



University of Groningen

Artificial Intelligence to Guide Empirical Antimicrobial Therapy-Ready for Prime Time?

van der Werf, Tjip S.

Published in: **Clinical Infectious Diseases**

DOI: 10.1093/cid/ciaa1585

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2021

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): van der Werf, T. S. (2021). Artificial Intelligence to Guide Empirical Antimicrobial Therapy-Ready for Prime Time? Clinical Infectious Diseases, 72(11), E856-E858. https://doi.org/10.1093/cid/ciaa1585

Copyright Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.



Artificial Intelligence to Guide Empirical Antimicrobial Therapy–Ready for Prime Time?

Tjip S. van der Werf

Department of Internal Medicine, Division of Infectious Diseases, University Medical Center Groningen University of Groningen, Groningen, The Netherlands

(See the Major Article by Lewin-Epstein et al on pages e848-55.)

Nosocomial infections by drug-resistant gram-negative bacteria and Staphylococcus aureus—especially methicillin-resistant S. aureus (MRSA)-impose a huge public health threat [1]. Mortality increases if initial antimicrobial therapy is inappropriate [2]. Doctors are aware of this daunting perspective, and fueled by their fear to leave such organisms uncovered, they often feel seduced to prescribe excessive, and sometimes, even inappropriate antimicrobials [3]. The ensuing cumulative antimicrobial pressure has long been recognized as the main driver of drug resistance [4, 5]. A policy to use reserve antimicrobials in a cycling pattern, rather than in a random, mixed pattern, does not help to reduce bacterial resistance in the high-risk environment of Intensive Care Units (ICU) [6]. In Dutch ICUs, oral and intestinal nonresorbable antimicrobials-colistin, tobramycin and amphotericin-combined with systemic cefotaxime for 4 consecutive days-resulted in a small but significant survival advantage [7]. However, in ICUs with a high pressure of resistant bacteria, selective digestive decontamination did not reduce blood stream infections caused by these bacteria, compared to standard care, chlorhexidine mouth and skin treatment, or oral selective decontamination [8]. Education on the risks of antimicrobial pressure, with instructions on updated guidelines for the treatment and prevention of nosocomial infections do not invariably address the emotional component of prescribers.

"Antimicrobial Stewardship," a term that has been used for more than 2 decades now [3], has been designed to address both cognitive, or intellectual, and emotional factors, that make prescribers change their behavior [9]. Antibiotic Stewardship Programs (APS) that included a direct feedback to prescribing clinicians, with information on culture and susceptibility results, epidemiological information, and updated guideline information, seem to work best. Two days after starting empirical antimicrobial treatment, by the time culture and susceptibility testing are available, a team of hospital pharmacists, clinical microbiologists, and infectious disease specialists consult with prescribers [10].

APS have been shown to reduce antimicrobial use, both in terms of reducing duration and choice of reserve antimicrobials [11]. These programs appear safe and cost-effective [12], and importantly, help to reduce the burden of high-risk pathogens, including resistant (extended spectrum beta-lactamase, ESBL) gram-negative Enterobacteriaceae, and MRSA [13]. These programs might be supplemented by acute-phase assessments, (eg, procalcitonin- or C-reactive protein-driven decisions to stop antimicrobial therapy if response to therapy seems favorable). Using an algorithm with procalcitonin resulted in a significant reduction of treatment duration by 2 days in one multicenter trial in ICU patients [14].

In one study among mechanically ventilated ICU patients with suspected ventilator-associated pneumonia (VAP), undergoing bronchoscopic bronchoalveolar lavage (BAL), researchers hypothesized that IL-1ß and IL-8 in BAL might be a good predictor of VAP; they advised stopping empirical antimicrobial treatment if biomarker levels were below a predefined threshold. This approach did not result in earlier discontinuation of antimicrobial treatment in the intervention group [15]; the authors suspected, that emotional factors driving antimicrobial use could not be addressed by this algorithm. The advice given was perhaps smart and safe, but prescribing behavior did not change; indeed, stopping a successful antimicrobial regimen ("never change a winning team") appears difficult. APS surely is an important approach to reduce antimicrobial pressure, and thereby, to fight ever-increasing antimicrobial resistance. Any time soon, we will be running out of antimicrobial options, with no novel antimicrobial products in the pipeline to salvage this problem [16]. A way to safely refrain from using reserve drugs in empirical antimicrobial treatment for our vulnerable patients is to use surveillance culture results in individual patients, collected earlier during hospital admission,

Received 6 October 2020; editorial decision 9 October 2020; published online 18 October 2020.

Correspondence: Tjip S van der Werf, Department of Internal Medicine, Division of Infectious Diseases, PO box 30.001, 9700 RB Groningen, The Netherlands (t.s.van.der. werf@umcq.nl).

Clinical Infectious Diseases[®] 2021;72(11):e856–8 © The Author(s) 2020. Published by Oxford University Press for the Infectious Diseases Society of America. All rights reserved. For permissions, e-mail: journals.permissions@oup.com. DOI: 10.1093/cid/ciaa1585

or general information on surveillance data. Using this information, an educated guess of the offending micro-organism for the current febrile episode can be made, to select empirical antimicrobials [17-19]. Could we improve the quality of advice on empirical antimicrobial treatment using machine learning with artificial intelligence (AI) software? Computer-aided pattern recognition has become an integral part of our everyday life. Indeed, AI is involved in prioritizing our likes by analyzing our pattern of internet searching using search engines. Even when I check my smart phone, it opens by recognizing my face, all the result of AI software, that recognizes my face under different angles, even with light only produced by the screen, using machine learning that requires only seconds. AI was perhaps first introduced by space scientists [20], but has become an integral part of our everyday life. AI has entered medicine over 3 decades ago [21], and has impacted on a vast array of diagnostic and therapeutic decisions [22].

In this issue of the Journal, Ohad Lewin-Epstein et al. report a machine-learning model to predict antimicrobial resistance to 5 commonly used antimicrobial agents-Ceftazidime, Gentamicin, Imipenemcilastin, Ofloxacin, and Cotrimoxazole [23]. To predict the presence of resistant bacteria causing infection to any of these drugs, they used a large database of culture and susceptibility testing from thousands of electronic in-patient records between May, 2013, and December, 2015, in the Rabin Medical Center in Israel. A large part of the data set was used to train the model; they next tested the model in a separate batch of medical records. Should the prediction models be perfect, the area under the receiver-operator curve (AUC) would be equal to 1. Unfortunately, the precision of their prediction model was less than what we need in the clinic; though better than usual care, AUC was only around 0.7. Using information of the typing of the offending organism, the AUC increased to 0.8, but using phenotypic culture-based data, this would result in a considerable delay to start effective

treatment. The authors argue that this delay might be overcome using molecular tools to identify the offending organism within hours, but this was not part of their work flow. The study followed the code of conduct to develop this machine learning tool, with large datasets to train, and to validate the model; what should happen next, is testing it prospectively. Could the model be flawed by the time elapsed between the development and the final prospective testing? Should perhaps more data be entered into the model, including comorbid conditions, surveillance data, epidemiological data, genomic, metabolomic, or microbiome data? Is the model, developed for this single medical center, applicable to other locales and settings? Although many questions remain unanswered, this study represents a hallmark in this novel approach, and deserves to be followed by similar approaches with even more sophistication. Perhaps, other technologies might be added, such as breath analysis of volatile organic compounds using machine learning and neural networks [24], an approach that eventually might be developed as a point-of-care test, as has been explored to detect tuberculosis [25, 26]. This work shows, that machine learning-although not yet ready for prime time-will one day help guide initial antimicrobial selection in nosocomial infections. One problem that needs to be addressed, is that clinical decisions are only partly driven by cognition-emotions are equally important. AI-driven machine learning may one day provide overwhelming scientific, cognitive evidence, but prescribers should feel comfortable about it, or else, they would not follow the evidence-driven advice. To incorporate these novel tools, interactive training is required to overcome these emotional barriers, to improve antimicrobial prescription behavior.

Notes

Author Contributions. The author has met the following 4 requirements: (a) significant contributions to the conception and design, acquisition of data, or analysis and interpretation of data; (b) drafting or revising the article for intellectual content; (c) final approval of the published

article; and (d) agreement to be accountable for all aspects of the article thus ensuring that questions related to the accuracy or integrity of any part of the article are appropriately investigated and resolved.

Potential conflicts of Interest. The author: No reported conflicts of interest. The author has submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest.

References

- Anderson M, Schulze K, Cassini A, Plachouras D, Mossialos E. A governance framework for development and assessment of national action plans on antimicrobial resistance. Lancet Infect Dis 2019; 19:e371–84.
- Raman G, Avendano E, Berger S, Menon V. Appropriate initial antibiotic therapy in hospitalized patients with gram-negative infections: systematic review and meta-analysis. BMC Infect Dis 2015; 15:395.
- Gould IM. Stewardship of antibiotic use and resistance surveillance: the international scene. J Hosp Infect 1999; 43(Suppl):S253–60.
- Goldmann DA, Weinstein RA, Wenzel RP, et al. Strategies to prevent and control the emergence and spread of antimicrobial-resistant microorganisms in hospitals. A challenge to hospital leadership. JAMA 1996; 275:234–40.
- Bell BG, Schellevis F, Stobberingh E, Goossens H, Pringle M. A systematic review and meta-analysis of the effects of antibiotic consumption on antibiotic resistance. BMC Infect Dis 2014; 14:13.
- van Duijn PJ, Verbrugghe W, Jorens PG, et al.; SATURN consortium. The effects of antibiotic cycling and mixing on antibiotic resistance in intensive care units: a cluster-randomised crossover trial. Lancet Infect Dis 2018; 18:401–9.
- de Smet AMGA, Kluytmans JAJW, Cooper BS, et al. Decontamination of the digestive tract and oropharynx in ICU patients. N Engl J Med 2009; 360:20–31.
- Wittekamp BH, Plantinga NL, Cooper BS, et al. Decontamination strategies and bloodstream infections with antibiotic-resistant microorganisms in ventilated patients: a randomized clinical trial. JAMA 2018; 320:2087–98.
- Schuts EC, Hulscher MEJL, Mouton JW, et al. Current evidence on hospital antimicrobial stewardship objectives: a systematic review and metaanalysis. Lancet Infect Dis 2016; 16:847–56.
- Dellit TH, Owens RC, McGowan JE Jr, et al; Infectious Diseases Society of America; Society for Healthcare Epidemiology of America. Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America guidelines for developing an institutional program to enhance antimicrobial stewardship. Clin Infect Dis 2007; 44:159–77.
- Honda H, Murakami S, Tagashira Y, et al. Efficacy of a postprescription review of broad-spectrum antimicrobial agents with feedback: a 4-year experience of antimicrobial stewardship at a tertiary care center. Open Forum Infect Dis 2018; 5:ofy314.
- Karanika S, Paudel S, Grigoras C, Kalbasi A, Mylonakis E. Systematic review and meta-analysis of clinical and economic outcomes from the implementation of hospital-based antimicrobial stewardship programs. Antimicrob Agents Chemother 2016; 60:4840–52.
- Baur D, Gladstone BP, Burkert F, et al. Effect of antibiotic stewardship on the incidence of infection and colonisation with antibiotic-resistant bacteria

and Clostridium difficile infection: a systematic review and meta-analysis. Lancet Infect Dis **2017**; 17:990–1001.

- 14. de Jong E, van Oers JA, Beishuizen A, et al. Efficacy and safety of procalcitonin guidance in reducing the duration of antibiotic treatment in critically ill patients: a randomised, controlled, open-label trial. Lancet Infect Dis 2016; 16:819–27.
- Hellyer TP, McAuley DF, Walsh TS, et al. Biomarkerguided antibiotic stewardship in suspected ventilator-associated pneumonia (VAPrapid2): a randomised controlled trial and process evaluation. Lancet Respir Med 2020; 8:182–91.
- Theuretzbacher U, Gottwalt S, Beyer P, et al. Analysis of the clinical antibacterial and antituberculosis pipeline. Lancet Infect Dis 2019; 19:e40–50.
- Depuydt P, Benoit D, Vogelaers D, et al. Outcome in bacteremia associated with nosocomial pneumonia and the impact of pathogen prediction by tracheal

surveillance cultures. Intensive Care Med 2006; 32:1773-81.

- Baba H, Nimmo GR, Allworth AM, et al. The role of surveillance cultures in the prediction of susceptibility patterns of Gram-negative bacilli in the intensive care unit. Eur J Clin Microbiol Infect Dis 2011; 30:739–44.
- Luna CM, Bledel I, Raimondi A. The role of surveillance cultures in guiding ventilator-associated pneumonia therapy. Curr Opin Infect Dis 2014; 27:184–93.
- Fleuren LM, Thoral P, Shillan D, Ercole A, Elbers PWG; Right Data Right Now Collaborators. Machine learning in intensive care medicine: ready for take-off? Intensive Care Med 2020; 46:1486–8.
- Schwartz WB, Patil RS, Szolovits P. Artificial intelligence in medicine. Where do we stand? N Engl J Med 1987; 316:685–8.

- 22. Rajkomar A, Dean J, Kohane I. Machine learning in medicine. N Engl J Med **2019**; 380:1347–58.
- Lewin-Epstein O, Baruch S, Hadany L, Stein GY, Obolski U. Predicting antibiotic resistance in hospitalized patients by applying machine learning to electronic medical records. Clin Infect Dis 2020.
- Palma SICJ, Traguedo AP, Porteira AR, Frias MJ, Gamboa H, Roque ACA. Machine learning for the meta-analyses of microbial pathogens' volatile signatures. Sci Rep 2018; 8:3360.
- Saktiawati AMI, Stienstra Y, Subronto YW, et . Sensitivity and specificity of an electronic nose in diagnosing pulmonary tuberculosis among patients with suspected tuberculosis. PLoS One 2019; 14:e0217963.
- 26. Saktiawati AMI, Putera DD, Setyawan A, Mahendradhata Y, van der Werf TS. Diagnosis of tuberculosis through breath test: a systematic review. EBioMedicine 2019; 46:202–14.