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Polygenic Risk Score Prediction for Endometriosis

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Endometriosis is a major health care challenge because many young women with endometriosis go undetected for an extended period, which may lead to pain sensitization. Clinical tools to better identify candidates for laparoscopy-guided diagnosis are urgently needed. Since endometriosis has a strong genetic component, there is a growing interest in using genetics as part of the clinical risk assessment. The aim of this work was to investigate the discriminative ability of a polygenic risk score (PRS) for endometriosis using three different cohorts: surgically confirmed cases from the Western Danish endometriosis referral Center (249 cases, 348 controls), cases identified from the Danish Twin Registry (DTR) based on ICD-10 codes from the National Patient Registry (140 cases, 316 controls), and replication analysis in the UK Biobank (2,967 cases, 256,222 controls). Patients with adenomyosis from the DTR (25 cases) and from the UK Biobank (1,883 cases) were included for comparison. The PRS was derived from 14 genetic variants identified in a published genome-wide association study with more than 17,000 cases. The PRS was associated with endometriosis in surgically confirmed cases [odds ratio (OR) = 1.59, $p = 2.57 \times 10^{-7}$] and in cases from the DTR biobank (OR = 1.50, $p = 0.0001$). Combining the two Danish cohorts, each standard deviation increase in PRS was associated with endometriosis (OR = 1.57, $p = 2.5 \times 10^{-11}$), as well as the major subtypes of endometriosis; ovarian (OR = 1.72, $p = 6.7 \times 10^{-5}$), infiltrating (OR = 1.66, $p = 2.7 \times 10^{-9}$), and peritoneal (OR = 1.51, $p = 2.6 \times 10^{-3}$). These findings were replicated in the UK Biobank with a much larger sample size (OR = 1.28, $p < 2.2 \times 10^{-16}$). The PRS was not associated with adenomyosis, suggesting that adenomyosis is not driven by the same genetic risk variants as endometriosis. Our results suggest that a PRS captures an increased risk of all types of endometriosis rather than an increased risk for endometriosis in specific locations. Although the discriminative accuracy is not yet sufficient as a stand-alone clinical utility, our data demonstrate that genetics risk variants

in form of a simple PRS may add significant new discriminatory value. We suggest that an endometriosis PRS in combination with classical clinical risk factors and symptoms could be an important step in developing an urgently needed endometriosis risk stratification tool.

Keywords: endometriosis, genetic association, ICD-10, polygenic risk score, subtypes

INTRODUCTION

Endometriosis affects 10% of fertile women and represents a major health care challenge (1). Early detection would be of value to avoid sensitization and development of chronic pelvic pain (2, 3). Since laparoscopy is needed for diagnosis, and symptoms overlap with common problems like primary dysmenorrhea (1), a high number of young girls with endometriosis today go undetected. Thus, there is a great need to develop clinical decision tools that will help the medical doctors to better identify cases where laparoscopy is warranted and/or medical treatment can be offered. Despite promising results with different types of circulating molecules (1, 4, 5), biomarkers have so far been of limited value because of modest sample sizes and lack of replication (6).

In recent years, there has been an increasing interest in, including genomic information, risk models. This development is driven by the increasing number of genetic risk variants for common complex diseases being identified through large-scale genome-wide association studies (GWAS) (7, 8), including also studies of endometriosis; a large study investigating association with common variants (9), a smaller study with rare coding variants (10), and a recent study in the Biobank Japan Project (11). Since individual risk loci identified in a GWAS usually only exercise small effect sizes (12–14), aggregating the effects of multiple genetic risk variants into a single score is an area of great interest. One of the most widely used scores is calculated as the sum of risk alleles weighted by single nucleotide polymorphism (SNP) effect sizes derived from GWAS, i.e., a polygenic risk score (PRS) (15, 16). Its correlation with genetic liability has led to widespread use of PRSs in biomedical research (17–19).

The predictive power of a PRS is most optimally studied in clinically well-defined cases, but to increase sample size, studies are also ongoing in large biobanks linked to health registries using phenotypes from for example the International Classification of Diseases (ICD) (20). Some of these efforts include, among others, BioBank Japan (21), FinnGen (22), and the UK Biobank project (23). Although ICD codes are convenient to use, the codes themselves can be suboptimal in defining diseases (24). In Denmark, the Danish National Patient Register (DNPR), containing ICD codes, is one of the world's oldest nationwide hospital registries and is used extensively for research (25), but no systematic studies validating endometriosis diagnoses in this register has been carried out.

The aim of this study was to evaluate the performance of a simple 14-SNP PRS to predict endometriosis in three different settings, namely surgically confirmed endometriosis cases (clinical cohort), cases with endometriosis defined from the Danish Twin Registry biobank (DTR cohort), and replication in

the UK Biobank (UKB). The 14 SNPs represent top hits from the largest GWAS metaanalysis of endometriosis published to date (9). The discriminative ability was tested for endometriosis, endometriosis subtypes, and adenomyosis for comparison. Adenomyosis was included because it shares features with endometriosis but is more and more considered its own disease entity (26). The goal was to uncover if genetics could be part of a risk predictor in the future.

MATERIALS AND METHODS

Clinical Cohort

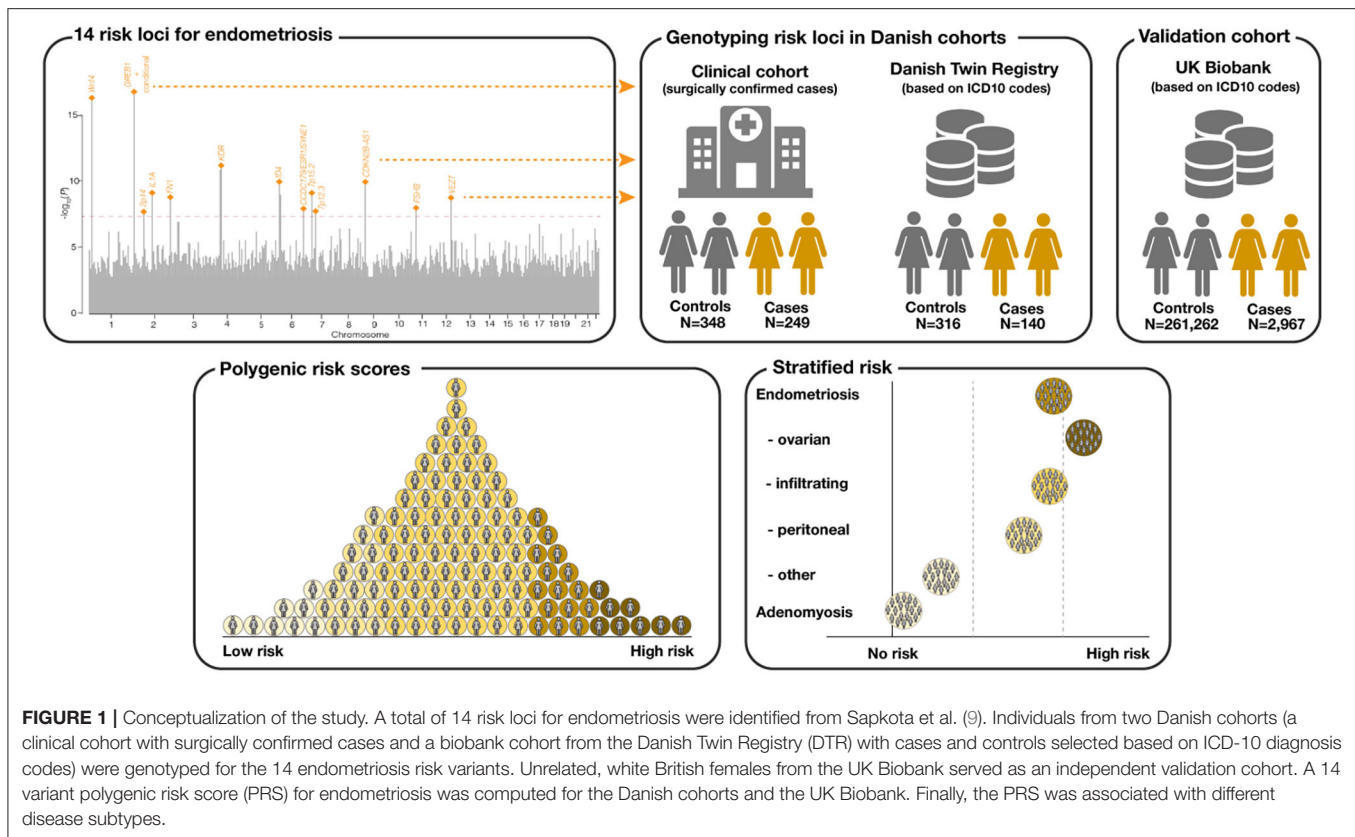
Patients with surgically confirmed endometriosis ($n = 249$) were enrolled from Department of Gynecology and Obstetrics, Aarhus University Hospital, Denmark. All cases were confirmed by laparoscopy, histological examination, and had undergone surgery for endometriosis. The ICD-10 codes for all surgically confirmed cases were extracted from medical records. All had ASRM stages II–IV. The control group consisted of Danish blood donors from the Aalborg University Hospital blood bank, Aalborg, Denmark, and were age-matched women without a diagnose of N80 ($n = 348$). Control samples were anonymized prior to genotyping. The study was approved by The Central Denmark Region Committees on Health Research Ethics (ID 25.736).

Danish Twin Registry (DTR) Biobank Cohort

Participants from the Danish Twin Registry were drawn from the Danish Twin Registry Infrastructure Study carried out from 2008 to 2011 (27). Unrelated women with an endometriosis diagnosis (N80.1–N80.9) in the Danish National Patient Registry (DNPR) (28) were selected as cases ($n = 140$). Four of the women had a twin sister with endometriosis; these sisters were not included. In addition, unrelated women with a diagnosis of adenomyosis (N80.0) were included for comparison ($n = 25$). Age-matched unrelated women without an N80 diagnose were selected as controls ($n = 316$). Individuals from the Danish Twin Registry were followed in DNPR until March 2014. The study was approved by the Regional Scientific Ethical Committees for Southern Denmark (project ID S-20170120) and registered at Research & Innovation Organization at University of Southern Denmark (registration number 10.584), who approves all scientific projects for University of Southern Denmark according to the Data Protection Regulation.

UK Biobank

For replication analysis, the 14 SNPs (see section “Assay Design”) were identified from imputed data (best guess genotypes) in the



UK Biobank (23). Among unrelated British Caucasian female individuals, cases with endometriosis (N80.1–N80.9) ($n = 2,967$), adenomyosis (N80.0, no N80.1–N80.9) ($n = 1,883$) and controls (no N80 diagnosis, and no self-reported endometriosis) ($n = 256,222$) were identified. ICD-10 codes were identified from main and secondary in-hospital ICD-10 records (data field 41270), while self-reported endometriosis was identified from data field 20002.

Endometriosis Subtypes From ICD-10 Diagnoses

Four major subgroups of endometriosis were constructed from the ICD-10 N80 diagnoses (Table 1). Cases with multiple different N80 sub-diagnoses were assigned to the most severe category with severity ranked from severe to mild in the following order: N80.5 (endometriosis of intestine), N80.4 (endometriosis of rectovaginal septum and vagina), N80.1 (endometriosis of ovary), N80.3 (endometriosis of pelvic peritoneum), N80.2 (endometriosis of fallopian tube), N80.9 (unspecified), N80.8 (endometriosis of thorax), N80.6 (endometriosis of cutaneous scar). Subjects with adenomyosis only (N80.0) were treated as a separate and distinct category.

Assay Design

The 14 genome-wide significant lead SNPs from an endometriosis GWAS comprising 17,045 endometriosis cases and 191,596 controls (9) were initially included for genotyping. One lead SNPs (rs760794) failed assay design, and we instead included rs77294520 which was region-wide associated

after conditioning on the index SNP in the *GREB1* locus (Supplementary Table 1). The 14-SNP multiplex genotyping assay was designed as a SNP type assay using the online Fluidigm D3 Assay tool (Supplementary Table 2).

Genotyping

Genomic DNA was extracted from whole blood using a manual (29) or a semiautomatic (Autopure, Qiagen, Hilden, Germany) salting out method. Genotyping was performed on a Fluidigm Biomark system (Fluidigm Corporation) with the standard SNP genotyping analysis user guide (PN 68000098 Q1), and the genotypes were called using the Fluidigm SNP Genotyping Analysis software, with rigorous manual inspection of all clusters. For quality control, 112 DNA samples were genotyped in duplicate in different runs yielding full (100%) consistency in genotype calls across all duplicates. For each genotyping, chip allele frequencies were compared to the CEU population to exclude allele swop. No significant deviations from Hardy-Weinberg equilibrium were found for any of the SNPs (p -value threshold 0.05). Missing genotypes (46 out of in total 15,092 genotypes, i.e., <0.5%) were imputed to the average of the clinical or the biobank cohort sample sets.

Polygenic Risk Score and Predictive Performance

The 14 SNPs were tested individually for association with endometriosis (N80.1–80.9) using logistic regression, with no covariates and including age-matched women as controls. A PRS for endometriosis was calculated as the weighted sum of risk

alleles; $PRS = \sum_{i=1}^{14} X_i \hat{b}_i$, where X_i is the i -th column of the genotype matrix containing allelic counts of the risk allele (0, 1, or 2) and \hat{b}_i is the estimated marker effect for the i -th variant; here the log odds ratio (OR), which were obtained from (9). The PRS was standardized within each of the three cohorts. For the combined Danish analysis (clinical and DTR biobank cohorts), the PRS was standardized to mean zero and variance 1 in the combined cohort.

Mean PRS between groups were compared using Student's t -test, and the discriminative ability of the PRS was determined using area under the receiver operating curve (AUC) and with Nagelkerke's variances explained (R_{Nag}^2) using the R package qgg (30).

The change in genetic risk with increasing PRS was investigated by binning individuals into 10 groupings according to the decile of the PRS, and the disease prevalence within each bin was determined. For each bin, we furthermore calculated OR using logistic regression with disease status as response variable and decile as predictor with the fifth decile as reference point. The strength of association for the PRS with endometriosis and endometriosis subtypes was estimated by calculating OR per standard deviation increase in standardized PRS.

RESULTS

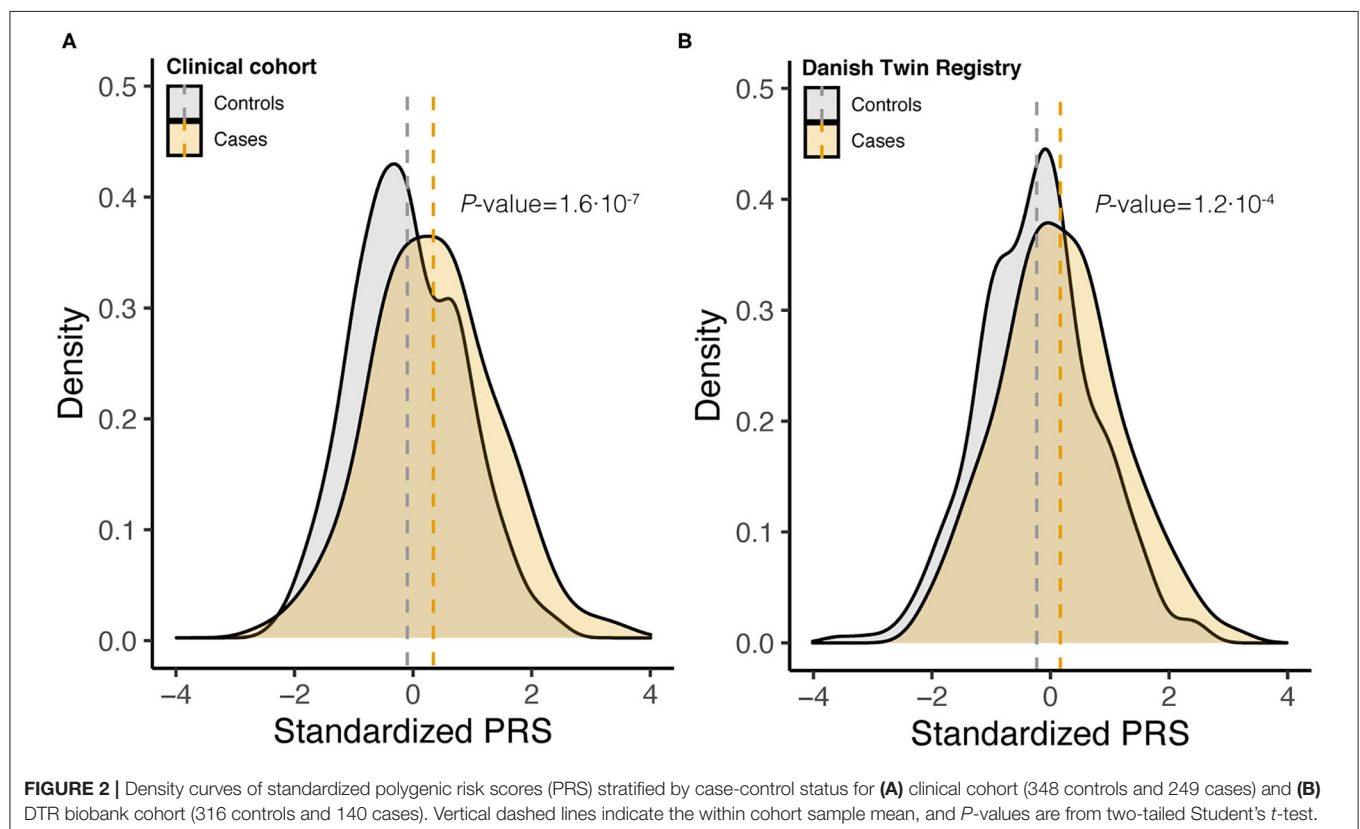
Single Variant Association

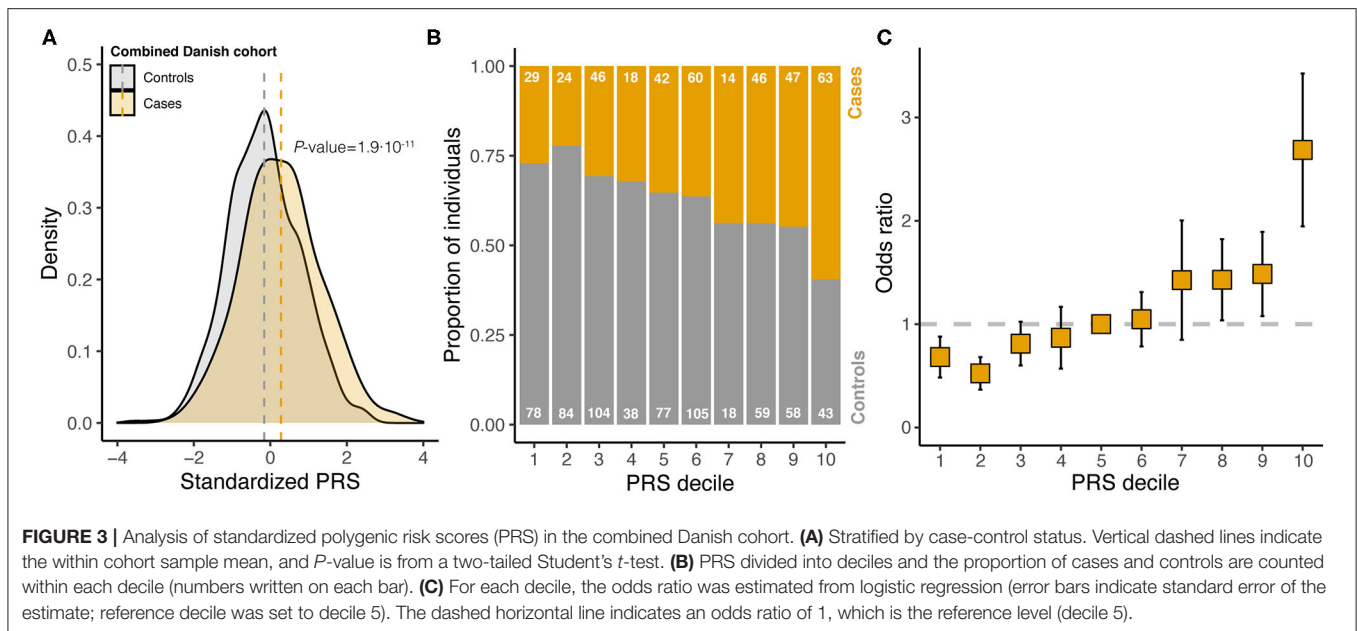
An initial single variant association analysis in the two Danish cohorts, the clinical cohort with surgically confirmed

endometriosis cases and the DTR biobank cohort with endometriosis cases identified from ICD-10 codes, revealed an overall directional consistency in the effect sizes compared with the discovery GWAS (**Supplementary Table 1**), indicating that most of the established risk variants also are risk variants in the Danish population.

PRS Association With Endometriosis

To assess the joint effect of the 14 risk variants, we computed the PRS for endometriosis (**Figure 1**). For both Danish cohorts, the PRS distribution approximated a normal distribution, with the density plot for the cases shifting toward a higher PRS (**Figure 2, Supplementary Table 3**). The average standardized PRS was significantly higher in endometriosis cases compared with controls in both the clinical cohort ($P = 1.6 \times 10^{-7}$) and within the DTR biobank cohort ($P = 1.2 \times 10^{-4}$; **Figure 2**). The PRS significantly predicted endometriosis in surgically confirmed cases (OR = 1.59, $P = 2.57 \times 10^{-7}$, AUC = 0.68) and in cases ascertained from DTR biobank (OR = 1.50, $P = 0.0001$, AUC = 0.61). No significant difference was found in average PRS between cases within the two cohorts, indicating that biobank cases identified using ICD-10 codes and surgically confirmed cases have a comparable genetic burden ($P = 0.15$). In addition, no significant difference in average PRS between control samples in the two cohorts was found ($P = 0.10$). Based on this finding, we combined the clinical and DTR biobank cohorts (combined Danish cohort hereafter) in subsequent analyses.





In the combined Danish cohort, a significantly increased PRS was found for cases (mean 0.28, SD 1.04) compared with controls (mean -0.16 , SD 0.94, $P = 1.9 \cdot 10^{-11}$) (Figure 3A). The discriminative ability of the 14-SNP PRS was $AUC = 0.64$ (Supplementary Table 4). Dividing the PRS into deciles, an increasing proportion of individuals with endometriosis was seen with increasing PRS (Figure 3B), with individuals in the top 10% decile having almost three times higher odds of disease compared with individuals with an average PRS (Figure 3C).

To investigate how PRS possibly varied across endometriosis subtypes, the ICD-10 codes were collapsed into major subtypes: infiltrating, ovarian, peritoneal, and other (Table 1). Adenomyosis was included as a distinct entity. The prevalence of the different subtypes varied largely between the different cohorts (Table 1), e.g., with infiltrating lesions making up a large proportion of samples in the surgically confirmed group, but only a minor proportion in the other cohorts, reflecting that the surgically confirmed cases were collected at the specialized surgical unit in Denmark treating the most severe and complicated forms of endometriosis. A significant predictive ability for the PRS was found for all subtypes, except for “other” (Figure 4, Supplementary Table 4). The best AUC among the endometriosis subtypes was found for ovarian endometriosis ($AUC = 0.64$). These findings were replicated within the UK Biobank, again with best AUC for ovarian endometriosis ($AUC = 0.60$), demonstrating that ovarian endometriosis may have a higher genetic burden. Both in the combined Danish cohort and in the UK Biobank the 14-SNP PRS had no predictive power for adenomyosis (Figure 4, Supplementary Table 4).

DISCUSSION

Endometriosis has a strong genetic component (31–33), and genomic information could provide value if incorporated into a clinical decision support tool to identify women in need for

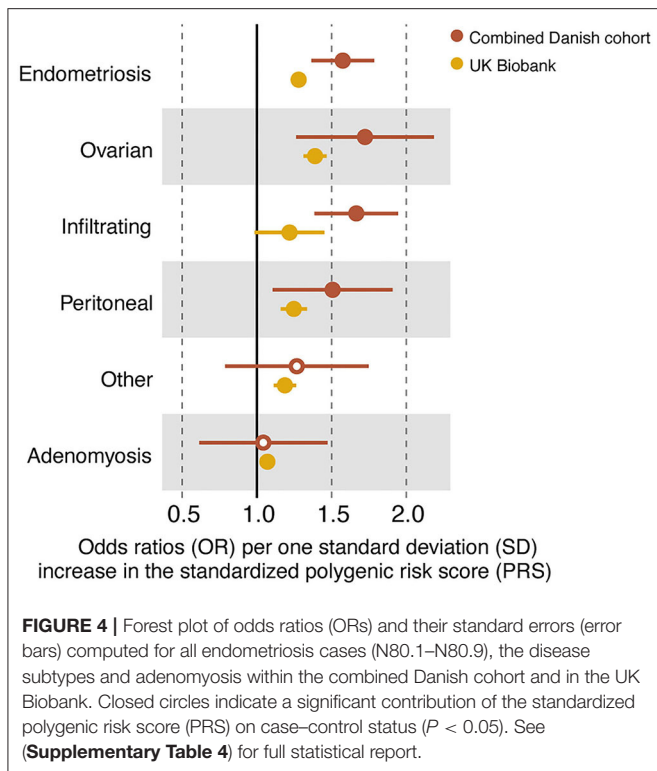
TABLE 1 | Distribution of endometriosis subtypes and adenomyosis subjects within the three cohorts.

Disease subtypes	Cohorts		
	Clinical	DTR	UK Biobank
Endometriosis (N80.1–N80.9)	249 (42%)	140 (29%)	2,967 (1.1%)
- infiltrating (N80.4, N80.5)	205 (34%)	5 (1%)	105 (0.04%)
- ovarian (N80.1)	21 (4%)	54 (11%)	1,158 (0.4%)
- peritoneal (N80.2, N80.3)	16 (3%)	44 (9%)	736 (0.3%)
- other (N80.6, N80.8, N80.9)	7 (1%)	37 (8%)	968 (0.4%)
Adenomyosis (N80.0)	0 (0%)	25 (5%)	1,883 (0.7%)

Percentages in parentheses indicate the case prevalence within each cohort.

laparoscopy. In this study, we constructed a PRS based on 14 established risk variants for endometriosis from the most recent published GWAS of endometriosis (9). We compared the PRS in two Danish case-control cohorts, representing two different and widely used designs, and found a comparable genetic burden between surgically confirmed cases and cases identified through health registries using ICD-10 codes. Since the mean PRS was not significantly different between cases in the two cohorts, our results suggest that ICD-10 codes are a reasonable tool to define cases for large GWAS studies or epidemiological studies of endometriosis.

Previously, it was shown that an increased genetic burden for endometriosis is associated with increasing severity (34, 35). As we did not have the severity score available to us, a formal replication of this finding was not possible. However, we believe our data are in good agreement with the previous finding as our surgically confirmed cases, ascertained at a surgical center specializing in severe and complicated endometriosis, had a slightly higher mean PRS than the remaining cases, although this was not statistically significant.



The finding of an AUC of around 0.64% for endometriosis is in line with other studies of PRS prediction in common complex diseases. Marigorta et al. (36) compared the predictive power of PRS for multiple common complex diseases and found that AUCs generally increase in the beginning when adding more SNPs to the PRS, but that the power of PRS seems to plateau around an AUC of ~60%. Due to the largely overlapping PRS distributions in cases and controls, and the relatively low discriminative accuracy of the PRS, it is not expected that the PRS alone will be useful for diagnostic or population screening purposes. It is, however, expected that genetics will become important for risk stratification in the future, when combined with information provided by other risk factors (37).

We observed in our study a lower effect size of the PRS in the UKB compared with the combined Danish cohort. A similar reduced effect size has been described for other variants in the UKB (38). This is likely because the UKB is enriched for healthy individuals, the so-called “healthy volunteer” selection bias (39), leading to an underestimation of penetrance (38). Despite the lower effect size all findings were replicated, demonstrating that a PRS has predictive abilities even in healthy populations.

Interestingly, we found that patients with adenomyosis had the same average PRS as the controls. This was observed in the combined Danish cohort and replicated in the UK Biobank. Endometriosis and adenomyosis represent different diseases but seem to share some pathophysiological pathways related to the junctional zone (40). Also, patients with severe endometriosis have a higher risk of adenomyosis (41). However, it is unknown if there are shared risk genes between the two conditions. Using the combined information from 14 risk SNPs for endometriosis, we found no discriminative ability for

women with adenomyosis without a diagnosis of endometriosis, supporting the clinical observations that adenomyosis is different from endometriosis (26).

A limitation of this study is that the subjects were not genotyped using SNP array, and therefore we were not able to test if the predictive ability could be improved by including additional SNPs below the genome-wide significance threshold or correct for hidden population structure on a genome-wide scale. As ethnicity was not available to us in the two Danish cohorts, we cannot exclude that these cohorts had a minor proportion of non-European individuals. In previous studies, it has been shown that polygenic scores derived from a European GWAS had lower predictive power in other populations (42). If this is the case, a small proportion of non-European individuals would have resulted in lower discriminative ability in our study making the odds ratios a conservative estimate. As we replicated the results in the UKB using only white-British individuals, we find it unlikely that our results are driven by hidden population structures in the two Danish cohorts. Another limitation is the use of imputed SNPs in the UKB validation set, which introduces the potential for some incorrect genotype calling. Because all SNPs had high info score (above 0.9), we find it unlikely that this has affected the results significantly.

As sample sizes for GWAS grow, more and more genome-wide significant loci are being discovered for endometriosis, which could be incorporated into a genome-wide polygenic score. Also, more sophisticated methods are being developed (and already exist) for calculating polygenic risk scores that include millions of variants below the genome-wide significant level (43) and from correlated traits (44, 45). The scope of our work was to test a simple PRS to provide a conservative estimate on what benefits can be achieved by including genetic variant. Taken together our results suggest that genetic risk variants could be an important component of a composite risk stratification tool aimed at women with symptoms of endometriosis to reduce diagnostic delay.

In summary, we found that a polygenic score composed of as little as 14 SNPs is associated with endometriosis across all anatomical sites. Our study is a first important step to investigate the value of using common genetic variants to predict endometriosis.

DATA AVAILABILITY STATEMENT

The standardized PRS for the combined Danish cohort can be found within **Supplementary Table S5**. Researchers can gain access to the UKB data by applying to www.ukbiobank.ac.uk.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by The Central Denmark Region Committees on Health Research Ethics and The Regional Committees on Health Research Ethics for Southern Denmark. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

MNye and LC planned the study. AF, RS, and LC was responsible for ascertainment of the patients and controls. RS, KK-M, and ST extracted DNA. KK-M and ST genotyped the samples and performed quality control. KK-M, ST, MNyg, CD, and PDR performed the statistical analysis. DR provided advise on the Danish health registries. MNye, MO, and LC supervised the study. All authors interpreted the results. MNye, KK, and PDR drafted the manuscript. All authors critically revised the manuscript, read, and approved the final manuscript.

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REFERENCES

- Zondervan KT, Becker CM, Missmer SA. Endometriosis. *N Engl J Med.* (2020) 382:1244–56. doi: 10.1177/1461444810365020
- Brawn J, Morotti M, Zondervan KT, Becker CM, Vincent K. Central changes associated with chronic pelvic pain and endometriosis. *Hum Reprod Update.* (2014) 20:737–47. doi: 10.1093/humupd/dmu025
- Morotti M, Vincent K, Brawn J, Zondervan KT, Becker CM. Peripheral changes in endometriosis-associated pain. *Hum Reprod Update.* (2014) 20:717–36. doi: 10.1093/humupd/dmu021
- Agrawal S, Tapmeier TT, Rahmioglu N, Kirtley S, Zondervan KT, Becker CM. The miRNA mirage: How close are we to finding a non-invasive diagnostic biomarker in endometriosis? *A systematic review. Int J Mol Sci.* (2018) 19:599. doi: 10.3390/ijms19020599
- Anastasiu CV, Moga MA, Neculau AE, Bălan A, Scărneciu I, Dragomir RM, et al. Biomarkers for the noninvasive diagnosis of endometriosis: state of the art and future perspectives. *Int J Mol Sci.* (2020) 21:1750. doi: 10.3390/ijms21051750
- Kimber-Trojanar Z, Pilszyk A, Niebrzydowska M, Pilszyk Z, Ruszała M, Leszczyńska-Gorzelak B. The potential of non-invasive biomarkers for early diagnosis of asymptomatic patients with endometriosis. *J Clin Med.* (2021) 10:2762. doi: 10.3390/jcm10132762
- Visscher PM, Wray NR, Zhang Q, Sklar P, McCarthy MI, Brown MA, et al. 10 Years of GWAS discovery: biology, function, and translation. *Am J Hum Genet.* (2017) 101:5–22. doi: 10.1016/j.ajhg.2017.06.005
- Buniello A, MacArthur JAL, Cerezo M, Harris LW, Hayhurst J, Malangone C, et al. The NHGRI-EBI GWAS Catalog of published genome-wide association studies, targeted arrays and summary statistics 2019. *Nucleic Acids Res.* (2019) 47:D1005–D1012. doi: 10.1093/nar/gky1120
- Sapkota Y, Steinhorsdottir V, Morris AP, Fassbender A, Rahmioglu N, de Vivo I, et al. Meta-analysis identifies five novel loci associated with endometriosis highlighting key genes involved in hormone metabolism. *Nat Commun.* (2017) 8:15539. doi: 10.1038/ncomms15539
- Sapkota Y, de Vivo I, de Steinhorsdottir V, Fassbender A, Bowdler L, Buring JE, et al. Analysis of potential protein-modifying variants in 9000 endometriosis patients and 150000 controls of European ancestry. *Sci Rep.* (2017) 7:1–11. doi: 10.1038/s41598-017-10440-9
- Masuda T, Low SK, Akiyama M, Hirata M, Ueda Y, Matsuda K, et al. GWAS of five gynecologic diseases and cross-trait analysis in Japanese. *Eur J Hum Genet.* (2020) 28:95–107. doi: 10.1038/s41431-019-0495-1

used in the presented study were obtained from the UKB Resource (project ID 31269).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/frph.2021.793226/full#supplementary-material>

Supplementary Figure 1 | Density curves of standardized polygenic risk scores (PRS) stratified by case-control status for UK Biobank (256,222 controls and 2,967 cases). Vertical dashed lines indicate the within cohort sample mean, and *P*-values are from two-tailed Student's *t*-test.

Supplementary Table 1 | Single SNP association for the 14 SNPs with endometriosis (N80.1-80.9) in Danish clinical and DTR cohorts, within the UK Biobank, and from the Sapkota et al. (9) GWAS.

Supplementary Table 2 | Primer sequences for the Fluidigm genotyping assay.

Supplementary Table 3 | Mean and standard deviation (SD) of the standardized polygenic risk score (PRS) for each Danish cohort, the combined Danish cohort and within UK Biobank.

Supplementary Table 4 | Discriminative power of the standardized polygenic risk score (PRS) within the Danish and UK Biobank for endometriosis (N80.1–N80.9), the disease subtypes, and adenomyosis (N80.0).

Supplementary Table 5 | The standardized PRS for the combined Danish cohort.

- Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, Nyholt DR, et al. Common SNPs explain a large proportion of the heritability for human height. *Nat Genet.* (2010) 42:565–9. doi: 10.1038/ng.608
- Dudbridge F. Power and predictive accuracy of polygenic risk scores. *PLoS Genet.* (2013) 9:e1003348. doi: 10.1371/journal.pgen.1003348
- Dudbridge F. Polygenic epidemiology. *Genet Epidemiol.* (2016) 40:268–72. doi: 10.1002/gepi.21966
- Choi SW, Mak TSH, O'Reilly PF. Tutorial: a guide to performing polygenic risk score analyses. *Nat Protoc.* (2020) 15:2759–72. doi: 10.1038/s41596-020-0353-1
- Lewis CM, Vassos E. Polygenic risk scores: From research tools to clinical instruments. *Genome Med.* (2020) 12:1–11. doi: 10.1186/s13073-020-00742-5
- Khera AV, Chaffin M, Aragam KG, Haas ME, Roselli C, Choi SH, et al. Genome-wide polygenic scores for common diseases identify individuals with risk equivalent to monogenic mutations. *Nat Genet.* (2018) 50:1219–24. doi: 10.1038/s41588-018-0183-z
- Duncan L, Shen H, Gelaye B, Meijns J, Ressler K, Feldman M, et al. Analysis of polygenic risk score usage and performance in diverse human populations. *Nat Commun.* (2019) 10:3328. doi: 10.1038/s41467-019-1112-0
- Richardson TG, Harrison S, Hemani G, Smith GD. An atlas of polygenic risk score associations to highlight putative causal relationships across the human phenome. *eLife.* (2019) 28:e43657. doi: 10.1101/467910
- Li R, Chen Y, Ritchie MD, Moore JH. Electronic health records and polygenic risk scores for predicting disease risk. *Nat Rev Genet.* (2020) 21:493–502. doi: 10.1038/s41576-020-0224-1
- Nagai A, Hirata M, Kamatani Y, Muto K, Matsuda K, Kiyohara Y, et al. Overview of the BioBank Japan project: study design and profile. *J Epidemiol.* (2017) 27:S2–8. doi: 10.1016/j.je.2016.12.005
- Locke AE, Steinberg KM, Chiang CWK, Service SK, Havulinna AS, Stell L, et al. Exome sequencing of Finnish isolates enhances rare-variant association power. *Nature.* (2019) 572:323–8. doi: 10.1038/s41586-019-1457-z
- Bycroft C, Elliott LT, Young A, Vukcevic D, Effingham M, Marchini J, et al. The UK Biobank resource with deep phenotyping and genomic data. *Nature.* (2018) 562:203–9. doi: 10.1038/s41586-018-0579-z
- Hripacsak G, Albers DJ. Next-generation phenotyping of electronic health records. *J Am Med Inform Assoc.* (2013) 20:117–21. doi: 10.1136/amiajnl-2012-001145
- Schmidt M, Schmidt SAJ, Sandegaard JL, Ehrenstein V, Pedersen L, Sørensen HT. The Danish National patient registry: a review of content,

- data quality, and research potential. *Clin Epidemiol.* (2015) 7:449–90. doi: 10.2147/CLEP.S91125
26. Tomassetti C, Johnson NP, Petrozza J, Abrao MS, Einarsson JI, Horne AW, et al. An international terminology for endometriosis, 2021. *Hum Reprod Open.* (2021). doi: 10.1093/hropen/hoab029. [Epub ahead of print].
 27. Pedersen DA, Larsen LA, Nygaard M, Mengel-From J, McGue M, Dalgård C, et al. The Danish twin registry: an updated overview. *Twin Res Hum Genet.* (2019) 22:499–507. doi: 10.1017/thg.2019.72
 28. Thygesen LC, Daasnes C, Thaulow I, Brønnum-Hansen H. Introduction to Danish (nationwide) registers on health and social issues: structure, access, legislation, and archiving. *Scand J Public Health.* (2011) 39:12–6. doi: 10.1177/1403494811399956
 29. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res.* (1988) 16:1215. doi: 10.1093/nar/16.3.1215
 30. Rohde PD, Fourie Sørensen I, Sørensen P. qgg: an R package for large-scale quantitative genetic analyses. *Bioinformatics.* (2020) 36:2614–5. doi: 10.1093/bioinformatics/btz955
 31. Treloar SA, O'Connor DT, O'Connor VM, Martin NG. Genetic influences on endometriosis in an Australian twin sample. *Fertil Steril.* (1999) 71:701–10. doi: 10.1016/S0015-0282(98)00540-8
 32. Saha R, Petterson HJ, Svedberg P, Olovsson M, Bergqvist A, Marions L, et al. Heritability of endometriosis. *Fertil Steril.* (2015) 104:947–52. doi: 10.1016/j.fertnstert.2015.06.035
 33. Matalliotakis M, Zervou MI, Matalliotaki C, Rahmioglu N, Koumantakis G, Kalogiannidis I, et al. The role of gene polymorphisms in endometriosis. *Mol Med Rep.* (2017) 16:5881–6. doi: 10.3892/mmr.2017.7398
 34. Painter JN, Anderson CA, Nyholt DR, MacGregor S, Lin J, Lee SH, et al. Genome-wide association study identifies a locus at 7p15.2 associated with endometriosis. *Nat Genet.* (2011) 43:51–4. doi: 10.1038/ng.731
 35. Sapkota Y, Attia J, Gordon SD, Henders AK, Holliday EG, Rahmioglu N, et al. Genetic burden associated with varying degrees of disease severity in endometriosis. *Mol Hum Reprod.* (2015) 21:594–602. doi: 10.1093/molehr/gav021
 36. Marigorta UM, Rodríguez JA, Gibson G, Navarro A. Replicability and prediction: lessons and challenges from GWAS. *Trends in Genetics.* (2018) 34:504–17. doi: 10.1016/j.tig.2018.03.005
 37. Wand H, Lambert SA, Tamburro C, Iacocca MA, O'Sullivan JW, Sillari C, et al. Improving reporting standards for polygenic scores in risk prediction studies. *Nature.* (2021) 591:211–9. doi: 10.1038/s41586-021-03243-6
 38. Wright CF, West B, Tuke M, Jones SE, Patel K, Laver TW, et al. Assessing the pathogenicity, penetrance, and expressivity of putative disease-causing variants in a population setting. *Am J Hum Genet.* (2019) 104:275–86. doi: 10.1016/j.ajhg.2018.12.015
 39. Fry A, Littlejohns TJ, Sudlow C, Doherty N, Adamska L, Sprosen T, et al. Comparison of sociodemographic and health-related characteristics of UK biobank participants with those of the general population. *Am J Epidemiol.* (2017) 186:1026–34. doi: 10.1093/aje/kwx246
 40. Benagiano G, Brosens I, Habiba M. Structural and molecular features of the endomyometrium in endometriosis and adenomyosis. *Hum Reprod Update.* (2014) 20:386–402. doi: 10.1093/humupd/dmt052
 41. Larsen SB, Lundorf E, Forman A, Dueholm M. Adenomyosis and junctional zone changes in patients with endometriosis. *Eur J Obstet Gynecol Reprod Biol.* (2011) 157:206–11. doi: 10.1016/j.ejogrb.2011.03.003
 42. Martin AR, Kanai M, Kamatani Y, Okada Y, Neale BM, Daly MJ. Clinical use of current polygenic risk scores may exacerbate health disparities. *Nat Genet.* (2019) 51:584–91. doi: 10.1038/s41588-019-0379-x
 43. Privé F, Arbel J, Vilhjálmsdóttir BJ. LDpred2: better, faster, stronger. *Bioinformatics.* (2020) 36:5424–31. doi: 10.1093/bioinformatics/btaa1029
 44. Krapohl E, Patel H, Newhouse S, Curtis CJ, von Stumm S, Dale PS, et al. Multi-polygenic score approach to trait prediction. *Mol Psychiatry.* (2018) 23:1368–74. doi: 10.1038/mp.2017.163
 45. Rohde PD, Nyegaard M, Kjolby M, Sørensen P. Multi-trait genomic risk stratification for type 2 diabetes. *Front Med.* (2021) 8:711208. doi: 10.3389/fmed.2021.711208
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