 mg formulation (fig. 3). The In vitro release kinetics studies of Rosuvastatin Calcium Polymeric Nanoparticles (PN7) were shown in table 2. The Rosuvastatin Calcium releases kinetic records for PN7 followed the zero-order release kinetic model in which the regression values (r²) were found to be 0.982 with shows good linearity. Rosuvastatin Calcium loaded PNs (PN7) followed zero order kinetics, from which the drug was released in a predetermined and controlled manner. It was confirmed as an ultimate model for the discharge of the drug in order to attain the required therapeutic action without toxic effects. Higuchi release kinetic pattern showed r²= 0.966, it confirmed diffusion type of drug release. Release exponent (n) value from peppas release kinetic data for PN7 formulation was found to be 0.502 (n = 0.45-0.89). It implied that the drug release from PNs followed a non-fickian diffusion mechanism ie. The drug was released from the polymer through polymer relaxation and diffusion [24, 25].

In vivo pharmacokinetic studies

The *in vivo* pharmacokinetic information is shown in table 3. From the data, it shows that there was a significant difference in 'p' value as<0.05 among the pharmacokinetic parameters of conventional Rosuvastatin Calcium formulation and Polymeric Nanoparticles (PN7) with T_{max} (h) of about 2 h and 6 h; and the maximum concentration of drug ($C_{max} \mu g/ml$) of 0.2634 $\mu g/ml$ and 0.1982 $\mu g/ml$. Area Under Curve (AUC $_{0-\alpha} \mu g/ml/h$) was about 7.542 $\mu g/ml/h$ and 44.2780 $\mu g/ml/h$, Mean Residence Time of the drug (MRT $_{0-\alpha}$ h) was shown as 8 h and 12 h. The *in vivo* pharmacokinetic data confirms that increase in AUC $_{0-\infty}$, T_{max} , MRT $_{0-\alpha}$ with reduce in C_{max} in PNs when compared to ARVAST conventional tablet. The relative bioavailability was calculated by considering conventional formulation as standard. It confirmed that PNs showed the enhancement of bioavailability of about 5.80 % than conventional dosage form [26, 27].

Stability studies

Short-term stability studies are done for the optimized polymeric nanoparticles (PN7) at 4 °C \pm 2 °C. The parameters were evaluated at three months' time interval. From the results of stability studies, it was observed that there was no drastic change in PS, ZP and PI of PN7. From the results (table 4 and fig. 4), it was confirmed that the drug-loaded PN7 were stable at stored temperature [28-30].

CONCLUSION

PNs show a significant enhancement of bioavailability by minimizing the dose-dependent adverse side effects of Rosuvastatin Calcium. From the above research, it was confirmed that the PNs shows control drug release pattern and also a potential drug delivery carrier for low soluble and poorly bioavailable drugs to enhance the bioavailability.

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AUTHORS OF CONTRIBUTIONS

We declare that this work was done by the all authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors.

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CONFLICT OF INTERESTS

No conflict of interest is associated with this work.

REFERENCES

- 1. Crucho Carina IC, Barros Maria Teresa. Polymeric nanoparticles: A study on the preparation variables and characterization methods. Mater Sci Eng C. 2017;80:771-84. doi: 10.1016/j.msec.2017.06.004.
- El-Say Khalid M, El-Sawy Hossam S. Polymeric nanoparticles: a promising platform for drug delivery. Int J Pharm. 2017;528(1-2):675-91. doi: 10.1016/j.ijpharm.2017.06.052, PMID 28629982.
- Kadian R. Nanoparticles: A promising drug delivery approach. Asian J Pharm Clin Res. 2018;11(1):30-5. doi: 10.22159/ajpcr.2017.v11i1.22035.
- Yadhav KS, Nagavarma BVN, Hemant. Different techniques for preparation of polymeric nanoparticles. Asian J Pharm Clin Res. 2012;5:16-23.
- Nska Aleksandra Zieli, Carreiro Filipa, Oliveira Ana M. Polymeric nanoparticles: production, characterization, toxicology and ecotoxicology. Molecules. 2020;25:1-20.
- Bisht Savita. Polymeric nanoparticles-encapsulated curcumin (nanocurcumin): a novel strategy for human cancer therapy. J Nanobiotechnology. 2007;5:1-18.
- Rajput Namita. Method of preparation of nanoparticles. IJAET. 2015;7:1806-11.
- Douglas D. Pharmaceutical nanotechnology: a therapeutic revolution. Int J Pharm Sci Dev Res. 2020;6(1):9-11. doi: 10.17352/ijpsdr.000027.
- 9. Pal Sovan Lal. Nanoparticles: an overview of preparation and characterization. J Appl Pharm Sci. 2011;1:228-34.
- 10. Lee Jonghwi, Chung Hesson. Methods of preparation of drug nanoparticles. Wiley online library; 2007. p. 14-9.
- 11. Iravani S, Korbekandi H, Mirmohammadi SV, Zolfaghari B. Synthesis of silver nanoparticles: chemical, physical and biological methods. Res Pharm Sci. 2014;9(6):385-406. PMID 26339255.
- Owens Donald E, Peppas Nicholas A. Opsonization, biodistribution, and pharmacokinetics of polymeric nanoparticles. Int J Pharm. 2006;307(1):93-102. doi: 10.1016/j.ijpharm.2005.10.010, PMID 16303268.
- Gudikandula Krishna, Charya Maringanti S. Synthesis of silver nanoparticles by chemical and biological methods and their antimicrobial properties. J Exp Nanosci. 2016;11(9):714-21. doi: 10.1080/17458080.2016.1139196.
- 14. Schmidt Helmut. Nanoparticles by chemical synthesis, processing to materials and innovative applications. Appl Organometal Chem. 2001;15(5):331-43. doi: 10.1002/aoc.169.
- You Xinru, Kang Yang, Hollett Geoffrey, Chen Xing, Zhao Wei, Gu Zhipeng, Wu Jun. Polymeric nanoparticles for colon cancer therapy: overview and perspectives. J Mater Chem B. 2016;4(48):7779-92. doi: 10.1039/c6tb01925k. PMID 32263770.

- Norouzi Banis H. Synthesis of silicide nanomaterials using chemical vapour deposition method. E Theses Dissertations. 2012;767:50-66.
- 17. Xia W, Su D, Birkner A, Ruppel L, Wang Y, Wöll C, Qian J, Liang C, Marginean G, Brandl W, Muhler M. Chemical vapor deposition and synthesis on carbon nanofibers: sintering of ferrocene-derived supported iron nanoparticles and the catalytic growth of secondary carbon nanofibers. Chem Mater. 2005;17(23):5737-42. doi: 10.1021/cm051623k.
- Hussain M, Sarma A, Rahman SS, Siddique AM, Eeswari TP. Formulation and evaluation of ethambutol polymeric nanoparticles. Int J App Pharm. 2020;12:207-17. doi: 10.22159/ijap.2020v12i4.36845.
- 19. Elbaz Nancy M. Core-shell silver polymeric nanoparticles-based combinational therapy against Breast cancer *in vitro*. Sci Rep 2016;6:1-9.
- Shibata Annemarie, McMullen Emily, Pham Alex, Belshan Michael, Sanford Bridget, Zhou You, Goede Michael, Date Abhijit A, Destache Christopher J. Polymeric nanoparticles containing combination antiretroviral drugs for HIV type 1 treatment. AIDS Res Hum Retroviruses. 2013;29(5):746-54. doi: 10.1089/aid.2012.0301, PMID 23289671.
- Parveen Suphiya, Sahoo Sanjeeb K. Polymeric nanoparticles for cancer therapy. J Drug Target. 2008;16(2):108-23. doi: 10.1080/10611860701794353, PMID 18274932.
- Khalid M. Polymeric nanoparticles: a promising platform for drug delivery. Int J Pharm. 2017;7:675-91.
- Gajra Balaram, Patel Ravi R, Dalwadi Chintan. Formulation, optimization and characterization of cationic polymeric nanoparticles of mast cell stabilizing agent using the boxbehnken experimental design. Drug Dev Ind Pharm. 2016;42(5):747-57. doi: 10.3109/03639045.2015.1093496, PMID 26559522.
- Dustgania Amir, Farahania Ebrahim Vasheghani, Imanib Mohammad. Preparation of chitosan nanoparticles loaded by dexamethasone sodium phosphate. Iran J Pharm Sci. 2008;4:111-4.
- Zhang Zhiping, Lee Sie Huey, Gan Chee Wee, Feng Si-Shen. *In vitro* and *in vivo* investigation on PLA-TPGS nanoparticles for controlled and sustained small molecule chemotherapy. Pharm Res. 2008;25(8):1925-35. doi: 10.1007/s11095-008-9611-6, PMID 18509603.
- Gabr Mai Mahmoud, Mortada Sana Mohamed, Sallam Marwa Ahmed. Hexagonal liquid crystalline nanodispersions proven superiority for enhanced oral delivery of rosuvastatin: *In vitro* characterization and *in vivo* pharmacokinetic study. J Pharm Sci. 2017;106(10):3103-12. doi: 10.1016/j.xphs.2017.04.060, PMID 28479357.
- Cooper Kelvin J, Martin Paul D, Dane Aaron L, Warwick Mike J, Schneck Dennis W, Cantarini Mireille V. Effect of itraconazole on the pharmacokinetics of rosuvastatin. Clin Pharmacol Ther. 2003;73(4):322-9. doi: 10.1016/s0009-9236(02)17633-8, PMID 12709722.
- Kanukula Raju, Salam Abdul, Rodgers Anthony, Kamel Bishoy. Pharmacokinetics of rosuvastatin: A systematic review of randomised controlled trials in healthy adults. Clin Pharmacokinet. 2021;60(2):165-75. doi: 10.1007/s40262-020-00978-9, PMID 33428168.
- Souto EB, Wissing SA, Barbosa CM, Muller RH, Soutoab SA, Wissinga CM Barbosab, RH Muller. Evaluation of the physical stability of SLN and NLC before and after incorporation into hydrogel formulations. Eur J Pharm Biopharm. 2004;58(1):83-90. doi: 10.1016/j.ejpb.2004.02.015, PMID 15207541.
- Makoni Pedzisai A, Wa Kasongo Kasongo, Walker Roderick B. Short-term stability testing of efavirenz-loaded solid lipid nanoparticle (SLN) and nanostructured lipid carrier (NLC) dispersions. Pharmaceutics. 2019;11(8):397. doi: 10.3390/pharmaceutics11080397, PMID 31398820.