

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL  
INSTITUTO DE CIÊNCIAS BÁSICAS DA SAÚDE  
CURSO DE GRADUAÇÃO EM BIOMEDICINA

Luiza Marques Prates Behrens

**ÁCIDOS GRAXOS DE CADEIA CURTA E SEU PAPEL NA PATOFISIOLOGIA DO  
HOSPEDEIRO**

Porto Alegre

2021

Luiza Marques Prates Behrens

**ÁCIDOS GRAXOS DE CADEIA CURTA E SEU PAPEL NA PATOFISIOLOGIA DO  
HOSPEDEIRO**

Trabalho de conclusão de curso de graduação apresentado ao Instituto de Ciências Básicas da Saúde da Universidade Federal do Rio Grande do Sul como requisito parcial para a obtenção do título de Bacharela em Biomedicina.

Orientador: Prof. Dr. José Cláudio Fonseca Moreira

Coorientador: MSc. Lucas dos Santos da Silva

Porto Alegre

2021

### CIP - Catalogação na Publicação

Behrens, Luiza  
Ácidos Graxos de Cadeia Curta e seu papel na  
Patofisiologia do Hospedeiro / Luiza Behrens. -- 2021.  
59 f.  
Orientador: José Cláudio Fonseca Moreira.

Coorientador: Lucas dos Santos.

Trabalho de conclusão de curso (Graduação) --  
Universidade Federal do Rio Grande do Sul, Instituto  
de Ciências Básicas da Saúde, Curso de Biomedicina,  
Porto Alegre, BR-RS, 2021.

1. Ácidos Graxos de Cadeia Curta. 2. Microbiota  
Intestinal. 3. Fisiologia do Hospedeiro. 4. Saúde  
Humana. 5. Comunicação entre órgãos. I. Moreira, José  
Cláudio Fonseca, orient. II. dos Santos, Lucas,  
coorient. III. Título.

Luiza Marques Prates Behrens

**ÁCIDOS GRAXOS DE CADEIA CURTA E SEU PAPEL NA PATOFISIOLOGIA DO  
HOSPEDEIRO**

Trabalho de conclusão de curso de graduação apresentado ao Instituto de Ciências Básicas da Saúde da Universidade Federal do Rio Grande do Sul como requisito parcial para a obtenção do título de Bacharela em Biomedicina.

Aprovado em: 27 de abril de 2021.

**BANCA EXAMINADORA**

---

Profa. Dra. Gertrudes Corção - UFRGS

---

Prof. Dr. Alex Sander da Rosa Araújo - UFRGS

---

Prof. Dr. José Cláudio Fonseca Moreira - UFRGS (orientador)

## AGRADECIMENTOS

De todos os desafios que eu imaginava que iria enfrentar durante a graduação, a pandemia foi o maior e mais inesperado deles. Foi muito difícil tentar manter a produtividade durante esse período, foram muitas mortes que poderiam ser evitadas, muito descaso com a ciência... Mas graças a algumas pessoas pude chegar até aqui, e não poderia deixar de agradecê-las.

Ao professor José Cláudio, agradeço por todo o aprendizado que me proporcionou. Um marco para mim foi o dia que o conheci, numa palestra sobre o eixo cérebro-intestino, da qual saí completamente fascinada e querendo atuar em pesquisas sobre o tema. O procurei e ele me deu a oportunidade de fazer Iniciação Científica o tendo como orientador, e essa oportunidade com certeza mudou o rumo da minha formação como biomédica. Muito obrigada por todas as dicas, conselhos, ensinamentos, puxadas de orelha, por tudo. Serei eternamente grata ao senhor por ter me apresentado o assunto que faz meu olho brilhar.

Graças a essa oportunidade, conheci pessoas incríveis no nosso amado Lab 32. E uma dessas pessoas foi o Lucas, uma das pessoas mais maravilhosas e dedicadas que já conheci. Obrigada por ter me treinado, por todas as horas que passamos juntos tomando um café no laboratório e fazendo experimentos. Você se tornou um grande amigo e quero te levar para toda a vida.

Agradeço também aos demais integrantes do Laboratório 32. Obrigada por terem me recebido, por todo o apoio, pelas conversas, pelos seminários semanais, fofocas e risadas. Adorei conhecer todos vocês e trabalhar com pesquisadores tão incríveis, e espero que nossos caminhos se cruzem muito no futuro.

Por fim, não posso deixar de agradecer aos meus amigos e família, que sempre me apoiaram e incentivaram não só durante a graduação, mas na vida. Obrigada por se preocuparem comigo e por terem me dado todo o suporte necessário para chegar até aqui. Eu não seria quem sou hoje sem vocês, e espero um dia poder retribuir tudo o que já fizeram por mim.

## RESUMO

Os ácidos graxos de cadeia curta (AGCC) são metabólitos resultantes da fermentação realizada pela microbiota intestinal e que possuem impacto relevante para a fisiologia do hospedeiro. Os principais AGCC microbianos são o butirato, acetato e propionato, e suas funções são decorrentes da sinalização gerada a partir da ligação com receptores acoplados à proteína G (GPCR) e modulação epigenética de histona deacetilases (HDAC). Os AGCC são absorvidos pelo epitélio intestinal e são utilizados como um importante substrato energético para os colonócitos, com um destaque para o butirato. O que não é consumido pelo epitélio intestinal entra na circulação sanguínea pelos capilares intestinais e migram rumo à veia porta, onde atingem o fígado. O propionato é utilizado como substrato para a gliconeogênese hepática, e dessa forma o acetato se torna o AGCC em maior concentração na circulação sanguínea após passar pelo fígado. Por meio da circulação, os AGCC então atingem o coração, de onde são distribuídos para os tecidos periféricos e para o cérebro, onde o acetato possui capacidade de atravessar a barreira hematoencefálica. A cada parte do organismo no qual os AGCC entram em contato, há a ativação de série de efeitos que são capazes de alterar a fisiologia do hospedeiro, podendo assim estar associados a patologias relacionadas a esses órgãos. Com o rápido desenvolvimento de tecnologias de sequenciamento em massa associadas a análises bioinformáticas mais robustas, pode-se ter maior compreensão acerca da microbiota, seus metabólitos e os mecanismos envolvidos na sua interação com o hospedeiro. Entretanto, novos estudos devem ser feitos para aprofundar o conhecimento acerca dos AGCC e aplicá-lo em prol do benefício humano.

Palavras-chave: Ácidos Graxos de Cadeia Curta. Microbiota Intestinal. Fisiologia do Hospedeiro. Saúde Humana. Comunicação entre órgãos.

## ABSTRACT

Short-chain fatty acids (SCFAs) are end products of gut bacteria and are known to impact host physiology significantly. The main microbial SCFA are butyrate, acetate, and propionate. Their functions are due the signaling generated from G protein-coupled receptors (GPCRs) and epigenetic modulation of histone deacetylases (HDACs). SCFAs are absorbed by the intestinal epithelium and are used as a crucial energetic substrate for colonocytes, emphasizing butyrate. SCFAs not consumed by the epithelium go to the bloodstream through intestinal capillaries and migrates towards the portal vein, where they reach the liver. Propionate is known as a substrate for hepatic gluconeogenesis, and then acetate becomes the SCFA find in greater concentrations in the bloodstream after passing through the liver. Through circulation, SCFAs can reach the heart and then are distributed to peripheral tissues and finally to the brain, where acetate can cross the blood-brain barrier (BBB). At each one of these organs, SCFAs can activate a series of effects that can influence the host's physiology and may be associated with pathologies related to these organs. With the accelerated development of mass sequencing technologies in association with bioinformatics analysis, we can understand more and more about the microbiota, its metabolites, and mechanisms involved in microbiota-host interaction. However, much remains to be discovered for SCFAs to be applied for human benefit in novel therapeutic strategies.

Keywords: Short-Chain Fatty Acids. Gut Microbiota. Host Physiology. Human Health. Inter-organ crosstalk.

## LISTA DE ABREVIATURAS

AGCC	Ácidos Graxos de Cadeia Curta
DII	Doença Inflamatória Intestinal
DT2	Diabetes do tipo 2
FFAR2	Receptor de ácidos graxos livres 2
FFAR3	Receptor de ácidos graxos livres 3
GLP-1	Peptídeo tipo-glucagon 1
GPCR	Receptor acoplado à proteína G
GPR109A	Receptor acoplado à proteína G 109A
GPR41	Receptor acoplado à proteína G 41
GPR43	Receptor acoplado à proteína G 43
HCAR2	Receptor de ácido hidroxicarboxílico 2
HDAC	Histona deacetilase
HMP	Human Microbiome Project
IL10	Interleucina 10
IMC	Índice de massa corporal
MCT1	Transportador de monocarboxilato 1
Olf558	Receptor olfatório 558
Olf78	Receptor olfatório 78
OR51E1	Receptor olfatório da família 51 e subfamília 1
OR51E2	Receptor olfatório da família 51 e subfamília 2
PYY	Peptídeo YY
SMCT1	Transportador de monocarboxilato acoplado à sódio 1
SNC	Sistema Nervoso Central
SNE	Sistema Nervoso Entérico
T2D	Diabetes do tipo 2
Treg	Célula T reguladora
5-HT	Serotonina



## LISTA DE FIGURAS

Figura 1 – Vias conhecidas da biossíntese de AGCC.....	14
Figura 2 – Esquema representativo dos receptores e transportadores de AGCC.....	18
Figura 3 – Esquema representativo da rota dos AGCC no corpo humano.....	38

## SUMÁRIO

<b>1</b>	<b>INTRODUÇÃO COMPREENSIVA</b> .....	<b>11</b>
1.1	Os Ácidos Graxos de Cadeia Curta .....	23
1.2	Receptores de AGCC.....	23
1.2.1	FFAR2/GPR43 .....	23
1.2.2	FFAR3/GPR41 .....	23
1.2.3	GPR109A/HCAR2 .....	23
1.2.4	OR51E1 e OR51E2 .....	23
1.3	Transporte de AGCC .....	23
1.4	Regulação Epigenética dos AGCC.....	23
1.5	AGCC e sinalização endócrina.....	19
1.6	AGCC e resposta imune .....	20
1.7	AGCC associados a patologias.....	20
<b>2</b>	<b>JUSTIFICATIVA</b> .....	<b>22</b>
<b>3</b>	<b>OBJETIVOS</b> .....	<b>23</b>
3.1	Objetivo geral .....	23
3.2	Objetivos específicos.....	23
<b>4</b>	<b>ARTIGO CIENTÍFICO</b> .....	<b>24</b>
<b>5</b>	<b>CONCLUSÕES E PERSPECTIVAS</b> .....	<b>38</b>
	<b>REFERÊNCIAS</b> .....	<b>40</b>
	<b>ANEXO A – NORMAS DE PUBLICAÇÃO DA REVISTA HUMAN MICROBIOME JOURNAL</b> .....	<b>47</b>

## 1 INTRODUÇÃO COMPREENSIVA

A frase “todas as doenças começam no intestino”, dita a mais de dois mil anos por Hipócrates, por mais que exagerada que tenha sido, só mais recentemente vem demonstrando o fundo de verdade que carrega. Com o desenvolvimento das tecnologias de sequenciamento associadas a análises bioinformáticas, dados cada vez mais sofisticados estão surgindo e auxiliando na compreensão acerca da microbiota humana, e a sua importância para diversos órgãos do hospedeiro.

Possuímos uma microbiota distinta em quase todos os nichos do nosso corpo, tais como: pele, vias aéreas, trato urogenital e olhos, sendo o trato gastrointestinal o responsável por abrigar a maior parte dos residentes microbianos do nosso organismo. O intestino abriga uma vasta população de fungos, archaeas e vírus, sendo a população bacteriana a mais abundante e que está melhor caracterizada atualmente pela literatura científica (ECKBURG *et al.*, 2005).

O trato intestinal humano abriga um ecossistema microbiano complexo e dinâmico, onde há mais de 100 trilhões de micróbios que somam 1-2 kg em um humano adulto (FORSYTHE e KUNZE, 2013), sendo um valor comparável ao peso do cérebro humano adulto (PARENT e CARPENTER, 1996). Além disso, mais de 99% dos genes presentes no nosso corpo são de origem microbiana, de forma que o microbioma humano supera em mais de 150 vezes a quantidade de genes presentes no genoma humano (QIN *et al.*, 2010). Com números de tanto destaque quanto esses, fica claro que o ser humano na verdade é um superorganismo e que é necessário estudá-lo levando a microbiota em consideração.

Projetos de sequenciamento microbiano em larga escala como o Human Microbiome Project (HMP) (TURNBAUGH *et al.*, 2007; HUMAN MICROBIOME PROJECT CONSORTIUM, 2012), o European MetaHIT (QIN *et al.*, 2010) e o projeto Eldermet (CLAESSON *et al.*, 2012) contribuíram para identificar a microbiota humana e revelar que esta consiste em pelo menos 40 mil cepas bacterianas de 1800 gêneros diferentes (LUCKEY, 1972; FRANK e PACE, 2008; FORSYTHE e KUNZE, 2013).

A microbiota é composta principalmente por organismos anaeróbicos estritos, que superam o número de bactérias anaeróbicas facultativas e aeróbicas em 2 a 3 vezes, aproximadamente (CLEMENTE *et al.*, 2012). Os filos presentes em maior abundância na microbiota são Firmicutes e Bacteroidetes, com menores números de Proteobacteria, Fusobacteria, Cyanobacteria, Verrucomicrobia e Actinobacteria (QIN, 2010). A microbiota intestinal em adultos pode ser dividida entre dois grandes enterotipos de acordo com o tipo de bactéria dominante, os quais estão fortemente associados com a dieta do hospedeiro (WU *et al.*,

2011). O enterótipo 1 possui as Bacteroides como população predominante, que são metabolizadoras de proteínas, enquanto o enterótipo 2 contém predominantemente Prevotella.

Embora haja variação interindividual na composição da microbiota, há um conjunto de funções conservadas entre o microbioma dos indivíduos (TURNBAUGH e GORDON, 2009), sugerindo que é a funcionalidade da microbiota que possui maior importância para o hospedeiro, e não a composição em si.

A microbiota pode ser influenciada por diversos fatores ambientais, como a dieta, estilo de vida ou habitat (MARQUES *et al.*, 2010), além de infecções, modo de nascimento (parto normal ou cesárea), uso de medicamentos (principalmente antibióticos), estressores ambientais e genética do hospedeiro (ROTHER e BLAUT, 2013; ZHANG *et al.*, 2015; COMPARE *et al.*, 2016)

Um dos mecanismos pelo qual a microbiota interfere na saúde e doença humana é a capacidade de produzir metabólitos a partir da dieta e que possuem capacidade de impactar diversos aspectos da fisiologia do hospedeiro. Uma das classes de metabólitos de maior destaque liberados no lúmen intestinal são os Ácidos Graxos de Cadeia Curta, que serão mais detalhados a seguir.

### 1.1 Os Ácidos Graxos de Cadeia Curta

Um dos principais tipos de metabólitos produzidos pela microbiota intestinal são ácidos graxos orgânicos que possuem entre 2 e 6 moléculas de carbono em sua estrutura, conhecidos como Ácidos Graxos de Cadeia Curta (AGCC). Os principais AGCC sintetizados pela microbiota intestinal são o butirato, acetato e propionato. (CUMMINGS *et al.*, 1987).

Em humanos, os AGCC são resultantes da fermentação de carboidratos complexos, fibras e amidos provenientes na dieta e que nossas enzimas digestivas não são capazes de decompor. Essas substâncias passam por um processo de fermentação bacteriana (principalmente ceco e no cólon) e os AGCC são formados em consequência (CUMMINGS *et al.*, 1987; MACFARLANE e MACFARLANE, 2012). Entretanto, quando esses substratos estão em escassez na dieta, os micróbios se adaptam e utilizam substratos menos favoráveis como proteínas e gorduras da dieta (CUMMINGS e MACFARLANE, 1991; WALL *et al.*, 2009).

Os gêneros bacterianos presentes na microbiota que possuem maior associação com a produção de AGCC são: *Akkermansia*, *Bacteroides*, *Bifidobacterium*, *Prevotella*,

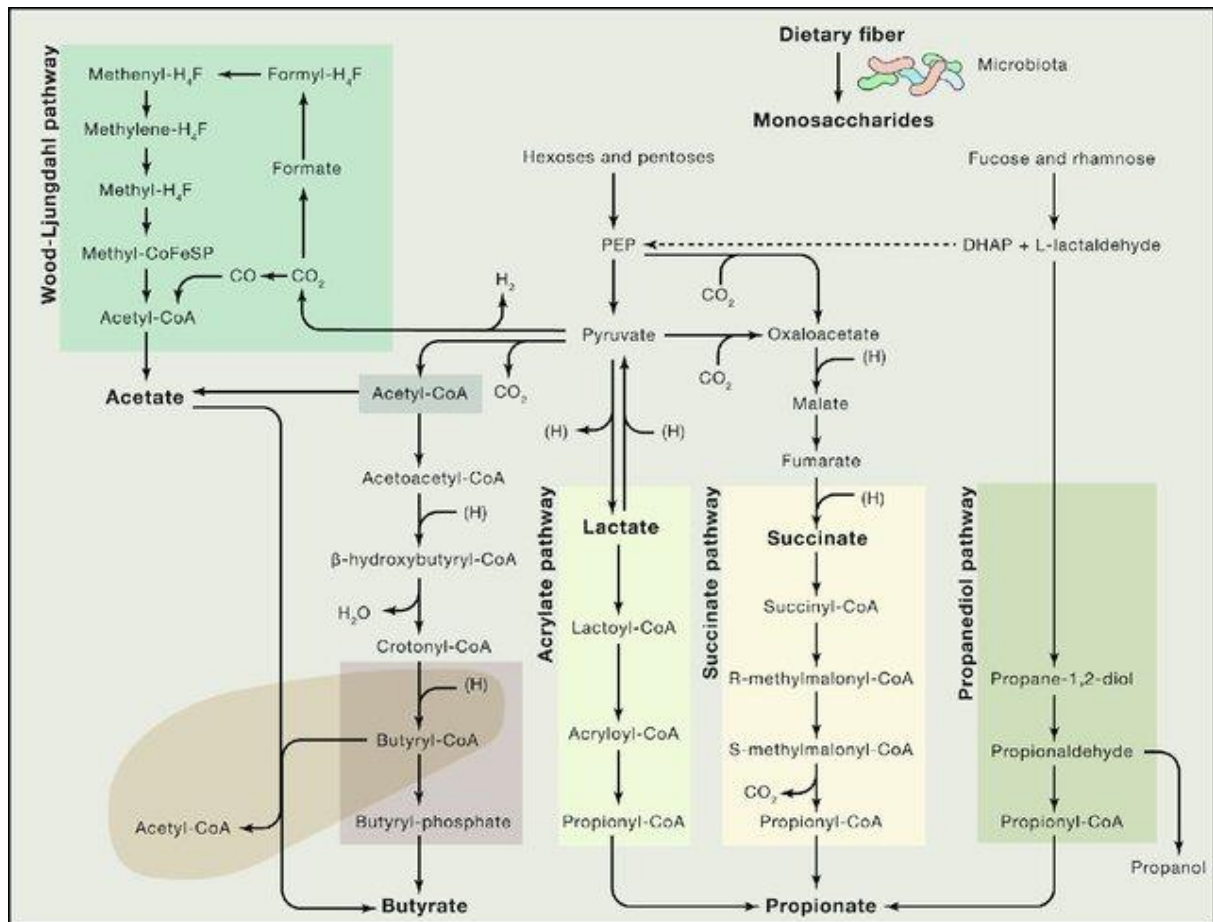
*Ruminococcus*, *Blautia*, *Chlostridium* e *Streptococcus* para o acetato (LOUIS *et al.*, 2014; REY *et al.*, 2010). Já para o acetato, são os gêneros *Bacteroides*, *Phascolarctobacterium*, *Dalister*, *Beilonella*, *Megasphaera*, *Coprococcus*, *Salmonella*, *Roseburia* e *Ruminococcus* (LOUIS *et al.*, 2014; SCOTT *et al.*, 2006). Por fim, em relação à síntese de butirato, os gêneros são: *Coprococcus*, *Canaerostripes*, *Eubacterium*, *Faecalibacterium* e *Roseburia* (DUNCAN *et al.*, 2002; LOUIS *et al.*, 2014).

A conversão de substratos provenientes da dieta em AGCC envolve diversas reações mediadas por enzimas bacterianas (Figura 1). O acetato, por exemplo, pode ser produzido a partir do piruvato por várias bactérias intestinais, seja via acetil-CoA ou pela rota Wood-Ljungdahl, na qual o acetato é sintetizado via redução do CO<sub>2</sub> em formato ou via redução de CO<sub>2</sub> para CO seguida da combinação com um grupo metil para produzir acetil-CoA (RAGSDALE e PIERCE, 2008).

Outro importante AGCC, o propionato, é sintetizado a partir da conversão do succinato em metilmalonil-CoA através da via do succinato. O propionato também pode ser sintetizado a partir do acrilato tendo o lactato como um precursor através da via do acrilato (HETZEL *et al.*, 2003) e pela via do propanodiol, na qual açúcares como fucose e ramnose servem como substratos (SCOTT *et al.*, 2006).

O terceiro principal AGCC, o butirato, é formado a partir da condensação de duas moléculas de acetil-CoA e uma subsequente redução em butiril-CoA, que pode ser convertido em butirato pelas enzimas fosfotransbutirilase e butirato quinase (LOUIS *et al.*, 2004). O butiril-CoA também pode ser transformado em butirato pela rota butiril-CoA/acetato CoA-transferase (DUNCAN *et al.*, 2002). Alguns micróbios também são capazes de utilizar lactato e acetato para a síntese de butirato, o que evita o acúmulo de lactato e estabiliza o meio intestinal. Há análises de metagenoma que sugerem que o butirato também pode ser sintetizado a partir de proteínas através da via da lisina (VITAL *et al.*, 2014), sugerindo que os micróbios intestinais podem se adaptar às mudanças nutricionais para manter a síntese de metabólitos essenciais.

### **Figura 1 – Vias conhecidas de biossíntese dos AGCC**



Fonte: Koh *et al.*, 2016.

A concentração de AGCC no intestino pode variar dependendo da composição da microbiota, do trânsito intestinal, da interação microbiota-hospedeiro e da dieta do indivíduo. Geralmente essa concentração se encontra na faixa de 20 a 140 mM no lúmen intestinal variando ao longo do comprimento do intestino, possuindo níveis mais elevados no ceco e cólon proximal e havendo diminuição em direção ao cólon distal (CUMMINGS *et al.*, 1987), numa proporção aproximada de 60:20:20 para acetato, propionato e butirato, respectivamente (GANAPATHY *et al.*, 2013).

Os AGCC estão mostrando cada vez mais o seu papel como peças-chave das interações entre microbiota e hospedeiro, possuindo um relevante impacto na saúde humana e no desenvolvimento de doenças. A seguir, serão descritos os mecanismos de sinalização pelos quais os AGCC agem na fisiologia.

## 1.2 Receptores de AGCC

Muitas das propriedades regulatórias dos AGCC ocorrem devido às vias de sinalização ativadas por receptores acoplados às proteínas G ou, em inglês, *G protein-coupled receptors* (GPCR), como o GPR43 (também conhecido como *free fatty acid receptor 2* – FFAR2), GPR41 (ou FFAR3) e GPR109A (também conhecido como *hydroxycarboxylic acid receptor 2* - HCAR2), moléculas que agem como receptores de AGCC no organismo (ROOKS e GARRETT, 2016). Existem também os receptores olfatórios da família 51 e subfamília 1 (OR51E1) e 2 (OR51E2), mas com menos dados na literatura. Uma representação dos receptores de AGCC pode ser vista na Figura 2.

Os GPCRs que se ligam aos AGCC são expressos não só em enterócitos intestinais, mas também em outros tipos celulares de outros órgãos e tecidos, como fígado, musculatura, neurônios entéricos e também em células do sistema imunológico, sendo um forte indício da amplitude de seus efeitos no organismo.

### 1.2.1 FFAR2/GPR43

O GPR43, também conhecido como FFAR2, foi identificado como um receptor de AGCC ativado principalmente por acetato e propionato, seguido por butirato (BROWN *et al.*, 2003; LE POUL *et al.*, 2003). FFAR2 é expresso pelas células L intestinais, onde estimulam a liberação de peptídeo YY (PYY) e peptídeo semelhante a glucagon 1 (*glucagon-like peptide-1*, GLP-1) quando ativado. Em consequência da liberação de GLP-1 no intestino, o acúmulo de gordura nos tecidos adiposos é suprimido, levando ao aumento da sensibilidade à insulina (TAZOE *et al.*, 2009). FFAR2 também está expresso em tecidos imunes, havendo evidências de que a microbiota intestinal e FFAR2 regulam a resposta inflamatória na colite (MASLOWSKI *et al.*, 2009). Como a resposta inflamatória também está relacionada com o desenvolvimento de obesidade e de diabetes do tipo 2 (DT2), a regulação da função imune via FFAR2 pode também estar relacionada com efeitos metabólicos benéficos dos AGCC.

### 1.2.2 FFAR3/GPR41

O GPR41 (ou FFAR3) também foi identificado como um receptor de AGCC. Esse receptor é ativado principalmente por propionato e butirato (BROWN *et al.*, 2003; LE POUL *et al.*, 2003) e também está expresso em células L secretoras de PYY e de GLP-1, indicando

seu envolvimento na homeostase energética (TAZOE *et al.*, 2009). FFAR3 também está expresso abundantemente nos gânglios neuronais simpáticos, em particular o gânglio cervical superior, que é responsável por controlar o gasto de energia por meio de efeitos neurais e hormonais sobre o metabolismo da glicose e gordura (KIMURA *et al.*, 2011). A ativação de FFAR3 pela interação com o propionato gera um aumento no ritmo cardíaco e no gasto de energia através da ativação simpática, além de levar à liberação de noradrenalina pelos neurônios simpáticos (INOUE *et al.*, 2012). Logo, há indícios de que FFAR3 regula a atividade simpática ao detectar o estado nutricional do indivíduo, mantendo assim a homeostase da energia corporal.

Também há evidência de que FFAR3 contribui para uma melhora da resistência à insulina pelas fibras dietéticas através a ativação do FFAR3 expresso nos nervos periféricos por meio dos AGCC produzidos pela microbiota (CUMMINGS *et al.*, 1987). Isso implica que o estímulo de FFAR3 gerado por AGCC exibe efeitos benéficos no metabolismo do hospedeiro por meio do sistema nervoso periférico e da secreção de hormônios no intestino. Assim como FFAR2, FFAR3 também afeta a resposta inflamatória. O propionato se mostrou capaz de afetar a hematopoiese da medula óssea de uma maneira dependente do FFAR3 em camundongos induzindo uma maior produção de macrófagos e precursores de células dendríticas, influenciando assim uma resposta inflamatória alérgica (AHMED *et al.*, 2009). Dessa forma, FFAR3 pode estar envolvido em efeitos benéficos dos AGCC no metabolismo do hospedeiro através da regulação de respostas imunes.

### 1.2.3 GPR109A/HCAR2

O GPR109A (ou HCAR2) foi primeiro identificado como um receptor para niacina, mas também é ativado por  $\beta$ -hidroxibutirato e butirato, porém não por acetato e propionato (AHMED *et al.*, 2009). O GPR109A está expresso nas células epiteliais do cólon e sua ativação por butirato é capaz de suprimir a inflamação no cólon e o processo de carcinogênese, promovendo propriedades anti-inflamatórias em macrófagos do cólon e células dendríticas, que induzem a diferenciação de células T reguladoras (Treg) e produtoras de interleucina 10 (IL10) (SINGH *et al.*, 2014). Além disso, o GPR109A também está expresso em tecidos adiposos e em macrófagos desses tecidos, tendo um papel na regulação da homeostase lipídica (AHMED *et al.*, 2009).



#### 1.2.4 OR51E1 e OR51E2

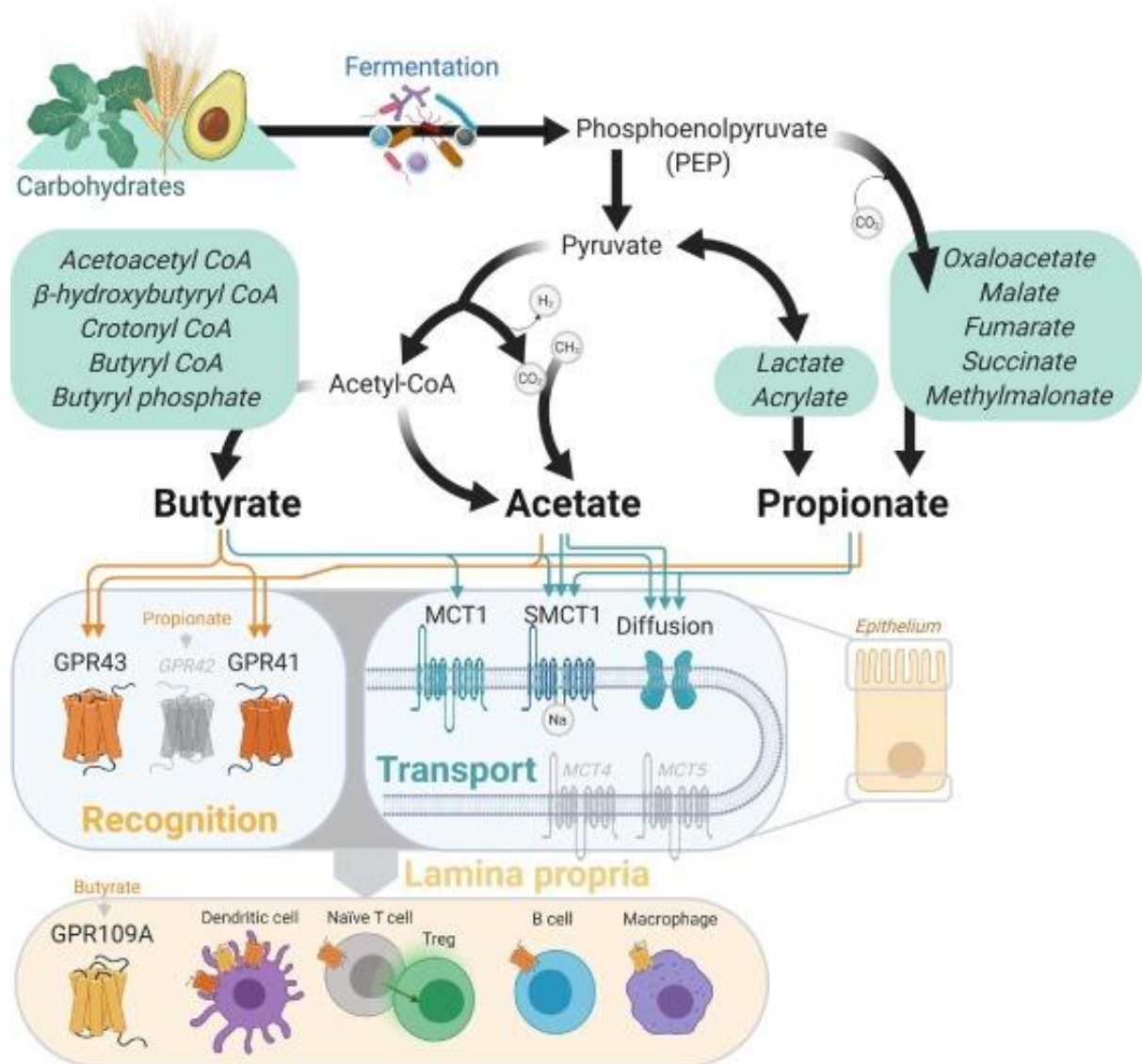
O receptor OR51E1 é equivalente ao receptor olfatório 558 (Olf558) presente em camundongos, e ambos são suscetíveis a ativação por meio do butirato. Entretanto, são necessárias concentrações mais altas para que a ativação ocorra, sendo improvável que o butirato circulante seja capaz de ativar esses receptores em tecidos além dos intestinais e hepáticos (PRIORI *et al.*, 2015).

Há também o OR51E2, o qual é ativado por acetato e propionato e que também possui um correspondente em camundongos, o Olf78 (PLUZNICK *et al.*, 2013). Ele está expresso em melanócitos humanos (GELIS *et al.*, 2016); entretanto, ainda é necessário obter mais dados acerca da contribuição desses receptores para a fisiologia do hospedeiro.

#### 1.3 Transporte de AGCC

AGCC podem ser absorvidos pelo epitélio intestinal de forma passiva mas a maior parte do transporte é realizado de forma ativa via transportador de monocarboxilato 1 (MCT1), e em menor quantidade pelo transportador de monocarboxilato acoplado à sódio 1 (SMCT1). (HALESTRAP e MEREDITH, 2004) Os AGCC, principalmente o butirato, após absorvidos podem ser metabolizados pelos colonócitos, servindo como uma rica fonte de energia (CLAUSEN e MORTENSEN, 1995). O que não for utilizado pelo epitélio é transportado pela membrana celular basolateral através de um mecanismo ainda não bem elucidado, que talvez seja realizado pelo MCT4 ou 5 (DEN BESTEN *et al.*, 2013). Na mucosa, os AGCC entram nos capilares sanguíneos e são direcionados para o fígado pela veia porta. O fígado utiliza o propionato como substrato para a gliconeogênese e o acetato acaba se tornando o AGCC mais presente na circulação sanguínea após a passagem do sangue pelo fígado. (BLOEMEN *et al.*, 2009). Os mecanismos de transporte dos AGCC estão representados na Figura 2.

#### **Figura 2 – Esquema representativo dos receptores e transportadores de AGCC**



Trends in Microbiology

Fonte: Hee e Wells, 2021.

#### 1.4 Regulação Epigenética dos AGCC

Os AGCC produzidos pela microbiota intestinal são capazes de induzir efeitos inibitórios na histona deacetilases (HDAC), funcionando como inibidores não competitivos de HDACs, e foi demonstrado que o butirato e o propionato inibem seletivamente HDAC1 e HDAC3. O butirato é considerado o inibidor mais potente, inibindo principalmente as HDAC das classes I e IIa (CLEOPHAS *et al.*, 2016; DAVIE, 2003).

Com relação à influência da inibição de HDAC pelos AGCC nas funções fisiológicas do hospedeiro, foi relatado que o propionato e o butirato produzidos por microorganismos

intestinais promovem a geração de células T regulatórias periféricas (ARPAIA *et al.*, 2013; FURUSAWA *et al.*, 2013). O butirato induz a diferenciação de células T reguladoras do cólon, aumentando a acetilação da histona H3 e reduzindo o desenvolvimento de colite (FURUSAWA *et al.*, 2013). Além disso, há evidência de que a diversidade da microbiota intestinal e o grau de metilação da região promotora do gene FFAR3 foram significativamente menores nos pacientes obesos e com diabetes do tipo 2 em comparação com indivíduos magros, demonstrando a existência de uma correlação entre o índice de massa corporal (IMC) mais alto e menor metilação de FFAR3 (REMELY *et al.*, 2014). Portanto, a regulação epigenética também pode estar relacionada aos efeitos benéficos dos AGCC no metabolismo do hospedeiro.

Recentemente, os AGCC foram associados à crotonilação de histonas, mas a relevância dessa alteração para a fisiologia do hospedeiro ainda precisa ser melhor investigada (FELLOWS *et al.*, 2018).

#### 1.4 AGCC e sinalização endócrina

Os AGCC são capazes de modular o sistema endócrino do hospedeiro por meio do estímulo à liberação de hormônios. Por exemplo, o butirato e o propionato estimulam a liberação do peptídeo tipo-glucagon 1 (glucagon-like peptide 1 – GLP1) e do hormônio intestinal peptídeo YY (PYY) regulador do apetite em células enteroendócrinas intestinais por meio de um mecanismo dependente de FFAR2 (PSICHAS *et al.*, 2015; TOLHURST *et al.*, 2012; KAJI *et al.*, 2014; CHAMBERS *et al.*, 2015). E ainda mais, bactérias produtoras de butirato foram associadas com uma maior secreção de GLP-1 e à maior expressão de genes envolvidos em sua síntese e excreção (YADAV *et al.*, 2013). Esses dados sugerem um papel dos AGCC derivados da microbiota intestinal na produção de hormônios endócrinos.

E, além disso, as células enteroendócrinas também são capazes de liberar GLP-2 em resposta à nutrição parenteral com butirato, aumentando as concentrações plasmáticas desse peptídeo. O GLP-2 é capaz de aumentar a área de superfície do epitélio do intestino delgado, aumentando a proliferação celular e inibindo a apoptose (TAPPENDEN *et al.*, 2003).

#### 1.5 AGCC e resposta imune

O sistema imune pode ser afetado pela microbiota pelo fato de que há muitas células imunes localizadas no trato gastrointestinal, o que significa que uma perturbação no equilíbrio do ecossistema intestinal também pode resultar em alterações do sistema imune. Os AGCC produzidos pela microbiota intestinal possuem propriedades anti-inflamatórias e podem modular a resposta imune (van de WOUW *et al.*, 2018). No intestino, os AGCC são capazes de influenciar a expressão de marcadores anti-inflamatórios, como a interleucina 10 (IL-10) em macrófagos e células dendríticas intestinais (SINGH *et al.*, 2014).

A sinalização de butirato através do receptor GPR109A demonstrou conferir propriedades anti-inflamatórias em macrófagos e células dendríticas presentes no cólon (SINGH *et al.*, 2014), sendo importantes para a manutenção da homeostase intestinal (WELLS *et al.*, 2011). Em outros estudos, os efeitos anti-inflamatórios do butirato e propionato nessas células mostraram ser independentes de GPCRs, mas dependentes da modulação das HDACs (ARPAIA *et al.*, 2013; CHANG *et al.*, 2014).

O butirato pode agir nas células imunes através dos GPRs (GPR41 e GPR43), que estão expressos nas células imunes, incluindo células polimorfonucleares, indicando um possível envolvimento desse AGCC na ativação dos leucócitos (MEIJER *et al.*, 2010). Além disso, o butirato é um importante regulador negativo de inflamação (MASLOWSKI *et al.*, 2009; KARLSSON *et al.*, 2012).

## 1.6 AGCC associado a patologias

A microbiota está constantemente em comunicação com os órgãos e sistemas do organismo do hospedeiro, incluindo a medula óssea (JOSEFSDOTTIR *et al.*, 2017), vasculatura (KARBACH *et al.*, 2016), rins (EVENEPOEL *et al.*, 2017), sistema imune (ROOKS e GARRETT, 2016), sistema nervoso autônomo (BERCIK *et al.*, 2011; BRAVO *et al.*, 2011) e o cérebro (DINAN e CRYAN, 2017). Essa complexa comunicação contribui para a homeostase e a saúde do hospedeiro, e desbalanços na microbiota (conhecida como disbiose) podem estar associados ao desenvolvimento de doenças. A disbiose possui um papel central em várias doenças crônicas, e atenuá-la pode ser uma estratégia em potencial para o controle dessas patologias (KONTUREK *et al.*, 2015).

AGCC geram diversos impactos em vários aspectos da fisiologia do hospedeiro, inclusive na suscetibilidade a doenças (EVANS *et al.*, 2013).

A influência dos AGCC na (pato)fisiologia do hospedeiro será abordada de forma mais extensiva no artigo presente neste trabalho.

## 2 JUSTIFICATIVA

Com o surgimento de tantos dados acerca da microbiota intestinal, se torna necessário organizar o conhecimento já adquirido para identificar as lacunas ainda existentes nas quais devem ser focadas novas pesquisas, buscando uma melhor compreensão sobre os AGCC, seus mecanismos de interação com o hospedeiro e seus efeitos no organismo.

Além disso, é necessário ressaltar o potencial de aplicação dos AGCC em estratégias clínica e terapêutica, visando atenuar o quadro patológico de diversas doenças humanas e contribuir para a busca de uma melhor qualidade de vida para o paciente.

## 3 OBJETIVOS

### 3.1 Objetivo geral

Realizar uma revisão compreensiva das evidências já presentes na literatura científica acerca dos ácidos graxos de cadeia curta de origem microbiana e sua influência na patofisiologia do hospedeiro, buscando uma maior compreensão de seus efeitos em escala sistêmica.

### 3.2 Objetivos específicos

3.2.1 Investigar o papel dos ácidos graxos de cadeia curta e as alterações fisiológicas que ocorrem no organismo em decorrência da sua produção pela microbiota intestinal.

3.2.2 Investigar a associação dos ácidos graxos de cadeia curta com o desenvolvimento de patologias humanas.

3.2.3 Investigar o efeito causado pelos ácidos graxos de cadeia curta com foco em órgãos cujas patologias possuem maior associação com a microbiota intestinal, como intestino, fígado, coração e cérebro.

3.2.4 Verificar possíveis interconexões entre os eixos microbiota-fígado, microbiota-coração e microbiota-cérebro.

3.2.5 Identificar lacunas ainda existentes no conhecimento acerca dos ácidos graxos de cadeia curta e que podem se tornar foco de investigações futuras.

## 4 ARTIGO CIENTÍFICO

### Microbial Short-Chain Fatty Acids and Host Pathophysiology

Luiza Behrens<sup>1</sup>; Lucas Santos<sup>1</sup>; José Cláudio Fonseca Moreira<sup>1\*</sup>

<sup>1</sup>Department of Biochemistry, Federal University of Rio Grande do Sul, 2600 Ramiro Barcelos, 90035003, Porto Alegre, Brazil

\*Corresponding author: [jcfm@ufrgs.br](mailto:jcfm@ufrgs.br)

#### Highlights

- Short-chain fatty acids (SCFAs) are speculated to have a critical role in host physiology
- SCFAs might interact with the host via G protein-coupled receptor or histone deacetylases
- SCFAs should be quantified in the systemic circulation and fecal samples to the development of specific biochemical assays
- SCFAs could be used in potential interventional strategies and novel therapeutic methods for clinical practice
- Further investigation is still needed to understand SCFAs mechanisms of action fully

#### Abstract

Short-chain fatty acids (SCFAs) are end products of gut bacteria and are known to impact host physiology significantly. The main microbial SCFA are butyrate, acetate, and propionate. Their functions are due the signaling generated from G protein-coupled receptors (GPCRs) and epigenetic modulation of histone deacetylases (HDACs). SCFAs are absorbed by the intestinal epithelium and are used as a crucial energetic substrate for colonocytes, emphasizing butyrate. SCFAs not consumed by the epithelium go to the bloodstream through intestinal capillaries and migrates towards the portal vein, where they reach the liver. Propionate is known as a substrate for hepatic gluconeogenesis, and then acetate becomes the SCFA find in greater concentrations in the bloodstream after passing through the liver. Through circulation, SCFAs can reach the heart and then are distributed to peripheral tissues and finally to the brain, where acetate can cross the blood-brain barrier (BBB). At each one of these organs, SCFAs can activate a series of effects that can influence the host's physiology and may be associated with pathologies related to these organs. With the accelerated development of mass sequencing technologies in association with bioinformatics analysis, we can understand more and more



about the microbiota, its metabolites, and mechanisms involved in microbiota-host interaction. However, much remains to be discovered for SCFAs to be applied for human benefit in novel therapeutic strategies.

**Keywords:** Short-Chain Fatty Acids. Gut microbiota. Host Physiology. Human Health. Interorgan crosstalk.

## 1 Introduction

The human gastrointestinal tract is home to most of the microbial residents of our body. The intestine is home to a vast bacterial population in a complex and dynamic microbial ecosystem, where more than 100 trillion microbes add up to 1-2 kg in a healthy adult<sup>1</sup>. Therefore, it is clear that the human being is a superorganism and that it is necessary to study it considering the microbiota as well. One of the mechanisms with which the microbiota interferes with human health and disease is through metabolites' production. One of the most prominent classes of metabolites released in the gut are short-chain fatty acids (SCFAs). The most studied SCFAs are butyrate, acetate, and propionate<sup>2</sup>. In humans, SCFAs are resultant of complex carbohydrates, non-digestible fibers and starches from diet. These substances undergo bacterial fermentation (mainly on the cecum and colon) and SCFAs are released as result<sup>2</sup>.

SCFAs are increasingly showing their crucial role in the microbiota-host interaction and their impact on human health and disease development. Many of the regulatory properties of SCFAs occur due to signaling pathways activated by G protein-coupled receptors (GPCR) such as GPR43 (also known as free fatty acid receptor 2, FFAR2), GPR41 (or FFAR3), and GPR109A (also known as hydroxycarboxylic acid receptor 2, HCAR2)<sup>3</sup>. There are also olfactory receptors of family 51 and subfamily 1 (OR51E1) and 2 (OR51E2), but there are less data about them. SCFAs can also induce inhibitory effects on histone deacetylases (HDACs), functioning as non-competitive inhibitors and, consequently, modulating the human epigenetic.

This is a comprehensive review of the role of microbial SCFAs on the physiology of the host, and our goal is to expand our view of what is already known about SCFAs mechanisms and how it could be used for human benefit, highlighting the high potential for novel therapeutic strategies based on the microbiota and its metabolites.

## 2 SCFAs and gut

It is believed that the microbiota can generate benefits in the intestinal barrier through SCFAs, which are an essential source of energy not only for the microbiota but also for cells of the intestinal epithelium itself. Moreover, in addition to being a substrate for energy production, SCFAs may affect several processes in the host organism, including host-microbiota signaling, colon pH, intestinal motility, cell epithelium proliferation, fluid and electrolyte absorption, hormone and cytokines secretion, and the maintenance of barrier function<sup>3</sup>.

SCFAs have multiple roles in the maintenance of intestinal homeostasis. As for ion absorption, SCFAs have a significant role in NaCl absorption and electrolyte balance. Butyrate, mainly, stimulates NaCl absorption and inhibits Cl<sup>-</sup> secretion<sup>4</sup>. Also, SCFAs participate in the control of the balance between proliferation and cell apoptosis on the intestinal epithelium<sup>5</sup>, induce antimicrobial peptides secretion<sup>6</sup> and differentiation of regulatory T cells (Treg) such as Th17<sup>7</sup>.

Between SCFAs, butyrate is the main energy source for colonocytes and is capable of inducing gut gluconeogenesis. Propionate is also used as substrate in the gut but to a lesser extent. There is evidence that intestinal gluconeogenesis enhances beneficial metabolic effects of butyrate and propionate via signalization for increased insulin sensitivity and glucose tolerance<sup>8</sup>.

In addition to being an energy substrate, SCFAs significantly influence in maintaining the integrity of the intestinal barrier. That barrier consists in the intestinal epithelium (which can be more or less permeable according to the expression of tight junctions) and a mucus layer full of antimicrobial peptides secreted by the epithelium itself. There is evidence that SCFAs-producing bacteria protects against infections caused by pathogenic bacteria, indicating that SCFAs plays a prominent role in maintaining the intestinal barrier's integrity, preventing the translocation of harmful bacteria and toxins from the lumen to the host<sup>9</sup>. On the other hand, the barrier must be permeable enough for fluids and nutrients absorption. To achieve both these goals, it is necessary to maintain a peaceful coexistence between microbiota and host. There is evidence that disturbances in this relationship can activate inflammatory processes and increase diseases' risk<sup>10</sup>. Reduction in SCFAs levels, especially butyrate, may influence the barrier dysfunction and facilitate the translocation of toxins and LPS from lumen to the bloodstream, activating inflammatory processes<sup>11</sup>. Thus, it reinforced the hypothesis that mechanisms of control of the intestinal barrier are essential for maintaining health, and that microbiota-related disorders can lead to disease development.

One of the mechanisms for the maintenance of the integrity of the gut barrier is the increase of mucin secretion by the goblet cells of the epithelium in response to SCFAs<sup>12</sup>,

increasing the mucus layer that protects the epithelium from direct contact with lumen microorganisms. Butyrate, significantly, can increase the expression of tight junction proteins, such as occludin, claudin, and zonula occludens<sup>13</sup> and reduce intestinal permeability, which prevents translocation of bacteria and toxins through the epithelium<sup>14</sup>. Through these mechanisms, the integrity of the gut barrier is preserved.

Likewise, patients who presents a deficit of butyrate-producing bacteria in the gut also shows a disruption of gut epithelium due to lesions in tight junctions (mainly related to claudin 1 and 2 proteins), which leads to an increase in intestinal permeability. This was seen in patients with inflammatory bowel disease (IBD), indicating that increased permeability may increase the host's susceptibility to inflammatory processes and that butyrate deficiency may be an indirect indicator of intestinal barrier function<sup>13</sup>. However, it would still be necessary to develop specific essays for butyrate evaluation to apply this knowledge in clinical practice.

Studies have shown that butyrate is reduced in CRC patients<sup>15</sup> and can have effects on colorectal carcinoma (CRC) cells, potentially reducing cell growth and inducing apoptosis in cancer cells<sup>16</sup>. One of the mechanisms involved is the inhibition of HDAC, preventing an imbalance in histone acetylation that can lead to genes involved in cell control and apoptosis<sup>17</sup>. Another mechanism would be through the GPR109A receptors of the colon, that are capable of activating the apoptosis in tumor cells<sup>18</sup>.

We can also mention the anti-inflammatory role of butyrate by inhibiting NF- $\kappa$ B in colon epithelial cells<sup>19</sup>, which may result from HDAC inhibition. NF- $\kappa$ B activity is often dysregulated in CRC patients and inflammatory bowel diseases (IBD) such as ulcerative colitis and Crohn's disease. In Crohn's disease, butyrate has been shown to reduce pro-inflammatory cytokines' expression by inhibiting NF- $\kappa$ B<sup>20</sup>.

Once produced SCFAs are absorbed by gut colonocytes or are eliminated in feces, which are great biological samples for microbiota analysis and are widely used to measure SCFAs production. The absorbed SCFAs that are not consumed by the gut epithelium as an energy source are transported to blood capillaries, migrates towards the portal vein, and reach the liver<sup>2</sup>.

#### **4 SCFAs and liver**

The gut-liver axis refers to a bidirectional relationship between the gut, its microbiota, and the liver. This reciprocal interaction occurs through the portal vein, which allows the transport of products derived from gut microbiota (such as SCFAs) to the liver and a hepatic

feedback response by bile and antibodies secretion. There is an interdependence between gut and liver, and disturbances in the intestinal barrier could result in an influx of bacteria and microbial products to the liver, resulting in inflammation processes and even developing certain diseases<sup>21</sup>.

There is evidence that SCFAs may play a role as a significant substrate for increasing triglyceride levels in the liver. Propionate is described as an essential substrate for hepatic gluconeogenesis<sup>8</sup>. On the other hand, butyrate can activate oxidation and thermogenesis of fatty acids by increasing the expression of PGC-1 $\alpha$  and phosphorylation of AMPK in liver tissues<sup>22</sup>. Thus, SCFAs promote energy storage and weight gain and may be related to pathological states that affect liver homeostasis.

Human studies comparing non-alcoholic fatty liver disease (NAFLD), non-alcoholic steatohepatitis (NASH) and healthy subjects show a higher fecal concentration of SCFAs in patients with NAFLD and NASH, concomitantly with an increase in SCFAs-bacterial producers. Furthermore, with the increase in fecal SCFAs and microbial signature in NASH, there is an associated reduction in Treg cells and a higher number of Th17 in peripheral blood, a systemic response observed in NASH. In both obesity and NAFLD, there is evidence of an association between clinical phenotypes and SCFAs attributed to a difference in the amount of individual SCFAs, and each one may have different effects on host metabolism<sup>23</sup>.

The microbiome of patients with Type 2 Diabetes mellitus (T2D) shows a reduction in the number of butyrate-producing bacteria<sup>23</sup>. Although the microbial dysbiosis in patients with T2D is moderate compared to the dysbiosis reported in obese patients, a study revealed that patients with T2D exhibit a lower number of butyrate-producing bacteria and more significant numbers of opportunistic pathogens. Enrichment of microbial functions associated with reduced sulfate and increased oxidative stress has also been reported, which is indicative of pathogenicity<sup>23</sup>. This promotes the idea that butyrate-producing bacteria protect the host against diseases. Therefore, it is worth considering butyrate as a potential biomarker for intestinal and hepatic health.

Diseases related to the consumption of alcohol can also be correlated with the microbiota. Liver damage from alcohol consumption is marked by a reduction in butyrate and propionate levels<sup>24</sup>. And higher levels of acetate (possibly because of the metabolism of ethanol in the intestinal lumen but mainly by liver metabolism). The reduction in the amount of butyrate is associated with a weakening of the intestinal tight junctions, related to gut permeability<sup>24</sup>.

For being an important substrate for liver metabolism, propionate is present in lower concentrations on peripheral tissues, leaving acetate as the most abundant SCFA in the

peripheral circulation<sup>2</sup>. Acetate, therefore, is the most present SCFA in the bloodstream when it reaches the heart.

### **5 SCFAs and heart**

After passing through the liver, SCFAs follow the bloodstream influx and eventually reach the heart and, from there, they can be distributed to the host's peripheral tissues. The concentration of SCFAs in a healthy adult's peripheral blood is estimated at 100-150  $\mu\text{mol/L}$  for acetate, 4-5  $\mu\text{mol/L}$  for propionate and 1-3  $\mu\text{mol/L}$  for butyrate<sup>2</sup>. The latter two are present in lower concentrations because they are metabolized in the gut and liver before reaching the bloodstream, as has already been shown.

As much as SCFAs reach the heart via circulation, they have a greater capacity to influence the cardiovascular system and its pathologies indirectly by signaling pathways from the gut. It has already been shown that in heart failure, for example, the microbiota's diversity is altered, showing depletion of butyrate-producing bacteria<sup>25</sup>. Moreover, this is inversely associated with high levels of soluble CD25 (alpha subunit of the IL-2 receptor) in plasma, a marker for activation of macrophages and T cells. These findings support the potential of the gut microbiota relevance and the modulation of inflammation through SCFAs production in heart disease.

It is hypothesized that when the intestinal barrier is disrupted, there is a more significant translocation of components from the microbiota to the bloodstream. It generates a systemic inflammatory condition that can have a negative impact on patients with heart failure<sup>26</sup>. Evidence is that bacterial translocation occurs in patients with heart failure because of changes in the gastrointestinal tract due to splenic congestion, as well as immunological abnormalities. Therefore, SCFAs are how the gut microbiota is involved with the regulation of blood pressure, especially butyrate<sup>27</sup>. Wilck and collaborators demonstrated that salt-related hypertension was associated with the depletion of SCFAs-producing bacteria strains, which is related with a lower induction of Th17 cells and hypertension<sup>28</sup>.

Pluznick and colleagues have shown that the gut microbiota can produce SCFAs and modulate blood pressure through the interaction of SCFAs with GPCRs. The stimulation of these receptors activates pathways that lead to renin secretion, and dysfunctions in this signaling process may be associated with the development of hypertension<sup>29</sup>. The receptors more associated with the development of hypertension through blood pressure changes are FFAR2, FFAR3, and OR51E2<sup>27,29</sup>.

In a pathological atherosclerosis context, it is noteworthy that SCFAs can play a crucial role in innate immunity and inflammatory processes through mechanisms involving HDAC and/or GPCR receptors<sup>30</sup>. *Eubacterium rectale*, a butyrate producer, is present at lower levels in atherosclerosis patients<sup>31</sup>. Researchers also observed lower levels of *Faecalibacterium prausnitzii*, other SCFA producer, in older patients with congestive heart failure<sup>32</sup>. Another group reported that the depletion of *Eubacterium* species and higher levels of soluble CD25 in plasma were associated with heart transplantation cases<sup>33</sup>.

Mechanisms by which SCFAs modulate cardiac function and physiology in humans still need to be further investigated. However, the measurement of SCFA levels in peripheral blood can be a potential marker for gut microbiota dysfunctions, the intestinal barrier's permeability, systemic inflammation, and predisposition to diseases, and can be used in clinical trials in the future.

## 6 SCFAs and brain

One of the communication mechanisms between the brain and microbiota are SCFAs bacterial fermentation products<sup>34</sup>. SCFAs can directly affect the central nervous system (CNS) by acetate present in the bloodstream, which can cross the blood-brain barrier (BBB) and reduce appetite through a central homeostatic mechanism related to the expression profiles of appetite regulating neuropeptides in the hypothalamus via activation of the TCA cycle<sup>35</sup>.

There is new evidence demonstrating that the microbiota has critical consequences for the host's physiological processes and affects neurodevelopment, behavior, cognition, and synthesis of neurotransmitters<sup>36</sup>. There are GPR43-dependent effects on the central nervous system, including maturation and function of microglia, activation of macrophages, and the maintenance of their homeostasis<sup>3</sup>. Also, AGCC can interact with neuron cells by stimulating the autonomic and sympathetic nervous system through GPR41<sup>37</sup> and GPR43<sup>38</sup> signaling.

Despite the low concentrations in bloodstream, propionate and butyrate can affect peripheral organs indirectly by activating the hormonal and nervous systems, consequently generating an important signalization through neural circuits to increase insulin sensitivity and activating intestinal gluconeogenesis via gut-brain axis. As result, metabolic benefits in body weight and glucose control are enhanced<sup>8</sup>.

The enteric nervous system (ENS) communicates bidirectionally with the brain by several mechanisms in addition to those mentioned above. One of them is through the vagus nerve, which sends sensory signals from gut to the nucleus of the solitary tract (NTS) in CNS.

Vagal afferents express receptors that are activated by SCFAs<sup>39</sup> and activation of this pathway is associated with glucose homeostasis<sup>40</sup>. Butyrate has also been shown to affect ENS and intestinal motility<sup>41</sup>.

Besides, it is estimated that 90% of the human body's serotonin (5-HT) is produced by enterochromaffin cells (EC) in the digestive tract<sup>42</sup>, and the functioning of these cells is known to be modulated by changes in the gut microbiome. One of the pathways in which microbiome disruption affects 5-HT production is through SCFAs; particularly butyrate and propionate have demonstrated the capacity to influence the synthesis of the enzyme tryptophan hydroxylase (a limiting enzyme for serotonin synthesis), aiding the synthesis of 5-HT by EC cells<sup>43</sup>. Recent studies demonstrate that gut microbiota influences the gut-brain axis and shapes symptoms associated to stress, such as anxiety and pain tolerance<sup>44</sup>. Some data suggests a lower visceral perception induced by butyrate via an increased release of 5-HT<sup>45</sup>, which can reduce discomfort in several intestinal diseases.

SCFAs are associated with the pathogenesis of several neurodegenerative diseases<sup>46</sup>. Many psychiatric diseases are associated with inflammation and an exacerbated immune response observed by the high level of cytokines<sup>47</sup>. This can be a result, among other causes, of a lower degree of SCFAs production. The interaction of the gut with environmental risk factors for psychiatric diseases, such as diet and stress (especially in early life) suggests that interventions targeting the gut microbiota could ameliorate psychiatric and neurodegenerative symptoms, enhancing the quality of life of the host.

We can also mention the association of SCFAs with the development of autism<sup>48</sup> and alterations in levels of fecal SCFAs seen in patients with Parkinson's Disease<sup>49</sup>. Some of the mechanisms of SCFAs signaling include the regulation of HDAC that can be part of the regulation of hypothalamus-pituitary-adrenal (HPA) axis, which has high association with stress, major depression and anxiety<sup>50</sup>.

If in one hand we already have evidence of the effects of SCFAs on CNS and its correlation with psychiatric and neurodegenerative diseases, on other hand there is still a long path to go to understand in depth the mechanisms underlying this signaling process and how we can use it for human benefit.

## **7 Conclusions and Prospective**

In the last few decades, the pace at which new studies on the human microbiota have emerged has increased significantly, revealing the most diverse pathways in which these

microorganisms are capable of impacting our lives. Currently, it is evident that the microbiota is a determining factor for health and disease, having a pivotal role in the regulation of human physiology, and SCFAs have a remarkable prominence as one of the primary mechanisms in which the host-microbiota interaction occurs.

It is already known the central role of SCFAs for maintenance intestinal epithelium homeostasis and being an essential source of energy and substrate for gluconeogenesis. Then, the SCFAs that are absorbed in the gut enter the bloodstream and migrate towards the portal vein, and reaching the liver, where part of them is also metabolized. From that point forward, SCFAs are transported to other peripheral tissues and can even reach the brain. In this process, SCFAs can influence the host physiology directly or indirectly and could be associated with the development of pathologies (or have a protective role against them).

It is still not entirely clear whether SCFAs induces beneficial effects by itself or if it is a consequence of combining them and other microbial metabolites in specific proportions and exact concentrations. However, the perception that gut microbiota can contribute to human health and the susceptibility to diseases promotes a new and broader perspective on understanding human pathologies.

The presence of SCFAs in venous and arterial blood, in addition to stool samples, allows the possibility for the dosage of these substances to be used as a prediction or diagnosis markers of diseases. However, it is still necessary to understand more about the potential that SCFAs have as biochemical markers.

As we acquire more profound knowledge about the microbiota, SCFAs, and their interactions with the host, we increasingly revealing potential therapeutic targets. Nevertheless, there are many gaps about SCFAs and their mechanisms to be revealed. These gaps should be targeted for future research to have therapeutic and preventive strategies based on microbiota one day. We face the challenge of identifying the exact role of SCFAs in the (patho)physiology of the host and need to acquire more knowledge about its mechanisms to use it in clinical practice.

### **Acknowledgements**

We thank our main affiliation with the Biochemistry Department of Federal University of Rio Grande do Sul and the Center of Studies on Oxidative Stress.



### Conflict of interest

The authors declare no conflict of interest.

### References

- [1] Forsythe P, Kunze WA. Voices from within: gut microbes and the CNS. *Cell Mol Life Sci* 2013;70:55-69. doi: 10.1007/s00018-012-1028-z.
- [2] Cummings JH, Pomare EW, Branch WJ, Naylor CP, Macfarlane GT. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut* 1987;28:1221-1227. doi: 10.1136/gut.28.10.1221.
- [3] Rooks MG, Garrett WS. Gut microbiota, metabolites and host immunity. *Nat Rev Immunol* 2016;16:341-352. doi:10.1038/nri.2016.42.
- [4] Kunzelmann K, Mall M. Electrolyte transport in the mammalian colon: mechanisms and implications for disease. *Physiol Rev* 2002;82:245-289. doi: 10.1152/physrev.00026.2001.
- [5] Donohoe DR *et al.* The Warburg effect dictates the mechanism of butyrate-mediated histone acetylation and cell proliferation. *Mol Cell* 2012;48:612-626. doi: 10.1016/j.molcel.2012.08.033.
- [6] Zeng X *et al.* Induction of porcine host defense peptide gene expression by short-chain fatty acids and their analogs. *PLoS ONE* 2013;8:e72922. doi: 10.1371/journal.pone.0072922.
- [7] Furusawa Y *et al.* Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 2013;504:446-450. doi: 10.1038/nature12721.
- [8] De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchampt A, Bäckhed F, Mithieux G. Microbiota-generated metabolites promote metabolic benefits via gut-brain neural circuits. *Cell* 2014;156:84-96. doi: 10.1016/j.cell.2013.12.016.
- [9] Fukuda S *et al.* Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature* 2011;469:543-547. doi: 10.1038/nature09646
- [10] Camilleri M, Madsen K, Spiller R, Greenwood-Van Meerveld B, Verne GN. Intestinal barrier function in health and gastrointestinal disease. *Neurogastroenterol Motil* 2012;2:503-512. doi: 10.1111/j.1365-2982.2012.01921.x.
- [11] Pedersen HK, Gudmundsdottir V, Nielsen HB *et al.* Human gut microbes impact host serum metabolome and insulin sensitivity. *Nature* 2016;535(7612):376-381. doi: 10.1038/nature18646.

[12] Willemsen LE, Koetsier MA, van Deventer SJ, van Tol EA. Short chain fatty acids stimulate epithelial mucin 2 expression through differential effects on prostaglandin E(1) and E(2) production by intestinal myofibroblast. *Gut* 2003;52:1442-1447. doi: 10.1136/gut.52.10.1442.

[13] Plöger S, Stumpff F, Penner GB, Schulzke J, Gäbel G, Martens H, Shen Z, Günzel D, Aschenbach JR. Microbial butyrate and its role for barrier function in the gastrointestinal tract. *Ann NY Acad Sci* 2012;1258:52-59. doi: 10.1111/i.1749-6632.2012.06553.x.

[14] Lewis K, Lutgendorff F, Phan V, Söderholm JD, Sherman PM, McKay DM. Enhanced translocation of bacteria across metabolically stressed epithelia is reduced by butyrate. *Inflamm Bowel Dis* 2010;16:1138-1148. doi: 10.1002/ibd.21177.

[15] Wang T *et al.* Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. *ISME J* 2012;6:320-329. doi: 10.1038/ismej.2011.109.

[16] Scheppach W, Weiler F. The butyrate story: old wine in new bottles? *Curr Opin Clin Nutr Metab Care* 2004;7:563-567. doi: 10.1097/00075197-200409000-00009.

[17] Canani RB, Di Costanzo M, Leone L, Pedata M, Meli R, Calignano A. Potential beneficial effects of butyrate in intestinal and extraintestinal diseases. *World J Gastroenterol* 2011;17:1519. doi: 10.3748/wjg.v17.i12.1519.

[18] Thangaraju M, Cresci GA, Liu K, Ananth S, Gnanaprakasam JP, Browning DD, Mellinger JD, Smith SB, Digby GJ, Lambert NA, *et al.* GPR109A is a G-protein-coupled receptor for the bacterial fermentation product butyrate and functions as a tumor suppressor in colon. *Cancer Res.* 2009;69:2826–2832. doi: 10.1158/0008-5472.CAN-08-4466.

[19] Inan MS, Rasoulpour RJ, Yin L, Hubbard AK, Rosenberg DW, Giardina C. The luminal short-chain fatty acid butyrate modulates NF-kappaB activity in a human colonic epithelial cell line. *Gastroenterology.* 2000;118:724–734. doi: 10.1016/s0016-5085(00)70142-9.

[20] Segain JP, Raingeard de la Blétière D, Bourreille A, Leray V, Gervois N, Rosales C, Ferrier L, Bonnet C, Blottière HM, Galmiche JP. Butyrate inhibits inflammatory responses through NFkappaB inhibition: implications for Crohn's disease. *Gut.* 2000;47:397–403. doi: 10.1136/gut.47.3.397.

[21] Tarao K *et al.* Relationship between endotoxaemia and protein concentration of ascites in cirrhotic patients. *Gut* 1979;20:205-210. doi: 10.1136/gut.20.3.205.

[22] Donohoe DR, Garge N, Zhang X, Sun W, O'Connell TM, Bunger MK, Bultman SJ. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. *Cell Metab.* 2011;13:517–526. doi: 10.1016/j.cmet.2011.02.018.

- [23] Qin J *et al.* A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* 2012;490:55-60. doi: 10.1038/nature11450.
- [24] Cresci GA *et al.* Prophylactic tributyrin treatment mitigates chronic-binge alcohol-induced intestinal barrier and liver injury. *J Gastroenterol Hepatol* 2017;32:1587-97. doi: 10.1111/jgh.13731.
- [25] Kummén M *et al.* Gut microbiota signature in heart failure defined from profiling of 2 independent cohorts. *J Am Coll Cardiol* 2018;71:1184-86. doi: 10.1016/j.jacc.2017.12.057.
- [26] Sandek A *et al.* Altered intestinal function in patients with chronic heart failure. *J Am Coll Cardiol* 2007;50:1561-69. doi: 10.1016/j.jacc.2007.07.016.
- [27] Yang T *et al.* Gut dysbiosis is linked to hypertension. *Hypertension* 2015;65:1331-40. doi: 10.1161/HYPERTENSIONAHA.115.05315.
- [28] Wilck N., Matus M.G., Kearney S.M., Olesen S.W., Forslund K., Bartolomeus H., Haase S., Mähler A., Balogh A., Markó L., *et al.* Salt-responsive gut commensal modulates TH17 axis and disease. *Nature*. 2017;551:585–589. doi: 10.1038/nature24628.
- [29] Pluznick JL, Protzko RJ, Gevorgyan H, Peterlin Z, Sipos A, Han J, Brunet I, Wan LX, Rey F, Wang T, Firestein SJ, Yanagisawa M, Gordon JI, Eichmann A, Peti-Peterdi J, Caplan MJ. Olfactory receptor responding to gut microbiota-derived signals plays a role in renin secretion and blood pressure regulation. *Proc Natl Acad Sci USA*. 2013; 110:4410–4415. doi: 10.1073/pnas.1215927110.
- [30] Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. *Cell*, 2016; 165: 1332-1345. doi: 10.1016/j.cell.2016.05.041.
- [31] Karlsson FH, *et al.* Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat Commun*. 2012;3:1245. doi: 10.1038/ncomms2266.
- [32] Kamo, T *et al.* Dysbiosis and compositional alterations with aging in the gut microbiota of patients with heart failure. *PLOS ONE* 2017;12:e0174099. doi: 10.1371/journal.pone.0174099.
- [33] Engels C, Ruscheweyh H J, Beerenwinkel N, Lacroix C, Schwab C. The common gut microbe *Eubacterium hallii* also contributes to intestinal propionate formation. *Front. Microbiol*. 2016;7:713. doi: 10.3389/fmicb.2016.00713.
- [34] Friedrich, MJ. Unraveling the influence of gut microbes on the mind. *JAMA* 2015;313:1699–1701, <https://doi.org/10.1001/jama.2015.2159>.
- [35] Frost G, Sleeth ML, Sahuri-Arisoylu M, Lizarbe B, Cerdan S, Brody L, Anastasovska J, Ghourab S, Hankir M, Zhang S, *et al.* The short-chain fatty acid acetate reduces

appetite via a central homeostatic mechanism. *Nat. Commun.* 2014;5:3611. doi: 10.1038/ncomms4611.

[36] Erny D, *et al.* Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci.* 2015;18(7):965–77. doi: 10.1038/nn.4020.

[37] Kimura I, Inoue D, Maeda T, Hara T, Ichimura A, Miyauchi S *et al.* Short-chain fatty acids and ketones directly regulate sympathetic nervous system via G proteincoupled receptor 41 (GPR41). *Proc Natl Acad Sci USA* 2011; 108: 8030–8035. doi: 10.1073/pnas.1016088108.

[38] Nohr MK, Pedersen MH, Gille A, Egerod KL, Engelstoft MS, Husted AS *et al.* GPR41/FFAR3 and GPR43/FFAR2 as cosensors for short-chain fatty acids in enteroendocrine cells vs FFAR3 in enteric neurons and FFAR2 in enteric leukocytes. *Endocrinology* 2013; 154: 3552–3564. doi: 10.1210/em.2013-1142.

[39] Lal, S., Kirkup, A. J., Brunnsden, A. M., Thompson, D. G. & Grundy, D. Vagal afferent responses to fatty acids of different chain length in the rat. *Am. J. Physiol. Gastrointest. Liver Physiol.* 2001;281:G907–G915. doi: 10.1152/ajpgi.2001.281.4.G907.

[40] Zadeh-Tahmasebi, M. *et al.* Activation of short and long chain fatty acid sensing machinery in the ileum lowers glucose production in vivo. *J. Biol. Chem.* 2016;291:8816–8824. doi: 10.1074/jbc.M116.718460.

[41] Soret R, Chevalier J, De Coppet P, Poupeau G, Derkinderen P, Segain JP, Neunlist M. Short-chain fatty acids regulate the enteric neurons and control gastrointestinal motility in rats. *Gastroenterology.* 2010;138:1772–1782. doi: 10.1053/j.gastro.2010.01.053.

[42] Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. *Curr Opin Endocrinol Diab Obes.* 2013;20(1):14–21. doi: 10.1097/MED.0b013e32835bc703.

[43] Silva YP, Bernardi A, Frozza RL. The role of short-chain fatty acids from gut microbiota in gut-brain communication. *Front Endocrinol.* 2020;11:25. doi: 10.3389/fendo.2020.00025.

[44] Cryan JF, O’Mahony SM. The microbiome-gut-brain axis: from bowel to behavior. *Neurogastroenterol Motil* 2011;23:187-192. doi: 10.1111/j.1365-2982.2010.01664.x.

[45] Kilkens TO, Honig A, van Nieuwenhoven MA, Riedel WJ, Brummer RJ. Acute tryptophan depletion affects brain-gut responses in irritable bowel syndrome patients and controls. *Gut.* 2004;53:1794–1800. doi: 10.1136/gut.2004.041657.

[46] Bourassa MW, Alim I, Bultman SJ, Ratan RR. Butyrate, neuroepigenetics and the gut microbiome: Can a high fiber diet improve brain health? *Neurosci Lett* 2016;625:56-63. doi: 10.1016/j.neulet.2016.02.009.

[47] Bauer ME, Teixeira AL. Inflammation in psychiatric disorders: what comes first? *Ann N Y Acad Sci* 2019;1437(1):57-67. doi: 10.1111/nyas.13712.

[48] de Theije CG, Wopereis H, Ramadan M, van Eijndthoven T, Lambert J, Knol J *et al.* Altered gut microbiota and activity in a murine model of autism spectrum disorders. *Brain Behav Immun* 2014; 37:197-206. doi: 10.1016/j.bbi.2013.12.005.

[49] Unger M *et al.* Short chain fatty acids and gut microbiota differ between patients with Parkinson's disease and age-matched controls. *Parkinsonism Relat Disord* 2016;32:66-72. doi: 10.1016/j.parkreldis.2016.08.019.

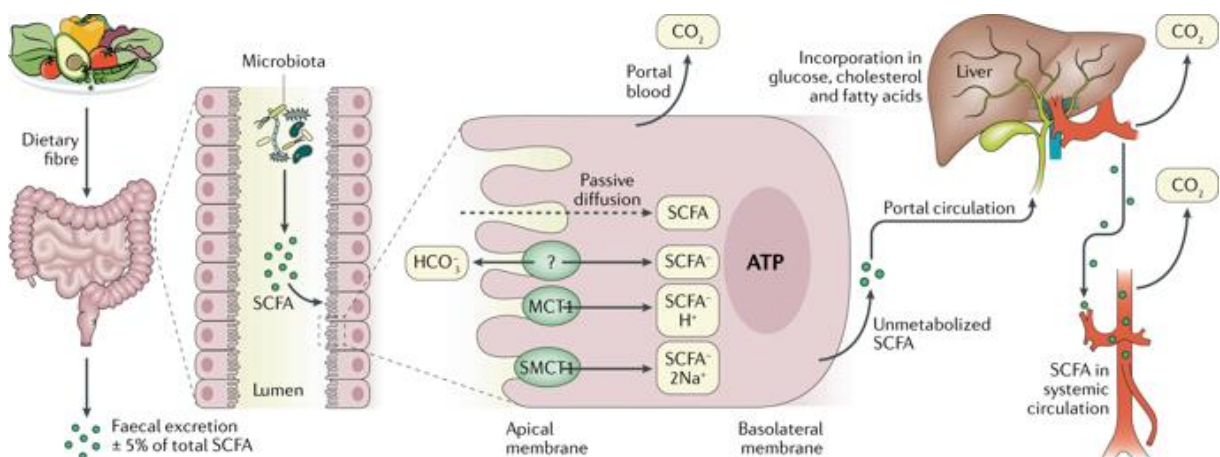
[50] Stilling RM, Dinan TG & Cryan JF. Microbial genes, brain & behaviour - epigenetic regulation of the gut-brain axis. *Genes Brain Behav* 2014;13:69–86. doi: 10.1111/gbb.12109.

## 5 CONCLUSÕES E PERSPECTIVAS

Nas últimas décadas o ritmo em que surgem novos estudos sobre a microbiota humana aumentou de forma significativa, revelando as mais diversas formas pelas quais esses microorganismos são capazes de impactar nossas vidas. Isso só foi obtido graças principalmente a estudos clínicos e translacionais de ponta. Atualmente é evidente que a microbiota é um fator determinante para a saúde e doença do hospedeiro, sendo uma peça-chave para a regulação da fisiologia humana, sendo que os AGCC possuem um grande destaque como um dos mecanismos de interação entre microbiota e hospedeiro.

Já se sabe da grande importância dos AGCC para o epitélio intestinal e manutenção da barreira, além de servirem como fonte de energia e substrato para a gliconeogênese intestinal. Os AGCC que são absorvidos entram na corrente sanguínea e migram em direção à veia porta, atingindo o fígado, onde parte também é metabolizada. A partir desse ponto, os AGCC são distribuídos para os demais tecidos do corpo, atingindo inclusive o cérebro e podendo alterar a fisiologia do hospedeiro de forma direta ou indireta e ter associação com o desenvolvimento de patologias. Um esquema representativo da rota dos AGCC no organismo do hospedeiro pode ser visto na Figura 3.

**Figura 3 – Esquema representativo da rota dos AGCC no corpo humano**



Fonte: Dalile *et al.*, 2019.

Ainda não está totalmente elucidado se os AGCC induzem efeitos benéficos de forma individual ou se seria uma combinação entre eles e outros metabólitos microbianos numa proporção específica, e quais as concentrações exatas para que esses efeitos sejam promovidos.

A percepção de que a microbiota intestinal pode contribuir tanto para a saúde humana quanto para a suscetibilidade de doenças promove uma nova e mais ampla perspectiva sobre o entendimento das doenças.

A presença dos AGCC no sangue venoso e arterial e em amostras de fezes nos permite considerar a possibilidade de que a dosagem dessa substância possa ser utilizada como predição ou diagnóstico de doenças. Entretanto, ainda é necessário um melhor entendimento do seu potencial como marcadores bioquímicos.

Esses, entre outros motivos, fizeram com que eu tivesse um grande interesse no estudo dos AGCC e, no laboratório no qual foi realizado o Estágio em Pesquisa que tornou possível realizar este Trabalho de Conclusão de Curso, me envolvi no desenvolvimento de um projeto que investiga o papel dos AGCC no sistema nervoso. O trabalho teve como foco o estudo da Depressão Maior em modelo animal com alto potencial translacional. Este trabalho está em andamento e possivelmente trará dados muito relevantes para a área.

Ao passo em que vamos adquirindo um conhecimento mais profundo acerca da microbiota, os AGCC e sua interação com o hospedeiro, revelamos novos alvos terapêuticos em potencial. Entretanto, ainda existem muitas lacunas acerca dos AGCC e seus mecanismos de ação, que devem ser alvo de futuras pesquisas e para que um dia o desenvolvimento de estratégias terapêuticas e preventivas baseada na microbiota seja possível.

Temos pela frente o desafio de identificar o papel exato dos AGCC na (pato)fisiologia do hospedeiro e pontuar precisamente os mecanismos pelos quais eles agem: um conhecimento que pode ser extremamente benéfico para a saúde humana e passível de ser utilizado na clínica.

## REFERÊNCIAS

AHMED, Kashan; TUNARU, Sorin; OFFERMANN, Stefan. GPR109A, GPR109B and GPR81, a family of hydroxyl-carboxylic acid receptors. **Trends in Pharmacological Sciences**, Cambridge, v. 30, n. 11, p. 557-562, Nov. 2009.

ARPAIA, Nicholas *et al.* Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. **Nature**, London, v. 404, p. 451-455, Nov 2013.

BERCIK, Premysl *et al.* The anxiolytic effect of *Bifidobacterium longum* NCC3001 involves vagal pathways for gut-brain communication. **Neurogastroenterology and motility: the official journal of the European Gastrointestinal Motility Society**, Leuven, v. 23, p. 1132-1139, Dec. 2011.

Den BESTEN, Gijs *et al.* The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. **Journal of Lipid Research**, Rockville, v. 54, p. 2325-2340, Sep. 2013.

BLOEMEN, Johanne *et al.* Short chain fatty acids exchange across the gut and liver in humans measured at surgery. **Clinical Nutrition**, Edinburgh, v. 28, n. 6, p. 657-661, Dec. 2009.

BRAVO, Javier *et al.* Ingestion of *Lactobacillus* strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. **Proceedings of the National Academy of Sciences of the United States of America**, Raleigh, v. 108, n. 38, p. 16050-16055, Sep. 2011.

BROWN, Andrew *et al.* The orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. **The Journal of Biological Chemistry**, Rockville, v. 278, n. 13, p. 11312-9, Mar. 2003.

CHAMBERS, Edward *et al.* Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. **Gut**, London, v. 64, n. 11, p. 1744-1754, Oct. 2015.

CHANG, Pamela *et al.* The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition **Proceedings of the National Academy of Sciences of the United States of America**, Raleigh, v. 111, n. 6, p. 2247-2252, Feb. 2014.



CLAESSION, Marcus *et al.* Gut microbiota composition correlates with diet and health in the elderly. **Nature**, London, v. 488, p. 178-184, July 2012.

CLAUSEN, Morten Rahr; MORTENSEN, Peter. Kinetic studies on colonocyte metabolism of short chain fatty acids and glucose in ulcerative colitis. **Gut**, London, v. 37, n. 5, p. 684-689, Nov. 1995.

CLEMENTE, Jose *et al.* The impact of the gut microbiota on human health: an integrative view. **Cell**, Cambridge, v. 148, n. 6, p. 1258-1270, Mar. 2012.

CLEOPHAS, Maartje *et al.* Suppression of monosodium urate crystal-induced cytokine production by butyrate is mediated by the inhibition of class I histone deacetylases. **Annals of the Rheumatic Diseases**, Kilchberg, v. 75, n. 3, p. 593-600, Mar. 2016.

COMPARE, Debora *et al.* The gut bacteria-driven obesity development. **Digestive Diseases**, Basel, v. 34, n. 3, p. 221-9, Mar. 2016.

CUMMINGS, John; MACFARLANE, George. The control and consequences of bacterial fermentation in the human colon. **The journal of applied bacteriology**, London, v. 70, n. 6, p. 443-459, Jun. 1991.

CUMMINGS, John *et al.* Short Chain fatty acids in human large intestine, portal, hepatic and venous blood. **Gut**, London, v. 28, n. 10, p. 1221-7, Oct. 1987.

DAVIE, James. Inhibition of histone deacetylase activity by butyrate. **The Journal of Nutrition**, Oxford, v. 133, suppl 7, p. 2485S-2493S, Jul. 2003.

DINAN, Timothy; CRYAN, John. Gut-brain axis in 2016: Brain-gut-microbiota axis – mood, metabolism and behavior. **Nature Reviews Gastroenterology and Hepatology**, London, v. 14, n. 2, p. 69-70, Feb. 2017.

DUNCAN, Sylvia *et al.* Acetate utilization and butyryl coenzyme A (CoA):acetate-CoA transferase in butyrate-producing bacteria from the human large intestine. **Applied and Environmental Microbiology**, Washington, v. 68, n. 10, p. 5186-5190, Oct. 2002.

ECKBURG, Paul *et al.* Diversity of the human intestinal microbial flora. **Science**, Washington, v. 308, n. 5728, p. 1635-8, Jun. 2005.

EVANS, James; MORRIS, Laura; MARCHESI, Julian. The gut microbiome: The role of a virtual organ in the endocrinology of the host. **Journal of Endocrinology**, Cambridge, v. 218, n. 3, p. R37-47, Aug. 2013.

EVENEPOEL, Peter; POESEN, Ruben; MEIJERS, Björn. The gut-kidney axis. **Pediatric Nephrology**, Kansas, v. 32, n. 11, p. 2005-2014, Nov. 2017.

FELLOWS, Rachel *et al.* Microbiota derived short chain fatty acids promote histone crotonulation in the colon through histone deacetylases. **Nature Communications**, London, v. 9, p. 105, 2018, Jan. 2018.

FORSHYTE, Paul; KUNZE, Wolfgang. Voices from within: gut microbes and the CNS. **Cellular and Molecular Life Sciences**, Basel, v. 70, n. 1, p. 55-69, Jan. 2013.

FRANK, Daniel; PACE, Norman. Gastrointestinal microbiology enters the metagenomics era. **Current Opinion in Gastroenterology**, Philadelphia, v. 24, n. 1, p. 4-10, Jan. 2008.

FURUSAWA, Yukihiro *et al.* Commensal microbiobe-derived butyrate induces the differentiation of colonic regulatory T cells. **Nature**, London, v. 504, n. 7480, p. 446-450, Dec. 2013.

GANAPATHY, Vadivel *et al.* Transporters and receptors for short-chain fatty acids as the molecular link between colonic bacteria and the host. **Current opinion in pharmacology**, Amsterdam, v. 13, n. 6, p. 869-874, Dec. 2013.

GELIS, Lian *et al.* Functional characterization of the odorant receptor 51E2 in human melanocytes. **Journal of Biological Chemistry**, Rockville, v. 291, n. 34, p. 17772-786, Aug. 2016.

HALESTRAP, Andrew; MEREDITH, David. The SLC16 gene family-from monocarboxylate transporters (MCTs) to aromatic amino acid transporters and beyond. **Pflügers Archiv**, New York, v. 447, n. 5, p. 619-628, Geb. 2004.

Van der HEE, Bart; WELLS, Jerry. Microbial Regulation of Host Physiology by Short-chain Fatty Acids. **Trends in Microbiology**, Cambridge, Mar. 2021.

HETZEL, Marc *et al.* Acryloyl-CoA reductase from *Clostridium propionicum*. **European Journal of Biochemistry**, New Jersey, v. 270, n. 5, p. 902-910, Mar. 2003.

HUMAN MICROBIOME PROJECT CONSORTIUM. Structure, function and diversity of the healthy human microbiome. **Nature**, London, v. 486, p. 207-214, June 2012.

INOUE, Daisuke *et al.* Short-chain fatty acid receptor GPR41-mediated activation of sympathetic neurons involves synapsin 2b phosphorylation. **FEBS Letter**, New Jersey, v. 586, n. 10, p. 1547-1554, May 2012.

JOSEFSDOTTIR, Kamilla *et al.* Antibiotic impair murine hematopoiesis by depleting the intestinal microbiota. **Blood**, Washington, v. 129, n. 6, p. 729-739, Feb. 2017.

KAJI, Izumi; KARAKI, Shin-ichiro; KUWAHARA, Atsukazu. Short-chain fatty acid receptor and its contribution to glucagon-like peptide-1 release. **Digestion**, Göttingen, v. 89, n. 1, p. 31-36, Jan. 2014.

KARBACH, Susanne *et al.* Gut microbiota promote angiotensin II-induced arterial hypertension and vascular dysfunction. **Journal of the American Heart Association**, Dallas, v. 5, n. 9, p. e003698, Aug. 2016.

KARLSSON, Fredrik *et al.* Symptomatic atherosclerosis is associated with an altered gut metagenome. **Nature Communications**, London, v. 3, n. 1245, Dec. 2012.

KIMURA, Ikuo *et al.* Short-chain fatty acids and ketones directly regulate sympathetic nervous system via G protein-coupled receptor 41 (GPR41). **Proceedings of the National Academy of Sciences of the United States of America**, Raleigh, v. 108, n. 19, p. 8030-35, May 2011.

KOH, Ara *et al.* From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. **Cell**, Cambridge, v. 165, n. 6, p. 1332-1345, Jun. 2019.

KONTUREK, Peter *et al.* Emerging role of fecal microbiota therapy in the treatment of gastrointestinal and extra-gastrointestinal diseases. **Journal of Physiology and Pharmacology**, Kraków, v. 66, n. 4, p. 483-491, Aug. 2015.

LE POUL, Emmanuel *et al.* Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. **Journal of Biological Chemistry**, Rockville, v. 278, n. 28, p. 25481-9, Jul. 2003.

LI, Junhua *et al.* An integrated catalog of reference genes in the human gut microbiome. **Nature Biotechnology**, London, v. 32, p. 824-841 July 2014.

LOUIS, Petra *et al.* Restricted Distribution of the Butyrate Kinase Pathway among Butyrate-Producing Bacteria from the Human Colon. **Journal of Bacteriology**, Washington, v. 186, n. 7, p. 2099-2106, Apr. 2004.

LOUIS, Petra; HOLD, Georgina; FLINT, Harry. The gut microbiota, bacterial metabolites and colorectal cancer. **Nature Reviews on Microbiology**, London, v. 12, p. 661-672, Sep. 2014.

LUCKEY, Thomas. Introduction to intestinal microecology. **American Journal of Clinical Nutrition**, Rockville, v. 25, n. 12, p. 1292-94, Dec. 1972.

MACFARLANE, George; MACFARLANE, Sandra. Bacteria, colonic fermentation, and gastrointestinal health. **Journal of AOAC International**, Oxford, v. 95, n. 1, p. 50-60, Jan./Feb. 2012.

MARQUES, Tatiana Milena *et al.* Programming infant gut microbiota: influence of dietary and environmental factors. **Current Opinion in Biotechnology**, Amsterdam, v. 21, n. 2, p. 149-156, Apr. 2010.

MASLOWSKI, Kendle *et al.* Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. **Nature**, London, v. 461, p. 1282-86, Oct. 2009.

MEIJER, Kees; De VOS, Paul; PRIEBE, Marion. Butyrate and other short-chain fatty acids as modulators of immunity: what relevance for health? **Current Opinion in Clinical Nutritional Metabolic Care**, Amsterdam, v. 13, n. 6, p. 715-721, Nov. 2010.

PARENT, André; CARPENTER, Malcolm. **Carpenter's Human Neuroanatomy**. Baltimore, MD: Williams & Wilkins. 1996.

PSICHAS, Arianna *et al.* The short chain fatty acid propionate stimulates GLP-1 and PYY secretion via free fatty acid receptor 2 in rodents. **International Journal of Obesity**, London, v. 39, n. 3, p. 424-429, Mar. 2015.

PLUZNICK, Jennifer *et al.* Olfactory receptor responding to gut microbiota-derived signals plays a role in renin secretion and blood pressure regulation. **Proceedings of the National Academy of Sciences of the United States of America**, Raleigh, v. 110, n. 11, p. 4410-15, Mar. 2013.

PRIORI, Davide *et al.* The Olfactory Receptor OR51E1 Is Present along the Gastrointestinal Tract of Pigs, Co-Localizes with Enteroendocrine Cells and Is Modulated by Intestinal Microbiota. **PLoS One**, v. 10, n. 6, p. e0129501, Jun. 2015.

RAGSDALE, Stephen; PIERCE, Elizabeth. Acetogenesis and the Wood-Ljungdahl Pathway of CO<sub>2</sub> Fixation. **Biochimica et Biophysica Acta**, Amsterdam, v. 1784, n. 12, p. 1873-1898, Dec. 2008.

REMELY, Marlene *et al.* Effects of short chain fatty acid producing bacteria on epigenetic regulation of FFAR3 in type 2 diabetes and obesity. **Gene**, Amsterdam, v. 537, n. 1, p. 85-92, Mar. 2014.

REY, Federico *et al.* Dissection the in vivo metabolic potential of two human gut acetogens. **Journal of Biological Chemistry**, Amsterdam, v. 285, n. 29, p. 22082-90, Jul. 2010.

ROOKS, Michelle; GARRETT, Wendy. Gut microbiota, metabolites and host immunity. **Nature Reviews on Immunology**, London, v. 16, p. 341-352, May 2016.

ROTHER, Monique; BLAUT, Michael. Evolution of the gut microbiota and the influence of diet. **Beneficial Microbes**, Wageningen, v. 4, n. 1, p. 31-7, Mar. 2013.

SCOTT, Karen *et al.* Whole-genome transcription profiling reveals genes upregulated by growth on fucose in the human gut bacterium "Roseburia inulinivorans". **Journal of Bacteriology**, New York, v. 188, n. 12, p. 4340-4349, Jun. 2006.

SINGH, Nagendra *et al.* Activation of Gpr109a, receptor for niacina and the comensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. **Immunity**, Cambridge, v. 40, n. 1, p. 128-139, Jan. 2014.

TAPPENDEN, Kelly *et al.* Glucagon-like peptide-2 and short-chain fatty acids: a new twist to an old story. **Journal of Nutrition**, Oxford, v. 133, n. 11, p. 3717-3720, Nov. 2003.

TAZOE, Hideaki *et al.* Expression of short-chain fatty acid receptor GPR41 in the human colon. **Biomedical Research**, Shangai, v. 30, n. 3, p. 149-156, Jun. 2009.

TOLHURST, Gwen *et al.* Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein coupled receptor FFAR2. **Diabetes**, Arlington, v. 61, n. 2, p. 364-371, Feb. 2012.

TURNBAUGH, Peter; GORDON, Jeffrey. The core gut microbiome, energy balance and obesity. **Journal of Physiology**, San Francisco, v. 587, n. 17, p. 4153-4158, Sep. 2009.

TURNBAUGH, Peter *et al.* The human microbiome project. **Nature**, London, v. 449, p. 804-810, Oct. 2007.

VITAL, Marius; HOWE, Adina Chuang; TIEDJE, James. Revealing the Bacterial Butyrate Synthesis Pathway by Analyzing (Meta)genomic Data. **mBio**, v. 5, n. 2, p. e00889-14, Mar. 2014.

QIN, Junjie *et al.* MetaHIT Consortium. A human gut microbial gene catalogue established by metagenomic sequencing. **Nature**, London, n. 464, p. 59-65, Mar. 2010.

WALL, Rebecca *et al.* Metabolic activity of the enteric microbiota influences the fatty acid composition of murine and porcine liver and adipose tissues. **American Journal of Clinical Nutrition**, Oxford, v. 89, n.5, p. 1393-1401, May 2009.

WELLS, Jerry *et al.* Epithelial crosstalk at the microbiota-mucosal interface. **Proceedings of the National Academy of Sciences of the United States of America**, Raleigh, v. 108, suppl. 1, p. 4607-4614, Mar. 2011.

Van de WOUW, Marcel *et al.* Short-chain fatty acids: microbial metabolites that alleviate stress-induced brain-gut axis alterations. **Journal of Physiology**, San Francisco, v. 596, n. 20, p. 4923-44, Oct. 2018.

WU, Gary *et al.* Linking long-term dietary patterns with gut microbial enterotypes. **Science**, Washington, v. 334, n. 6052, p. 105-108, Oct. 2011.

YADAV, Hariom *et al.* Beneficial metabolic effects of a probiotic via butyrate-induced GLP-1 hormone secretion. **Journal of Biological Chemistry**, Rockville, v. 288, n. 35, p. 25088-25097, Aug. 2013.

ZHANG, Chenhong *et al.* Dietary modulation of gut microbiota contributes to alleviation of both genetic and simple obesity in children. **EBioMedicine**, Amsterdam, v. 2, n. 8, p. 968-84, Jul. 2015.

## ANEXO A – NORMAS DE PUBLICAÇÃO DA REVISTA HUMAN MICROBIOME JOURNAL

### **For a review article:**

Unstructured abstract (up to 250 words)

5-10 keywords

3000 words maximum

3 to 4 tables/figures maximum (optional colour)

50 references maximum

### **Ensure that the following items are present:**

One author has been designated as the corresponding author with contact details:

- E-mail address
- Full postal address

All necessary files have been uploaded:

#### *Manuscript:*

- Include keywords
- All figures (include relevant captions)
- All tables (including titles, description, footnotes)
- Ensure all figure and table citations in the text match the files provided
- Indicate clearly if color should be used for any figures in print

*Graphical Abstracts / Highlights files* (where applicable)

*Supplemental files* (where applicable)

#### Further considerations

- Manuscript has been ‘spell checked’ and ‘grammar checked’
- All references mentioned in the Reference List are cited in the text, and vice versa
- Permission has been obtained for use of copyrighted material from other sources (including the Internet)

- A competing interests statement is provided, even if the authors have no competing interests to declare
- Journal policies detailed in this guide have been reviewed
- Referee suggestions and contact details provided, based on journal requirements

### **Ethics in publishing**

The publication of an article in a peer-reviewed journal is an essential building block in the development of a coherent and respected network of knowledge. It is a direct reflection of the quality of the work of the authors and the institutions that support them. Peer-reviewed articles support and embody the scientific method. It is therefore important to agree upon standards of expected ethical behavior. Ethics topics to consider when publishing:

*Authorship of the paper:* Authorship should be limited to those who have made a significant contribution to the conception, design, execution, or interpretation of the reported study. Transparency about the contributions of authors is encouraged, for example in the form of a CRediT (Contributor Roles Taxonomy) author statement.

*Originality and plagiarism:* The authors should ensure that they have written entirely original works, and if the authors have used the work and/or words of others, that this has been appropriately cited or quoted.

*Data access and retention:* Authors may be asked to provide the raw data in connection with a paper for editorial review, and should be prepared to provide public access to such data.

*Multiple, redundant or concurrent publication:* An author should not in general publish manuscripts describing essentially the same research in more than one journal or primary publication. Elsevier does not view the following uses of a work as prior publication: publication in the form of an abstract; publication as an academic thesis; publication as an electronic preprint. Note: some society-owned titles and journals that operate double-blind review have different policies on prior publication. Information on prior publication is included within each Elsevier journal's guide for authors.

*Acknowledgement of sources:* Proper acknowledgment of the work of others must always be given.

*Disclosure and conflicts of interest:* All submissions must include disclosure of all relationships that could be viewed as presenting a potential conflict of interest.

*Fundamental errors in published works:* When an author discovers a significant error or inaccuracy in his/her own published work, it is the author's obligation to promptly notify the journal editor or publisher and cooperate with the editor to retract or correct the paper.



*Reporting standards:* Authors of reports of original research should present an accurate account of the work performed as well as an objective discussion of its significance.

*Hazards and human or animal subjects:* Statements of compliance are required if the work involves chemicals, procedures or equipment that have any unusual hazards inherent in their use, or if it involves the use of animal or human subjects.

*Use of patient images or case details:* Studies on patients or volunteers require ethics committee approval and informed consent, which should be documented in the paper.

### **Declaration of competing interest**

All authors must disclose any financial and personal relationships with other people or organizations that could inappropriately influence (bias) their work. Examples of potential conflicts of interest include employment, consultancies, stock ownership, honoraria, paid expert testimony, patent applications/registrations, and grants or other funding. Authors should create a declaration of competing interest statement using Elsevier Declaration of Interests Tool and upload to the submission system at the Attach Files step. This statement will be published within the article if accepted.

### **Submission declaration and verification**

Submission of an article implies that the work described has not been published previously (except in the form of an abstract, a published lecture or academic thesis), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright-holder. To verify originality, your article may be checked by the originality detection service Crossref Similarity Check.

### **Use of inclusive language**

Inclusive language acknowledges diversity, conveys respect to all people, is sensitive to differences, and promotes equal opportunities. Content should make no assumptions about the beliefs or commitments of any reader; contain nothing which might imply that one individual is superior to another on the grounds of age, gender, race, ethnicity, culture, sexual orientation, disability or health condition; and use inclusive language throughout. Authors should ensure that writing is free from bias, stereotypes, slang, reference to dominant culture and/or cultural

assumptions. We advise to seek gender neutrality by using plural nouns (“clinicians, patients/clients”) as default/wherever possible to avoid using “he, she”, or “he/she”. We recommend avoiding the use of descriptors that refer to personal attributes such as age, gender, race, ethnicity, culture, sexual orientation, disability or health condition unless they are relevant and valid. These guidelines are meant as a point of reference to help identify appropriate language but are by no means exhaustive or definitive.

### **Author contributions**

For transparency, we encourage author to submit an author statement file outlining their individual contributions to the paper using the relevant CRediT roles: Conceptualization; Data curation; Formal analysis; Funding acquisition; Investigations; Methodology; Project administration; Resources; Software; Supervision; Validation; Visualization; Roles/Writing – original draft; Writing – review & editing. Authorship statements should be formatted with the names of authors first and CRediT role(s) following.

### **Authorship**

All authors should have made substantial contributions to all of the following: (1) the conception and design of the study, or acquisition of data, or analysis and interpretation of data, (2) drafting the article or revising it critically for important intellectual content, (3) final approval of the version to be submitted.

### **Changes to authorship**

Authors are expected to consider carefully the list and order of authors before submitting their manuscript and provide the definitive list of authors at the time of the original submission. Any addition, deletion or rearrangement of author names in the authorship list should be made only before the manuscript has been accepted and only if approved by the journal Editor. To request such a change, the Editor must receive the following from the corresponding author: (a) the reason for the change in author list and (b) written confirmation (e-mail, letter) from all authors that they agree with the addition, removal or rearrangement. In the case of addition or removal of authors, this includes confirmation from the author being added or removed. Only in exceptional circumstances will the Editor consider the addition, deletion or rearrangement of authors after the manuscript has been accepted. While the Editor considers the request, publication of the manuscript will be suspended. If the manuscript has already been published in an online issue, any requests approved by the Editor will result in a corrigendum.

**Article transfer service**

This journal is part of our Article Transfer Service. This means that if the Editor feels your article is more suitable in one of our other participating journals, then you may be asked to consider transferring the article to one of those. If you agree, your article will be transferred automatically on your behalf with no need to reformat. Please note that your article will be reviewed again by the new journal.

**Copyright**

Upon acceptance of an article, authors will be asked to complete a 'License Agreement'. Permitted third party reuse of open access articles is determined by the author's choice of user license.

**Role of the funding source**

Upon acceptance of an article, authors will be asked to complete a 'License Agreement' (see more information on this). Permitted third party reuse of open access articles is determined by the author's choice of user license.

**Open access**

This journal is a peer reviewed, open access journal. All articles published open access will be immediately and permanently free for everyone to read, download, copy and distribute. As an open access journal with no subscription charges, a fee is payable by the author or research funder to cover the costs associated with publication. This ensures your article will be immediately and permanently free to access by everyone. The gold open access publication fee for this journal is USD 2400, excluding taxes. We automatically apply Article Publishing Charge waivers or discounts to those articles in gold open access journals for which all authors are based in a country eligible for the Research4Life program. Elsevier has established agreements with funding bodies, including Wellcome Trust and Research Councils UK. This ensures authors can comply with funding body open access policies and may also be reimbursed for their publication fees. For open access publishing this journal uses an exclusive licensing agreement. Authors will retain copyright alongside scholarly usage rights and Elsevier will be granted publishing and distribution rights.

**Elsevier Researcher Academy**

Researcher Academy is a free e-learning platform designed to support early and mid-career researchers throughout their research journey. The "Learn" environment at Researcher Academy offers several interactive modules, webinars, downloadable guides and resources to guide you through the process of writing for research and going through peer review. Feel free to use these free resources to improve your submission and navigate the publication process with ease.

### **Language** (usage and editing services)

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Authors who feel their English language manuscript may require editing to eliminate possible grammatical or spelling errors and to conform to correct scientific English may wish to use the English Language Editing service available from Elsevier's Author Services.

### **Submission**

Our online submission system guides you stepwise through the process of entering your article details and uploading your files. The system converts your article files to a single PDF file used in the peer-review process. Editable files (e.g., Word, LaTeX) are required to typeset your article for final publication. All correspondence, including notification of the Editor's decision and requests for revision, is sent by e-mail.

### **Use of word processing software**

It is important that the file be saved in the native format of the word processor used. The text should be in single-column format. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing the article. In particular, do not use the word processor's options to justify text or to hyphenate words. However, do use bold face, italics, subscripts, superscripts etc. When preparing tables, if you are using a table grid, use only one grid for each individual table and not a grid for each row. If no grid is used, use tabs, not spaces, to align columns. The electronic text should be prepared in a way very similar to that of conventional manuscripts. Note that source files of figures, tables and text graphics will be required whether or not you embed your figures in the text. To avoid unnecessary errors you are strongly advised to use the 'spell-check' and 'grammar-check' functions of your word processor.

## **Article structure**

### *Subdivision - numbered sections*

Divide your article into clearly defined and numbered sections. Subsections should be numbered 1.1 (then 1.1.1, 1.1.2, ...), 1.2, etc. (the abstract is not included in section numbering). Use this numbering also for internal cross-referencing: do not just refer to 'the text'. Any subsection may be given a brief heading. Each heading should appear on its own separate line.

## **Essential title page information**

- *Title*. Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.
- *Author names and affiliations*. Please clearly indicate the given name(s) and family name(s) of each author and check that all names are accurately spelled. You can add your name between parentheses in your own script behind the English transliteration. Present the authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lowercase superscript letter immediately after the author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name and, if available, the e-mail address of each author.
- *Corresponding author*. Clearly indicate who will handle correspondence at all stages of refereeing and publication, also post-publication. This responsibility includes answering any future queries about Methodology and Materials. Ensure that the e-mail address is given and that contact details are kept up to date by the corresponding author.
- *Present/permanent address*. If an author has moved since the work described in the article was done, or was visiting at the time, a 'Present address' (or 'Permanent address') may be indicated as a footnote to that author's name. The address at which the author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes.

## **Highlights**

Highlights are optional yet highly encouraged for this journal, as they increase the discoverability of your article via search engines. They consist of a short collection of bullet points that capture the novel results of your research as well as new methods that were used

during the study (if any). Highlights should be submitted in a separate editable file in the online submission system. Please use 'Highlights' in the file name and include 3 to 5 bullet points (maximum 85 characters, including spaces, per bullet point).

### **Graphical abstract**

Although a graphical abstract is optional, its use is encouraged as it draws more attention to the online article. The graphical abstract should summarize the contents of the article in a concise, pictorial form designed to capture the attention of a wide readership. Graphical abstracts should be submitted as a separate file in the online submission system. Image size: Please provide an image with a minimum of  $531 \times 1328$  pixels (h  $\times$  w) or proportionally more. The image should be readable at a size of  $5 \times 13$  cm using a regular screen resolution of 96 dpi. Preferred file types: TIFF, EPS, PDF or MS Office files. Authors can make use of Elsevier's Illustration Services to ensure the best presentation of their images and in accordance with all technical requirements.

### **Keywords**

Immediately after the abstract, provide a maximum of 6 keywords, using British spelling and avoiding general and plural terms and multiple concepts (avoid, for example, 'and', 'of'). Be sparing with abbreviations: only abbreviations firmly established in the field may be eligible. These keywords will be used for indexing purposes.

### **Acknowledgements**

Collate acknowledgements in a separate section at the end of the article before the references and do not, therefore, include them on the title page, as a footnote to the title or otherwise. List here those individuals who provided help during the research (e.g., providing language help, writing assistance or proof reading the article, etc.).

### **Electronic artwork**

#### *General points*

- Make sure you use uniform lettering and sizing of your original artwork.
- Embed the used fonts if the application provides that option.
- Aim to use the following fonts in your illustrations: Arial, Courier, Times New Roman, Symbol, or use fonts that look similar.

- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Provide captions to illustrations separately.
- Size the illustrations close to the desired dimensions of the published version.
- Submit each illustration as a separate file.
- Ensure that color images are accessible to all, including those with impaired color vision.

### *Formats*

If your electronic artwork is created in a Microsoft Office application (Word, PowerPoint, Excel) then please supply 'as is' in the native document format. Regardless of the application used other than Microsoft Office, when your electronic artwork is finalized, please 'Save as' or convert the images to one of the following formats (note the resolution requirements for line drawings, halftones, and line/halftone combinations given below): EPS (or PDF): Vector drawings, embed all used fonts. TIFF (or JPEG): Color or grayscale photographs (halftones), keep to a minimum of 300 dpi. TIFF (or JPEG): Bitmapped (pure black & white pixels) line drawings, keep to a minimum of 1000 dpi. TIFF (or JPEG): Combinations bitmapped line/half-tone (color or grayscale), keep to a minimum of 500 dpi. Please do not: (1) supply files that are optimized for screen use (e.g., GIF, BMP, PICT, WPG); these typically have a low number of pixels and limited set of colors, (2) supply files that are too low in resolution, or (3) submit graphics that are disproportionately large for the content.

### *Illustration services*

Elsevier's Author Services offers Illustration Services to authors preparing to submit a manuscript but concerned about the quality of the images accompanying their article. Elsevier's expert illustrators can produce scientific, technical and medical-style images, as well as a full range of charts, tables and graphs. Image 'polishing' is also available, where our illustrators take your image(s) and improve them to a professional standard.

### *Figure captions*

Ensure that each illustration has a caption. Supply captions separately, not attached to the figure. A caption should comprise a brief title (not on the figure itself) and a description of

the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

### **Tables**

Please submit tables as editable text and not as images. Tables can be placed either next to the relevant text in the article, or on separate page(s) at the end. Number tables consecutively in accordance with their appearance in the text and place any table notes below the table body. Be sparing in the use of tables and ensure that the data presented in them do not duplicate results described elsewhere in the article. Please avoid using vertical rules and shading in table cells.

### **References**

#### *Citation in text*

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these references are included in the reference list they should follow the standard reference style of the journal and should include a substitution of the publication date with either 'Unpublished results' or 'Personal communication'. Citation of a reference as 'in press' implies that the item has been accepted for publication.

#### *Data references*

This journal encourages you to cite underlying or relevant datasets in your manuscript by citing them in your text and including a data reference in your Reference List. Data references should include the following elements: author name(s), dataset title, data repository, version (where available), year, and global persistent identifier. Add [dataset] immediately before the reference so we can properly identify it as a data reference. The [dataset] identifier will not appear in your published article.

#### *References in a special issue*

Please ensure that the words 'this issue' are added to any references in the list (and any citations in the text) to other articles in the same Special Issue.

#### *Reference style*



*Text:* Indicate references by number(s) in square brackets in line with the text. The actual authors can be referred to, but the reference number(s) must always be given. List: Number the references (numbers in square brackets) in the list in the order in which they appear in the text.

*Examples:*

Reference to a journal publication:

[1] Van der Geer J, Hanraads JAJ, Lupton RA. The art of writing a scientific article. *J Sci Commun* 2010;163:51–9. <https://doi.org/10.1016/j.Sc.2010.00372>. Reference to a journal publication with an article number:

[2] Van der Geer J, Hanraads JAJ, Lupton RA. The art of writing a scientific article. *Heliyon*. 2018;19:e00205. <https://doi.org/10.1016/j.heliyon.2018.e00205> Reference to a book:

[3] Strunk Jr W, White EB. *The elements of style*. 4th ed. New York: Longman; 2000. Reference to a chapter in an edited book:

[4] Mettam GR, Adams LB. How to prepare an electronic version of your article. In: Jones BS, Smith RZ, editors. *Introduction to the electronic age*, New York: E-Publishing Inc; 2009, p. 281–304. Reference to a website:

[5] Cancer Research UK. Cancer statistics reports for the UK, <http://www.cancerresearchuk.org/aboutcancer/statistics/cancerstatsreport/>; 2003 [accessed 13 March 2003].

Reference to a dataset:

[dataset] [6] Oguro M, Imahiro S, Saito S, Nakashizuka T. Mortality data for Japanese oak wilt disease and surrounding forest compositions, Mendeley Data, v1; 2015. <https://doi.org/10.17632/xwj98nb39r.1>.

Note shortened form for last page number. e.g., 51–9, and that for more than 6 authors the first 6 should be listed followed by '*et al.*' For further details you are referred to 'Uniform Requirements for Manuscripts submitted to Biomedical Journals' (*J Am Med Assoc* 1997;277:927–34).

## **Video**

Elsevier accepts video material and animation sequences to support and enhance your scientific research. Authors who have video or animation files that they wish to submit with their article are strongly encouraged to include links to these within the body of the article. This can be done in the same way as a figure or table by referring to the video or animation content and noting in the body text where it should be placed. All submitted files should be properly labeled so that they directly relate to the video file's content. In order to ensure that your video

or animation material is directly usable, please provide the file in one of our recommended file formats with a preferred maximum size of 150 MB per file, 1 GB in total. Video and animation files supplied will be published online in the electronic version of your article in Elsevier Web products, including ScienceDirect. Please supply 'stills' with your files: you can choose any frame from the video or animation or make a separate image. These will be used instead of standard icons and will personalize the link to your video data. Note: since video and animation cannot be embedded in the print version of the journal, please provide text for both the electronic and the print version for the portions of the article that refer to this content.

### **Supplementary material**

Supplementary material such as applications, images and sound clips, can be published with your article to enhance it. Submitted supplementary items are published exactly as they are received (Excel or PowerPoint files will appear as such online). Please submit your material together with the article and supply a concise, descriptive caption for each supplementary file. If you wish to make changes to supplementary material during any stage of the process, please make sure to provide an updated file. Do not annotate any corrections on a previous version. Please switch off the 'Track Changes' option in Microsoft Office files as these will appear in the published version.

### **Research data**

This journal encourages and enables you to share data that supports your research publication where appropriate, and enables you to interlink the data with your published articles. Research data refers to the results of observations or experimentation that validate research findings. To facilitate reproducibility and data reuse, this journal also encourages you to share your software, code, models, algorithms, protocols, methods and other useful materials related to the project. Below are a number of ways in which you can associate data with your article or make a statement about the availability of your data when submitting your manuscript. If you are sharing data in one of these ways, you are encouraged to cite the data in your manuscript and reference list. Please refer to the "References" section for more information about data citation.

#### *Data linking*

If you have made your research data available in a data repository, you can link your article directly to the dataset. Elsevier collaborates with a number of repositories to link articles

on ScienceDirect with relevant repositories, giving readers access to underlying data that gives them a better understanding of the research described. There are different ways to link your datasets to your article. When available, you can directly link your dataset to your article by providing the relevant information in the submission system. For supported data repositories a repository banner will automatically appear next to your published article on ScienceDirect. In addition, you can link to relevant data or entities through identifiers within the text of your manuscript, using the following format: Database: xxxx (e.g., TAIR: AT1G01020; CCDC: 734053; PDB: 1XFN).

#### *Mendeley Data*

This journal supports Mendeley Data, enabling you to deposit any research data (including raw and processed data, video, code, software, algorithms, protocols, and methods) associated with your manuscript in a free-to-use, open access repository. During the submission process, after uploading your manuscript, you will have the opportunity to upload your relevant datasets directly to Mendeley Data. The datasets will be listed and directly accessible to readers next to your published article online.

#### *Data statement*

To foster transparency, we encourage you to state the availability of your data in your submission. This may be a requirement of your funding body or institution. If your data is unavailable to access or unsuitable to post, you will have the opportunity to indicate why during the submission process, for example by stating that the research data is confidential. The statement will appear with your published article on ScienceDirect.