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*Epichloid* endophytes confer resistance to the smut *Ustilago bullata* in the wild grass *Bromus auleticus* (Trin.)

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- 1 Epichloid endophytes confer resistance to the smut Ustilago bullata in the
- 2 wild grass Bromus auleticus (Trin.)
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14

#### 15 Abstract

- In this work it was studied for the first time whether asexual *Epichloë* (*Neotyphodium*)
- endophytes of *Bromus auleticus*, protect their host plants against the pathogenic fungus
- 18 Ustilago bullata.
- 19 Seeds of two different ecotypes of *B. auleticus*, one of them infected with the endophyte
- 20 Neotyphodium pampeanum (NpE+) and the other infected with the endophyte N.
- 21 tembladerae (NtE+) and their respectively endophyte-free (NpE-/NtE-) counterparts were
- used. Seeds of each ecotype and endophytic status were superficially disinfected and were
- 23 randomly assigned to different treatments named: S+ (smut fungus inoculated) and S-
- 24 (mock-inoculated). It was evaluated the effect of *Ustilago bullata* infection on plant
- characteristics in every stage of their life cycle: seedling emergence, vegetative growth,
- 26 mortality and smut symptoms in the florets.
- 27 In NtE+ infected plants, smut disease was almost completely suppressed, whereas in their
- 28 endophyte-free counterparts (NpE-) the incidence of smut symptoms reached 64%. In
- 29 NpE+ infected plants smut incidence was significantly lower (7%) than in endophyte-free
- plants (39%). Although *U. bullata* infection decreased the emergence rate of both
- 31 endophyte-infected and endophyte-free plants, neutral or protective effects of the

32	endophytes were observed in seedling development and survival. The survival during the
33	first year of NtE+ plants was higher than in their NtE- counterparts.
34	These results indicate a strong beneficial effect of vertically transmitted endophytes against
35	this pathogen.
36	
37	Keywords
38	Defensive mutualism; Grass-endophytes; Neotyphodium; Smut disease; Ustilago bullata
39	
40	1. Introduction
41	Some cool-season grasses (subfamily Pooideae) establish symbiotic associations with
42	endophytic fungi of the genus Epichloë Tul. and their asexual derivatives Neotyphodium
43	Glenn, Hanlin & Bacon (Clavicipictaceae, Hypocreales, Ascomycota). This association is
44	quite specific and so each endophytic species is able to colonize one or a few host species.
45	These fungi colonize the plant shoot meristems where they grow systemically in the
46	apoplast of developing leafs and culms obtaining nutrients (Kuldau and Bacon, 2008).
47	Since its growth is synchronized with the growth of the host plant and does not require the
48	degradation of cell walls of the host, no noticeable symptoms of endophytic infection are
49	produced (Christensen et al., 2008; Christensen and Voisey, 2007). Epichloë species
50	produce stromata with perithecia in the culms of reproductive tillers avoiding the
51	development of the flowers, causing total or partial sterility of the host plant (choke
52	disease). Ascospores produced in the perithecia are forcibly discharged and are responsible
53	for the infection of new plants. Some Epichloë species and most of Neotyphodium species
54	do not produce stromata. In these asexual species, hyphae colonize meristems of the
55	developing flowers and remain visible, in the mature seeds, between the aleurone cell layer
56	and the seed coat (Schardl et al., 2004; White, 1993). Thus, these endophytes are vertically
57	transmitted through the seeds of the host plant.
58	The associations between grasses and epichloid endophytes, mainly those established with
59	vertically transmitted endophytes, are considered in general as mutualists (Clay and
60	Schardl, 2002; Müller and Krauss, 2005; Schardl et al., 2004). The plant provides
61	photosynthates and shelter to the endophytes and they provide several benefits to the host

- 62 plant. Among these benefits, the most important are protection against herbivores, mediated
- by the production of different fungal alkaloids including loline and peramine, mainly toxic
- to insects, and lolitrems and ergot alkaloids that affect primarily cattle (Bacon, 1977; Clay
- and Schardl, 2002; Lane et al., 2000; Latch, 1993; Panaccione et al., 2006; Popay et al.,
- 66 2009; Schardl et al., 2007; 2004; Schardl and Phillips, 1997; Torres et al., 2008). Increased
- 67 growth and drought resistance have also been attributed to these endophytes in agronomic
- and native wild grasses (Clay, 1987; Iannone and Cabral, 2006; Novas et al., 2003).
- However, the endophyte may be detrimental under some environmental conditions and in
- some host species (Cheplick and Faeth, 2009; Faeth et al., 2004).
- Endophytes seem to protect their host against some fungal pathogens (Bonos et al., 2005;
- 72 Clarke et al., 2006; Gwinn and Gavin, 1992; Nan and Li, 2000; Yue et al., 2000) and also
- 73 to modulate positively or negatively the interaction between their hosts and arbuscular
- mycorrhizal fungi (AM) (Chu-Chou et al., 1992; Guo et al., 1992; Liu et al., 2011; Mack
- 75 and Rudgers, 2008; Müller, 2003; Novas et al., 2005; 2009; Omacini et al., 2006).
- 76 Smut fungi (Ustilaginales, Basidiomycota) are common pathogens of cereals and are
- studied because of their impact on agriculture worldwide (Agrios, 2005; Wilcoxson et al.,
- 78 1996). These pathogens cause diseases and losses in crops (Martínez-Espinoza et al., 2002;
- 79 Wilcoxson et al., 1996) and also infect wild grasses, such as Festuca and Lolium (Durán
- and Fischer, 1961; Vánky, 1994).
- 81 Although several smut fungi species may present differences in their life cycles, all of them
- 82 cause sterility in their hosts. The ovary of the infected plants is replaced by the pathogen
- that produces masses of spores, known as teliospores, in the sori within host tissues
- 84 (Martínez-Espinoza et al., 2002). Teliospores are resting spores that are spread by wind and
- remain in the soil or attached to the lemma and palea or to the cariopses coat (Agrios,
- 86 2005). Dikariotic teliospores that undergo karyogamy, germinate along with the seed
- 87 forming a germ tube (promycelia) (Alexopoulos et al., 1996; Meyer et al., 2001). The
- 88 diploid nucleus migrates to the promycelium and undergoes meiosis forming four haploid
- 89 basidiospores. Basidiospores can either unite as compatible mating types producing the
- 90 infection hypha, or they can proliferate mitotically to produce sporidia. Sporidia of
- ompatible mating types may then fuse to penetrate the host as a dikaryotic hypha (Agrios,
- 92 2005).

93	Ustilago bullata, the causal organism of head smut of grasses, is a highly polymorphic and
94	systemic smut fungus that infects its host soon after the emergence of the coleoptile from
95	the seed (Falloon, 1979; Fischer, 1940). The presence of the fungus in their host becomes
96	apparent at anthesis when the glumes and ovary of infected hosts are destroyed, being
97	replaced by a dark black mass of teliospores (Falloon and Hume, 1988). The effects of U.
98	bullata on Bromus spp, invasive species in USA, or forage species have been extensively
99	studied by Falloon (1976; 1979); Falloon and Hume (1988); García-Guzmán et al. (1996);
100	Hirschhorn (1986); Meyer et al. (2001).
101	Bromus auleticus Trin., is a native perennial grass that inhabits grasslands of Argentina,
102	Uruguay and southern Brazil. In Argentina, B. auleticus is infected by two species of
103	endophytes with a frequency of infection higher than 95% in most of the studied
104	populations (Iannone et al., 2009). This grass has been reported as host of the smut $U$ .
105	bullata Berk (Astiz Gassó and Molina, 2010; Traverso, 2001). Field surveys carried out in
106	Argentina indicate that, infection of B. auleticus by U. bullata has not been very commonly
107	observed in nature, but in field assays, studying endophyte-free plants, smut symptoms
108	produced by <i>U. bullata</i> are usually observed (De Battista, personal communication).
109	In grasses infected simultaneously by vertically transmitted epichloid endophytes and smut
110	fungi, both fungi compete for the colonization of the ovary and require, in a different way,
111	the flower for their reproduction and dissemination. If the endophyte is able to avoid the
112	replacement of the ovary by the smut fungus, leading to the development of a normal seed,
113	both the host plant and the endophyte will be able to reproduce and disperse.
114	The triple interaction host plant-epichloid endophyte-smut fungus represents an interesting
115	model to study the effect of endophyte on pathogenic fungi that remains to be explored.
116	Thus, the aim of this study was to establish whether vertically transmitted endophyte
117	species confer resistance to the smut fungus in the pathosystem Ustilago bullata-Bromus
118	auleticus Trin.
119	
120	2. Materials and methods
121	2.1. Plant and smut fungus material

4

122	Endophyte infected $(E+)$ and endophyte-free $(E-)$ seeds of two different ecotypes of $B$ .
123	auleticus, originally from Intendente Alvear, La Pampa province (LP ecotype), Argentina,
124	infected with Neotyphodium pampeanum Iannone & Cabral and from El Palmar (EP
125	ecotype), Entre Ríos province, Argentina associated with Neotyphodium tembladerae
126	Cabral & White (Iannone et al., 2009) were used. Endophyte-free seeds of each ecotype
127	were obtained in 2007 by loss of endophyte viability in long term stored seeds. Since 2007,
128	E+ and E- plants of each ecotype were grown in the field and seeds are collected every
129	year. Seeds used for all the experiments described below were collected during December
130	from the previous year to each experiment described below.
131	Teliospores of <i>U. bullata</i> were collected from infected <i>Bromus catharticus</i> plants in
132	December 2008 and 2009. Diseased florets exhibiting fully ripen sori were collected and
133	mildly ground in a mortar and a pestle to release the teliospores. The powder containing
134	teliospores and pieces of vegetal tissues was sieved in a 1mm sieve to remove plant tissues.
135	Teliospores were kept dry at 4 °C and were used during the first 12 months after the
136	collection. For the taxonomic identification of Ustilago bullata, ITS region was amplified
137	by PCR accordingly to White et al. 1990. PCR product was purified and sequenced in an
138	ABI 3730xl DNA Analyzer. Identification of the smut fungus was performed by means of
139	BLAST on the GenBank database and followed by phylogenetic analyses using Maximum
140	Parsimony (Winclada v0.9.9) (Nixon, 1999) and MrBayes algorithms (Mr. Bayes 3.2)
141	(Ronquist et al., 2012) (not shown).
142	
143	2.2. Endophyte detection
144	The endophytic status of the seed lots and plants was established by the examination of the
145	endophyte in seeds previous to each experiment and in the seedlings or plants at the end of
146	each experiment. To confirm the presence of the endophyte in seeds, caryopses were
147	soaked for 5 h in a 10 % v/v aqueous solution of sodium hydroxide at room temperature
148	(22–24°C), and then rinsed and stained with aniline blue (0.1% aqueous) (Clark et al.,
149	1983). Endophytic mycelia were visualized in parenchymal tissues within the culm pith or
150	in the parenchyma of peeled sheaths, aniline blue stained as mentioned and observed under
151	a light microscope. Plants were considered as endophyte infected if a mass of dark blue

152	hyphae was observed between the aleurone cell layer and the seed coat or when
153	characteristic unbranched hyphae were observed in parenchymal tissues.
133	characteristic unbranched hyphae were observed in parenchymai tissues.
154	
155	2.3. Treatments
156	For all the experiments discussed below, seeds of each ecotype and endophytic status (N.
157	pampeanum-infected (NpE+); N. pampeanum-free (NpE-); N. tembladerae-infected (NtE+)
158	and N. tembladerae-free (NtE-)) were superficially disinfected by consecutive washes as
159	follows: ethanol 50%, 1 minute; sodium hypochlorite 2%, 5 minutes and ethanol 50%, 1
160	minute. E+ and E- seeds of each ecotype were randomly assigned to the different
161	treatments named: S+ (smut fungus inoculated) and S- (mock-inoculated). To achieve this,
162	seeds assigned to S+ treatments were placed in Petri dishes and a powder of teliospores
163	(0.15 mg teliospores.seed <sup>-1</sup> ) was poured on them. For control treatments, a mock-
164	inoculation with heat inactivated teliospores (180°C for 4 hours) was done. The Petri dishes
165	were closed and gently shaken for 5 minutes to obtain a homogeneous spore distribution on
166	the seeds. In this way, 4 treatments were established for each ecotype named as follows:
167	NpE+S+; NpE+S-; NpE-S+; NpE-S-; NtE+S+; NtE+S-; NtE-S+ and NtE-S Before the
168	inoculation, the viability of the teliospores was evaluated by preparing a suspension of
169	teliospores in water $(1.5 \times 10^8 \text{ spores.ml}^{-1})$ . Fifty $\mu l$ of the solution were spread in Petri
170	dishes with water agar 2% and incubated 6 hours in darkness at 24°C. Spores able to
171	germinate (producing a germinating tube) were considered as viable and the percentage of
172	germination was registered. Teliospores viability ranged between 30 and 60%, and in those
173	inactivated for the S- treatments the percentage of germination was zero (even when the
174	inactivated teliospores were re-checked after 72 hours of incubation).
175	
176	2.4. Effect of Ustilago bullata on seedling emergence and plant development
177	In order to determine the effect of infection by <i>U. bullata</i> on <i>B. auleticus</i> seedling
178	emergence and development, 150 seeds of each treatment and ecotype were sown in ten
179	Petri dishes (ten replicates with fifteen seeds/dish in each treatment) filled with sterilized
180	sand and incubated in a growing chamber at 22 °C under 12 hours photoperiod. The
181	percentage of seed germination was recorded and shoot length was measured after 15 days

182	from the sowing. Results were analyzed by a two way ANOVA (p<0.05) for each ecotype
183	where the inoculation with the smut fungus and the endophytic status were the main
184	factors. All data analyses were performed using the Infostat software (Di Rienzo et al.,
185	2011).
186	
187	2.5. Evaluation of plant survival and smut symptoms development
188	One hundred and fifty seeds of each ecotype and endophytic status were inoculated with
189	teliospores as described above. Seeds of each treatment were germinated in trays filled with
190	sterilized sand in a growth chamber at 22 °C under 12 hours photoperiod. Two-month-old
191	seedlings were transplanted individually to 25 cm deep x 15 cm in diameter pots, filled with
192	commercial garden soil: sand: perlite 3:1:1 and transferred outdoors to the experimental
193	field of the Facultad de Ciencias Exactas y Naturales, University of Buenos Aires where
194	they were allowed to grow and produce flowers. During this period, the survival of the two-
195	month-old seedlings (before being transplanted to pots in the field), plant survival before
196	flowering and the incidence of the disease in NpE+/NtE+ or NpE-/NtE- plants were
197	evaluated. The incidence of the disease in each treatment was evaluated as the number of
198	plants with symptoms (flowers with sori/number of flowered plants). For each ecotype, the
199	differences among treatments in seedling and plant survival and disease incidence were
200	compared by means of a Chi-square test of homogeneity of proportions and the Marascuilo
201	procedure was used to make comparisons between all pairs of groups (Marascuilo and
202	McSweeney, 1977).
203	
204	2.6. Vertical transmission of the endophyte via seeds
205	In those plants that produced seeds, the transmission of the endophyte was evaluated by
206	checking the presence of endophyte in the seeds, as previously described.
207	
208	3. Results
209	3.1. Seedling emergence

210 The inoculation with teliospores of *U. bullata* in seeds decreased the overall percentage of emergence of B. auleticus (in LP ecotype  $F_{1:36}$ =63.46 P<0.0001 and in EP  $F_{1:36}$ =5.28 211 212 P=0.0275). The presence of the endophyte did not affect seedling emergence (LP:  $F_{1:36}$ =0.40 P=0.5293; EP:  $F_{1:36}$ =2.98 P=0.0930) (Fig.1). 213 In LP ecotype the seedling emergence in NpE+S+ treatment was 49% lower than in 214 215 NpE+S- treatment, whereas in NpE-S+ treatment was 35% lower than in NpE-S- treatment, but the difference in the emergence between NpE+S+ and NpE-S- treatment was not 216 statistically significant (Fig. 1A). In EP ecotype seedling emergence decreased 16% in 217 NtE+ seeds while in NtE- seeds the germination was 23% lower than in the control (Fig. 218 1B). 219 220 3.2. Seedling growth 221 No significant differences were observed in the shoot length between E+ and E- plants of 222 each ecotype (LP:  $F_{1:36}$ =1.31, P=0.2592; EP:  $F_{1:36}$ =2.43, P=0.1278) (Fig. 2). However, in 223 both ecotypes, seedlings were negatively affected by the presence of the smut fungus (LP: 224  $F_{1:36}$ =138.14, P<0.0001; EP:  $F_{1:36}$ =39.46, P<0.0001). Ustilago bullata effects were more 225 evident in LP ecotype where NpE+S+ and NpE-S+ plants were 46% and 43% smaller 226 227 respectively than their S- counterparts (Fig. 2A). In EP ecotype, NtE-S+ seedlings were 228 39% smaller than the NtE-S- ones, whereas NtE+S+ seedlings were only 29% smaller than their NtE+S- counterparts (Fig. 2B), but this difference was not statistically significant. 229 230 3.3. Plant survival 231 232 The inoculation of seeds with teliospores of *U. bullata* decreased the seedlings survival of both ecotypes, during the first 60 days of growth (LP:  $\chi^2_{0.95;3}$ =32.02; P<0.0001and EP: 233  $\chi^2_{0.95;3}$ =61.04; P<0.0001) (Fig. 3A and B). However, while in LP ecotype no differences 234 were observed due to the endophytic status among the smut inoculated plants, in NtE+S+ 235 seedlings was significantly higher than in the NtE- ones. 236 Among the plants grown to evaluate the development of the disease at the flowering time, 237 the percentage of survival during the first year of growth in the field was significantly 238

239	higher in plants grown from S- seeds (Fig. 3C and D) (LP: $\chi^2_{0.95;3}$ =74.67; $P$ <0.0001 and
240	EP: $\chi^2_{0.95;3}$ =78.32; $P$ <0.0001).
241	In LP ecotype, even though only the 15% of the NpE-S+ plants survived, this value was not
242	significantly different from the 31% of survival presented by the NpE+S+ ones (Fig. 3C).
243	On the other hand in EP ecotype the 65.8 % of survival presented by the NtE+S+ plants
244	was significantly higher than that observed in the NtE-S+, where only the 3% of the plants
245	survived (Fig. 3D).
246	
247 248	3.4. Development of smut symptoms in field
240	
249	The presence of smut disease symptoms in the florets was evaluated in one or two year old
250	plants grown in pots at field conditions. Disease incidence was almost totally suppressed or
251	significantly diminished in E+ plants of both ecotypes (LP: $\chi^2_{0.95;l}$ =12.67; $P$ =0.0004 and
252	EP: $\chi^2_{0.95;i}$ =78.21; $P$ <0.0001) (Fig. 4). None of the control plants (mock-inoculated)
253	presented symptoms of disease (not shown in figure 4). In the plants that presented smut
254	symptoms all the flowers were destroyed by the pathogen.
255	
256	3.5. Vertical transmission of the endophyte
257	None of the NpE+ or NpE- smut-symptomless plants (from LP ecotype) produce fully ripen
258	seeds. In EP ecotype 11 plants produced fully ripen seeds, but only two to five seeds were
259	produced by each plant. The analysis of the presence of the endophytes in the seeds showed
260	that all the seeds were endophyte infected; indicating that in EP ecotype, the inoculation
261	with Ustilago bullata did not affected the transmission of the endophyte to the seeds.
262	
263	4. Discussion
264	The present work, to our knowledge, is the first report of protective effect of <i>Neotyphodium</i>
265	endophytes against a systemic pathogen like U. bullata that produces castration of the

266	plants. Our findings suggest that plants of Bromus auleticus associated with Neotyphodium
267	tembladerae or N. pampeanum were more resistant to the "head smut" of grasses produced
268	by Ustilago bullata than endophyte-free plants.
269	In this work we found that, whereas in endophyte-free plants the incidence of the disease
270	reached 39 to 64%, in endophyte-infected plants disease incidence ranged from 1 to 7%. In
271	those plants that presented smut symptoms seed production was totally suppressed
272	producing sterility in the affected plants. Thus, our results show that the endophytes prevent
273	castration of the host plant, ensuring sexual reproduction of the host. Although the amount
274	of fully ripen seeds produced by control or symptomless plants, in the S+ treatment, and
275	checked for endophyte infection, was not enough to evaluate accurately the efficiency of
276	the transmission of the endophyte through the seeds; our results also showed that the
277	vertical transmission of the endophyte is not affected by the inoculation of the smut fungus.
278	In vitro assays, performed in our laboratory, showed that teliospore germination is inhibited
279	by N. pampeanum and N. tembladerae (Iannone et al., 2012b). Protective effects of
280	epichloid endophytes against plant fungal pathogens such as Laetisaria fuciformis (Bonos
281	et al., 2005), Alternaria alternata, Fusarium (Nan and Li, 2000), Cercospora,
282	Cryphonectria parasitica (in vitro) (Yue et al., 2000), Sclerotinia homeocarpa (Clarke et
283	al., 2006), Rhizoctonia zeae (Gwinn and Gavin, 1992) have been also reported. All
284	together, these results are in agreement with the hypothesis of the defensive mutualism
285	suggested for the grass-endophyte associations (Clay, 1988; 1989; Saikkonen et al., 2010).
286	In spite of the beneficial effects observed in E+ plants with respect to prevention of smut
287	disease development, the endophytes had neutral effects on seedling emergence and
288	growth, since these variables where similarly (negatively) affected by the presence of the
289	smut fungus both in the E+ as in the E- treatments. Considering that <i>U. bullata</i> requires
290	flower production for its dissemination, negative effects on plant survival and development
291	should not be expected. However, these kind of effects produced by this pathogen on its
292	host plants were also reported in Bromus catharticus (Falloon, 1976; García-Guzmán et al.,
293	1996). In addition, we consider that the amount of teliospores used in each experiment was
294	significantly higher than that expected to be found in nature since after the inoculation the
295	seeds remained totally covered by a black coat of spores. Thus, detrimental effects of the

296	smut fungus could have been enhanced and some of the protective effects of the endophyte
297	could have been masked in our experiments. Protective effects of the endophyte could be
298	even more important in natural conditions where the charge (inoculum) of teliospores is
299	expected to be lower.
300	Different behaviors were observed between plants of different ecotypes, whereas smut
301	development was almost totally suppressed in NtE+ plants (EP ecotype; N. tembladerae-
302	infected), in NpE+ plants (LP ecotype; N. pampeanum-infected) the incidence of the
303	disease was diminished but not so drastically as in NtE+ plants. In the presence of the
304	pathogen, survival of NtE+ plants was higher than in the NtE- ones, but no differences were
305	observed between NpE+ and NpE- plants. These differences observed in plant survival and
306	disease incidence between the E+ plants of each ecotype seem to indicate that the
307	protective effects of N. tembladerae against this pathogen are stronger than those conferred
308	by N. pampeanum. However, we cannot discard that the observed differences could be due
309	to differences in the susceptibility of each plant ecotype. Supporting our hypothesis of a
310	higher protective capacity of <i>N. tembladerae</i> , there are <i>in vitro</i> studies that showed that <i>N</i> .
311	tembladerae presented the highest inhibitory capacity against several fungal plant
312	pathogens with respect to other Epichloë/Neotyphodium species (Yue et al., 2000) and
313	against U. bullata (Iannone et al., 2012b).
314	The protective effects shown in the E+ plants against the head smut of grasses disease
315	could be due to, 1) the endophytes preventing the infection by <i>U. bullata</i> at seedling stage
316	or 2) the endophytes preventing the colonization of the ovary by the pathogen. The
317	detrimental effects of smut fungus inoculation observed on seedling emergence, seedling
318	survival and development in E+ and E- treatments would support hypothesis 2, indicating
319	that the smut fungus is able to infect the seedlings of B. auleticus irrespectively of their
320	endophytic status.
321	The higher survival and resistance of endophyte-infected plants to <i>U. bullata</i> , in addition to
322	other beneficial properties observed in endophyte infected plants (Iannone et al., 2012a)
323	could explain the higher incidence of endophytes in populations (smut-symptomless) of this
324	host in nature. Endophyte infected plants produced more seeds than E- ones (Iannone et al.,
325	2012a) and seed production was suppressed in E- plants when infected with <i>U. bullata</i> .
326	However, considering that <i>B. auleticus</i> is a highly perennial plant, more long term studies

327	are necessary in order to evaluate the importance of <i>Ustilago bullata</i> and the effects of both
328	symbionts on the dynamics of the populations of this host.
329	Although more research should be done in order to establish the mechanism through which
330	both fungal symbionts interact in the host plant so that the incidence of the disease is lower
331	in E+ plants; our findings are relevant for a better understanding of the biology of the grass-
332	endophyte symbiosis and could be also applied in grass breeding programs. Currently,
333	studies are being conducted in our laboratory in order to evaluate the effect of the
334	endophyte-smut fungus interactions in plant competition, and seed production in field.
335	Finally, O'Hanlon et al. (2012) stated that more attention should be paid to dissecting the
336	potential of fungal endophytes as biological control agents against cereal pathogens. In this
337	sense, our studies and results on smut resistance should be expanded to other endophyte-
338	infected grasses, mainly wild barley species as Hordeum bogdanii, H. brevisubulatum and
339	H.comosum.
340	
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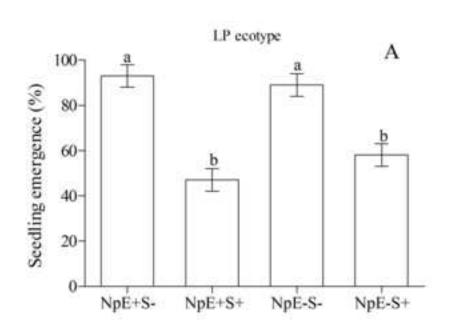
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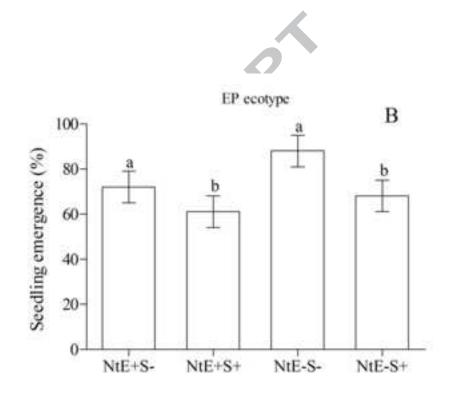
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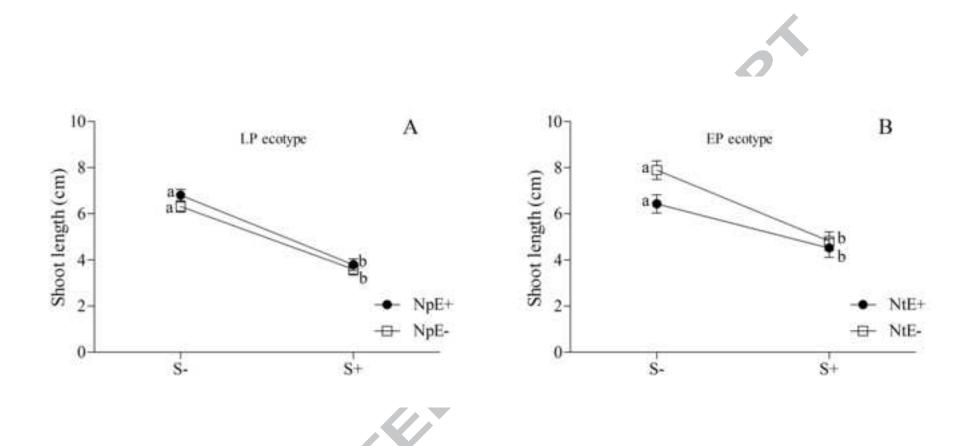
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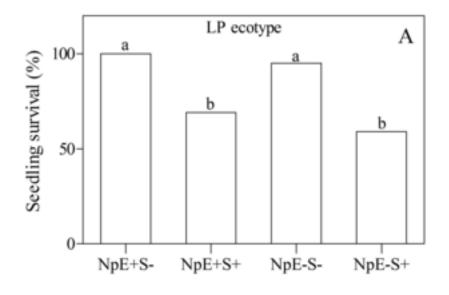
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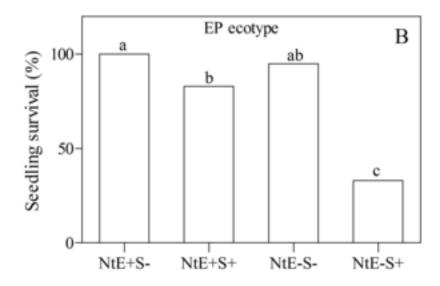
496	
497	Fig. 1. Effect of Ustilago bullata and Neotyphodium pampeanum (Np) (A) or N. tembladerae (Nt)
498	(B) endophytic status on seedling emergence of Bromus auleticus. Endophyte infected
499	(NpE+/NtE+) and endophyte free (NpE-/NtE-) seeds, inoculated (S+) or mock-inoculated (S-) with
500	Ustilago bullata. Different letters indicate significant differences (P<0.05).
501	
502	Fig. 2. Effect of Ustilago bullata and Neotyphodium pampeanum (Np) (A) or N. tembladerae (Nt)
503	(B) on Bromus auleticus seedlings shoot length (cm). Endophyte infected (NpE+/NtE+) and
504	endophyte free (NpE-/NtE-) seeds, inoculated (S+) or mock-inoculated (S-) with <i>Ustilago bullata</i> .
505	Data are means; SE. Different letters indicate significant differences ( <i>P</i> <0.05).
506	
507	Fig. 3. Effect of Ustilago bullata and Neotyphodium pampeanum (Np) (A, C) or N. tembladerae
508	(Nt) (B, D) on Bromus auleticus seedlings survival during the first two months of growth (A, B)
509	and plants survival during the first year of growth under field conditions (C, D). Endophyte infected
510	(NpE+/NtE+) and endophyte free (NpE-/NtE-) seeds, inoculated (S+) or mock-inoculated (S-) with
511	Ustilago bullata. Different letters indicate significant differences (P<0.05).
512	
513	Fig. 4. Percentage of Bromus auleticus plants inoculated with Ustilago bullata with smut symptoms
514	in the florets. NpE+: Neotyphodium pampeanum infected; NpE-: N. pampeanum free; NtE+: N.
515	tembladerae infected and NtE-: N. tembladerae free. Different letters indicate significant
516	differences within each ecotype ( $P$ <0.05). The plants in those treatments inoculated with inactive
517	teliospores (S-) did not present smut disease symptoms (not shown in the figure).
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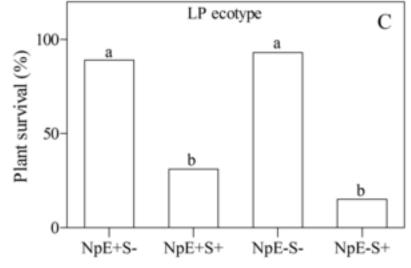


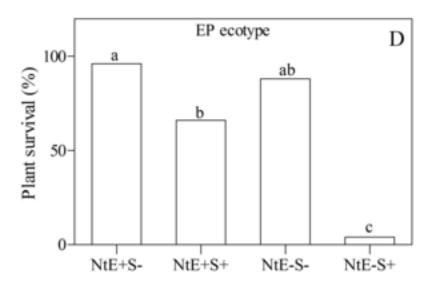




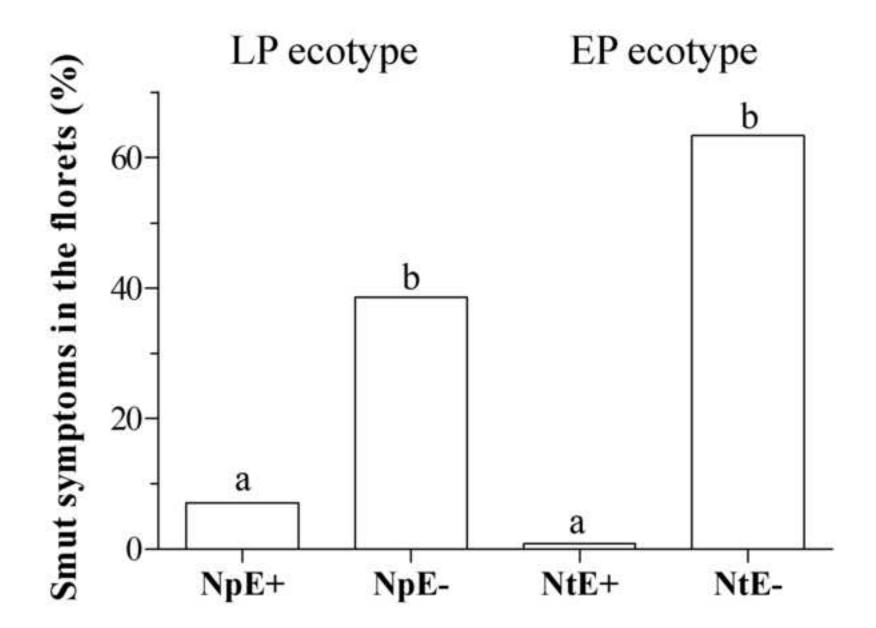




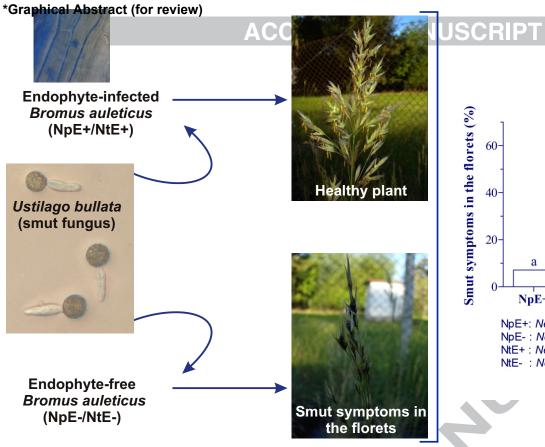


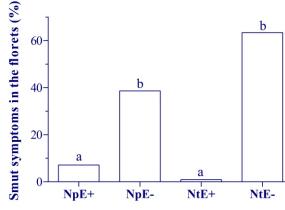












NpE+: Neotyphodium pampeanum-infected NpE-: Neotyphodium pampeanum-free NtE+: Neotyphodium tembladerae-infected NtE-: Neotyphodium tembladerae-free

525	Highlights
526	Some grasses are usually co-infected by smut fungi and mutualist epichloid endophytes.
527	Endophytes are transmitted via seeds and smut fungi replace the seeds with teliospores.
528	The endophyte and the smut fungus compete in a race for the colonization of the ovary.
529	The effect of <i>Neotyphodium</i> spp. against head smut of grasses was evaluated.
530	Disease incidence was diminished in endophyte-infected <i>Bromus auleticus</i> plants.
531	