Do microplastics mediate the effects of chemicals on aquatic

2	organisms?					
3	Danielle J. Marchant ¹ ,*, J. Iwan Jones ¹ , Gabriela Zemelka ¹ , Ozge Eyice ¹ ,					
4	Pavel Kratina ¹					
5						
6						
7	¹ School of Biological and Behavioural Sciences, Queen Mary University of London, Mile					
8	End Road, London E1 4NS, United Kingdom					
9	Pavel Kratina orcid: 0000-0002-9144-7937					
10						
11						
12	Running Head: Combined impacts of microplastic and chemical pollution					
13	Type of Article: Full Research Paper					
14	Abstract: 262 words Main text: 5495 words Figures: 5 References: 79					
15						
16						
17	*Corresponding Author:					
18	Danielle J. Marchant (d.marchant@qmul.ac.uk)					
19						
20	Conflict of interest: The authors have no competing interests.					
21						
22						

Abstract

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

Microplastics are ubiquitous in both marine and freshwater ecosystems, where they can act as a physical contaminant, as well as interact with chemicals present in the environment. It has been suggested that chemical contaminants can sorb to microplastics, such that microplastics act as a vector for chemicals into aquatic biota and enhance their negative effects. It has been repeatedly suggested that the main factors underpinning the binding of chemicals to microplastics are hydrophobic partitioning and the size of microplastic particles. Therefore, we used the hydrophobicity of chemicals, as log K_{ow}, as well as the size of microplastic particles to conduct a quantitative analysis of published results to evaluate the influence of microplastics on chemical toxicity. We collated data from 39 laboratory studies that assessed the effects of microplastics, chemicals and their combination on several ecotoxicological responses of freshwater and marine organisms. Each chemical was assigned the relevant octanol / water partition coefficient ($\log K_{OW}$) as a measure of its hydrophobicity, and the mean size of microplastics particles used in each study was recorded. We found no effect of log K_{OW} or the size of microplastic particles on the interaction between microplastics and chemicals with regards to any of the relevant ecotoxicological responses (behaviour, growth, survival and cellular) considered in this study. These findings are significant in showing that the effect of microplastics on the toxicity of chemicals is more complex than just considering hydrophobicity of chemicals and size of microplastics. We call for more mechanistic experiments to motivate a robust risk assessment and mitigation of microplastic toxicity in the environment.

44

- Keywords: Aquatic organisms, chemical toxicity, microplastic pollution, multiple stressors,
- 46 hydrophobicity.

1. INTRODUCTION

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

Currently, around 76% of total plastics produced have been disposed of as waste (Geyer et al., 2017) and the mass littering of plastics into aquatic ecosystems has become a major cause for concern. Microplastics, defined as particles within the size range $1\mu m - 5$ mm (Frias and Nash, 2019; Thompson, 2004), have been found in almost all marine (Barnes et al., 2009), freshwater (Eriksen et al., 2013; Su et al., 2016), estuarine (Sadri and Thompson, 2014) and terrestrial (Browne et al., 2011) environments. Although there is compelling evidence that microplastics can negatively impact aquatic biota (Cole et al., 2011), an overall effect remains inconclusive (Foley et al., 2018). Their physical characteristics can cause harm to organisms (e.g. blockages in the digestive tract) that can lead to reduced food intake and starvation (Galloway et al., 2017; Wright et al., 2013). Exposure of individual organisms to microplastics can also have other adverse impacts, including reduced growth, fecundity, survival and metabolism (Au et al., 2015; Kratina et al., 2019; Rist et al., 2017), hepatic stress, loss of energetic reserves, oxidative stress, genotoxicity, immunotoxicity and neurotoxicity (Barboza et al., 2018). In addition to their independent impacts, microplastics can interact with chemical contaminants present in the environment originating from wastewater, urban runoff, and landfill leachate. Many studies have noted high concentrations of chemicals such as hydrophobic organic compounds (Hirai et al., 2011; Mato et al., 2001; Rochman et al., 2013a), pharmaceutical and personal care products (Wu et al., 2016), and heavy metals (Holmes et al., 2012) associated with microplastic particles. Microplastics also act as persistent reactive surfaces, with chemicals used as starting materials (e.g. bisphenol-A) and additives for improving plastic properties (e.g. flame retardants, heat stabilizers and plasticizers) bound to

the polymer matrix (Zhang and Chen, 2014). Once ingested, chemicals sorbed to the surface

of microplastics, as well as additives, can leach from the polymer matrix and negatively

impact aquatic biota. These chemicals may have carcinogenic and mutagenic properties (Teranishi et al., 1975), and can cause endocrine disruption by mimicking endogenous hormones (Oehlmann et al., 2009). Thus, there have been concerns that microplastics can act as a vector for the introduction of these harmful chemicals into aquatic biota (Avio et al., 2015; Oliveira et al., 2013). However, this vector effect remains poorly understood, due to a lack of consistency in effects reported in the literature.

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

Chemical contaminants can form strong and often irreversible bonds with microplastic particles. Depending on the strength of adsorption of chemicals onto microplastic surfaces, microplastic particles can make chemical contaminants more bioavailable, by acting as a vector for the introduction of chemical contaminants into aquatic organisms (Rochman et al., 2013b) and enhance the negative effects of chemicals. Microplastics can also alleviate the toxicity of chemicals on aquatic organisms and may exert a positive effect, by forming strong, irreversible bonds with chemical contaminants, rendering them less bioavailable (Rehse et al., 2018a; Yang et al., 2020). There is also evidence to suggest that microplastics do not alter the toxicity of chemicals to aquatic organisms (Gerdes et al., 2019; Magara et al., 2019). Finding no effect of microplastics on chemical toxicity could suggest that microplastics are harmful to aquatic organisms due to their physical characteristics (e.g. morphology and size) rather than their capacity to leach additives (Oliviero et al., 2019) or act as vectors for the introduction of chemicals into aquatic organisms (Ašmonaitė et al., 2018). The influence of microplastics on chemical toxicity may also differ among ecotoxicological responses, where sublethal responses such as feeding and reproduction can be more sensitive to combined microplastic and chemical exposure compared with mortality (Bartonitz et al., 2020).

The interaction between microplastics and chemicals is complex, likely depending on the environmental conditions, plastic characteristics, and chemical properties. For instance, ions present in seawater can interact with different surface groups of plastics and change their overall charge (Cole and Galloway, 2015; Paul-Pont et al., 2018), altering the sorption capacity of chemicals onto microplastics (Zhao et al., 2020a; Zuo et al., 2019). This suggests that interactions between microplastics and chemicals in marine ecosystems, and thus their effect on aquatic biota, may be different from those occurring in freshwater ecosystems. The size of microplastic particles is another important consideration because it determines their uptake, retention, and effects within aquatic organisms. Smaller particles are more likely to be ingested by a wider range of aquatic organisms, and particles < 5µm are potentially more harmful due to their ability to enter biological tissues (Browne et al., 2008). These smaller microplastics can thus alter cellular and molecular pathways and impact immunological responses, antioxidant system, neurotoxicity, genotoxicity and gene expression (Lu et al., 2016; von Moos et al., 2012). Smaller microplastics have a larger surface area (per unit mass) to interact with environmental chemicals. The sorption capacity of chemicals onto nanoplastics is generally larger than for microplastics (Wang and Wang, 2018; Zhang et al., 2019) due to their large surface area and increased sorption sites.

Similarly, chemical properties are crucial in determining how chemicals and microplastics interact in the environment. Although there are many mechanisms that bind chemicals and microplastics together, hydrophobic partitioning is a major mechanism investigated in many published studies (Horton et al., 2018; Tourinho et al., 2019; Wang et al., 2015). During hydrophobic partitioning, chemicals partition between an aqueous phase and a hydrophobic solvent (here, microplastics), a process largely determined by the solubility of the chemical. The octanol / water partition coefficient (log K_{OW}) is a measure of the extent to which a chemical remains dissolved in aqueous solution (i.e. a measure of its hydrophobicity). Hydrophobic chemicals, with a higher log K_{OW}, have low water solubility and preferentially sorb to organic particles present in the aquatic environment (Elzerman and Coates, 1987; Lee et al., 2014), subsequently enhancing the ability of microplastics to act as

vectors for these chemicals into aquatic biota (Horton et al., 2018; Koelmans et al., 2013). Chemicals with lower log K_{OW} are described as hydrophilic and rather remain dissolved in aqueous solution (Wang et al., 2018), reducing the ability of microplastics to influence chemical toxicity. It has been shown that chemicals with a higher log K_{OW} also have a higher distribution coefficient, log K_d (Šunta et al., 2020; Zhao et al., 2020b). This suggests that more hydrophobic chemicals become more strongly sorbed to microplastics, something that has been directly observed in several experiments (Bakir et al., 2012; Teuten et al., 2009).

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

Here, we performed a quantitative review of published studies to examine the evidence for the combined effects of microplastics and chemicals on ecotoxicological responses of growth, survival, and behaviour of aquatic organisms. We also pooled responses at the cellular level, such as gene transcription and expression, oxidative stress, neurotoxicity, immunotoxicity and genotoxicity and refer to these as "cellular responses". Quantitative reviews allow for the combined analysis of different results from a range of studies to assess the overall effect of a given treatment (here, the influence of microplastics on chemical toxicity). Specifically, we used log K_{OW} of chemicals and the size of microplastic particles as key characteristics to determine whether and how microplastics influence chemical toxicity. We included nanoplastics in our definition of "microplastics", even though they are smaller than 1 µm. We focused on this broader range of particle sizes due to the growing evidence that nanoplastics can interact with and cross membranes, with significant ecotoxicological effects (Tallec et al., 2018). We hypothesized that (i) chemicals with a higher value of log K_{OW} would exert stronger negative effects on aquatic organisms than those chemicals with a lower value of log K_{OW}. We also hypothesized that (ii) smaller plastic particles would enhance the negative effects of chemical toxicity compared with larger particles. To the best of our knowledge, this is the first quantitative review investigating the two key mechanisms that underpin the interaction between microplastics and the toxicity of chemical contaminants. The

evidence provided by this synthesis will advance our understanding of how microplastics interact with other stressors in aquatic environments.

2. MATERIALS AND METHODS

2.1 Literature search

We selected studies published prior to July 2020 via a search of several online databases including ISI Web of Science, Google Scholar, Science Direct, Scopus and PubMed. A combination of key words (Table S1) was used to obtain studies focused on the interactive ecotoxicological effects of microplastics and chemical contaminants on aquatic organisms. The searches identified 3449 evidence sources, of which, 39 were retained for the final analysis (Fig. 1). Sources included in the analysis were those that i) investigated how microplastics affect at least one chemical and at least one ecotoxicological response in either freshwater or marine organisms and ii) included both independent and combined treatments of microplastics and chemicals. We excluded studies that only assessed the sorption kinetics of chemicals onto plastics and those that used heavy metals as they could not be assigned a value for log Kow.

Studies that investigated how microplastics affect the toxicity of chemicals encompassed a variety of different methods of introducing chemicals during the test exposure. Chemicals were either adsorbed onto microplastics prior to exposure of the study organism, added to the water during the test exposure, or the study organisms were exposed to the chemical prior to the introduction of microplastics. Some studies also assessed the toxicity of microplastic leachates on aquatic organisms. For this analysis, we did not discriminate between the various methods of exposure used. Because these different methods were not sufficiently comparable it was not possible to undertake a traditional meta – analysis. We assigned and compared the direction of effects (either positive, negative, or neutral), hereafter

referred to as outcome, to each individual record (Tourinho et al., 2019). Direction of effect was based on whether the presence of microplastics increased, decreased, or had no effect (positive, negative, or neutral, respectively) on the toxicity of the chemical as measured in the relevant ecotoxicological response. A response was considered an effect when the toxicity of chemicals was significantly different in the presence of microplastics (indicated by effective and lethal concentrations). Where effective and lethal concentrations were not reported, particularly for cellular responses, we considered a response an effect when there was a significant difference between microplastic, chemicals and their combined treatment on the ecotoxicological response (e.g. significant difference in the transcription levels of genes in the combined treatment compared with microplastics and chemical contaminants considered independently).

From each study, we extracted information about ecosystem type (freshwater or marine), the study organisms, properties of the microplastic used in the experiment (polymer type, mean size and morphology, such as bead, mixture, flake, or fibre), the chemical used and the measured ecotoxicological response. These responses included feeding rate, individual growth (change in weight and/or length), reproduction (e.g., number of offspring, fecundity), survival (number of organisms alive at the end of the experiment), metabolism (e.g. disturbed metabolism of lipids analysed via biomarker responses) and behavioural changes, such as inhibition of mobility or impairment of swimming ability. For each chemical tested we obtained information on their octanol / water partition coefficient (log K_{OW}) either from direct reporting within evidence sources, or from the PubChem website (https://pubchem.ncbi.nlm.nih.gov). Some studies used multiple treatments to assess the effect of microplastics on chemical toxicity to aquatic organisms (e.g., the response was analysed over more than one size of microplastic particles, or more than one chemical). For these studies we created individual records. Some studies also measured the effect of

microplastics on chemical toxicity to aquatic organisms using more than one ecotoxicological response (e.g. growth and metabolism) and individual records were also included from each experiment.

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

200

198

199

2.2 Reliability scoring

To ensure that this review was based on robust evidence, the reliability of the evidence provided from each study was assessed using the CRED (Criteria for Reporting and evaluating Ecotoxicity Data) method (Moermond et al., 2016). A full list of the criteria and descriptors can be found in the supplementary material (Table S2). In brief, a quality assessment of each source obtained for the final analysis was carried out. Each source was assessed on fifteen criteria that cover various aspects of the experimental design (e.g. validity criteria, adequate controls, appropriate statistical methods) and reporting of the results (e.g. identity and source of test organisms, duration of test clearly stated). For each criterion, the source under review was scored between 0-2. A score of 0 for a criterion indicates that the information provided from the evidence source under review was unreliable, 1 indicates that the evidence was reliable but with restrictions and the highest score of 2 indicates that the information provided by the evidence source was reliable without restrictions. Information from studies was considered "reliable" when the overall reliability score met the median score of 20. Because the CRED method does not define a threshold for which sources are considered reliable, we considered the top scoring 50% of studies (those with higher than the median reliability score) as "reliable" for our analysis. To determine whether reliability score had an influence on the results, we performed separate statistical analyses on the reliable datasets here we had sufficient sample size. Statistical analyses showed qualitatively similar results when analysing both datasets (Figs. S1 - S2). Therefore, to increase the robustness of the statistical analyses, we included all evidence sources regardless of reliability score in our final analyses. Due to the unequal number of studies for freshwater and marine biota, and a possible influence of salinity, we performed separate analyses on the two types of aquatic organisms. We also performed separate analyses each of the ecotoxicological responses, again due to the unequal number of studies. For the freshwater data, we analysed changes in behaviour, growth, and survival. For the marine data, there were enough studies to analyse changes in growth, survival, and cellular responses. The mean size of microplastics used in experimental studies were log10 transformed prior to the analysis, to improve the assumption of normality.

2.3 Statistical analyses

Multinomial logistic regression modelling was used to determine the relative importance of the log $K_{\rm OW}$ of chemicals and mean size of microplastic particles in predicting whether microplastics have a positive, negative, or neutral impact on the toxicity of chemicals on ecotoxicological responses of aquatic organisms. Multinomial logistic regression is an extension of binomial logistic regression that allows for a dependent variable with two or more categories (i.e. positive, negative or neutral). Multinomial logistic models are composed of k-1 equations that contrast the odds of one outcome level k compared with a reference level (Aziz et al., 2016). The reference group was first defined as the outcome level that had the most observations and was compared with each of the other two outcome levels (hereafter, comparison group). Due to the number of statistical tests that were carried out, the value at which we accepted significance was Bonferroni adjusted. Multinomial logistic regression models were built with the function "multinom" in the "nnet" package in R Statistical Software (Venables and Ripley, 2002).

One of the assumptions of logistic regression is an absence of multicollinearity amongst independent variables, as correlated variables present within the same model could result in weakened statistical power of the model. Linear relationships between the two continuous variables, mean size of microplastic particles and log K_{OW} of chemicals were assessed using Pearson's product moment correlation in R statistical software (R Core Team., 2020). There was a weak, positive correlation between the log K_{OW} of the chemical and the mean size of microplastic particles used in studies investigating how microplastics and chemical contaminants interact (Pearson's correlation, r = 0.31, n = 103, p = 0.001; Fig. S2). Therefore, we constructed separate models for microplastic particle size and log K_{OW} of chemicals to avoid multicollinearity in the analysis.

For ecotoxicological responses where only binary outcomes were observed, we applied generalized linear models (GLM) to examine the relationship between log K_{OW} or mean size of microplastic particles and observed outcomes. Due to the binomial nature of the response variable, the GLM's used a binomial family distribution. Data were modelled using the function "glm" in the "lme4" package (Bates et al., 2015) for R Statistical Software (R Core team., 2020). Odds ratios, 95% confidence intervals and p values were calculated to evaluate the outcome for each multinomial and binomial logistic model. Generally, an odds ratio of 1 indicates that the predictor variable in question is not associated with the outcome being in either the comparison or reference group. An odds ratio > 1 indicates that the outcome is more likely to be in the referent group. Whereas an odds ratio < 1 indicates that the outcome is more likely to be in the referent group than the comparison group.

Finally, to determine whether differences among individual studies may confound the effects observed by using log K_{OW} and mean size of microplastic particles as predictor variables in the logistic models, mixed effects logistic models were built in SAS/STAT $_{\hat{a}}$ using the PROC GLIMMIX procedure. Here, individual studies were included as random effect, whereas log K_{OW} , mean size of microplastic particles and ecotoxicological response were included as fixed effects. There was no difference between the model with and without the

random effect, and thus below we present the results from models without the random effect.

3. RESULTS

3.1 Evidence sources

Out of a total 3449 evidence sources identified from initial searches, a total of 156 studies were retained for meeting the inclusion criteria (Fig. 1). A total of 39 evidence sources comprising 88 observations were identified for the analysis of the effects of microplastics on chemical toxicity to aquatic organisms (Table S3). Studies that investigated the impacts of microplastics on the toxicity of chemicals affecting the feeding, reproduction and metabolism of aquatic organisms were excluded from the analysis due to insufficient observations within each category to perform statistical analysis. We also excluded studies that considered the sorption kinetics of chemicals onto microplastics and those that included heavy metals which could not be assigned a value of log $K_{\rm OW}$. Chemicals investigated in the retained studies included hydrophobic organic compounds, insecticides, pesticides, fungicides, plastic additives and several PPCP's (such as roxithromycin and 17b – oestradiol). Log $K_{\rm OW}$ of the chemicals studied ranged from – 5.4 to 9.37 and size of microplastic particles used in experiments ranged from 0.05 - 3000 μ m.

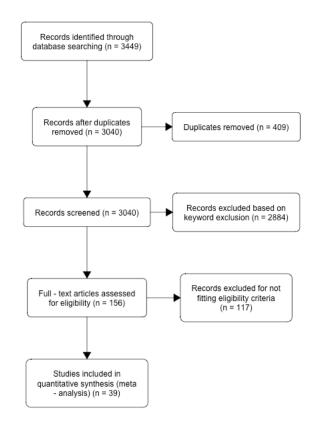


Figure 1. Evidence map of literature search and the selection process for collecting evidence relevant for the quantitative analysis of the interactive impact of microplastics and chemicals on aquatic biota.

3.3 The effect of $\log K_{OW}$

To test our first hypothesis, that microplastics influence the toxicity of hydrophobic chemicals, the log K_{OW} of the chemical used in the studies was used as a predictor in each logistic model. Overall, there was no relationship between log K_{OW} and the effect of microplastics on chemical toxicity (Fig. 2). There was a trend towards increasing probability of a neutral outcome (i.e. no influence of microplastics on the toxicity of the chemical tested) with increasing log K_{OW} (Fig. S3) for the survival of freshwater biota exposed to microplastics and chemicals. However, the likelihood of observing a neutral outcome compared with a

positive (p = 0.038) or negative (p = 0.018) outcomes was not statistically significant (significance accepted at p < 0.004).

For each of the other ecotoxicological responses tested, behavioural, growth and cellular changes, there was no significant relationship between the log K_{OW} of the chemical investigated and the effect of microplastics on chemical toxicity (Fig. 2). To determine whether environmental salinity influences the way that chemicals interact with microplastics, freshwater and marine studies were considered separately. The log K_{OW} of chemicals did not significantly affect the influence of microplastics on chemical toxicity on the behaviour, growth, survival, or cellular responses of organisms in freshwater ecosystems (Fig. 2). Similarly, log K_{OW} did not have a significant effect on the likelihood of microplastics influencing the effect of chemicals on the growth, survival, and cellular responses of organisms in marine ecosystems (Fig. 3). Neither outcome category (positive, negative, or neutral) was more likely for any given value of log K_{OW} (Figs. S4 – 10), rejecting our first hypothesis.

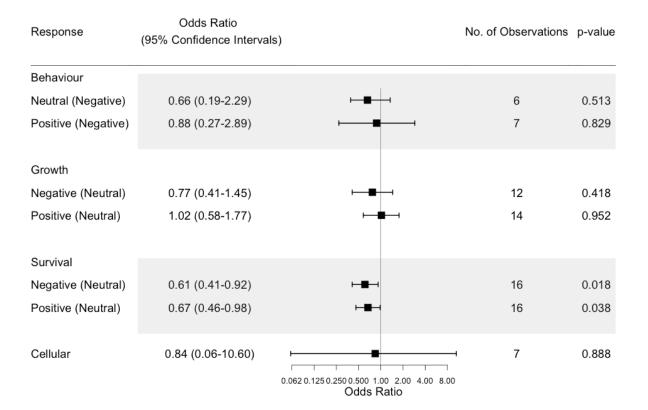


Figure 2. The effect of microplastics on chemical toxicity, determined by mean odds ratios (illustrated as filled squares) with 95% confidence intervals (horizontal bars), for the four different response categories of freshwater biota exposed to microplastics and chemicals along a gradient of log $K_{\rm OW}$. Vertical line that intercepts the value of 1 indicates no difference in the response variable. An odds ratio > 1 indicates that the outcome is more likely to be in the comparison group than the reference group (appears in parentheses). Whereas an odds ratio < 1 indicates that the outcome is more likely to be in the reference group than the comparison group. Although the mean odds ratio and confidence intervals for survival do not overlap the vertical line, this effect was marginally non-significant after the Bonferroni correction (alpha level of p < 0.004).

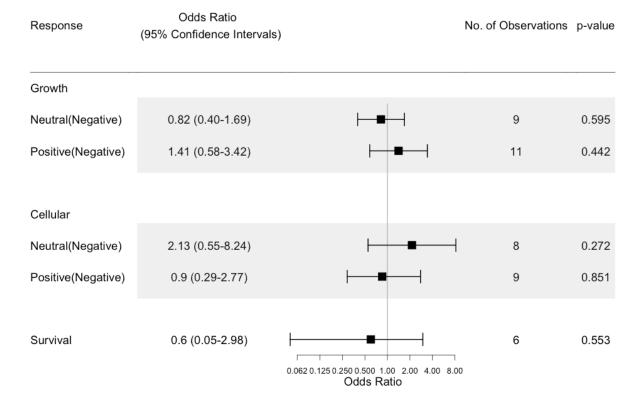


Figure 3. The effect of microplastics on chemical toxicity, determined by mean odds ratios (illustrated as filled squares) with 95% confidence intervals (horizontal bars), for the three different response categories of marine biota exposed to microplastics and chemicals along a gradient of log $K_{\rm OW}$. Vertical line that intercepts the value of 1 indicates no difference in the response variable. An odds ratio > 1 indicates that the outcome is more likely to be in the comparison group than the reference group (appears in parentheses). Whereas an odds ratio < 1 indicates that the outcome is more likely to be in the reference group (appear in parentheses) than the comparison group. Statistical significance was accepted at p < 0.004.

3.4 The effect of microplastic particle size

To test the second hypothesis that smaller microplastic particles enhance the negative effects of chemicals on aquatic biota, the mean size of microplastic particles used in experimental studies was used as a predictor in each logistic regression model. There was no effect of mean microplastic size on the likelihood of a positive, negative, or neutral effect of microplastics

on chemical toxicity (Figs S4 - 10) for any of the responses of freshwater (Fig. 4) or marine (Fig. 5) organisms, rejecting our second hypothesis.



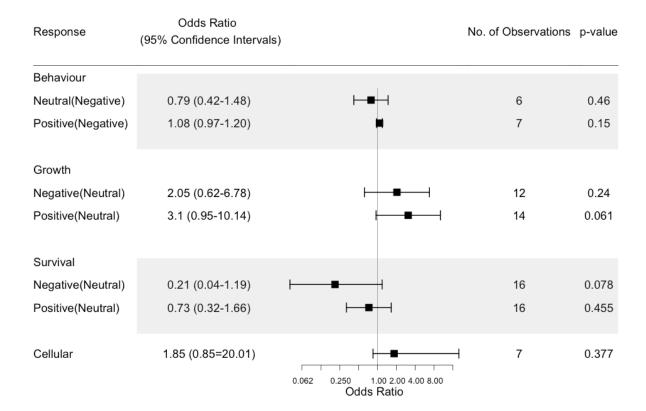


Figure 4. The effect of microplastic particle size on chemical toxicity, determined by mean odds ratios (illustrated as filled squares) with 95% confidence intervals (horizontal bars), for the four different responses of freshwater biota to chemicals along a gradient of microplastic size. Vertical line that intercepts the value of 1 indicates no difference in the response variable. An odds ratio > 1 indicates that the outcome is more likely to be in the comparison group than the referent group (appears in parentheses). Whereas an odds ratio < 1 indicates that the outcome is more likely to be in the reference group (appear in parentheses) than the comparison group. Statistical significance was accepted at p < 0.004.

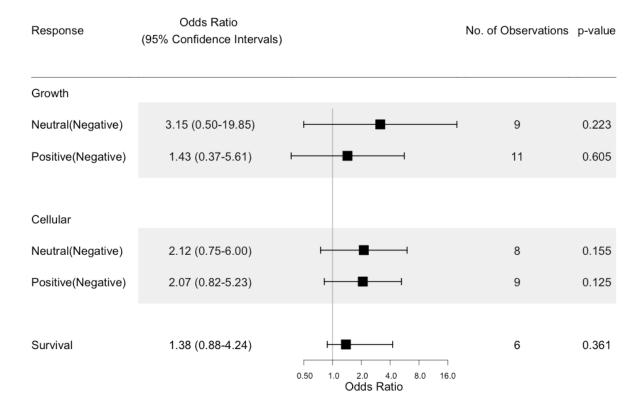


Figure 5. The influence of microplastic particle size on chemical toxicity, determined by mean odds ratios (illustrated as filled squares) with 95% confidence intervals (horizontal bars), for the three different responses of marine biota to chemicals along a gradient of microplastic size. Vertical line that intercepts the value of 1 indicates no difference in the response variable. An odds ratio > 1 indicates that the outcome is more likely to be in the comparison group than the reference group (appears in parentheses). Whereas an odds ratio < 1 indicates that the outcome is more likely to be in the referent group (appear in parentheses) than the comparison group. Statistical significance was accepted at p < 0.004.

4. DISCUSSION

One of the greatest knowledge gaps in understanding the toxicity of microplastics in aquatic environments is whether or not microplastic particles mediate the toxicity of environmental contaminants (e.g. hydrophobic organic pollutants). Our findings do not support assertions

that microplastics mediate the toxicity of chemicals (Besseling et al., 2013; Brandts et al., 2018; Rochman et al., 2013b). Results across all ecotoxicological endpoints considered (behaviour, growth, survival and cellular) were consistent. Neither the octanol / water partition coefficient of chemicals (log K_{OW}) nor the mean size of microplastic particles used in experiments had a significant influence on observed outcome (Fig. 2 - 5). We applied odds ratios to estimate the effect sizes and to determine the association between log K_{OW}, size of microplastics used and observed outcome in organismal responses. Odds ratios and 95% confidence intervals for all but one ecotoxicological response contained a value of 1, which indicates that the evidence available to date does not support any influence of log K_{OW} or mean size of microplastic particles on the responses of aquatic organisms.

The strongest response to the combined effects of microplastics and chemicals was observed in the survival of freshwater organisms. There was a trend towards increasing probability of a neutral outcome for chemicals with higher values $\log K_{\rm OW}$, and subsequently an increasing probability of a negative outcome for chemicals with lower values of $\log K_{\rm OW}$. However, this effect was marginally non-significant. We would not expect this trend given our first hypothesis that chemicals with a higher $\log K_{\rm OW}$ would exert stronger negative effects on aquatic organisms than those with a lower $\log K_{\rm OW}$. Although, there is some evidence suggesting that microplastics can alter the bioavailability and toxicity of chemicals with a lower $K_{\rm OW}$ (Zocchi and Sommaruga, 2019), possibly due to other factors influencing their interaction. The survival response of the freshwater organisms was also the response where we had the highest number of observations (n = 21), suggesting that stronger responses may be detected for other ecotoxicological variables, as more observations accumulate.

The findings that microplastics may not alter the toxicity of chemicals agree with individual studies that observed no effects of microplastics on chemical toxicity (Batel et al., 2016; Magara et al., 2019). This is a significant result, contributing to the recent debate

suggesting that microplastics are toxic because of their chemical counterparts, rather than their physical characteristics (Oliviero et al., 2019; Rehse et al., 2018b). The results shown in the current research may be attributed to chemicals not being adsorbed onto plastic particles in sufficient concentrations to cause strong effects (Schmieg et al., 2020), as well as other mechanisms that influence the sorption and desorption of chemicals onto microplastics. A variety of interaction mechanisms, including electrostatic and pi-pi interactions, hydrogen bonding and van der Waals forces (Tourinho et al., 2019) contribute to the binding of chemicals and microplastics. Despite this, one of the main mechanisms reported in the literature is hydrophobic partitioning, where molecules remain dissolved whilst they are partitioned into a sorbing matrix (Hartmann et al., 2017). Our analysis focused on the hydrophobicity of chemicals, its influence on the binding of chemicals to microplastics and, thus, the contribution of hydrophobicity to a positive, negative, or neutral effect of microplastics on chemical toxicity. The reason that no consistent effect of microplastics on chemical toxicity to aquatic biota was observed may have been due to the simplified nature of using hydrophobic partitioning, as log K_{OW}, to describe the sorption of chemicals onto microplastics. Considering additional mechanisms that influence the binding of chemicals and microplastics, as well as other characteristics of the microplastics and the environment, remains a promising venue for future research.

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

The adverse effects of microplastics and associated chemicals could be reduced by rapid egestion of microplastic particles, which in turn may be influenced by the physical characteristics of microplastics such as shape and size. Spherical microplastics are likely to be egested more quickly compared with microplastic fibres (Au et al., 2015) and smaller microplastics can cross biological barriers and therefore remain within the organism for a longer period, allowing for desorption of chemical contaminants (Rist et al., 2017). The size of microplastics in this analysis did not influence observed outcome level, in contrast to our

second hypothesis that smaller microplastic particles would exert stronger negative effects due to their larger surface area. We were not able to consider microplastic morphology in this analysis due to insufficient data, although morphology was unlikely to influence the results since many sources retained for the final analysis used spherical microplastic particles.

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

Polymer type is also an important characteristic and could influence the vector function of microplastics. Polymer type has been shown to influence the sorption of chemicals, where polyethylene and polystyrene had greater affinity for the sorption of polycyclic aromatic hydrocarbons than polypropylene (Lee et al., 2014; Rochman et al., 2013a). The concept adopted in this study was that microplastic particles are nonpolar and, therefore, attract nonpolar chemicals present in the environment. However, not all plastic polymers are nonpolar, hydrophilic microplastics such as polyamides have been shown to have higher sorption affinity for hydrophilic chemicals (Li et al., 2018; Liu et al., 2019). Many biodegradable materials, such as poly (butylene adipate co-terephthalate) (PBAT) and polyurethane (PLU) are also polar, due to the presence of oxygen-containing functional groups that facilitate their degradation (Zhao et al., 2020b). Polarity can also be altered by ions present in seawater and, thus, salinity may influence the interaction between microplastics and chemicals. There were two ecotoxicological responses (growth and survival) where we had sufficient observations to determine the effects of microplastics on chemical toxicity to both freshwater and marine organisms. Despite the possible influence of salinity, microplastics did not influence the growth or survival of either freshwater or marine organisms.

Other species - dependent factors could also influence how multiple stressors interact. A recent meta-analysis has shown that the physical effects of microplastics may differ amongst taxonomic groups (Foley et al., 2018). Whereas some species show selectivity against microplastic particles in the presence of food (Aljaibachi and Callaghan, 2018), other

species only ingests microplastic particles within a certain size range (Straub et al., 2017). Crustaceans and fish appeared to be the most frequently used organisms within evidence sources relating to the interactive effects of microplastics and chemicals on aquatic biota. This was also the case for a recent review assessing the ecological impacts of microplastics on freshwater and estuarine organisms (Jones et al., 2019). Future research should thus identify the effects of microplastics and chemical contaminants on a wider range of species within different taxonomic groups, to partition any species-specific effects.

There were not enough studies to analyse the interactive effects of microplastics and chemicals on other sublethal responses such as reproduction, feeding and metabolism. Such sublethal effects could have broader ramifications within an ecosystem, leading to altered population densities, community structure and ecosystem functioning (Ward et al., 2016). The evidence for the effects of microplastics at the ecosystem level are generally lacking regarding both physical (Green, 2016; Green et al., 2016; Seeley et al., 2020) and chemical characteristics of microplastic particles. All studies considered for this analysis only investigated the effects of microplastics on chemical toxicity impacting the health and fitness of individual species, rather than on communities and ecosystems. Given the physicochemical properties of microplastics and their movement in the natural environment, perhaps one of the biggest challenges in microplastic research is scaling up from laboratory tests on individual organisms to the entire ecosystems. Such studies are urgently needed, considering the effects observed at the individual level can influence ecosystem structure and function.

Microplastics present in the environment are one of many stressors acting on aquatic organisms. Whereas previous work has summarized the potential hazards of microplastics and associated chemicals (Koelmans et al., 2016; Wang et al., 2018), this is the first quantitative analysis of the factors (hydrophobicity of chemicals and size of microplastics) that underpin the interaction between these two major pollutants of aquatic ecosystems.

Studies assessing the interaction between microplastics and other environmental stressors (e.g. habitat degradation, climate warming, eutrophication) are becoming increasingly important in understanding the overall effect that microplastics have in the environment (Kratina et al., 2019). We did not find evidence to show that the log K_{OW} of chemicals and the mean size of microplastics influence how microplastics affect chemical toxicity. Despite assertions that these two factors influence the interaction between microplastics and chemical contaminants (Horton et al., 2018; Ma et al., 2016), the interaction between microplastics and chemicals remains complex, with numerous factors potentially having an influence on the sorption / desorption of chemicals onto microplastics. A better understanding of these mechanisms would foster a robust risk assessment and directly assist restoration and conservation of both marine and freshwater ecosystems.

Acknowledgements

This research was funded by the EU INTERREG France (Channel) England project "Preventing Plastic Pollution" co-financed by the European Regional Development Fund. PK acknowledges funding from the Royal Society, Newton Advanced Fellowship (grant no. NAF/R2/180791). We thank John F. Murphy, Amanda Arnold, James L. Pretty, Kate Spencer, Adriaan A. Markus and. Andre D. Vethaak for their help with data collection and Marina Resmini, Kate Heppell and Mark Trimmer for comments on an earlier draft.

References

- Aljaibachi, R., Callaghan, A., 2018. Impact of polystyrene microplastics on Daphnia magna
- 493 mortality and reproduction in relation to food availability. PeerJ 6.
- 494 https://doi.org/10.7717/peerj.4601
- 495 Ašmonaitė, G., Larsson, K., Undeland, I., Sturve, J., Carney Almroth, B., 2018. Size Matters:
- Ingestion of Relatively Large Microplastics Contaminated with Environmental
- 497 Pollutants Posed Little Risk for Fish Health and Fillet Quality. Environmental Science
- 498 & Technology 52, 14381–14391. https://doi.org/10.1021/acs.est.8b04849
- 499 Au, S.Y., Bruce, T.F., Bridges, W.C., Klaine, S.J., 2015. Responses of Hyalella azteca to
- acute and chronic microplastic exposures. Environmental Toxicology and Chemistry 34,
- 501 2564–2572. https://doi.org/10.1002/etc.3093
- Avio, C.G., Gorbi, S., Milan, M., Benedetti, M., Fattorini, D., d'Errico, G., Pauletto, M.,
- Bargelloni, L., Regoli, F., 2015. Pollutants bioavailability and toxicological risk from
- microplastics to marine mussels. Environmental Pollution 198, 211–222.
- 505 https://doi.org/10.1016/j.envpol.2014.12.021
- Aziz, N.A.A., Ali, Z., Nor, N.M., Baharum, A., Omar, M., 2016. Modeling multinomial
- logistic regression on characteristics of smokers after the smoke-free campaign in the
- area of Melaka. https://doi.org/10.1063/1.4954625
- Bakir, A., Rowland, S.J., Thompson, R.C., 2012. Competitive sorption of persistent organic
- pollutants onto microplastics in the marine environment. Marine Pollution Bulletin 64.
- 511 https://doi.org/10.1016/j.marpolbul.2012.09.010
- Barboza, L.G.A., Vieira, L.R., Branco, V., Figueiredo, N., Carvalho, F., Carvalho, C.,
- Guilhermino, L., 2018. Microplastics cause neurotoxicity, oxidative damage and
- energy-related changes and interact with the bioaccumulation of mercury in the
- European seabass, Dicentrarchus labrax (Linnaeus, 1758). Aquatic Toxicology 195.
- 516 https://doi.org/10.1016/j.aquatox.2017.12.008
- Barnes, D.K.A., Galgani, F., Thompson, R.C., Barlaz, M., 2009. Accumulation and
- fragmentation of plastic debris in global environments. Philosophical Transactions of
- 519 the Royal Society B: Biological Sciences 364, 1985–1998.
- 520 https://doi.org/10.1098/rstb.2008.0205
- Bartonitz, A., Anyanwu, I.N., Geist, J., Imhof, H.K., Reichel, J., Graßmann, J., Drewes, J.E.,
- Beggel, S., 2020. Modulation of PAH toxicity on the freshwater organism G. roeseli by

microparticles. Environmental Pollution 260. 523 https://doi.org/10.1016/j.envpol.2020.113999 524 Batel, A., Linti, F., Scherer, M., Erdinger, L., Braunbeck, T., 2016. Transfer of benzo a 525 pyrene from microplastics to Artemia nauplii and further to zebrafish via a trophic food 526 web experiment: CYP1A induction and visual tracking of persistent organic pollutants. 527 Environmental Toxicology and Chemistry 35, 1656–1666. 528 https://doi.org/10.1002/etc.3361 529 Bates, D., Mächler, M., Bolker, B., Walker, S., 2015. Fitting Linear Mixed-Effects Models 530 Using lme4. Journal of Statistical Software 67. https://doi.org/10.18637/jss.v067.i01 531 Besseling, E., Wegner, A., Foekema, E.M., van den Heuvel-Greve, M.J., Koelmans, A.A., 532 2013. Effects of Microplastic on Fitness and PCB Bioaccumulation by the Lugworm 533 Arenicola marina (L.). Environmental Science & Technology 47, 593-600. 534 https://doi.org/10.1021/es302763x 535 Brandts, I., Teles, M., Gonçalves, A.P., Barreto, A., Franco-Martinez, L., Tvarijonaviciute, 536 A., Martins, M.A., Soares, A.M.V.M., Tort, L., Oliveira, M., 2018. Effects of 537 nanoplastics on Mytilus galloprovincialis after individual and combined exposure with 538 carbamazepine. Science of Environment The Total 643. 539 https://doi.org/10.1016/j.scitotenv.2018.06.257 540 Browne, M.A., Crump, P., Niven, S.J., Teuten, E., Tonkin, A., Galloway, T., Thompson, R., 541 2011. Accumulation of Microplastic on Shorelines Woldwide: Sources and Sinks. 542 Environmental Science & Technology 45, 9175-9179. 543 https://doi.org/10.1021/es201811s 544 Browne, M.A., Dissanayake, A., Galloway, T.S., Lowe, D.M., Thompson, R.C., 2008. 545 Ingested Microscopic Plastic Translocates to the Circulatory System of the Mussel, 546 Mytilus edulis (L.). Environmental Science & Technology 42, 5026–5031. 547 https://doi.org/10.1021/es800249a 548 Cole, M., Galloway, T.S., 2015. Ingestion of Nanoplastics and Microplastics by Pacific 549 49. 550 Ovster Larvae. Environmental Science & Technology 14625–14632. https://doi.org/10.1021/acs.est.5b04099 551 Cole, M., Lindeque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants 552

A review.

Marine

Pollution

Bulletin

62.

environment:

https://doi.org/10.1016/j.marpolbul.2011.09.025

in

553

- Elzerman, A.W., Coates, J.T., 1987. Hydrophobic Organic Compounds on Sediments:
- Equilibria and Kinetics of Sorption. pp. 263–317. https://doi.org/10.1021/ba-1987-
- 557 0216.ch010
- Eriksen, M., Mason, S., Wilson, S., Box, C., Zellers, A., Edwards, W., Farley, H., Amato, S.,
- 559 2013. Microplastic pollution in the surface waters of the Laurentian Great Lakes. Marine
- Pollution Bulletin 77, 177–182. https://doi.org/10.1016/j.marpolbul.2013.10.007
- Foley, C.J., Feiner, Z.S., Malinich, T.D., Höök, T.O., 2018. A meta-analysis of the effects of
- exposure to microplastics on fish and aquatic invertebrates. Science of The Total
- 563 Environment 631–632, 550–559. https://doi.org/10.1016/j.scitotenv.2018.03.046
- Frias, J.P.G.L., Nash, R., 2019. Microplastics: Finding a consensus on the definition. Marine
- Pollution Bulletin 138. https://doi.org/10.1016/j.marpolbul.2018.11.022
- Galloway, T.S., Cole, M., Lewis, C., 2017. Interactions of microplastic debris throughout the
- marine ecosystem. Nature Ecology & Evolution 1. https://doi.org/10.1038/s41559-017-
- 568 0116
- Gerdes, Z., Ogonowski, M., Nybom, I., Ek, C., Adolfsson-Erici, M., Barth, A., Gorokhova,
- E., 2019. Microplastic-mediated transport of PCBs? A depuration study with Daphnia
- magna. PLOS ONE 14, e0205378. https://doi.org/10.1371/journal.pone.0205378
- Geyer, R., Jambeck, J.R., Law, K.L., 2017. Production, use, and fate of all plastics ever made.
- 573 Science Advances 3, e1700782. https://doi.org/10.1126/sciadv.1700782
- Green, D.S., 2016. Effects of microplastics on European flat oysters, Ostrea edulis and their
- associated benthic communities. Environmental Pollution 216, 95–103.
- 576 https://doi.org/10.1016/j.envpol.2016.05.043
- 577 Green, D.S., Boots, B., Sigwart, J., Jiang, S., Rocha, C., 2016. Effects of conventional and
- biodegradable microplastics on a marine ecosystem engineer (Arenicola marina) and
- sediment nutrient cycling. Environmental Pollution 208, 426–434.
- 580 https://doi.org/10.1016/j.envpol.2015.10.010
- Hartmann, N.B., Rist, S., Bodin, J., Jensen, L.H., Schmidt, S.N., Mayer, P., Meibom, A.,
- Baun, A., 2017. Microplastics as vectors for environmental contaminants: Exploring
- sorption, desorption, and transfer to biota. Integrated Environmental Assessment and
- Management 13. https://doi.org/10.1002/ieam.1904
- Hirai, H., Takada, H., Ogata, Y., Yamashita, R., Mizukawa, K., Saha, M., Kwan, C., Moore,
- 586 C., Gray, H., Laursen, D., Zettler, E.R., Farrington, J.W., Reddy, C.M., Peacock, E.E.,
- Ward, M.W., 2011. Organic micropollutants in marine plastics debris from the open

- ocean and remote and urban beaches. Marine Pollution Bulletin 62, 1683–1692.
- 589 https://doi.org/10.1016/j.marpolbul.2011.06.004
- Holmes, L.A., Turner, A., Thompson, R.C., 2012. Adsorption of trace metals to plastic resin
- 591 pellets in the marine environment. Environmental Pollution 160, 42–48.
- 592 https://doi.org/10.1016/j.envpol.2011.08.052
- Horton, A.A., Vijver, M.G., Lahive, E., Spurgeon, D.J., Svendsen, C., Heutink, R., van
- Bodegom, P.M., Baas, J., 2018. Acute toxicity of organic pesticides to Daphnia magna
- is unchanged by co-exposure to polystyrene microplastics. Ecotoxicology and
- Environmental Safety 166, 26–34. https://doi.org/10.1016/j.ecoenv.2018.09.052
- Jones, J., Murphy, J., Arnold, A., Pretty, J., Spencer, K., Markus, A., Vethaak, A., 2019.
- Evidence Reviews on Analysis, Prevalence & Impact of Microplastics in Freshwater and
- Estuarine Environments. Evidence Review 3: What is/are the impact(s) of microplastics
- in freshwater and marine biota? London.
- Koelmans, A.A., Bakir, A., Burton, G.A., Janssen, C.R., 2016. Microplastic as a Vector for
- 602 Chemicals in the Aquatic Environment: Critical Review and Model-Supported
- Reinterpretation of Empirical Studies. Environmental Science & Technology 50, 3315–
- 3326. https://doi.org/10.1021/acs.est.5b06069
- Koelmans, A.A., Besseling, E., Wegner, A., Foekema, E.M., 2013. Plastic as a Carrier of
- POPs to Aquatic Organisms: A Model Analysis. Environmental Science & Technology
- 47. https://doi.org/10.1021/es401169n
- Kratina, P., Watts, T.J., Green, D.S., Kordas, R.L., O'Gorman, E.J., 2019. Interactive effects
- of warming and microplastics on metabolism but not feeding rates of a key freshwater
- 610 detritivore. Environmental Pollution 255, 113259.
- https://doi.org/10.1016/j.envpol.2019.113259
- Lee, H., Shim, W.J., Kwon, J.-H., 2014. Sorption capacity of plastic debris for hydrophobic
- organic chemicals. Science of The Total Environment 470–471, 1545–1552.
- https://doi.org/10.1016/j.scitotenv.2013.08.023
- Li, J., Zhang, K., Zhang, H., 2018. Adsorption of antibiotics on microplastics. Environmental
- Pollution 237. https://doi.org/10.1016/j.envpol.2018.02.050
- Liu, F., Liu, G., Zhu, Z., Wang, S., Zhao, F., 2019. Interactions between microplastics and
- phthalate esters as affected by microplastics characteristics and solution chemistry.
- Chemosphere 214, 688–694. https://doi.org/10.1016/j.chemosphere.2018.09.174
- 620 Lu, Y., Zhang, Y., Deng, Y., Jiang, W., Zhao, Y., Geng, J., Ding, L., Ren, H., 2016. Uptake
- and Accumulation of Polystyrene Microplastics in Zebrafish (Danio rerio) and Toxic

- Effects in Liver. Environmental Science & Technology 50.
- 623 https://doi.org/10.1021/acs.est.6b00183
- Ma, Y., Huang, A., Cao, S., Sun, F., Wang, L., Guo, H., Ji, R., 2016. Effects of nanoplastics
- and microplastics on toxicity, bioaccumulation, and environmental fate of phenanthrene
- 626 in fresh water. Environmental Pollution 219, 166–173.
- 627 https://doi.org/10.1016/j.envpol.2016.10.061
- Magara, G., Khan, F.R., Pinti, M., Syberg, K., Inzirillo, A., Elia, A.C., 2019. Effects of
- combined exposures of fluoranthene and polyethylene or polyhydroxybutyrate
- microplastics on oxidative stress biomarkers in the blue mussel (Mytilus edulis).
- Journal of Toxicology and Environmental Health, Part A 82, 616–625.
- https://doi.org/10.1080/15287394.2019.1633451
- Mato, Y., Isobe, T., Takada, H., Kanehiro, H., Ohtake, C., Kaminuma, T., 2001. Plastic resin
- pellets as a transport medium for toxic chemicals in the marine environment.
- Environmental Science and Technology 35, 318–324.
- https://doi.org/10.1021/es0010498
- Moermond, C.T.A., Kase, R., Korkaric, M., Ågerstrand, M., 2016. CRED: Criteria for
- reporting and evaluating ecotoxicity data. Environmental Toxicology and Chemistry 35,
- 639 1297–1309. https://doi.org/10.1002/etc.3259
- Oehlmann, J., Schulte-Oehlmann, U., Kloas, W., Jagnytsch, O., Lutz, I., Kusk, K.O.,
- Wollenberger, L., Santos, E.M., Paull, G.C., Van Look, K.J.W., Tyler, C.R., 2009. A
- critical analysis of the biological impacts of plasticizers on wildlife. Philosophical
- Transactions of the Royal Society B: Biological Sciences 364, 2047–2062.
- https://doi.org/10.1098/rstb.2008.0242
- Oliveira, M., Ribeiro, A., Hylland, K., Guilhermino, L., 2013. Single and combined effects
- of microplastics and pyrene on juveniles (0+ group) of the common goby
- Pomatoschistus microps (Teleostei, Gobiidae). Ecological Indicators 34, 641–647.
- https://doi.org/10.1016/j.ecolind.2013.06.019
- Oliviero, M., Tato, T., Schiavo, S., Fernández, V., Manzo, S., Beiras, R., 2019. Leachates of
- 650 micronized plastic toys provoke embryotoxic effects upon sea urchin Paracentrotus
- lividus. Environmental Pollution 247. https://doi.org/10.1016/j.envpol.2019.01.098
- Paul-Pont, I., Tallec, K., Gonzalez-Fernandez, C., Lambert, C., Vincent, D., Mazurais, D.,
- Zambonino-Infante, J.-L., Brotons, G., Lagarde, F., Fabioux, C., Soudant, P., Huvet, A.,
- 654 2018. Constraints and Priorities for Conducting Experimental Exposures of Marine

- Organisms to Microplastics. Frontiers in Marine Science 5.
- https://doi.org/10.3389/fmars.2018.00252
- Rehse, S., Kloas, W., Zarfl, C., 2018a. Microplastics Reduce Short-Term Effects of
- Environmental Contaminants. Part I: Effects of Bisphenol A on Freshwater Zooplankton
- Are Lower in Presence of Polyamide Particles. International Journal of Environmental
- Research and Public Health 15. https://doi.org/10.3390/ijerph15020280
- Rehse, S., Kloas, W., Zarfl, C., 2018b. Microplastics Reduce Short-Term Effects of
- Environmental Contaminants. Part I: Effects of Bisphenol A on Freshwater Zooplankton
- Are Lower in Presence of Polyamide Particles. International Journal of Environmental
- Research and Public Health 15. https://doi.org/10.3390/ijerph15020280
- Rist, S., Baun, A., Hartmann, N.B., 2017. Ingestion of micro- and nanoplastics in Daphnia
- 666 magna Quantification of body burdens and assessment of feeding rates and
- reproduction. Environmental Pollution 228, 398–407.
- https://doi.org/10.1016/j.envpol.2017.05.048
- Rochman, C.M., Brookson, C., Bikker, J., Djuric, N., Earn, A., Bucci, K., Athey, S.,
- Huntington, A., McIlwraith, H., Munno, K., De Frond, H., Kolomijeca, A., Erdle, L.,
- 671 Grbic, J., Bayoumi, M., Borrelle, S.B., Wu, T., Santoro, S., Werbowski, L.M., Zhu, X.,
- Giles, R.K., Hamilton, B.M., Thaysen, C., Kaura, A., Klasios, N., Ead, L., Kim, J.,
- Sherlock, C., Ho, A., Hung, C., 2019. Rethinking microplastics as a diverse contaminant
- suite. Environmental Toxicology and Chemistry 38, 703–711.
- https://doi.org/10.1002/etc.4371
- Rochman, C.M., Hoh, E., Hentschel, B.T., Kaye, S., 2013a. Long-Term Field Measurement
- of Sorption of Organic Contaminants to Five Types of Plastic Pellets: Implications for
- Plastic Marine Debris. Environmental Science & Technology 130109073312009.
- 679 https://doi.org/10.1021/es303700s
- Rochman, C.M., Hoh, E., Kurobe, T., Teh, S.J., 2013b. Ingested plastic transfers hazardous
- chemicals to fish and induces hepatic stress. Scientific Reports 3.
- https://doi.org/10.1038/srep03263
- Sadri, S.S., Thompson, R.C., 2014. On the quantity and composition of floating plastic debris
- entering and leaving the Tamar Estuary, Southwest England. Marine Pollution Bulletin
- 685 81, 55–60. https://doi.org/10.1016/j.marpolbul.2014.02.020
- Schmieg, H., Huppertsberg, S., Knepper, T.P., Krais, S., Reitter, K., Rezbach, F., Ruhl, A.S.,
- Köhler, H.-R., Triebskorn, R., 2020. Polystyrene microplastics do not affect juvenile

- brown trout (Salmo trutta f. fario) or modulate effects of the pesticide methiocarb.
- Environmental Sciences Europe 32, 49. https://doi.org/10.1186/s12302-020-00327-4
- 690 Seeley, M.E., Song, B., Passie, R., Hale, R.C., 2020. Microplastics affect sedimentary
- 691 microbial communities and nitrogen cycling. Nature Communications 11, 2372.
- 692 https://doi.org/10.1038/s41467-020-16235-3
- 693 Straub, S., Hirsch, P.E., Burkhardt-Holm, P., 2017. Biodegradable and Petroleum-Based
- Microplastics Do Not Differ in Their Ingestion and Excretion but in Their Biological
- 695 Effects in a Freshwater Invertebrate Gammarus fossarum. International Journal of
- Environmental Research and Public Health 14. https://doi.org/10.3390/ijerph14070774
- 697 Su, L., Xue, Y., Li, L., Yang, D., Kolandhasamy, P., Li, D., Shi, H., 2016. Microplastics in
- Taihu Lake, China. Environmental Pollution 216, 711–719.
- 699 https://doi.org/10.1016/j.envpol.2016.06.036
- 500 Šunta, U., Prosenc, F., Trebše, P., Bulc, T.G., Kralj, M.B., 2020. Adsorption of acetamiprid,
- chlorantraniliprole and flubendiamide on different type of microplastics present in
- alluvial soil. Chemosphere 261. https://doi.org/10.1016/j.chemosphere.2020.127762
- Tallec, K., Huvet, A., di Poi, C., Gonzalez-Fernandez, C., Lambert, C., Petton, B., le Goic,
- N., Bercel, M., Soudant, P., Paul-Pont, I., 2018. Nanoplastics impaired oyster free living
- stages, gametes and embryos. Environmental Pollution 242.
- 706 Teranishi, K., Hamada, K., Hiromu, W., 1975. Quantitative relationship between
- carcinogenecity and mutagenicity of polyaromatic hydrocarbons in Salmonnella
- 708 typhimurium mutants. Mutation Research/Environmental Mutagenesis and Related
- 709 Subjects 31. https://doi.org/10.1016/0165-1161(75)90071-0
- 710 Teuten, E.L., Saquing, J.M., Knappe, D.R.U., Barlaz, M.A., Jonsson, S., Björn, A., Rowland,
- S.J., Thompson, R.C., Galloway, T.S., Yamashita, R., Ochi, D., Watanuki, Y., Moore,
- C., Viet, P.H., Tana, T.S., Prudente, M., Boonyatumanond, R., Zakaria, M.P.,
- Akkhavong, K., Ogata, Y., Hirai, H., Iwasa, S., Mizukawa, K., Hagino, Y., Imamura,
- A., Saha, M., Takada, H., 2009. Transport and release of chemicals from plastics to the
- environment and to wildlife. Philosophical Transactions of the Royal Society B:
- Biological Sciences 364. https://doi.org/10.1098/rstb.2008.0284
- Thompson, R.C., 2004. Lost at Sea: Where Is All the Plastic? Science 304, 838–838.
- 718 https://doi.org/10.1126/science.1094559
- 719 Tourinho, P.S., Kočí, V., Loureiro, S., van Gestel, C.A.M., 2019. Partitioning of chemical
- contaminants to microplastics: Sorption mechanisms, environmental distribution and

- 721 effects on toxicity and bioaccumulation. Environmental Pollution 252.
- 722 https://doi.org/10.1016/j.envpol.2019.06.030
- Venables, W.N., Ripley, B.D., 2002. Modern Applied Statistics with S. Springer New York,
- New York, NY. https://doi.org/10.1007/978-0-387-21706-2
- von Moos, N., Burkhardt-Holm, P., Köhler, A., 2012. Uptake and Effects of Microplastics on
- 726 Cells and Tissue of the Blue Mussel Mytilus edulis L. after an Experimental Exposure.
- Environmental Science & Technology 46. https://doi.org/10.1021/es302332w
- Wang, F., Shih, K.M., Li, X.Y., 2015. The partition behavior of perfluorooctanesulfonate
- 729 (PFOS) and perfluorooctanesulfonamide (FOSA) on microplastics. Chemosphere 119.
- 730 https://doi.org/10.1016/j.chemosphere.2014.08.047
- Wang, Fen, Wong, C.S., Chen, D., Lu, X., Wang, Fei, Zeng, E.Y., 2018. Interaction of toxic
- chemicals with microplastics: A critical review. Water Research 139.
- 733 https://doi.org/10.1016/j.watres.2018.04.003
- Wang, W., Wang, J., 2018. Comparative evaluation of sorption kinetics and isotherms of
- pyrene onto microplastics. Chemosphere 193, 567–573.
- 736 https://doi.org/10.1016/j.chemosphere.2017.11.078
- Ward, T.D., Algera, D.A., Gallagher, A.J., Hawkins, E., Horodysky, A., Jørgensen, C.,
- Killen, S.S., McKenzie, D.J., Metcalfe, J.D., Peck, M.A., Vu, M., Cooke, S.J., 2016.
- Understanding the individual to implement the ecosystem approach to fisheries
- management. Conservation Physiology 4. https://doi.org/10.1093/conphys/cow005
- Wright, S.L., Thompson, R.C., Galloway, T.S., 2013. The physical impacts of microplastics
- on marine organisms: A review. Environmental Pollution 178, 483–492.
- 743 https://doi.org/10.1016/j.envpol.2013.02.031
- Wu, C., Zhang, K., Huang, X., Liu, J., 2016. Sorption of pharmaceuticals and personal care
- products to polyethylene debris. Environmental Science and Pollution Research 23,
- 746 8819–8826. https://doi.org/10.1007/s11356-016-6121-7
- 747 Yang, W., Gao, X., Wu, Y., Wan, L., Tan, L., Yuan, S., Ding, H., Zhang, W., 2020. The
- combined toxicity influence of microplastics and nonylphenol on microalgae Chlorella
- 749 pyrenoidosa. Ecotoxicology and Environmental Safety 195.
- 750 https://doi.org/10.1016/j.ecoenv.2020.110484
- Zhang, X., Chen, Z., 2014. Observing Phthalate Leaching from Plasticized Polymer Films at
- the Molecular Level. Langmuir 30. https://doi.org/10.1021/la500476u
- 753 Zhang, X., Zheng, M., Yin, X., Wang, L., Lou, Y., Qu, L., Liu, X., Zhu, H., Qiu, Y., 2019.
- Sorption of 3,6-dibromocarbazole and 1,3,6,8-tetrabromocarbazole by microplastics.

755	Marine	Pollution	Bullet	in 13	8, 458	3–463.	
756	https://doi.org/10.1016/j.marpolbul.2018.11.055						
757	Zhao, L., Rong, L., Xu, J., Lian, J., Wang, L., Sun, H., 2020a. Sorption of five organic						
758	compounds by	polar and	nonpolar	microplastics.	Chemosphere	257.	
759	https://doi.org/10.1016/j.chemosphere.2020.127206						
760	Zhao, L., Rong, L., Xu, J., Lian, J., Wang, L., Sun, H., 2020b. Sorption of five organic						
761	compounds by	polar and	nonpolar	microplastics.	Chemosphere	257.	
762	https://doi.org/10.1016/j.chemosphere.2020.127206						
763	Zocchi, M., Sommaruga, R., 2019. Microplastics modify the toxicity of glyphosate on						
764	Daphnia maga	na. Science	of T	The Total	Environment	697.	
765	https://doi.org/10.1016/j.scitotenv.2019.134194						
766	Zuo, LZ., Li, HX., Lin, L., Sun, YX., Diao, ZH., Liu, S., Zhang, ZY., Xu, XR., 2019.						
767	Sorption and desorption of phenanthrene on biodegradable poly(butylene adipate co-						
768	terephtalate)	microp	lastics.	Chemosp	ohere	215.	
769	https://doi.org/10.1016/j.chemosphere.2018.09.173						
770							