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DATA NOTE

The genome sequence of the European golden eagle, *Aquila chrysaetos chrysaetos* Linnaeus 1758 [version 1; peer review: 3 approved]

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Abstract
We present a genome assembly from an individual female *Aquila chrysaetos chrysaetos* (the European golden eagle; Chordata; Aves; Accipitridae). The genome sequence is 1.23 gigabases in span. The majority of the assembly is scaffolded into 28 chromosomal pseudomolecules, including the W and Z sex chromosomes.

Keywords
Aquila chrysaetos, European golden eagle, genome sequence, chromosomal

This article is included in the Tree of Life gateway.

Open Peer Review

Reviewer Status ✓ ✓ ✓

	Invited Reviewers		
	1	**2**	**3**
version 1 14 May 2021	✓ report	✓ report	✓ report

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Any reports and responses or comments on the article can be found at the end of the article.

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Author roles: Mead D: Conceptualization, Investigation, Project Administration, Writing – Review & Editing; **Ogden R**: Formal Analysis, Investigation, Resources, Writing – Review & Editing; **Meredith A**: Formal Analysis, Investigation, Resources, Writing – Review & Editing; **Peniche G**: Data Curation, Formal Analysis, Investigation, Resources, Writing – Review & Editing; **Smith M**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Corton C**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Oliver K**: Formal Analysis, Investigation, Supervision, Writing – Review & Editing; **Skelton J**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Betteridge E**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Doulcan J**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Holmes N**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Wright V**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Loose M**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Quail MA**: Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **McCarthy SA**: Data Curation, Formal Analysis, Investigation, Software, Writing – Review & Editing; **Howe K**: Data Curation, Formal Analysis, Investigation, Software, Supervision, Validation, Writing – Review & Editing; **Chow W**: Data Curation, Formal Analysis, Investigation, Software, Validation, Writing – Review & Editing; **Torrance J**: Data Curation, Formal Analysis, Investigation, Software, Validation; **Collins J**: Data Curation, Formal Analysis, Investigation, Software, Validation, Visualization, Writing – Review & Editing; **Challis R**: Data Curation, Formal Analysis, Investigation, Software, Validation, Visualization, Writing – Review & Editing; **Durbin R**: Data Curation, Funding Acquisition, Supervision, Writing – Review & Editing; **Blaxter M**: Data Curation, Funding Acquisition, Project Administration, Supervision, Writing – Original Draft Preparation, Writing – Review & Editing

Species taxonomy

Eukaryota; Metazoa; Chordata; Vertebrata; Aves; Accipitri-formes; Accipitridae; Accipitrinae; Aquila; *Aquila chrysaetos* subspecies *chrysaetos* Linnaeus 1758 (NCBI:txid223781).

Introduction

The golden eagle, *Aquila chrysaetos*, is an apex predator with a range that spans the Holarctic. It has been divided into six subspecies, with the nominate European subspecies, *A. chrysaetos chrysaetos* found across Europe, except for the Iberian peninsula, and extending eastwards in Russia as far as western Siberia. However, mitochondrial sequence and microsatellite analyses suggest that only two major clades exist within the species, a globally distributed northern clade and a distinct Mediterranean clade (Nebel *et al.,* 2015; Nebel *et al.,* 2019; Sato *et al.,* 2017). Formerly widespread, *A. chrysaetos chrysaetos* is now confined to wilderness areas. Once found throughout Britain and Ireland, the golden eagle was extirpated from England and Wales by 1850 and in Ireland by 1912. The golden eagle was particularly badly impacted by bioaccumulating pesticides in the late 20th century (Watson *et al.,* 2010). A single pair nested in the English Lake District from 1969–2004, but this has not led to a sustained recolonisation. There is ongoing monitoring of the remaining population in Scotland, where deliberate persecution is thought to be a major threat (Fielding *et al.,* 2006).

Genome sequence report

The genome was sequenced from a single female *A. chrysaetos chrytsaetos* collected by Gabriela Peniche under UK Home Office project licence PB8A1D5C7. A total of 46-fold coverage in Pacific Biosciences single-molecule long reads (N50 19 kb) and 47-fold coverage in 10X Genomics read clouds (from molecules with an estimated N50 of 68 kb) were generated. Primary assembly contigs were scaffolded with chromosome conformation HiC data. The HiC scaffolds were validated using BioNano long-range restriction maps (140-fold effective coverage in molecules of N50 310 kb). The final assembly has a total length of 1.23 Gb in 145 sequence scaffolds with a scaffold N50 of 46.9 Mb (Table 1). The majority, 99.0%, of the assembly sequence was assigned to 28 chromosomal-level scaffolds representing 26 autosomes (numbered by sequence length), and the W and Z sex chromosomes (Figure 1–Figure 4; Table 2). The assembly has a BUSCO (Simão *et al.,* 2015) completeness of 97.4% using the aves_odb10 reference set. While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited. The *A. chrysaetos chrysaetos* assembly has equivalent span and scaffold-level contiguity to an assembly of *A. chrysaetos canadiensis* (NCBI:txid216574) produced by the Erez Lieberman Aiden lab as part of the DNAZoo project.

As the Hi-C data was sourced from a male bird, it was not possible to fully construct the W chromosome for the female sample bAquChr1. Scaffolds identified as belonging to W have

therefore been submitted as unordered fragments. The largest of these fragments has been designated as the W Chromosome and all other W scaffolds labelled as W_unloc.

Table 1. Genome data for *Aquila chrysaetos chrysaetos* bAquChr1.4.

Project accession data	
Assembly identifier	bAquChr1.4
Species	*Aquila chrysaetos chrysaetos*
Specimen	GE037-17
NCBI taxonomy ID	223781
BioProject	PRJEB27699
Biosample ID	SAMEA994725
Isolate information	Female, heart muscle tissue
Raw data accessions	
PacificBiosciences SEQUEL I	ERR2980431, ERR2980432, ERR2980435, ERR2980436, ERR2980437, ERR2980438, ERR2980439, ERR2980440, ERR2980448, ERR2980449, ERR2980450, ERR2980451, ERR2990043, ERR2990044, ERR3013207, ERR3013208
10X Genomics Illumina	ERR3316065, ERR3316066, ERR3316067, ERR3316068
Hi-C Illumina	ERR3312497
BioNano	ERZ1392826
Genome assembly	
Assembly accession	GCA_900496995.4
Accession of alternate haplotype	GCA_902153765.2
Span (Mb)	1,233
Number of contigs	373
Contig N50 length (Mb)	22
Number of scaffolds	145
Scaffold N50 length (Mb)	47
Longest scaffold (Mb)	85
BUSCO* genome score	C:97.4%[S:96.7%,D:0.7%], F:0.6%,M:2.1%,n:8338

* BUSCO scores based on the aves_odb10 BUSCO set using v5.0.0. C= complete [S= single copy, D=duplicated], F=fragmented, M=missing, n=number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/Aquila%20chrysaetos%20chrysaetos/dataset/UFQG04/busco.

Scaffold statistics

- ☐ Log10 scaffold count (total 150)
- ☐ Scaffold length (total 1.2G)
- ☐ Longest scaffold (88M)
- ☐ N50 length (47M)
- ☐ N90 length (23M)

BUSCO aves_odb10 (8338)

- ☐ Complete (97.4%) ☐ Fragmented (0.6%)
- ☐ Duplicated (0.7%) ☐ Missing (2.1%)

Scale

🕐 1.2G
🕐 88M

Dataset: UFQG04

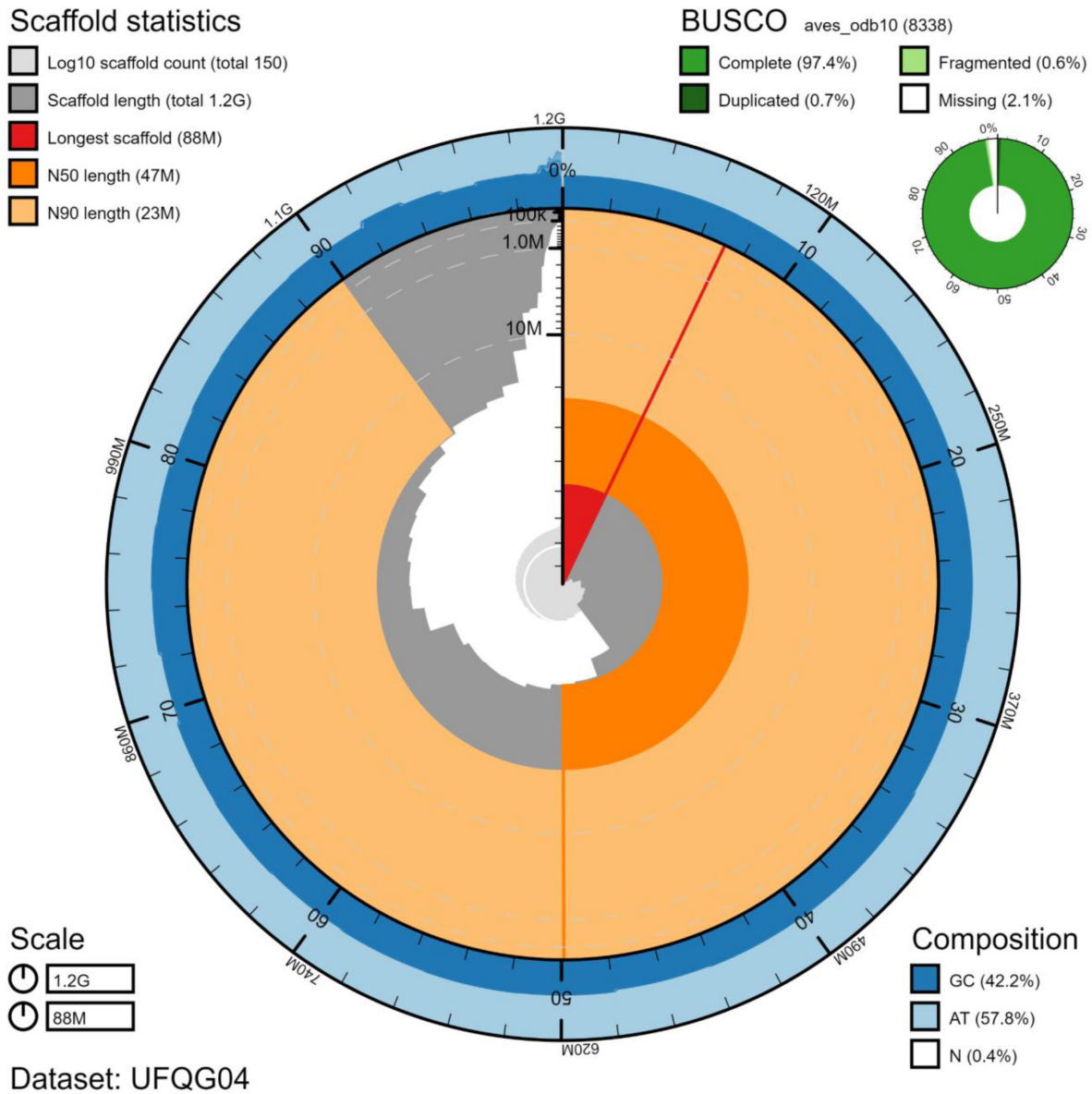## Composition

- ☐ GC (42.2%)
- ☐ AT (57.8%)
- ☐ N (0.4%)

Figure 1. Genome assembly of *Aquila chrysaetos chrysaetos* bAquChr1.4. BlobToolKit Snailplot. The plot shows N50 metrics for bAquChr1.4 and BUSCO scores for the Aves set of orthologues. Interactive version available at https://blobtoolkit.genomehubs.org/view/Aquila%20chrysaetos%20chrysaetos/dataset/UFQG04/snail.

Methods

The golden eagle specimen was collected, following death by natural causes, from an area 15 km from the Highland village of Fort Augustus, Scotland, under UK Home Office project licence no. PB8A1D5C7. The heart was dissected out during autopsy. The specimen is preserved frozen at the University of Edinburgh.

DNA was extracted from heart tissue following the BioNano protocol. Pacific Biosciences CLR long read and 10X Genomics read cloud sequencing libraries were constructed according to manufacturers' instructions. Sequencing was performed by the Scientific Operations core at the Wellcome Sanger Institute on Pacific Biosciences SEQUEL I and Illumina HiSeq X instruments. Hi-C data were generated using the Dovetail HiC library preparation kit at WSI.

BioNano data were generated in DeepSeq, University of Nottingham. High Molecular Weight genomic DNA (HMW gDNA) was extracted from an agarose plug (bAquChr (Golden Eagle); Plug 2) that had been prepared and shipped according to the Bionano Agarose Plug Shipping Instructions. The Bionano Prep Animal Tissue DNA Isolation Soft Tissue Protocol (Document Number: 30077; Document Revision: C) was used to
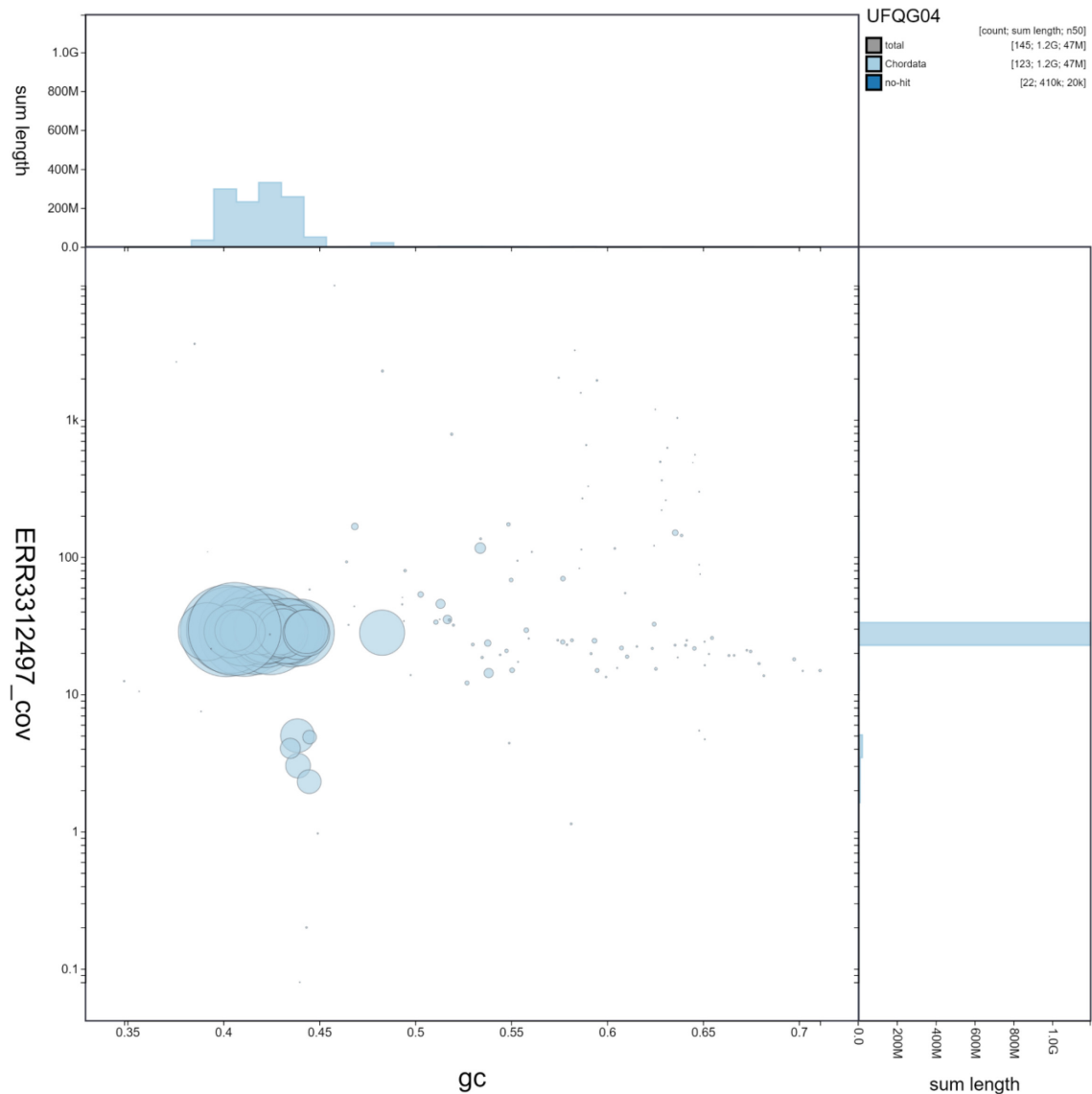
Figure 2. Genome assembly of *Aquila chrysaetos chrysaetos* bAquChr1.4. BlobToolKit GC-coverage plot. Interactive version available at https://blobtoolkit.genomehubs.org/view/Aquila%20chrysaetos%20chrysaetos/dataset/UFQG04/blob?plotShape=circle.

complete HMW gDNA extraction. DNA quantitation, using the Qubit Fluorometer and the Qubit dsDNA BR kit (ThermoFisher; Q32853), gave a mean concentration of 117 ng/ul (CV = 0.118). Labelling was performed with an input of 750 ng of HMW gDNA, using the DLS DNA Labeling Kit (Bionano: 80003) and the Bionano Prep Direct Label and Stain (DLS) Protocol (Document Number: 30206; Document Revision: D). The labelled sample was quantified by Qubit Fluorometer and the Qubit dsDNA HS Assay Kit (ThermoFisher: Q32854). The average concentration of the labelled sample was 4.37 ng/ul (CV = 0.042). The labelling reaction was run over one flowcell of a Bionano Saphyr Chip (Bionano: 20319) on the Bionano Saphyr (Bionano; 60239) running software versions - Bionano Access: 1.2.2; Bionano Tools: 7921; Bionano Solve:

Solve3.2.2_08222018; RefAligner: 7782.7865rel; HybridScaffold/SVMerge/VariantAnnotation: 08222018. For analysis, the molecule file was used to generate a *de novo* assembly using the default Bionano Access settings. This assembly was used to generate a hybrid scaffold from the reference ufqg01.fasta. The hybrid scaffold was constructed using default settings; conflict resolution was set to 'Resolve Conflicts' for both the Bionano assembly and sequence assembly.

Assembly was carried out using Falcon-unzip (falcon-kit 1.1.1) (Chin *et al.,* 2016), haplotypic duplication was identified and removed with purge_dups (Guan *et al.,* 2020) and a first round of scaffolding carried out with 10X Genomics read clouds using scaff10x. Hybrid scaffolding was performed
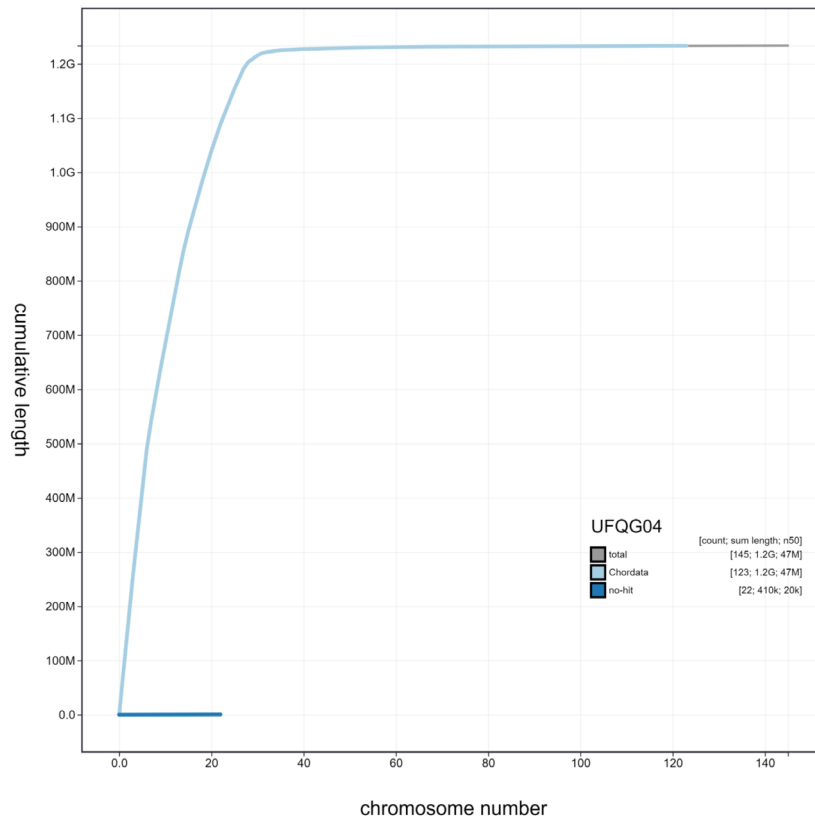
Figure 3. Genome assembly of *Aquila chrysaetos chrysaetos* bAquChr1.4. BlobToolKit Cumulative sequence plot. Interactive version available at https://blobtoolkit.genomehubs.org/view/Aquila%20chrysaetos%20chrysaetos/dataset/UFQG04/cumulative.
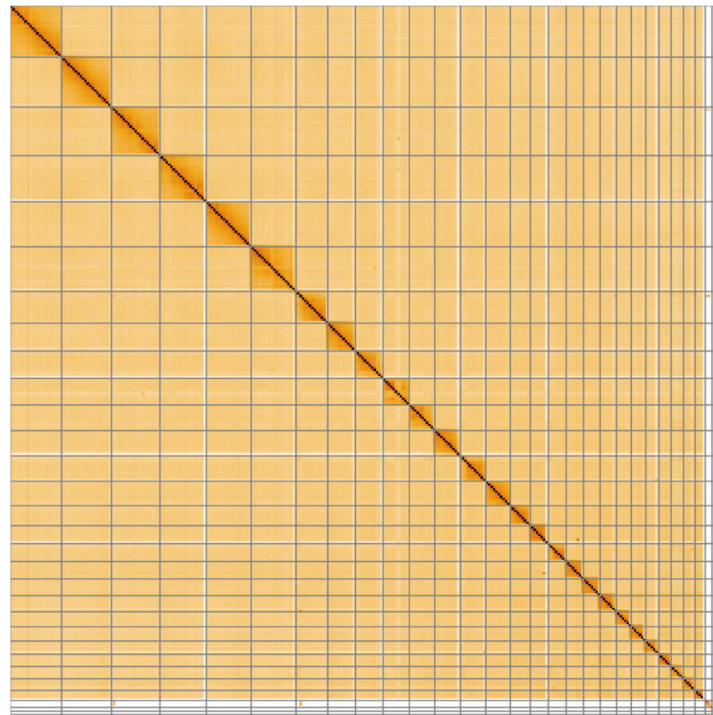

Figure 4. Genome assembly of *Aquila chrysaetos chrysaetos* bAquChr1.4. Hi-C contact map. Hi-C contact map of the bAquChr1.4 assembly, visualized in HiGlass.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Aquila chrysaetos chrysaetos* bAquChr1.4.

ENA accession	Chromosome	Size (Mb)	GC%
LR606181.1	1	85.46	40.2
LR606182.1	2	83.00	41.1
LR606183.1	3	79.38	41.8
LR606184.1	4	77.27	40.4
LR606185.1	5	76.62	42.5
LR606186.1	6	54.40	42.2
LR606187.1	7	47.78	41
LR606188.1	8	46.94	43.6
LR606189.1	9	45.24	44.1
LR606190.1	10	43.95	43.5
LR606191.1	11	43.76	42.2
LR606192.1	12	43.48	43.3
LR606193.1	13	41.79	42.3
LR606194.1	14	34.34	39.2
LR606195.1	15	30.98	42.7
LR606196.1	16	30.61	43.9
LR606197.1	17	29.70	42.6
LR606198.1	18	28.56	40.4
LR606199.1	19	27.98	41.9
LR606200.1	20	25.31	43.1
LR606201.1	21	24.76	43.1
LR606202.1	22	22.51	44.3
LR606203.1	23	21.01	41.1
LR606204.1	24	20.99	48.3
LR606205.1	25	19.84	44.3
LR606206.1	26	17.72	40.7
HG999777.1	W	11.73	44.1
LR606180.1	Z	88.22	40.7
-	unplaced	30.39	48.4

using the BioNano DLE-1 data and Bionano Solve v3.3. The Hi-C scaffolded assembly was polished with arrow using the PacBio data, then polished with the 10X Genomics Illumina data by aligning to the assembly with longranger align, calling variants with freebayes (Garrison & Marth, 2012) and applying homozygous non-reference edits using bcftools consensus. Two rounds of the Illumina polishing were applied. The

assembly was checked for contamination and manually corrected using the gEVAL system (Chow *et al.*, 2016; Howe *et al.*, 2021). This reduced the sequence length by 2.2% and the scaffold count by 44.7% whilst increasing the scaffold N50 by 4.4%. The genome was analysed within the BlobToolKit environment (Challis *et al.*, 2020). Software versions are given in Table 3.

Table 3. Software tools used.

Software tool	Version	Source
Falcon-unzip	falcon-kit 1.2.2	(Chin *et al.,* 2016)
purge_dups	1.0.0	(Guan *et al.,* 2020)
SALSA2	2.2	(Ghurye *et al.,* 2019)
scaff10x	4.2	https://github.com/wtsi-hpag/Scaff10X
arrow	GenomicConsensus 2.3.3	https://github.com/PacificBiosciences/GenomicConsensus
longranger align	2.2.2	https://support.10xgenomics.com/genome-exome/software/pipelines/latest/advanced/other-pipelines
freebayes	v1.1.0-3-g961e5f3	(Garrison & Marth, 2012)
bcftools consensus	1.9	http://samtools.github.io/bcftools/bcftools.html
HiGlass	1.11.6	(Kerpedjiev *et al.,* 2018)
PretextView	0.0.4	https://github.com/wtsi-hpag/PretextView
gEVAL	N/A	(Chow *et al.,* 2016)
BlobToolKit	2.5	(Challis *et al.,* 2020)

Data availability

Underlying data
European Nucleotide Archive: Aquila chrysaetos chrysaetos (European golden eagle) genome assembly, Accession number PRJEB33202.

The genome sequence is released openly for reuse. The *A. chrysaetos chrysaetos* genome sequencing initiative is part of the Wellcome Sanger Institute's "25 genomes for 25 years" project. It is also part of the Vertebrate Genome Project (VGP) ordinal references programme and the Darwin Tree of Life (DToL) project. The specimen has been preserved at Edinburgh University and will be deposited in CryoArks. All raw data and the assembly have been deposited in the ENA. The genome will be annotated and presented through the Ensembl pipeline at the European Bioinformatics Institute. Raw data and assembly accession identifiers are reported in Table 1.

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Publisher Full Text

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Reference Source

Open Peer Review

Current Peer Review Status: ✓ ✓ ✓

Version 1

Reviewer Report 15 November 2021

https://doi.org/10.21956/wellcomeopenres.18335.r46304

✓   **Xiang-jiang Zhan** (iD)

Key Lab of Animal Ecology and Conservation Biology, Institute of Zoology, Chinese Academy of Sciences, Beijing, China

Zhongru Gu (iD)

Key Lab of Animal Ecology and Conservation Biology, Institute of Zoology, Chinese Academy of Sciences, Beijing, China

This data note presents the chromosomal genome assembly of the European golden eagle, *Aquila chrysaetos chrysaetos.* Although it was not the first golden eagle genome assembly reported*,* the authors denovo sequenced a new individual using hybrid techniques including PacBio, 10X Genomics, Hi-C and BioNano. It will facilitate the comparative and population genomics analysis for golden eagle in future. In general, the report was clearly written and the authors did a good job for this top predator. I have only two comments:

1. There are several inconsistencies in the description of software. The version of Falcon-unzip is inconsistent between the method part (falcon-kit 1.1.1) and Table 3 (1.2.2). The SALSA2 listed in Table 3 is not mentioned in the method. The Bionano Solve (v3.3) is mentioned in the method, but not in Table 3.

2. The authors could increase the font size in figure 2 and 3.

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Partly

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Conservation genetics of raptorial birds

We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 07 October 2021

https://doi.org/10.21956/wellcomeopenres.18335.r46301

Carles Lalueza-Fox

Institute of Evolutionary Biology (CSIC-Universitat Pompeu Fabra), University of Barcelona, Barcelona, Spain

This is a comprehensible and well-written report on the generation of a golden eagle female specimen. Golden eagles have ecological importance for being top predators among birds, but also as religious and political icons along history in many different regions and countries. Their genome can provide the basis for further research on their specific adaptations. The details of the sequencing effort and the quality of the assembly and annotation are sound; overall, I think this would be a valuable resource for the scientific community.

I only have a minor comment; maybe the authors should cite a previous work (Doyle *et al.* 2014[1]) where a male from another subspecies (*A. c. canadiensis*) from Sierra Nevada (USA) was sequenced in 2,552 scaffolds. Obviously, due to technological improvements, the current annotation is much better, but future researchers could likely use the previous one to explore inter subspecies diversity.

Also, as a minor comment, Linnaeus 1758 should be between brackets.

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Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Evolutionary Genomics, Paleogenomics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 10 June 2021

https://doi.org/10.21956/wellcomeopenres.18335.r43937

✔   **Bengt Hansson** 🆔
Department of Biology, Lund University, Lund, Sweden

I think this is a well-written Data Note, which presents an important resource for further genetic analyses of the golden eagle and related species. I have only two comments.
1. First, the focus on UK and Ireland in the Introduction seems a bit arbitrary for a general readership. Similar stories about population declines can be applied to several countries. Perhaps, simply, a well-placed "For example" (or similar) would be enough?

2. Secondly, the HiC individual is not included in Table 1, which then provides incorrect information.

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Population genomics and genetics of birds

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.