

# Identification and Characterization of a Fimbrial Gene Cluster of *Edwardsiella tarda* Expressing Mannose-resistant Hemagglutination

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**ABSTRACT**—We determined the nucleotide sequence of a 8.6 kb DNA region containing *etfA* encoding a putative fimbrial major subunit from chromosomal DNA of *Edwardsiella tarda* KG8401 which expresses mannose-resistant hemagglutination (MRHA). This region contained three novel genes, *etfBCD*, at the downstream region of *etfA*. The deduced amino acid sequences of EtfABCD contained conserved fimbrial domains; fimbrial protein, fimbrial chaperone, fimbrial usher and fimbrial protein, respectively. *Escherichia coli* transformed with the cloned *etf* operon expressed MRHA and fimbriae that reacted with rabbit antiserum against the fimbrial major subunit of *E. tarda*, showing the implication of the fimbriae in the hemagglutination of *E. tarda*.

**Key words:** *Edwardsiella tarda*, mannose-resistant hemagglutination, fimbriae, gene cluster

*Edwardsiella tarda* has a wide host range, having been isolated from a variety of animals including fish, birds, mammals and reptiles (White *et al.*, 1973; Owens *et al.*, 1974; van Damme and Vandepitte, 1980; Goldstein *et al.*, 1981). Edwardsiellosis in Japanese flounder *Paralichthys olivaceus*, red sea bream *Pagrus major* and Japanese eel *Anguilla japonica* is characterized by skin lesions and formation of abscesses and granulomas in internal organs such as liver, kidney and spleen (Miyazaki and Kaige, 1985; Rashid *et al.*, 1997) and causes serious damage to their aquaculture industry (Wakabayashi and Egusa, 1973; Nakatsugawa, 1983; Miyazaki and Kaige, 1985; Kusuda and Salati, 1993).

Pathogenicity of *E. tarda* seems to be multifactorial. Several virulence properties and factors have been reported, namely dermonecrotic and lethal toxins (Ullah and Arai, 1983; Suprpto *et al.*, 1996), anti-phagocyte killing (Ainsworth and Chen, 1990; Iida and Wakabayashi, 1993), hemolysins (Janda and Abbott, 1993; Chen *et al.*, 1996; Hirono *et al.*, 1997; Strauss *et al.*, 1997), siderophore (Igarashi *et al.*, 2002), serum resistance and the ability to invade epithelial cells (Janda *et al.*, 1991; Ling *et al.*, 2000). However, very little is known

about the roles of these factors in disease occurrence and the bacterial invasion process, and also the portal of entry of *E. tarda* into hosts has not been shown precisely.

For many pathogenic bacteria, adherence to host is mediated by adhesin existing on the tip of fimbriae (Hoepelman and Tuomanen, 1992; Kuehn *et al.*, 1992; Jones *et al.*, 1995). Although *E. tarda* has fimbriae that exhibit mannose-resistant hemagglutination against guinea pig erythrocytes, hemagglutinin has not been identified (Nowotarska and Mulczyk, 1977). In the previous paper, we identified a gene, *etfA*, encoding a 19.3 kDa protein that was associated with the possession of hemagglutinating activity among *E. tarda* strains (Sakai *et al.*, 2003). The predicted amino acid sequence of EtfA has a significant homology with type-1 fimbrial major subunits of *Serratia marcescens* (Nichols *et al.*, 1990) and *Escherichia coli* O157:H7 (Perna *et al.*, 2001) and long polar fimbrial major subunit of *Salmonella typhimurium* (Bäumler and Heffron, 1995). Type-1 and P fimbriae of *E. coli* are constructed with several thousands copies of the major subunit, forming helical filament, and several minor subunits, and carbohydrate-binding adhesin is located at the distal end of the fimbria (Lindberg *et al.*, 1987; Gong and Makowski, 1992; Kuehn *et al.*, 1992; Jones *et al.*, 1995). Genes encoding fimbrial subunits and accessory proteins participating

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in the formation of fimbriae constitute an operon, and the gene encoding adhesin is located downstream in a fimbrial gene cluster (Boyd and Hartl, 1998).

In this study, we determined the DNA sequence of a fimbrial gene cluster, *etfABCD*, from chromosomal DNA of *E. tarda* and showed that *E. coli* harbored a DNA fragment containing *etfABCD* expressed mannose-resistant hemagglutination against guinea pig erythrocytes.

## Materials and Methods

### *Bacterial strains and growth conditions*

*E. tarda* KG8401 isolated from Japanese eel was cultured at 28°C for 24 h on yeast extract agar, consisted of 1% polypepton, 0.5% Bacto-yeast extract (Difco Laboratories, USA), 0.5% NaCl and 1.5% agar at pH 7.2. *E. coli* JM109 was grown in Luria-Bertani (LB) medium (Sambrook *et al.*, 1989).

### *Hemagglutination and hemagglutination inhibition test*

Hemagglutination test and hemagglutination inhibition test were performed according to Sakai *et al.* (2003). Briefly, a 3% (v/v) formalin-fixed erythrocyte suspension in phosphate-buffered saline containing 1% bovine serum albumin (PBS-BSA) was added to serial twofold dilution of bacterial cells suspended in PBS-BSA at 0.1 g (wet wt) bacteria/mL. Hemagglutination titer was defined as the reciprocal of the highest dilution of a test sample that showed complete hemagglutination. For hemagglutination inhibition test, serial twofold dilution of bacterial cells was prepared in PBS-BSA contained 1% D-mannose or 1% fetuin (Sigma Aldrich, USA).

### *DNA manipulation*

Preparation of chromosomal DNA, restriction endonuclease digestion, ligation, transformation and DNA electrophoresis were performed as described by Sambrook *et al.* (1989). Plasmid DNA was purified using Quantum Prep Plasmid Miniprep Kit (Bio-Rad Laboratories, USA).

Southern hybridization and colony hybridization were performed according to the DIG systems user's guide (Roche Applied Science, Germany). The oligonucleotide probe was end-labeled with digoxigenin (DIG) using DIG Oligonucleotide 3'-End Labeling Kit (Roche Applied Science, Germany).

Nucleotide sequences were determined by the dideoxy chain termination method with BigDye Terminator Cycle Sequencing Kit and ABI 377 DNA sequencer (Applied Biosystems, USA).

### *Computerized sequence analysis*

The nucleotide sequence data was analyzed using the DNASIS program (Hitachi Software, Japan) and the GENETYX sequence analysis program (Software Development, Japan). Analysis of a signal sequence of the

deduced amino acid sequence was performed with the SOSUI program available at the Tokyo University of Agriculture and Technology website (<http://sosui.proteome.bio.tuat.ac.jp/sosuimenu0.html>). Homology searching was performed using the FASTA program served by Genome Net (Bioinformatics Center, Institute for Chemical Research, Kyoto University, <http://fasta.genome.ad.jp/SIT/FASTA.html>). The conserved domain database (CDD) and CD-Search service at the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/Structure/cdd/cdd.shtml>) were used to identify the conserved domains present in protein sequences.

### *Expression of EtfABCD in E. coli JM109*

A DNA fragment containing *etfABCD* was PCR amplified with sense primer, 5'-CACTTTCCGCAACC-ATGATC-3' (positions 1574 to 1593 in Fig. 2), and antisense primer, 5'-CTCTCCTTGTCACAATAACGC-3' (positions 7290 to 7310 in Fig. 2). PCR was performed with TaKaRa LA PCR Kit (TaKaRa Bio Inc., Japan) according to the following amplification protocol: after an initial denaturation step (94°C, 2 min), 30 cycles of denaturation (94°C, 30 s), annealing (65°C, 30 s) and extension (72°C, 7 min), followed by a final extension step (72°C, 10 min). The PCR product was ligated into a pGEM-T Easy vector (Promega, USA) and transformed into *E. coli* JM109. Hemagglutinating activity and fimbrial formation of cells transformed with recombinant plasmid DNA were then examined.

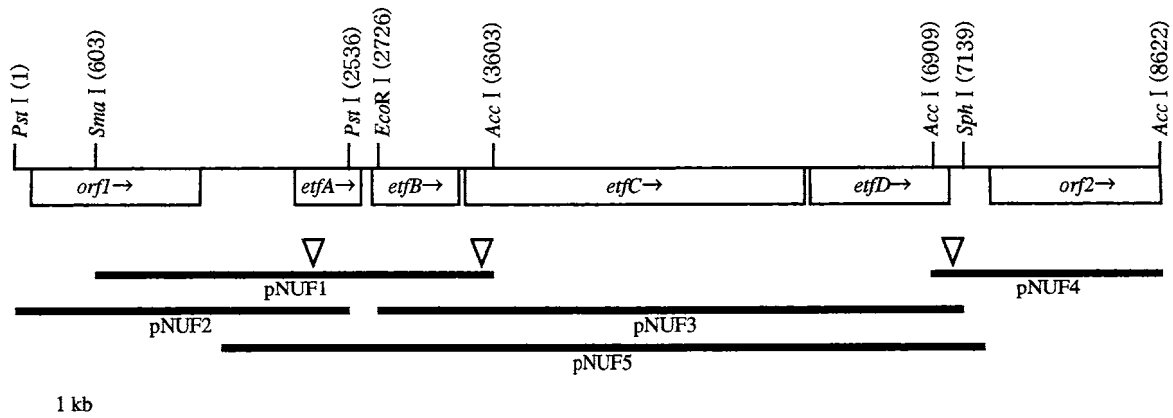
### *Immunogold electron microscopy*

Bacterial cells were incubated for 1 h at room temperature in rabbit anti-19.3-kDa protein (EtfA) serum (Sakai *et al.*, 2003) diluted to 1:100 in PBS-BSA at 10 mg (wet wt) bacteria/mL. After washed thrice with PBS-BSA, bacterial cells were incubated for 1 h at room temperature in goat anti-rabbit IgG-gold (10 nm; Sigma USA) diluted to 1:100 in PBS-BSA. After washed thrice with 2% ammonium acetate, bacterial cells were negatively stained with 1% sodium phosphotungstate (pH 7.0) and examined under a JEM 100S electron microscope (JEOL, Japan) at 80 kV.

## Results and Discussion

### *Cloning and sequencing of the etf operon*

Two oligonucleotide probes were designed based on the nucleotide sequence of a 3.0-kb *Sma*I-*Acc*I fragment containing *etfA* (Sakai *et al.*, 2003), pNUF1 in Fig. 1. Hybridized 2.5-kb *Pst*I and 4.4-kb *Eco*R I-*Sph*I fragments from chromosomal DNA were each subcloned into pUC18, designated pNUF2 and pNUF3 (Fig. 1), respectively, and sequenced. And also, a 1.7-kb *Acc*I fragment hybridized with an oligonucleotide probe designed from the nucleotide sequence of the *Eco*R I-



**Fig. 1.** Restriction and genetic map of the *E. tarda* KG8401 chromosome DNA surrounding *etfA*. Boxes and arrows indicate the open reading frames and the direction of transcription, respectively. The positions of the DNA fragments cloned in 5 recombinant plasmid DNA derivatives (pNUF1, pNUF2, pNUF3, pNUF4 and pNUF5) are given at the bottom of the figure. Triangles indicate the site of the oligonucleotide probes.

*Sph* I fragment was cloned into pUC18, designated pNUF4 (Fig. 1). The nucleotide sequence of a 8,627-bp DNA region consisted of these overlapping fragments was determined (Fig. 2). This region contained 4 open reading frames, designated *etfABCD*, and a complete open reading frame, designated *orf1*, and a partial open reading frame, designated *orf2*, were found at the sides of the *etf* genes (Figs. 1 and 2). Start and stop codons of the *etf* genes were separated by only a few base pairs. Furthermore, putative ribosome binding sites (Shine and Dalgarno, 1975) were identified at the upstream regions of each gene from *etfA* to *etfD* (Fig. 2). And also, putative promoter sequences located at the upstream regions of *etfA* and *orf2* (Fig. 2). It is suggested that the *etf* genes constitute an operon. The positions of *etfABCD* and sizes of the predicted polypeptides are shown in Table 1 and Fig. 1. All predicted polypeptides were found to contain acceptable signal sequences.

#### Homology of *EtfABCD* to other fimbrial proteins

The deduced amino acid sequences of all 4 *etf* genes showed homology to proteins from various bacterial fimbrial systems. *EtfABCD* revealed homology with type 1 fimbrial major subunit (FimA) of *S. marcescens* (68.9% identity; E value, 4.7e-42) (Nichols *et al.*, 1990), a putative long polar fimbrial chaperone protein (LpfB) of *E. coli* (38.4% identity; E value, 2.7e-26), F1C fimbrial usher protein (FocD) of *E. coli* (39.2% identity; E value, 3.3e-99) (Welch *et al.*, 2002) and a fimbrial protein of *E. coli* (26.3% identity; E value, 2.5) (Hayashi *et al.*, 2001), respectively. The deduced amino acid sequences of *Orf1* and *Orf2* revealed homology with inosine kinase of *Yersinia pestis* (85.7% identity; E value, 1.5e-155) (Parkhill *et al.*, 2001) and UDP-sugar hydrolase of *Photobacterium luminescens* subsp. *laumondii* (72.4% identity; E value, 2.7e-135) (Duchaud *et al.*, 2003),

respectively. And also, the domains of fimbrial proteins participating in the formation of fimbriae were shown in the deduced amino acid sequences of mature *EtfABCD* on the CD-search (Table 2). It is suggested that *EtfA*, *EtfB*, *EtfC* and *EtfD* function as a fimbrial major subunit, fimbrial chaperone, fimbrial usher and fimbrial subunit, respectively. It is known that long polar fimbriae and F1C fimbriae of *E. coli* are synthesized by chaperone-usher fimbrial biosynthesis system (Soto and Hultgren, 1999). Fimbriae of *E. tarda* may consist of fimbrial subunits, *EtfA* and *EtfD*, mediated by fimbrial chaperone, *EtfB*, and fimbrial usher, *EtfC*.

#### Electron microscopic observation and hemagglutination of *E. coli* harbored the *etf* fimbrial operon

Immunogold-labeling electron microscopy revealed that gold particles attached to the fimbrial structure of *E. coli* JM109 harbored plasmid DNA containing the *etf* operon (pNUF5) (Fig. 3A). On the contrary, no gold particles were observed on the fimbrial structure of *E. coli* JM109 lacking pNUF5 (Fig. 3B).

*E. coli* JM109 harbored pNUF5 acquired hemagglutinating activity, which was not inhibited by D-mannose but was strongly inhibited by fetuin like *E. tarda* KG8401 (Table 3). It is suggested that fimbriae encoded in the *etf* gene cluster participate in the mannose-resistant hemagglutination of *E. tarda*.

Adherence of pathogenic bacteria to host is considered to be the first step in infection. Although adherence of *E. tarda* on the gill and body surface has been observed in experimentally infected fish (Ling *et al.*, 2001), the factors associated with the adherence were not identified. For many pathogenic bacteria, it has been shown that adherence to host is mediated by fimbriae possessing hemagglutinating activity (Beachey, 1981). In this study, we identified an *etf* gene cluster associated with the formation of fimbriae expressing

1 CTGCAGCAGTGGTCCGGTACCCGCGGTAGGATGCCCGTACGCGGCGGGGGAGTGTGCTATTCTAGCCCGCTCCCTTTTCTTTTGGCGATAACACAGATGAAATCCCCGGT 120  
M K F P G

121 CAACGCAAGTCCAAACTATTCCCGGTTAATGCCCCGCGATCCGCTGCTGCAAAAGACGCGCATAAATAAAGCTACGCTCGTGGTATCGATCAGACGCTGGTGGATATT 240  
Q R K S K H Y F P V N A R D P L L Q K T Q H N E I D K A Y V V G I D Q T L V D I

241 GAGGCCAAGTGGATGAGCGTTTATCGCCGCTACGGCCTGAGCCAGGGCAGCTCGTGGTATCGAGGATGATGTCCGCGAAGCGCTGTACCAGGAGCTGTGTGACAGTAAGCTGATT 360  
E A K V D E A K F I A R Y G L S Q G H S L V I E D D V A E A L Y Q E L C D S K L I

361 ACCCATGAGTTTGGCGGGGCGATCGGCAATACCCCTGCACAACTACTCCGTGCTGGCGGAGATCGCTCGGTTCTGCTGGGGGTGATGTGCGAAAACGTCAAAATCGGCAGCTATGCC 480  
T H E F A G G T I G N T L H N Y S V L A D D R S V L L G V M C E N V K I G S Y A

481 TACCGCTATCTGTGTAACACCTCAAGCCGACCGATCTGGACCAITTTGCAGGCGTTCGATGGCCATCGGGCGCTGTTTACCCTGATCACCAGTAAACGGCGAAGCCACCTTTGCCATT 600  
Y R Y L C N T A S S R T D L D H L Q G V D P I G R C P T L I T D N G E R T F A I

601 AGCCCGGGCATGATGAACAGCTCGCCCGGAAAGCATTCCCGAGGCGGTGATCGCCGCTGCTCGGCGTGGTCTGACCGCCTATCTGGTGGCTTAAGCCGGGCGAGCCGATGCCG 720  
S P G M M N Q L R P E S I P E A V I A G A S A L V L T A Y L V R C K P G E P M R

721 GATGCGACCATGCGGCGATCGCCTATGCAAAAAGCACCAGTTCGCGTGGTGTGACCCGCGCACCAGTATGATGCGCGACGATCCGCGAGTCTGCGCGGATTTCTCGCGCAT 840  
D A T M Q V I A Y A K K H H V P V L T L T G K Y V I A D D P Q F W R D P L R D

841 CACGTCACCAITGTGGGATGAAGAAGAGGCTCAGGAGCTACCGGGCTVTCGATCGCTGGCGCCTCGGACAGGGCGCTGGAGTGGTGGATCTGGTCTGTCACCGCGGG 960  
H V T I V A M N E E E A Q E L T G L S D P L A A S D R A L E W V D L V L C T A G

961 CCGGTGCGCTGTATGCGGGGTATACCGAAGTACGCGCAAGCCKAAKCCAGCATCGCTGCTCGCGGGGTGATCCCTGATCAACCTGTACAGTATTAGCCGGGCGAGTCCGCT 1080  
P V G L Y M T S R T E D D G K R K A C T Q H P L G V I P E F N L Y E F S R A M R

1081 CGCGCCATGTGCGAGCGCGCTGCGGATCTACTCGCATATTGCGCCCTATATGGGGGCGCGAAGATCATGAACACCACGGCGGAGATGGCGCGCTCTCGGCTGCTGCGAT 1200  
R A M C D A P C R I Y S H I A P Y M G G P E K I M N T N G A G D G A L S A L L H

1201 GATATCGCGCCAAATGAGTTTACCCTGACCAATGTGCTAACTCCAGTAAGCATCAGCGTAGCTCTGACTTACTCCTCCCTGGCGAGGTGTGCAAAATGATGTAACCCGCTCAGCTAT 1320  
D I A A N T N V P N S K H Q R S Y L T Y S S L A Q V C K Y A N R V S Y

1321 CAGGTGCTCAACGACACTCGCCGCTGACCCCGGCTGCGCGAGCGGGAAGACAGCTGGAAGATCTACTCGGAGCGCTGAATAGTGTCTCGCCCGTGGCAGGTGAGATGAT 1440  
Q V L N Q H S P R L T R G L P E R E D S L E E S Y W E R \*

1441 CTTACGGAGATACCGCGGCGAGCGCGCGGTTTCTTTCTCGCGCTCGCGGTTATCTGCGCGAGTCCGTTGGCTAAAGAGGGGCGAAAAGGCTAGAACAATCTCAITTTCCCTTAAGCG 1560  
1561 GAATACCCCTTTTCACTTTCCGCAACCATGATCAAGAGCAATTTCTTGATATTGGTAATGTCATACAAAATAAATAATATGATGTAACCAATGAATTTCTTTTTCGATTGA 1680  
1681 TCAAAATCTAATAAATACTATCTTAAATTAATCAATATTGTTTTTATCAGTTTATGATTTACTGTTTCTCGCGCAATTAATCTTTGCGCAAGTTTTCACMTAATA 1800  
1801 AAATAAATATCTTCTGTAATATCAGTGAAGTATATGCGCTGACAATATTAATCAATTTGGGATTCGCAATTAATAGCGCAGTGCCTATTTTGTCTATTTAAATAA 1920

1921 ATCCTTACTCGATTGGATTTATGATTTCCGCAATAATCCATCATTTTATGCGCTGCGGAATAAAGATTTCTCGCTCGGCTCTCTAAGCGAAATATATTCGACCGCTCGTTCCGC 2040  
SD effA  
M K K I L L P V M A L A A A V S G Q A S A A N

2041 GCCAACCTAGCGGTGGATTTTTAATGATAAACGGGTTATGATAAAGAAAATTTACTGCTGTTATGGCTCTGGCGCGCGCGCTGTGAGTGGTCCAGGCAAGCGCAGCTAAC 2160  
M K K I L L P V M A L A A A V S G Q A S A A N

2161 GGTAAGTTGAGTTTACGGGTGAAATCGTTAACTCCACCTGCGAGTAAAGCAGCGACTCTCAAATATCAACGTGATTTGGGTAATATCCGACTCTGCAITTTAAGCGGTTGGCGAT 2280  
K V E F T G E I N S T C V S S D S Q N I N V Y L G S K Y P T S A F K A V G D

2281 AAGTCGCACTCCAGGCTTCCAGATCAACCTGGAGCAGTGTGAGCGGGAGCTACACCGTACGTTTCGATGGCAATACCGTTGCGCGTCACTGACCTGCTGGCGCTCCAGCAGC 2400  
K S A S K A F Q I N L E Q C E P G S Y T V R F D G N T V A G H P D L L A V S S S

2401 GGTGCCACCGCGCGGCGAAAAGGCTGGTATCGAAATACGGACGTTAAGCGCAAGCGGTTCCGATCGCCAGTACAGACCGAGGCGATGTGCCACCGTACGGCTGTAGCGATAAG 2520  
G A T D A V A K G V I E I T D V S S Q D Q S Q G D V P T V T V S D K

2521 AAGCGATATTTAACCTGAGCGCGCTATCGCTCTTACGCCAACCGGTTACGCGGCTGGCCAACCGGACCGCGTTACCATCGAATATAAATAATTTATCGTCCGGAATATTC 2640  
K A I F N L Q A R Y R S Y A N T V T A G L A N A T S P P T I E Y K \*

2641 GCCAGGCGCGTCCGTTATAGGAAATAGATGAAAAGAAATCGTTCCGCAATATTACTGGTGGGCGATTGTGCCAACCGGAAATCAGGTTGATGCTACGCGGTTATTTATAAC 2760  
SD effB  
M K R I V A A L L V S A F A H A G I Q V D A T R V I Y N

2761 GGCGATGAAAAGTCAAGCTCACTGCTTCCACATGACAGCGCGGATCGCTATATGGTGCAGACTGGCTGGATAAAGGGGATAGCACCAAGGTAGGAAATAGTTACGATGTTGTG 2880  
G D E K S A S L P I H N D S A D A Y M V Q T W L D K G D S T K V E N K L P M V V

2881 GTGCGCGGATTTAAGCTGGATGCGCAAAAATCGGCAATTTACGCTCATCTATTCTGGCAGGGTTTTCCGCGAGATCAGGAAACGCTGCTGTGGGTCAAGCTCAGGAAATCCG 3000  
V P P I V K L I D Q K S A I L R F I Y S G G P P Q D Q E T L L W V N V Q E I P

3001 CCGACGCCAAGCAAGAAAAGTATACAGGTTGCGGTCGCGCAGCGGATTAAGCTGTTTTCGCGGAGTTCGCTGCATACCACTGGATGAGCAGGTCATAGCTGCGCTGGCAG 3120  
P T P K Q E N V L Q V A V R T R I K L F Y R P S S L H T T L D E Q V H K L R W Q

3121 AAAGCGGTAATCAATGTGCGCATCAACGCGGGCGCTACACATCACTTTGGCTCTGTACAGATGCGGGCAGCAACGGCAAAACCCATCGCGTGCATGCAACATGGTGTACCC 3240  
K A G N Q L L I D G P L L A I N D G P L H I T F G S I G Q M Q L A F A G S N G K P I A V D A N M V D P

3241 CACAGCCGCTGGGATTCGCCATTCCGCGGGGCTCGGTCGCAATACAGTCTCCTTTCAGTCAATCAATGACTTTGGAGGTGGGACTGAGTTAAGGACCGGATAGTACTACGCGG 3360  
H S R L A I A I P A G A R V G N T V S F S Y I N D F G G R T E V K D A I V H \*  
SD

3361 AGAGAAGCGGATGGCGCAACCAACCGGACGATCACTGAAGACAGGCGCAGCTGGATGCTGCGGTTTCCGCGCGTACCTGCTGCTGGGTGACGCGATCGGCTGG 3480  
effC  
M A Q Q Q R T V S L K T E F T R W M C L A F A A G S L S L G C S A S A W

3481 CCGAGTTCAACATGTCATTCGTTCCGCGCAGGAAACGTCACCGCGCTCATGCGGTGCGAGGGCAACCGCTGCGCGCGGAGATCCGTTCCGATTTACCTGAACAGCAGC 3600  
A E F N M S F V H G S E N V N A A H A V A E G N A L P P G E Y P F A I Y L N K Q

3601 AGTCCGCAACAAGTCACTCCGCTTACCGCGGAAAGAGGGCGCGCGCCAGCCGCTGTCGACGTTGAGATCTGCTTAACATGCGGTGAAATCGCGGATACGCTCAGCGGGCA 3720  
Q V D N K V I R F T A K E G A A A Q P C L T V E D L L L N Y G V K L L P D T L T P G

3721 GCTGCGTCGAGTGGCGCGCGCTCAATGGCGGAGCGCTCCTTCGATCCGCGGCTGCGAGCAGTACGCTTCTGTCGCGCAGGTCATGCTGGCGCAGCGCGCGGCGCCATT 3840  
S C V D V A A A V N G A T L S F D P A V Q Q I D L S V P Q V M L A Q H A R G A I

3841 CGACGACCTGTACGATCAGGCGATCAACCGCGCTTTAGCAACTACAGCTGAACTATAACCGTAACGACGCGCGCGGAGCGCGCAACAGCGAATATACCTTCTCCCTGGA 3960  
P T T L Y D Q G I N A A F S N Y S L L N Y N R N S Q R Q S G Q N S E Y T F L S L

3961 ATAACGCTTAACCTGGAATGCGCTGCGCAATACATGACCATGAATGAATGCGGTCAGGCGGCAAGTGGGCAATATCGGACCTGGCGGAGACCGATTTGCGTCT 4080  
N N G L N L W N R L R N N M T M N K V S G Q A A Q W D N I A T W A E T D I A S

4081 GCGCAGTGCAGCTGCGCGCGGCGCAGCACAATATACCGTTTTTATAGCATTCAGTTTGGCGGTCGCCAGCTTTCCAGCGTGGATGACATGCTGCCGCAAGGCTGCGCGGCT 4200  
W R S R L Q L G Q A S T N N T V P N S I Q F R G A Q L S S V D D M L P E S L R G

4201 ATGCGCGGTTGTCGCGGCTCGCCGACCAACCGCGCTGCACATTCAGAACCGGCTGGTGTGACAGCACCAAGCTGGCGCGGCGCCCTTGGATCCGCGATCTATC 4320  
Y A P V V G V A A T N A R V D I R Q N G Y V V Y S T N V A P G P F E I R D I Y

4321 CGCACTCCACAGCGTGACCTGCAAGTAAACGCTCAATGAGGCGGATGGTACCGCTTAACGCTTTTACTCTCTATCGCCACATGCTGCGCAAGGATCTGCAACTATC 4440  
P H S N S G D L Q V T V N E A D G T R K H F T V S Y S S I A N M L R E G I W N Y

4441 AGCTGACGCGGCTCCTACCGAAGCGCGATACCCGCTATACCCCTAATTTATGCGAGGCGGCTGCGCGCGCATGCCCTATGCTGACGCCCTATGCGCGGCTGATGCGGCGG 4560  
Q L T Y A G D T R Y T P N F M Q G T L A R G I A Y G L T P Y G G L I V A



**Table 1.** Size and positions of genes and gene products of the *etf* operon

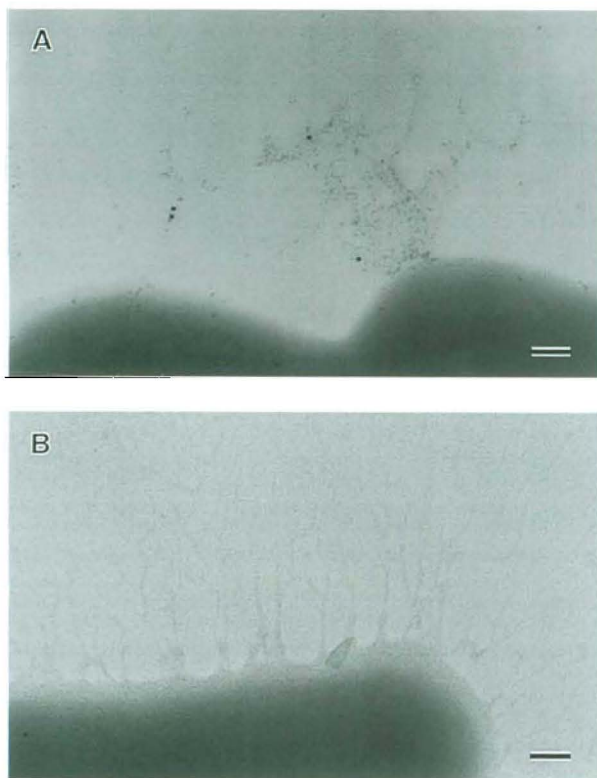
ORF	Position (bp)*	Length of polypeptide (amino acids)	Signal sequence (amino acids)	Calculated mass of mature protein
<i>etfA</i>	2089-2622	177	1-22	16,246
<i>etfB</i>	2671-3357	228	1-18	23,162
<i>etfC</i>	3372-5963	863	1-35	90,336
<i>etfD</i>	5976-7046	356	1-26	35,045

\* Position in the nucleotide sequence shown in Fig. 1.

**Table 2.** Protein domain conserved in the deduced amino acid sequences of EtfABCD

Protein	Position (amino acids)	Conserved domain		
		Domain name (CDD* accession no.)	Function	E-value
EtfA	17-169	Fimbrial (981)	Fimbrial protein	4e-30
EtfB	19-221	FimC (12460)	P pilus assembly protein	7e-47
EtfC	38-841	Usher (16770)	Fimbrial usher protein	2e-172
EtfD	228-356	Fimbrial (981)	Fimbrial protein	0.43

\* Conserved domain database served by NCBI.



**Fig. 3.** Immunogold electron micrographs showing *E. coli* (pNUF5) (A) and *E. coli* JM109 (B). Bars; 100 nm.

mannose-resistant hemagglutination. Further studies are needed to determine the roles of fimbriae in disease occurrence and the bacterial invasion process in piscine edwardsiellosis.

**Table 3.** Hemagglutinating activity of *E. coli* harbored recombinant plasmid, pNUF5, and *E. tarda* KG8401

Bacteria	Hemagglutination titer (log <sub>2</sub> ) of cells with		
	None	D-Mannose (10 mg/mL)	Fetuin (10 mg/mL)
<i>E. tarda</i> KG8401	13	13	7
<i>E. coli</i> (pNUF5)	9	9	3
<i>E. coli</i> JM109	< 2	NT*	NT

\* NT; not tested.

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