

1 **Evaluation of a novel rapid TRC assay for the detection of influenza using nasopharyngeal swabs**
2 **and gargle samples**

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20 Running Head: Evaluation of a novel rapid TRC assay for Flu

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27

28 **Abstract**

29 We evaluated a novel transcription-reverse transcription concerted reaction (TRC) assay that can detect
30 influenza A and B within 15 min using nasopharyngeal swab and gargle samples obtained from patients
31 with influenza-like illness, between January and March, 2018, and between January and March, 2019.
32 Based on the combined RT-PCR and sequencing results, in the nasal swabs, the sensitivity and specificity
33 of TRC for detecting influenza were calculated as 1.000 and 1.000, respectively. In the gargle samples, the
34 sensitivity and specificity of TRC were 0.946 and 1.000, respectively. The TRC assay showed comparable
35 performance to RT-PCR in the detection of influenza viruses.

36

37 **Keywords:** rapid detection; influenza; TRC method; RT-PCR

38

39 All authors meet the ICMJE authorship criteria.

40

41 **Introduction**

42 The transcription-reverse transcription concerted reaction (TRC) method is a combination of direct
43 and rapid isothermal RNA amplification and real-time identification using an intercalation-activating
44 fluorescence (INAF) probe [1,2]. In Europe, Japan, and Vietnam, TRC ready-to use reagents have been
45 used for the diagnosis of tuberculosis [3], nontuberculous mycobacterial infections, *Chlamydia* infection,
46 gonorrhoea, and mycoplasma pneumonia. Recently, a novel TRC assay that can detect influenza A and B
47 within 15 min was developed [4,5]. In this study we evaluated the efficacy of the novel rapid TRC assay
48 for detecting influenza viruses in nasopharyngeal swab and gargle samples obtained from patients with
49 influenza-like illness.

50

51 **Materials and Methods**

52 *Ethics*

53 This study was approved by the ethics committee of Nagasaki University Hospital (approval numbers:
54 17121822 and 18111919) and was registered at the UMIN Clinical Trials Registry (reference numbers:
55 UMIN000032395 and UMIN000034545). Written informed consent for participation in and publication of
56 this study was obtained from all participants before sample collection.

57

58

59 *Study design*

60 A prospective observational study was conducted in period 1, between January and March, 2018, and period
61 2, between January and March, 2019. Patients who visited or were hospitalized at the Department of
62 Respiratory Medicine, Japanese Red Cross Nagasaki Genbaku Hospital, with influenza-like illness (ILI)
63 [6] were included in this study. Patients were excluded if they were administered anti-influenza agents
64 within one month prior to the day on which they were sampled. Nasopharyngeal swabs were collected from
65 patients in both periods using two swabs; one was used for antigen testing using silver amplification
66 immunochromatography (FUJI DRI-CHEM IMMUNO AG Cartridge FluAB, Fujifilm, Kanagawa, Japan)
67 [7] at Nagasaki Genbaku Hospital and the other was stored at -20°C until further analysis. Gargle samples
68 were collected from patients in period 2. For gargle sampling, the patients gargled their throat with 20 mL
69 distilled water for 10 sec. The physicians determined the clinical diagnosis with history taking, physical
70 findings, and results of the influenza antigen test. All information, such as clinical report forms and results
71 of the TRC and RT-PCR, was summarized and analyzed at Nagasaki University Hospital.

72

73 *Sample analysis*

74 We performed a rapid TRC assay based on the protocol described in Japan-patent (JP,2017-195871, A)
75 at Nagasaki University Hospital. In summary, the procedure was performed as follows. A nasopharyngeal
76 swab or gargle swab was mixed in 1 mL extraction buffer containing surfactant and incubated at 52°C for

77 1 min. Thirty microliters of the sample was mixed with dry reagent containing enzymes, substrates, primers,
78 and INAF probes. The mixture was incubated at 46 °C and the fluorescence was monitored. RT-PCR and
79 sequencing were performed as a gold standard at Tosoh Corporation based on the Influenza Diagnosis
80 Manual [8–10]. Total RNA was isolated from 140 µL of TRC extraction buffer mixed with a
81 nasopharyngeal swab or 140 µL of a gargle specimen using the Qiagen RNeasy kit. RT-PCR was performed
82 using the One Step PrimeScript™ RT-PCR Kit (TAKARA BIO, Shiga, Japan). TRC assay and RT-PCR
83 were performed independently, and the results of them were combined at Nagasaki University Hospital
84 after all analysis was completed. If the results of the TRC and RT-PCR were different, the samples were
85 analyzed by sequencing.

86

87 *Statistical analysis*

88 All statistical analyses were performed with EZR version 1.41 (Saitama Medical Center, Jichi Medical
89 University, Saitama, Japan) [11], which is a graphical user interface for R (the R Foundation for Statistical
90 Computing, Vienna, Austria; version 3.6.3). Fisher's exact test was used to compare categorical variables,
91 and the statistical significance level was set at <0.05. The sensitivity (Se), specificity (Sp), positive
92 predictive value (PPV), and negative predictive value (NPV) of the TRC against the combined results of
93 the RT-PCR and sequencing were calculated with 95% confidence intervals (95% CI).

94

95 **Results**

96 *Patient characteristics*

97 During the study period, a total of 188 patients were evaluated, comprising 92 patients in period 1 and
98 96 patients in period 2. Patient characteristics are shown in Table 1. Of the patients, 95 (50.5%) visited a
99 hospital within 24 h of the onset of symptoms. The percentage of patients with fatigue was significantly
100 lower in period 1 (37.0%) than in period 2 (71.9%, $P = <0.001$). The influenza antigen test using silver
101 amplification immunochromatography detected influenza A and B in 38 (20.2%) and 39 (20.7%) patients,
102 respectively. The percentage of influenza A was significantly lower, while that of influenza B was
103 significantly higher, in period 1 than in period 2 (Table 1).

104

105 *Comparison of TRC and RT-PCR results*

106 The results of the RT-PCR and TRC are shown in Table 2. In the nasal swabs, influenza A and B were
107 detected using RT-PCR in 36 (19.1%) and 39 (20.7%) patients, respectively, and were detected using TRC
108 in 38 (20.2%) and 40 (21.3%) patients, respectively (Table 2). Of all patients, three tested negative for
109 influenza with RT-PCR, but positive with TRC. Influenza A was detected in two of these three patients and
110 influenza B was detected in one by sequencing. Based on the combined RT-PCR and sequencing results,
111 the Se, Sp, PPV, and NPV of the TRC were 1.000, 0.973, 0.962, and 1.000, respectively (Table 3).

112 In the gargle samples, influenza A was detected by RT-PCR and TRC in 37 (38.5%) and 35 (36.5%)

113 patients, respectively (Table 2). Of all patients in period 2, two tested positive for influenza A with RT-PCR,
114 but negative with TRC. Influenza A was detected in these two patients by sequence analysis. Based on the
115 combined RT-PCR and sequence analysis results, the Se, Sp, PPV, and NPV of the TRC in the gargle
116 samples were 0.946, 1.000, 1.000, and 0.967, respectively (Table 3).

117

118 *Comparison of results between nasopharyngeal swabs and gargle samples*

119 In the RT-PCR testing, five patients tested negative for influenza A in their nasopharyngeal swabs, but
120 tested positive for influenza A in their gargle samples (Fig. 1). The Se, Sp, PPV, and NPV of the RT-PCR
121 in the gargle samples were 1.000, 0.922, 0.865, and 1.000, respectively. The Se, Sp, PPV, and NPV of the
122 TRC in the gargle samples were 0.912, 0.935, 0.886, and 0.951, respectively.

123

124 **Discussion**

125 The novel rapid TRC assay showed great sensitivity and specificity in nasopharyngeal swabbing in
126 both periods 1 and 2. Based on the results of the antigen test, period 1 was considered to be the influenza
127 B epidemic season and period 2 was considered the influenza A epidemic season (Table 1). The
128 sensitivity and specificity of the rapid TRC assay was 1.000 and 1.000, respectively, based on the
129 combined RT-PCR and sequencing results for both periods. There are several rapid RT-PCR assays for
130 detection of influenza, such as the ID Now influenza A & B 2 assay (ID Now), Cobas influenza A/B

131 nucleic acid test (Liat), and Xpert Xpress Flu assay (Xpert). The previous studies reported that the
132 sensitivity and specificity of these methods for detecting influenza A/B was 0.932 to 1.000/0.917 to 1000
133 and 0.977 to 1.000/0.976 to 0.998, respectively [12–14]. These results indicate that the performance of
134 the rapid TRC assay is comparable to that of rapid RT-PCR assays.

135 In the present study, the rapid TRC assay also showed great sensitivity and specificity for gargle
136 sampling. The rapid TRC assay and RT-PCR detected influenza in more patients from gargle samples
137 than from nasopharyngeal swabs. The sensitivity and specificity of the Rapid TRC assay were 0.946 and
138 1.000, respectively. The previous studies reported that the sensitivity of Xpert and Liat for gargle
139 sampling was 0.917 and 1.000, respectively, in comparison with in-house RT-PCR [15,16]. Although
140 there are no data on the specificity of rapid RT-PCR assays, the rapid TRC assay seems to be comparable
141 to rapid RT-PCR assays for gargle samples. In the diagnosis of influenza, nasopharyngeal swabbing is the
142 major sampling type, but these samples are difficult to obtain and the procedure is uncomfortable for
143 patients [17]. In addition, due to the novel coronavirus (SARS-CoV-2) pandemic, healthcare workers
144 must wear personal protective equipment when they obtain nasopharyngeal swabs. Therefore, it is vital to
145 develop a diagnostic method, such as the rapid TRC assay, using gargle samples, that is easy to perform,
146 non-invasive, material saving, and safe for healthcare workers [18].

147 There are some limitations in this study. First, the samples were obtained from one community hospital
148 in Nagasaki, which might have limited the generalizability of the findings. Second, we used an equipment

149 of the novel TRC assay under development. Accordingly, we are conducting a multicenter study for the
150 rapid TRC assay using production version. Third, the rapid TRC assay was not compared with other rapid
151 RT-PCR assays. Since these assays have not yet been approved by the Japanese Pharmaceuticals and
152 Medical Devices Agency, we will conduct a comparative study in the future.

153 In conclusion, the novel rapid TRC assay showed comparable performance to RT-PCR in the detection
154 of influenza viruses. In addition, because it could detect influenza viruses using gargle samples, the rapid
155 TRC assay could contribute to the diagnosis of influenza during the SARS-CoV-2 pandemic.

156 **Ethical approval:** This study was approved by the ethics committee of Nagasaki University Hospital

157 (approval numbers: 17121822 and 18111919)

158 **Consent to participate:** Written informed consent for participation in and publication of this study was

159 obtained from all participants before sample collection.

160 **Consent to publish:** Written informed consent for publication of this study was obtained from all

161 participants before sample collection.

162 **Authors contributions:** All authors meet the ICMJE authorship criteria.

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165 **Competing interests:** The authors have no conflict of interest directly relevant to the content of this article.

166 **Availability of data and materials:** Raw data were generated at Nagasaki University Hospital. Derived

167 data supporting the findings of this study are available from the corresponding author on request.

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253

254 **Figure legends**

255 **Fig. 1. Comparison of gargle sample and nasopharyngeal swab results.**

256 The results obtained from gargle samples and nasopharyngeal swabs during period 2 were compared.

257 Period 2 was between January 1 and March 31, 2019.

258 **Tables**259 **Table 1. Patient characteristics.**

Characteristic	Overall (N=188)		Period 1 (N=92)		Period 2 (N=96)		P value
	N	(%)	N	(%)	N	(%)	
Age (average \pm S.D)	50.5	\pm 19.4	51.3	\pm 19.5	49.6	\pm 19.5	NS
Gender = female	103	(54.8%)	58	(63.0%)	45	(46.9%)	0.029
Underlying diseases	94	(50.0%)	48	(52.2%)	46	(47.9%)	NS
Time since onset of symptoms							
0–12 h	48	(25.5%)	23	(25.0%)	25	(26.0%)	NS
12–24 h	47	(25.0%)	23	(25.0%)	24	(25.0%)	NS
24–48 h	39	(20.7%)	17	(18.5%)	22	(22.9%)	NS
48–72 h	19	(10.1%)	7	(7.6%)	12	(12.5%)	NS
72+ h	26	(13.8%)	14	(15.2%)	12	(12.5%)	NS
Unknown	9	(4.8%)	8	(8.7%)	1	(1.0%)	0.017
Symptoms							
Fever	143	(76.1%)	66	(71.7%)	77	(80.2%)	NS
Cough	124	(66.0%)	60	(65.2%)	64	(66.7%)	NS
Fatigue	103	(54.8%)	34	(37.0%)	69	(71.9%)	<0.001

Sore throat	100 (53.2%)	46 (50.0%)	54 (56.3%)	NS
Nasal discharge	99 (52.7%)	47 (51.1%)	52 (54.2%)	NS
Headache	70 (37.2%)	35 (38.0%)	35 (36.5%)	NS
Arthralgia	56 (29.8%)	23 (25.0%)	33 (34.4%)	NS
Myalgia	56 (29.8%)	22 (23.9%)	34 (35.4%)	NS
Diarrhea	11 (5.9%)	3 (3.3%)	8 (8.3%)	NS
Nausea	9 (4.8%)	3 (3.3%)	6 (6.3%)	NS

Results of influenza antigen test using silver amplification immunochromatography

Influenza A	38 (20.2%)	3 (3.3%)	35 (36.5%)	<0.001
Influenza B	39 (20.7%)	38 (41.3%)	1 (1.0%)	<0.001
Negative	111 (59.0%)	50 (54.3%)	60 (62.5%)	NS

S.D. = standard deviation; NS = not significant.

261 **Table 2. Results of RT-PCR and TRC.**

Item	Overall (N=188)		Period 1 (N=92)		Period 2 (N=96)		P value
	N	(%)	N	(%)	N	(%)	
RT-PCR in nasopharyngeal swab							
Influenza A	36	(19.1%)	4	(4.3%)	32	(33.3%)	<0.001
subtype H1	9	(4.8%)	0	(0.0%)	9	(9.4%)	0.003
subtype H3	27	(14.4%)	4	(4.3%)	23	(24.0%)	0.001
Influenza B	39	(20.7%)	39	(42.4%)	0	(0.0%)	<0.001
Negative	113	(60.1%)	49	(53.3%)	64	(66.7%)	NS
TRC in nasopharyngeal swab							
Influenza A	38	(20.2%)	4	(4.3%)	34	(35.4%)	<0.001
Influenza B	40	(21.3%)	40	(43.5%)	0	(0.0%)	<0.001
Negative	110	(58.5%)	48	(52.2%)	62	(64.6%)	NS
RT-PCR in gargle sample							
Influenza A	N/A		NCW		37	(38.5%)	N/A
subtype H1	N/A		NCW		11	(11.5%)	N/A
subtype H3	N/A		NCW		26	(27.1%)	N/A
Influenza B	N/A		NCW		0	(0.0%)	N/A

Negative	N/A	NCW	59 (61.5%)	N/A
TRC in gargle sample				
Influenza A	N/A	NCW	35 (36.5%)	N/A
Influenza B	N/A	NCW	0 (0.0%)	N/A
Negative	N/A	NCW	61 (63.5%)	N/A

NS = not significant; N/A = not applicable; NCW = not complied with.

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263 **Table 3. Performance of TRC assay for detection of influenza.**

	Overall	Period 1	Period 2
Nasopharyngeal swab			
TP	78	44	34
TN	110	48	62
FP	0	0	0
FN	0	0	0
Se (95% CI)	1.000 (0.931-1.000)	1.000 (0.882-1.000)	1.000 (0.851-1.000)
Sp (95% CI)	1.000 (0.951-1.000)	1.000 (0.891-1.000)	1.000 (0.915-1.000)
PPV (95% CI)	1.000 (0.931-1.000)	1.000 (0.882-1.000)	1.000 (0.851-1.000)
NPV (95% CI)	1.000 (0.951-1.000)	1.000 (0.891-1.000)	1.000 (0.915-1.000)

Gargle samples

TP	35	N/A	35
TN	59	N/A	59
FP	0	N/A	0
FN	2	N/A	2
Se (95% CI)	0.946 (0.818-0.993)	N/A	0.946 (0.818-0.993)
Sp (95% CI)	1.000 (0.911-1.000)	N/A	1.000 (0.911-1.000)
PPV (95% CI)	1.000 (0.855-1.000)	N/A	1.000 (0.855-1.000)
NPV (95% CI)	0.967 (0.887-0.996)	N/A	0.967 (0.887-0.996)

Period 1 = between January 1 and March 31, 2018; period 2 = between January 1 and March 31, 2019;

TP = true positive; TN = true negative; FP = false positive; FN = false negative; CI = confidence

interval; N/A = not applicable.

RT-PCR				TRC			
Nasopharyngeal swab				Nasopharyngeal swab			
		Pos	Neg			Pos	Neg
Gargle	Pos	32	5	Gargle	Pos	31	4
	Neg	0	59		Neg	3	58

Se, Sp, PPV, and NPV in gargle samples

Se	1.000	(0.842-1.000)	0.912	(0.763-0.981)
Sp	0.922	(0.827-0.974)	0.935	(0.843-0.982)
PPV	0.865	(0.712-0.955)	0.886	(0.733-0.968)
NPV	1.000	(0.911-1.000)	0.951	(0.863-0.990)

Fig. 1. Comparison of results in gargle samples with nasopharyngeal swabs

The results obtained from gargle samples and nasopharyngeal swabs during period 2 were compared. Period 2 was between January 1 and March 31, 2019.