



## UvA-DARE (Digital Academic Repository)

### Anti-neutrophil cytoplasm autoantibodies (ANCA). Clinical and functional studies

Reumaux, D.

**Publication date**  
2002

[Link to publication](#)

#### **Citation for published version (APA):**

Reumaux, D. (2002). *Anti-neutrophil cytoplasm autoantibodies (ANCA). Clinical and functional studies.*

#### **General rights**

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

#### **Disclaimer/Complaints regulations**

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: <https://uba.uva.nl/en/contact>, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.

## CHAPTER V

### Pro-Myeloperoxidase: A Target Antigen for Anti-Neutrophil Cytoplasm Autoantibodies (ANCA)

*Dominique REUMAUX*<sup>1</sup>  
*Patrick DUTHILLEUL*<sup>1</sup>  
*William M. NAUSEEF*<sup>2</sup>  
*Dirk ROOS*<sup>3</sup>

<sup>1</sup> Département d'Hématologie-Immunologie-Cytogénétique, Centre Hospitalier de Valenciennes, Valenciennes, France

<sup>2</sup> Department of Medicine, Veterans Administration Medical Center and the University of Iowa, Iowa City, USA

<sup>3</sup> Sanquin Research at Central Laboratory of the Netherlands Blood Transfusion Service (CLB); and Landsteiner Laboratory, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands

**Pro-Myeloperoxidase: A Target Antigen for Anti-Neutrophil Cytoplasm Autoantibodies (ANCA)**

*To the Editor:*

We read with interest the article by Russel *et al.* regarding the relationship between anti-neutrophil cytoplasm autoantibodies (ANCA) reacting with the pro-form of proteinase-3 (PR3) and disease activity in patients with Wegener's granulomatosis and microscopic polyangiitis [1]. The authors reported that PR3-ANCA subsets reactive with epitopes accessible on recombinant pro-PR3 are better correlated with disease activity than are subsets reactive with epitopes accessible only on recombinant mature PR3. Proteinase-3 and myeloperoxidase (MPO) are the two main target antigens of ANCA in systemic vasculitides [2-4]. Both proteins are located in the azurophil granules of neutrophils.

The aim of the current study was to assess the antigen specificity of MPO-ANCA by testing MPO-ANCA-positive sera in the standardized ANCA indirect immunofluorescence (IIF) assay on MPO-deficient neutrophils from three healthy, unrelated donors (CV, DC and JB). These neutrophils lacked the heavy and light subunits of the MPO molecule but contained the pro-MPO precursor protein, as previously described [5]. We now report that some MPO-ANCA-positive sera react with pro-MPO.

MPO deficiency was defined spectrally, enzymatically, and immunochemically. The donor subject DC has been previously described [6]. Azurophil granules from control neutrophils and MPO-deficient neutrophils (CV) were solubilized 1:1 with 4% reduced *Laemmli* sample buffer and electrophoresed into a 10% (w/v) acrylamide gel. Thereafter, separated proteins were transferred to nitrocellulose membranes, and the blots were probed with a rabbit anti-human MPO (Dako, Glostrup, Denmark). The immunoblot analysis of azurophil granules from control neutrophils showed the MPO subunits 59 kDa and 13.5 kDa, as well as the 39-kDa and 24-kDa proteins related to the 59-kDa and the 13.5-kDa proteins, respectively [7]. In addition, a ~50-kDa protein was found that may be a precursor form of the 59-kDa subunit. In contrast, the azurophil granules from the MPO-deficient neutrophils (CV) lacked the mature subunits (59 kDa and 13.5 kDa) and the related proteins (39 kDa and 24 kDa), but contained the pro-MPO form (90 kDa) and the ~50-kDa precursor protein (**Figure 1**).

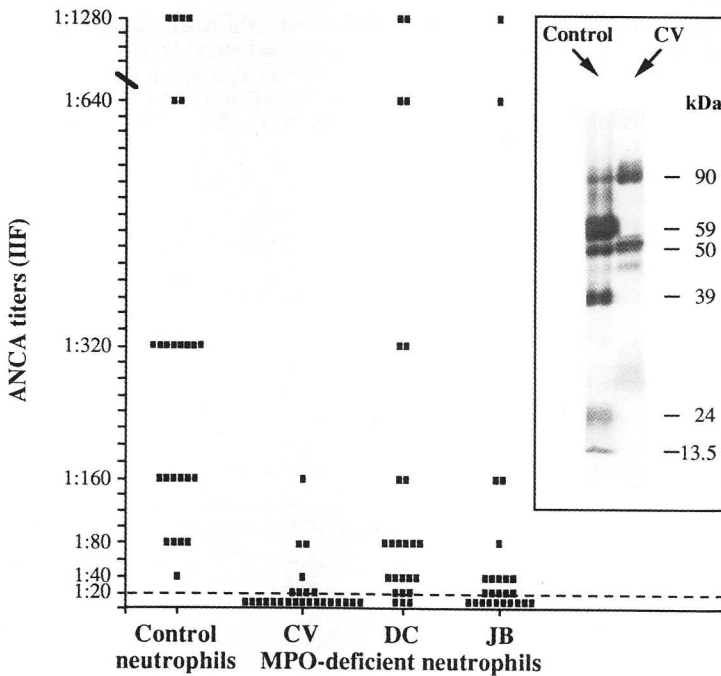
The mutations in the MPO-deficient donor (CV) were characterized as a splice-site mutation at the 3' end of intron 11 (A-2→G) and a deletion of 14 nucleotides from A1555-C1568 in exon 9 of the MPO gene. Subject DC is a compound heterozygote for the R569W missense mutation in exon 10 causing MPO deficiency [8]. Subject JB has a normal exon 10, but the genetic basis for the MPO deficiency has not been defined.

Determination of ANCA by the standardized IIF assay [9] on control *versus* MPO-deficient neutrophils from CV, DC and JB was performed with 25 sera of high MPO-ANCA reactivity from patients with microscopic polyangiitis [10]. The 25 MPO-ANCA sera were perinuclear ANCA-positive in the IIF assay on control neutrophils. MPO-ANCA were detected by standardized enzyme-linked immunosorbent assay using human purified MPO from Calbiochem (La Jolla, CA, USA). When the fluorescence was positive (1:20 dilution), serum titration was carried out in two-fold dilutions to 1:1,280.

IIF titers of these 25 MPO-ANCA sera on control *versus* MPO-deficient neutrophils are shown in **Figure 1**. ANCA were positive in 32%, 88%, and 60% of sera tested on neutrophils from

subjects CV, DC, and JB, respectively. These sera exhibited either a diffuse cytoplasmic or a perinuclear fluorescence pattern. IIF titers obtained with the MPO-deficient neutrophils were somewhat lower than IIF titers obtained with control neutrophils. Therefore, the sera that react with the MPO-deficient neutrophils might contain not only anti-pro-MPO but also anti-mature MPO. Three of 8, 12 of 22, and 6 of 15 ANCA-positive sera with MPO-deficient neutrophils from subjects CV, DC, and JB, respectively, displayed similar IIF titers as with control neutrophils. This suggests that these sera contain mainly reactivity with pro-MPO.

In conclusion, the polyclonality of MPO-ANCA described in systemic vasculitides [11] may be relevant to the disparity of the ANCA-IIF results on these MPO-deficient neutrophils. The pro-MPO, which resides in the endoplasmic reticulum, is hereby described for the first time as a potential target antigen for MPO-ANCA-positive sera. It is conceivable that some MPO-ANCA gain access to pro-MPO *in vivo*, as was recently suggested for PR3-ANCA toward pro-PR3, for instance at times of accelerated neutrophil apoptosis, *i.e.*, during active inflammation [1].



**Figure 1.** Titers of anti-neutrophil cytoplasm autoantibodies (ANCA) by the standardized indirect immunofluorescence (IIF) assay on control neutrophils *versus* myeloperoxidase (MPO)-deficient neutrophils from CV, DC and JB, performed with 25 MPO-ANCA-positive sera. Boxed area shows immunoblot analysis of azurophil granules from control neutrophils *versus* MPO-deficient neutrophils from subject CV.

## REFERENCES

1. Russell KA, Fass DN, Specks U. Antineutrophil cytoplasmic antibodies reacting with the pro form of proteinase 3 and disease activity in patients with Wegener's granulomatosis and microscopic polyangiitis. *Arthritis Rheum* 2001, 44:463-8.
2. Lüdemann J, Utecht B, Gross WL. Anti-neutrophil cytoplasm antibodies in Wegener's granulomatosis recognize an elastolytic enzyme. *J Exp Med* 1990, 171:357-62.
3. Jenne DE, Tschopp J, Lüdemann J, Utecht B, Gross WL. Wegener's autoantigen decoded [letter]. *Nature* 1990, 346:520.
4. Falk RJ, Jennette JC. Anti-neutrophil cytoplasmic autoantibodies with specificity for myeloperoxidase in patients with systemic vasculitis and idiopathic necrotizing and crescentic glomerulonephritis. *N Engl J Med* 1988, 318:1651-7.
5. Nauseef WM, Root RK, Malech HL. Biochemical and immunologic analysis of hereditary myeloperoxidase deficiency. *J Clin Invest* 1983, 71:1297-307.
6. Nauseef WM, Brigham S, Cogley M. Hereditary myeloperoxidase deficiency due to a missense mutation of arginine 569 to tryptophan. *J Biol Chem* 1994, 269:1212-6.
7. Nauseef WM, Malech HL. Analysis of the peptide subunits of human neutrophil myeloperoxidase. *Blood* 1986, 67:1504-7.
8. Nauseef WM, Cogley M, Bock S, Petrides PE. Pattern of inheritance in hereditary myeloperoxidase deficiency associated with the R569W missense mutation. *J Leukoc Biol* 1998, 63:264-9.
9. Wiik A. Delineation of a standard procedure for immunofluorescence detection of ANCA. *APMIS* 1989, 6 (Suppl 1):12-3.
10. Jennette JC, Falk RJ, Andrassy K, Bacon PA, Churg J, Gross WL, et al. Nomenclature of systemic vasculitides: Proposal of an international consensus conference. *Arthritis Rheum* 1994, 37:187-92.
11. Tomizawa K, Mine E, Fujii A, Ohashi YY, Yamagoe S, Hashimoto Y, et al. A panel set for epitope analysis of myeloperoxidase (MPO)-specific antineutrophil cytoplasmic antibody MPO-ANCA using recombinant hexamer histidine-tagged MPO deletion mutants. *J Clin Immunol* 1998, 18:142-52.

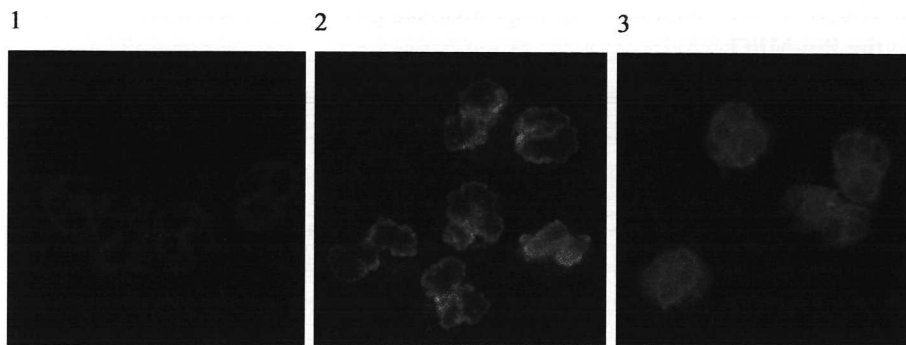
**ADDENDUM: Assessment of Myeloperoxidase (MPO)-ANCA-Positive Sera Reacting with the Pro-MPO****ANCA patterns with myeloperoxidase-deficient neutrophils**

MPO-ANCA-positive sera were obtained from three selected patients (1, 2, and 3) among the aforementioned 25 patients with microscopic polyangiitis, and were tested by the standardized IIF assay on MPO-deficient neutrophils from donor CV (**Figure 2**). The serum from patient 1 gave a negative ANCA pattern, and thus does not react with pro-MPO. In contrast, the two other sera from patients 2 and 3, gave a perinuclear ANCA (P-ANCA) pattern and an atypical ANCA pattern, respectively. The atypical pattern exhibits some weak perinuclear fluorescence and a diffuse cytoplasmic fluorescence without any granular fluorescence or staining of the central part of the cells. The sera did not react with PR3, azurocidin, bactericidal/permeability-increasing protein, cathepsin G, lactoferrin, lysozyme or elastase (not shown). Hence, the two sera from patients 1 and 2 might react with pro-MPO.

MPO-ANCA antibodies normally produce a P-ANCA pattern on ethanol-fixed neutrophils with the IIF assay [1]. Baslund *et al.* have described sera from some patients that are MPO-positive in ELISA but produce a cytoplasmic pattern, which is normally seen in conjunction with antibodies to PR3 [2]. These authors concluded that not all MPO relocates after ethanol fixation. The current study suggests that the diffuse cytoplasmic pattern observed with some MPO-ANCA-positive sera may be due to reactivity with pro-MPO, which resides in the endoplasmic reticulum.

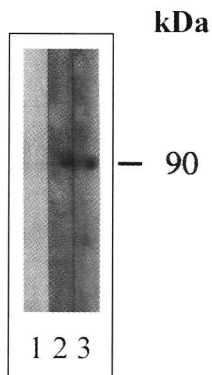
**Immunoblot analysis of myeloperoxidase-related peptides**

We assessed whether MPO-ANCA-positive sera truly react with the pro-MPO protein, by means of immunoblotting (**Figure 3**). We purified immunoglobulin G (IgG) from sera 1, 2, and 3 with MPO-ANCA by passage over HiTrap protein-G sepharose (Amersham Biosciences, Piscataway, NJ, USA). Purity of the IgG preparations as determined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis was always greater than 95%. Azurophil granules from MPO-deficient neutrophils from donor CV were solubilized 1:1 with 4% reduced *Laemmli* sample buffer and electrophoresed onto a 10% (w/v) acrylamide gel. Thereafter, separated proteins were transferred to nitrocellulose membranes, and the blots were probed with purified IgG from the three MPO-ANCA-positive sera. The immunoblot in **Figure 3** shows no band in lane 1, but it does show a reaction in lanes 2 and 3 at 90 kDa, the molecular mass of pro-MPO. Hence, we directly prove that MPO-ANCA-positive sera from patients 2 and 3, which gave a P-ANCA and an atypical ANCA pattern by the IIF assay on MPO-deficient neutrophils, do react with the pro-MPO protein.



**Figure 2.** ANCA patterns of myeloperoxidase (MPO)-ANCA-positive sera from three different patients with microscopic polyangiitis tested with the indirect immunofluorescence assay on MPO-deficient neutrophils from donor CV.

(1) Negative ANCA pattern, (2) P-ANCA pattern, and (3) an atypical ANCA pattern.



**Figure 3.** Immunoblot analysis of myeloperoxidase (MPO)-related peptides from MPO-deficient neutrophils (donor CV) probed with purified immunoglobulin G from the three MPO-ANCA-positive sera 1, 2, and 3.

#### ACKNOWLEDGMENTS

The authors gratefully thank the MPO-deficient donors for the donation of their blood and Daniel Dehay for his technical assistance.

**REFERENCES**

1. Falk RJ, Jennette JC. Anti-neutrophil cytoplasmic autoantibodies with specificity for myeloperoxidase in patients with systemic vasculitis and idiopathic necrotizing and crescentic. *N Engl J Med* 1988, 318:1651 -7.
2. Segelmark M, Baslund B, Wieslander J. Some patients with anti-myeloperoxidase autoantibodies have a C-ANCA pattern. *Clin Exp Immunol* 1994, 96:458-65.



