

# **Spontaneous Remission of a Non-small Cell Lung Cancer Possibly Caused by Anti-NY-ESO-1 Immunity**

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## **Abstract**

Spontaneous remission of malignant tumors is rare and the biological mechanism of such remission has not been addressed. We report the case of a 71-year-old Japanese patient with non-small cell lung cancer with a right hilar tumor and pleural dissemination that spontaneously regressed. NY-ESO-1 is a cancer/testis antigen that can elicit specific immune responses in patients with cancer. Strong anti-NY-ESO-1 immunity was detected in this patient. His tumor cells expressed NY-ESO-1 and MHC class I molecules. Anti-NY-ESO-1 immunity might have contributed to spontaneous remission in this patient.

## 1. Introduction

Spontaneous remission is extremely rare in patients with non-small cell lung cancer (NSCLC). Any underlying mechanism of this remission remains unclear [1]. A systemic reaction, such as an immune response to tumors, seemed to be a possible causative mechanism. However, there have been no reports indicating immunoreaction-mediated spontaneous remission in patients with NSCLC.

NY-ESO-1 was originally identified in esophageal cancer by serological expression cloning using autologous patient serum and found to be a cancer/testis antigen that is expressed in cancer and testis, but not in normal adult somatic tissues [2, 3]. This antigen has made one of the fastest transitions from molecular, cellular, and immunological descriptions to a vaccine and immunotherapy candidate. NY-ESO-1 has already been tested in various formulations in more than 30 clinical trials worldwide, and its main characteristic resides in its capacity to elicit spontaneous antibody and T-cell responses in a proportion of patients with cancer [4]. Here, we present the case of a patient with NSCLC that spontaneously regressed, possibly mediated by anti-NY-ESO-1 immunity.

## **2. Case report**

A 71-year-old man was referred to Goto Central Hospital, Nagasaki, Japan in November 2004 for further examination of abnormal shadows on his chest x-ray. A chest computed tomography (CT) revealed a right hilar tumor, measuring 3 x 3 cm, and right multiple focal pleural thickenings (Fig. 1A). The patient underwent a thoracoscopy and tumor specimens were collected from the right pleural thickening. The pathological diagnosis was poorly differentiated adenocarcinoma with positive staining for cytokeratin 7 and negative staining for cytokeratin 20 (Fig. 1B). The clinical diagnosis was c-T4N0M0 stage IIIB NSCLC. The patient refused to receive any treatment at that time. A follow-up chest CT showed the disappearance of pleural dissemination and shrinkage of the right hilar tumor (Fig. 1A).

## **3. Materials and Methods**

Blood was drawn from the patient with informed consent. Collected serum samples were frozen until use. Peripheral blood mononuclear cells (PBMC) were isolated by density gradient centrifugation. CD4 and CD8 T-cells were obtained from PBMC using CD4 and CD8 microbeads, respectively, with columns and

magnetic devices (Miltenyi Biotec, Auburn, CA). Residual cells were used as CD4- and CD8-depleted cells. These cells were stored in liquid N<sub>2</sub> until use.

Antibody responses to cancer/testis antigens were evaluated by enzyme-linked immunosorbent assay (ELISA) as described elsewhere [5]. NY-ESO-1 is composed of 180 amino acids [2]. Twenty-eight 18-mer NY-ESO-1 overlapping peptides, spanning 1-173 amino acids of N-terminal NY-ESO-1, and one 30-mer C-terminal peptide, spanning 151-180 amino acids, were synthesized with standard solid-phase methods using a Multiple Peptide Synthesizer (AMS422; ABIMED, Langenfeld, Germany) [6]. To detect T-cell response to NY-ESO-1, CD4 and CD8 T-cells ( $2 \times 10^6$ ) were cultured with irradiated (30 Gy) CD4- and CD8-depleted cells ( $2 \times 10^6$ ) in the presence of 28 18-mer NY-ESO-1 overlapping peptides and a 30-mer C-terminal peptide (1  $\mu$ g/ml for each peptide) in AIM-V (Invitrogen, Carlsbad, CA) with 5% heat-inactivated pooled human serum with 10 units/ml IL-2 (Takeda Chemical Industry, Osaka, Japan) and 10 ng/ml IL-7 (Peprotech, London, UK) in a 24-well plate at 37°C in a 5% CO<sub>2</sub> atmosphere for 12 days. IFN $\gamma$  secretion assays with  $2 \times 10^5$  cells were performed according to manufacturer's protocol [6]. Immunohistochemistry was performed as described elsewhere [7].

#### 4. Results

Antibody responses to cancer/testis antigens were determined by enzyme-linked immunosorbent assay using 1  $\mu$ g of NY-ESO-1 ( $\circ$ ), SSX-2 ( $\bullet$ ), SSX-4 ( $\blacktriangle$ ), and XAGE-1 ( $\blacksquare$ ) recombinant proteins because of their high expression in lung cancer and strong immunogenicity in patients with NSCLC. A high titer of IgG antibody specific to NY-ESO-1 was detected and observed throughout the period starting July 2006 (Fig. 2A). We also observed strong CD4 and CD8 T-cell responses specific to NY-ESO-1 in an assay for interferon gamma ( $\text{IFN}\gamma$ ) secretion. Thus, integrated anti-NY-ESO-1 immunity consisting of antibody with CD4 and CD8 T-cell responses was detected in the patient (Fig. 2B).

Immunohistochemical staining was performed for NY-ESO-1, MHC class I, and CD8<sup>+</sup> T-cells. Cytoplasmic expression of NY-ESO-1 was observed in 50-60% cancer cells (Fig. 3A). MHC class I was stained on the cell surface of 30-40% cancer cells (black arrows) (Fig. 3B). CD8<sup>+</sup> T-cells were observed in the interface between the stromal and tumor tissues (black arrows) and also within the tumor tissue (white arrows) (Fig. 3C). CD8<sup>+</sup> T-cells in tumor tissue were counted using a 40x objective lens in 10 fields and more than 30 cells were observed in each

field. In addition, double staining of CD25 (brown) and FOXP3 (red) showed that CD25<sup>+</sup> FOXP3<sup>+</sup> T-cells (black arrows) were mainly distributed in stromal tissue and that CD25<sup>+</sup> FOXP3<sup>-</sup> T-cells (white arrows) were observed in tumor tissue in this patient (Fig. 3D).

As shown in Figure 1A, a follow-up chest CT scan in February 2006 showed the disappearance of pleural dissemination, while the right hilar tumor had increased to 4.5 x 3 cm. Once the patient agreed to receive treatment, radiation (a total of 60 Gy in 30 fractions) against the tumor was started in March, 2006; this was effective and resulted in a partial response [5]. As of September 2007, the patient was doing well and no recurrence of pleural dissemination had been observed.

## **Discussion**

We detected a high titer of IgG antibody specific to NY-ESO-1 from a patient with NSCLC that spontaneously regressed. We also observed anti-NY-ESO-1 CD4 and CD8 T-cell responses from his lymphocytes. Integrated anti-NY-ESO-1 immunity was elicited by tumor cells expressing NY-ESO-1.

Immunohistochemical staining of tumor-infiltrating lymphocytes (TIL) showed a

high number of CD8 TIL at the tumor sites. Although it was not verified that CD8 TILs were NY-ESO-1-specific T cells, systemic NY-ESO-1 immunity evoked in this patient might have contributed to tumor regression.

The mechanism of spontaneous remission remains unclear. However , it was suggested that systemic immunity against NY-ESO-1 contributed to the tumor regression in this case . More analysis regarding tumor microenvironment could provide the exact mechanism of tumor remission in NSCLC.

No authors have any potential conflicts of interest regarding this manuscript.



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## Figure Legends

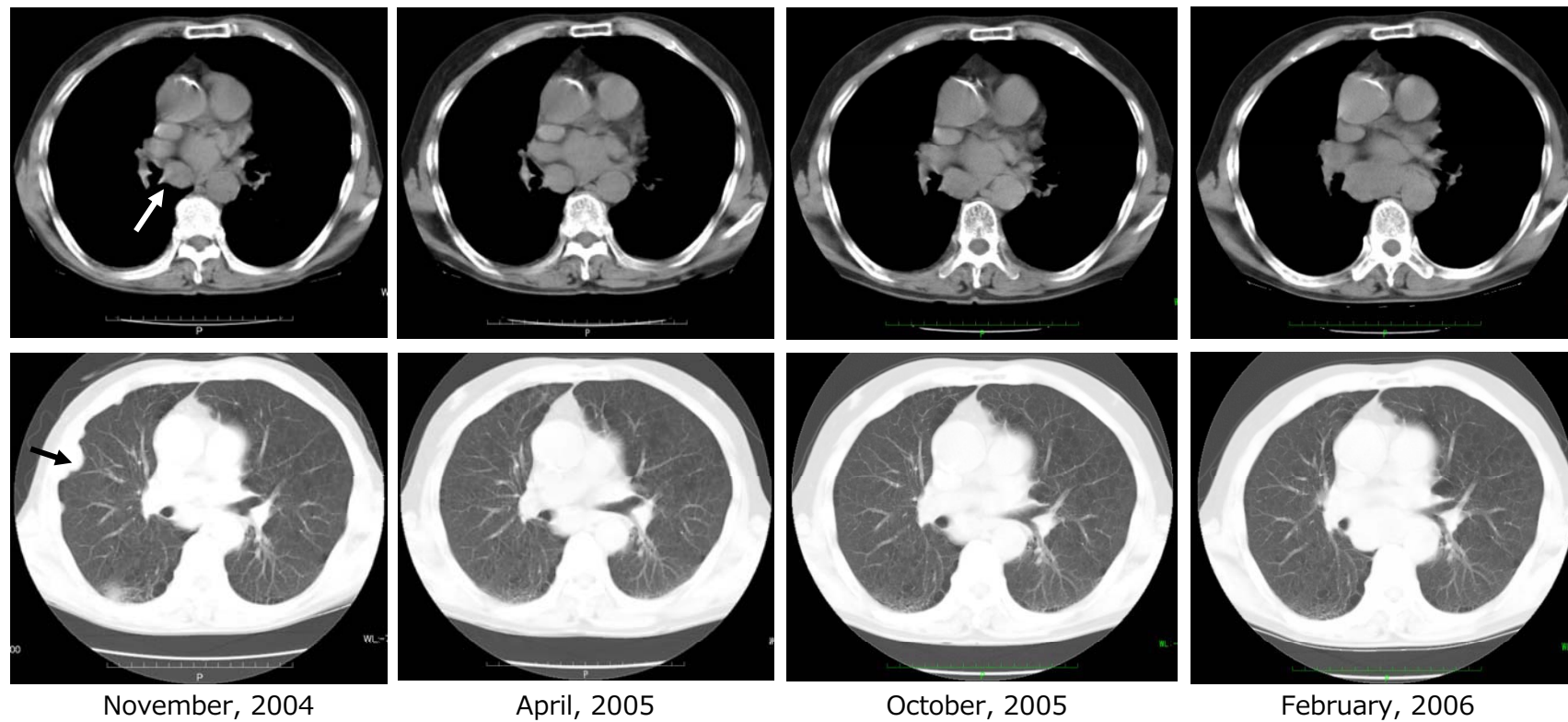
**Fig. 1.** (A) Chest computed tomography. White arrows indicate the right hilar tumor and black arrows indicate pleural disseminations. (B) Staining of a biopsy sample of pleural metastasis. The pathological diagnosis was poorly differentiated adenocarcinoma with positive staining for cytokeratin 7 and negative staining for cytokeratin 20.

**Fig. 2.** Immune response to NY-ESO-1 in the patient. (A) Enzyme-linked immunosorbent assay (ELISA). Antibody response to cancer/testis antigens was determined by ELISA using 1  $\mu\text{g}$  of NY-ESO-1 ( $\circ$ ), SSX-2 ( $\bullet$ ), SSX-4 ( $\blacktriangle$ ), and XAGE-1 ( $\blacksquare$ ) recombinant proteins. (B) Interferon gamma ( $\text{IFN}\gamma$ ) secretion assays. CD4 and CD8 T-cells ( $2 \times 10^6$ ) purified from peripheral blood mononuclear cells (PBMC) using magnetic cell sorting were cultured for 12 days with NY-ESO-1 peptides. The cells ( $2 \times 10^5$ ) were assayed for  $\text{IFN}\gamma$  secretion in response to PFA-treated autologous CD4- and CD8-depleted PBMC ( $2 \times 10^5$ ) pre-pulsed or unpulsed with NY-ESO-1 peptides using fluorescence-activated cell sorting. Values higher than 0.1% were considered to be significant.

**Fig. 3.** Immunohistochemistry of a biopsy sample from pleural metastasis. (A) NY-ESO-1. Cytoplasmic staining was observed. (B) MHC class I. Surface staining was observed as indicated by black arrows. (C) CD8<sup>+</sup> T-cells were observed in the interface between stromal and tumor tissues (black arrows) and also within the tumor tissue (white arrows). (D) Double staining of CD25 and FOXP3. Regulatory T-cells (black arrows) were detected by double staining of CD25 (brown, cell surface staining) and FOXP3 (red, nuclear staining). White arrows indicate CD25<sup>+</sup> FOXP3<sup>-</sup> T-cells. A bold line indicates the border between the stroma and the tumor.

Figure1

(A)



(B)

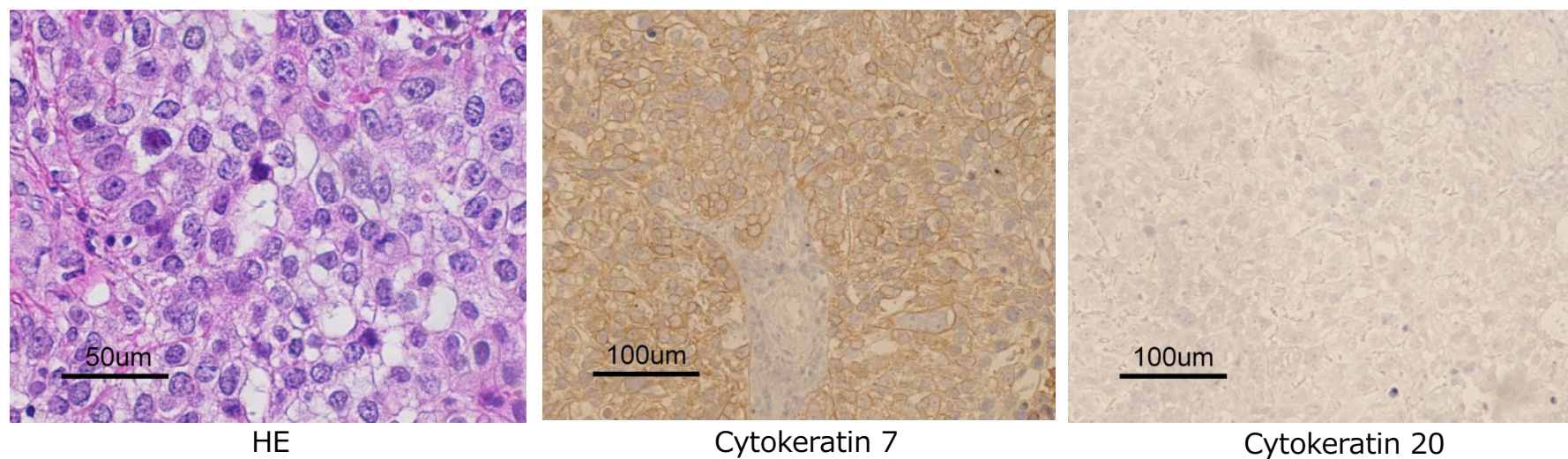
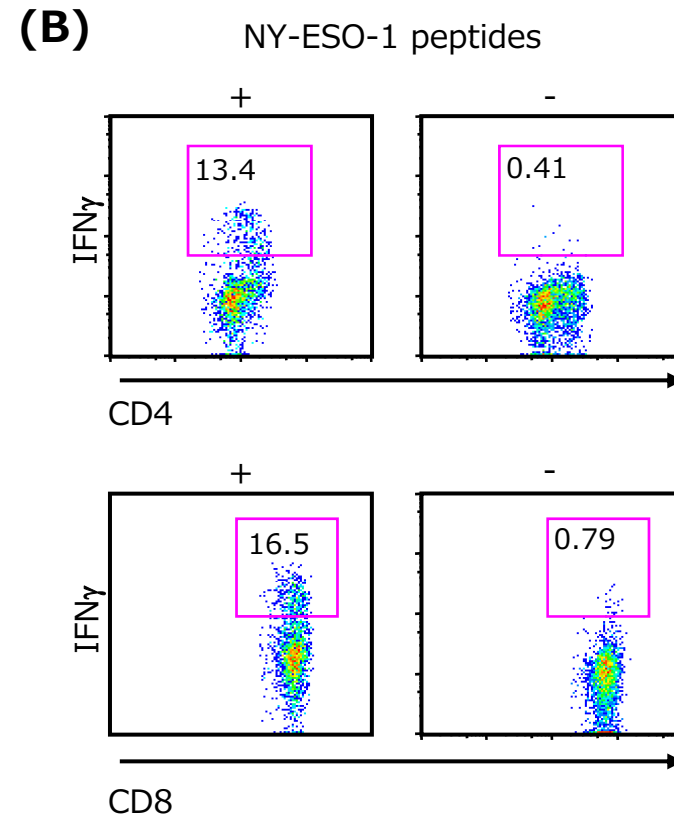
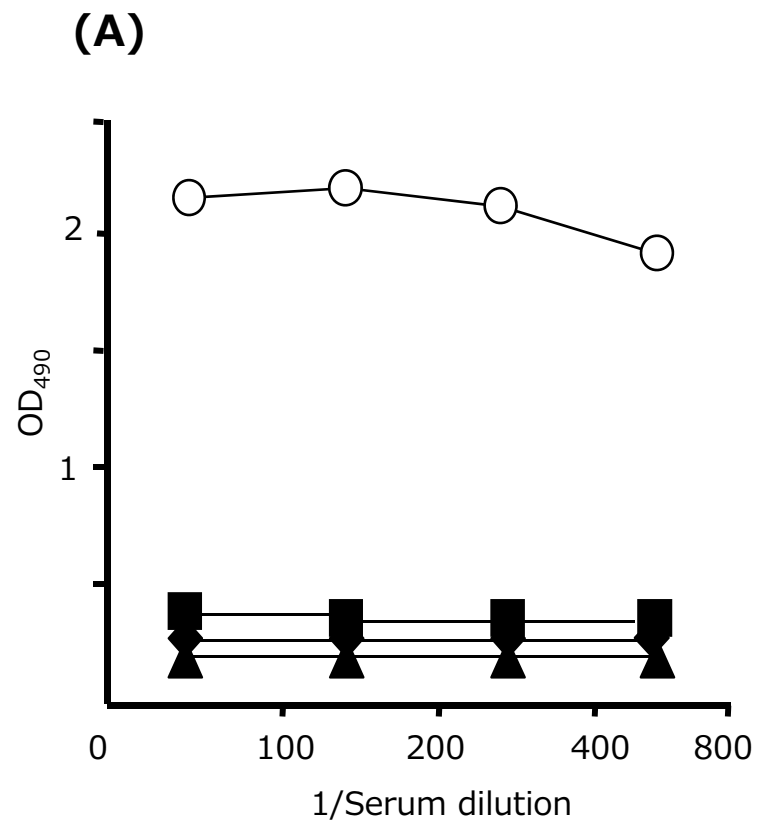
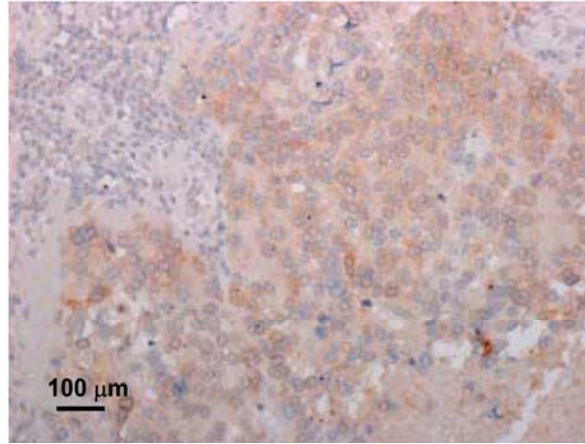


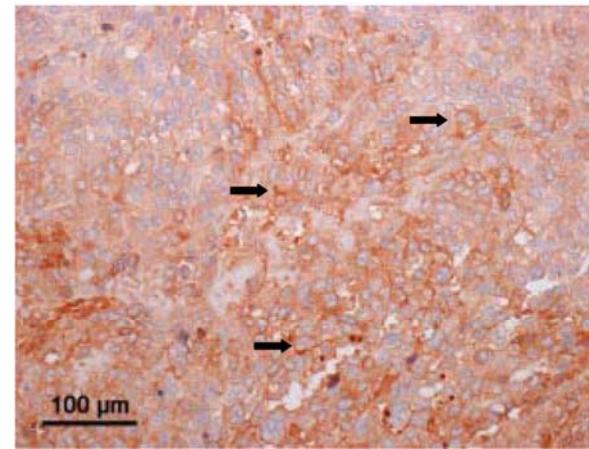
Figure2



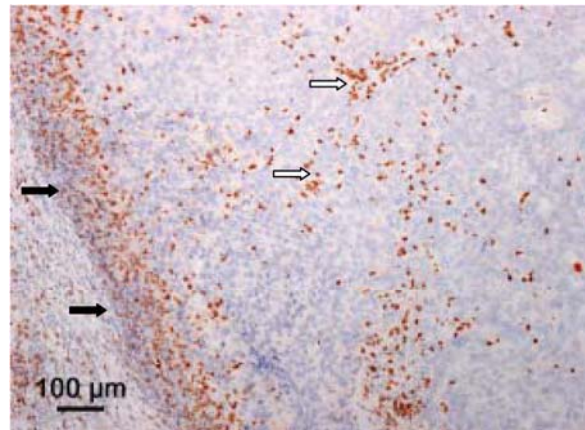
**(A)** NY-ESO-1



**(B)** MHC class I



**(C)** CD8



**(D)** CD25/FOXP3

