

RESEARCH

Open Access

Preventive effect of permethrin-impregnated long-lasting insecticidal nets on the blood feeding of three major pyrethroid-resistant malaria vectors in western Kenya

Hitoshi Kawada^{1*}, Kazunori Ohashi², Gabriel O Dida^{1,3}, George Sonye⁴, Sammy M Njenga⁵, Charles Mwandawiro⁶ and Noboru Minakawa^{1,7}

Abstract

Background: Since the World Health Organization (WHO) adopted the use of long-lasting insecticidal nets (LLINs) as a principal strategy for effective malaria prevention and control, pyrethroids have been the only class of insecticides used for LLINs. The dramatic success of insecticide-treated nets (ITNs) and LLINs in African countries, however, has been threatened by the rapid development of pyrethroid resistance in vector mosquitoes. ITNs and LLINs are still used as effective self-protection measures, but there have been few studies on the effectiveness of ITNs and LLINs in areas where vector mosquitoes are pyrethroid-resistant.

Methods: To investigate the behavioral pattern of mosquitoes in the houses where LLINs were used, indoor mosquito trappings of *Anopheles gambiae* s.s., *An. arabiensis*, and *An. funestus* s.s. were performed with Centers for Disease Control and Prevention (CDC) miniature light trap equipped with a collection bottle rotator at 2-hour intervals between 4:00 PM and 8:00 AM. The trapped female mosquitoes were identified and classified as unfed, blood fed, and gravid. The abdominal contents of fed female mosquitoes were used for DNA extractions to identify the blood source.

Results: A large proportion of human blood feeding of *An. arabiensis* and *An. funestus* s.s. (but not *An. gambiae* s.s.) took place during the time people were active outside LLINs. However, during the hours when people were beneath LLINs, these provided protective efficacy as indicated by reduced human blood feeding rates.

Conclusion: LLINs provided effective protection against pyrethroid-resistant malaria vector populations during bedtime hours. However, protection of LLINs was insufficient during the hours when people were active outside of the bed nets. Such limitation of LLINs will need to be intensively addressed in African countries in the near future.

Keywords: Permethrin, Resistance, *Anopheles gambiae* s.s., *Anopheles arabiensis*, *Anopheles funestus*, LLIN, Kenya

Background

One of the most successful breakthroughs in the development of pesticides of natural origin has been the discovery of pyrethrum and the successful development of synthetic pyrethroids. Pyrethroids are the predominant insecticides and used in various formulations for mosquito control. Globally, pyrethroids comprise 40% of the insecticides used annually for indoor residual spraying

against malaria vectors [1]. The use of ITNs as a simple and inexpensive self-protection measure against malaria has reduced the morbidity of children (<5 years old) by 50% and global child mortality by 20%–30% [2,3]. However, ITNs required impregnation and reimpregnation with pyrethroids [4], thus the use of LLINs, in which insecticides are incorporated into synthetic fibers and have long residual effects, might be preferable [5]. Since the WHO adopted the use of LLINs as a principal strategy for effective malaria control in the Roll Back Malaria Partnership [6], pyrethroid has been the only class of insecticide used for LLINs [1,7].

* Correspondence: vergiss@nagasaki-u.ac.jp

¹Department of Vector Ecology & Environment, Institute of Tropical Medicine, Nagasaki University, Nagasaki, Japan

Full list of author information is available at the end of the article

Pyrethroids are safe for mammals and have unique modes of action such as fast knockdown and excito-repellent effects [8]. Although the use of untreated bed nets was already common in some tropical and sub-tropical countries before the introduction of ITNs [9], several studies have demonstrated the greater effectiveness of ITNs compared with untreated nets [10-13]. Five deltamethrin-coated (or -incorporated) nets, 4 alpha-cypermethrin-coated (or -incorporated) nets, and 2 permethrin-incorporated nets have been listed as WHO-recommended LLINs.

The dramatic success of ITNs and LLINs in African countries, however, has been countered by the rapid development of pyrethroid resistance in vector mosquitoes over the past decade [14]. ITN coverage might be one of the major factors causing such resistance [15], as well as agricultural usage of pyrethroids, and mainly attributable to the fact that malaria vector control currently depends on a single class of insecticides (pyrethroids) [14].

ITNs and LLINs are still used as effective self-protection measures, but there have been few studies on the effectiveness of ITNs and LLINs in areas where vector mosquitoes are pyrethroid-resistant. Personal protection of the two LLINs (Olyset[®] Net and Permanet[®]) was evaluated in a rice-growing area of south-western Burkina Faso, where *kdr*-governed pyrethroid resistance in *An. gambiae* s.s. was reported, and significantly lower numbers of mosquitoes were collected in LLIN-intervention houses compared to the control houses, indicating the additional deterrent effect of the LLINs [16]. Significantly higher mortality of *An. epiroticus* Linton & Harbach was observed with Permanet[®] 2.0 and 3.0 despite the fact that the mosquito population showed metabolic resistance to pyrethroids [17]. The aim of this study was to investigate the difference in the reaction of vector mosquitoes to pyrethroid-impregnated bed nets in the area where multimodal pyrethroid resistance has developed, and the effect of LLINs on these malaria vectors is discussed.

Methods

Study site

The study area was the Mbita and Suba districts of Nyanza Province in western Kenya. The rainfall pattern in the area is bimodal, with the long rainy season occurring from March to May and the short rainy season occurring in November and December. Malaria infection rates rise steadily between September and February and peak briefly in June, following the long rains [18]. The Mbita and Suba districts have been identified as high vector transmission areas in Kenya, and more than 50% of the population is exposed to malaria at a rate of $\geq 40\%$ PfPR₂₋₁₀ (*Plasmodium falciparum* parasite rate corrected to a standard age range of 2 to <10 y) [19]. Kawada *et al.* [20,21] reported that the 3 major malaria vectors, *Anopheles gambiae* Giles

(*An. gambiae* sensu stricto [s.s.]), *An. arabiensis* Patton, and *An. funestus* Giles (*An. funestus* s.s.), in the study area were highly resistant to pyrethroids and that resistance mechanisms were multimodal, including the knockdown resistance (*kdr*) caused by the point mutation of the voltage-gated sodium channel (L1014S) in *An. gambiae* s. s. and cytochrome P450-related metabolic factors in both *An. arabiensis* and *An. funestus* s.s. Recently Kawada *et al.* [22] also reported the insecticide resistance status of the three malaria vectors in the present study area. *Anopheles gambiae* s.s., *An. arabiensis*, and *An. funestus* s.s. showed high resistance to both permethrin and deltamethrin. The KT_{50s} of the 3 species for permethrin and deltamethrin were >60 minutes except for deltamethrin in *An. arabiensis* (KT₅₀, 29.5 minutes), indicating low knockdown activities of the 2 pyrethroids against these mosquitoes as well as low killing activities. In contrast, lethal activities of fenitrothion and propoxur against the three species were higher (95–100% mortality) than those of the 2 pyrethroids.

Indoor collection of mosquitoes by miniature light traps equipped with collection bottle rotators in the houses using LLIN

To investigate the behavioral pattern of mosquitoes, indoor mosquito trapping of *An. gambiae* s.s., *An. arabiensis* and *An. funestus* s.s. was performed with 6 sets of the CDC miniature light trap (model 512) equipped with a collection bottle rotator (model 1512) (John W. Hock Co., FL, USA) (Figure 1). Indoor trapping of *An. gambiae* s.s. was performed in 6 houses on Mfangano Island from May 16 to June 1, 2011. Indoor trapping of *An. arabiensis* and *An. funestus* s.s. was conducted in 6 houses in Nyaroya Village from October 4 to 31, 2011, and July 11 to 18, 2012, and in 6 houses in Ungoe Village from February 21 to March 6, 2012 (Figure 2). The residents of the houses were informed about the study, and their written consent was obtained before mosquito collection. The houses used for indoor mosquito collection were of standard construction for the area; they had mud walls and eaves, traditional opening structures between the roof and the walls, and had 1–2 bedrooms and a living room. Before the start of the study, each house was inspected and untreated bed nets and LLINs were exchanged for new permethrin-incorporated LLINs (Olyset[®] Nets; Sumitomo Chemical Co., Ltd., Tokyo, Japan). Additional new Olyset[®] Nets were provided for use in the living room when the residents (most were children >5 years) slept in the living room with no bed nets. Tables 1 and 2 show the number of Olyset[®] Nets, the number of persons who slept in the bedroom(s) and living room, and the approximate number of livestock for each house.

The collection bottle rotator was programmed to collect active mosquitoes at 2-hour intervals between 4:00 PM

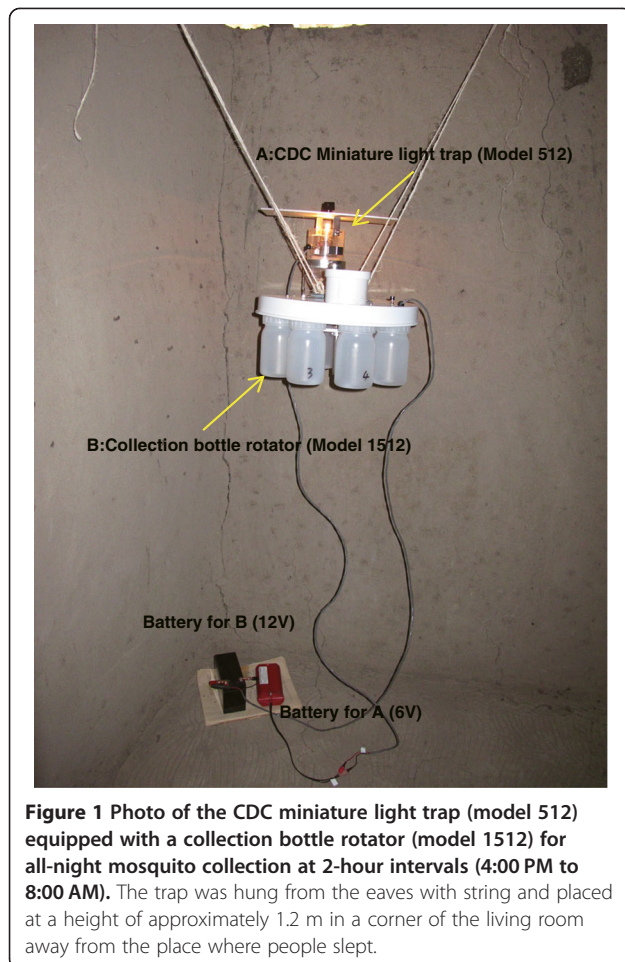


Figure 1 Photo of the CDC miniature light trap (model 512) equipped with a collection bottle rotator (model 1512) for all-night mosquito collection at 2-hour intervals (4:00 PM to 8:00 AM). The trap was hung from the eaves with string and placed at a height of approximately 1.2 m in a corner of the living room away from the place where people slept.

and 8:00 AM into 8 different plastic bottles. The trap was hung from the ceiling with strings and placed at a height of approximately 1.2 m in a corner of the living room away from where people slept. The trapped female mosquitoes were identified and classified as unfed, blood fed, and gravid. The abdominal contents of fed female mosquitoes were used for DNA extractions to identify the blood source. One to 6 replicates in each house (total of 21 replicates) on Mfangano Island, 2 to 16 replicates in each house (total of 43 replicates) in Nyaroya Village, and 2 to 15 replicates in each house (total of 88 replicates) in Nyaroya and Ungoe villages were performed for collection of *An. gambiae* s.s., *An. arabiensis*, and *An. funestus* s.s., respectively.

Species identification

Adult mosquitoes were examined microscopically to distinguish *An. gambiae* s.l. and *An. funestus* s.l. from other anophelines based on identification keys developed by Gillies and Coetzee [23]. Multiplex polymerase chain reaction (PCR) methods described by Scott *et al.* [24] and Koekemoer *et al.* [25] were used for the species identification.

Identification of meal sources in blood-fed mosquitoes

Mosquitoes collected in the field were individually placed in 1.5-mL plastic tubes containing silica gel desiccant and stored at -10°C until processed. Engorged abdomens were separated from the rest of the body for blood source identification. DNA was extracted from the blood in each abdomen using the REExtract-N-Amp Tissue PCR Kit (Sigma, MO, USA) per the manufacturer's instructions. Species identification of the host blood source was performed by multiplex PCR as described by Kent and Norris [26] using published cytochrome B primers for human (human741F, GGCTTAC TTCTCTTCATTCTCTCCT) and cow (cow121F, CATCG GCACAAATTTAGTCG) and a universal reverse primer (UNREV1025, GGTTGTCCCTCCAATTCATGTTA). A 20- μL cocktail consisting of 0.5 $\mu\text{mol/L}$ of each primer and previously diluted (300 times) 1 μL of the DNA template was placed in a 0.2-mL cell containing AccuPower™ PCR Premix (Bioneer, Daejeon, Korea). PCR was performed under the following conditions: a hot start at 95°C for 5 min; 35 cycles of template denaturing at 95°C for 1 min, primer annealing at 54°C for 1 min, and amplicon extension at 72°C for 1 min; and a final extension at 72°C for 7 min.

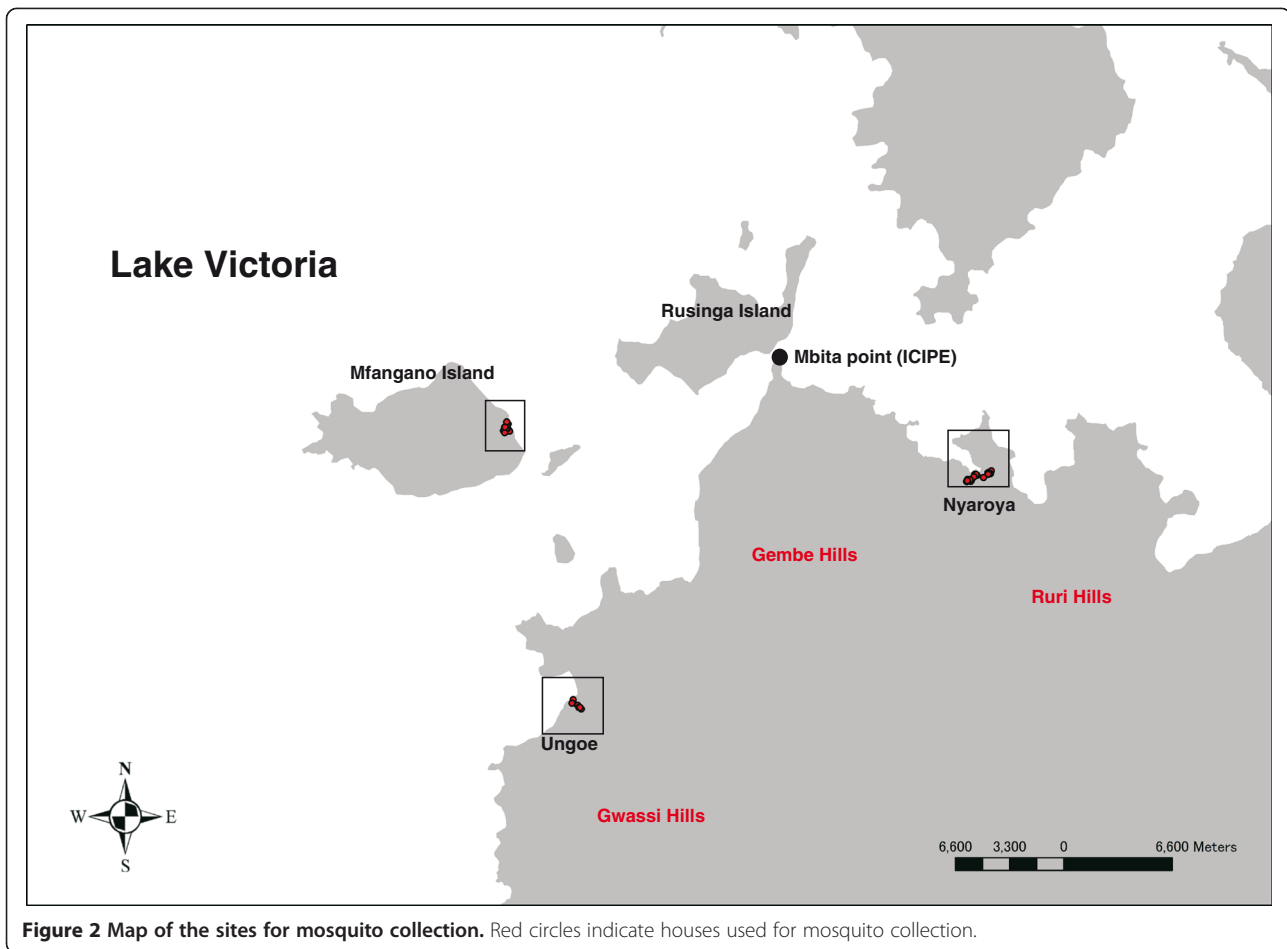
Detection of point mutations in the voltage-gated sodium channel

PCR and direct DNA sequencing were performed to identify point mutations at 1014 L in the field-collected mosquitoes according to the method by Kawada *et al.* [20,21]. Direct DNA sequencing was performed using the 3730 DNA Analyzer (Applied Biosystems). The electropherogram of the targeted amino acid replacement was analyzed using MEGA 4.0 public domain software (<http://www.megasoftware.net/>). The unique DNA haplotype sequences were deposited into GenBank.

Data analysis

A digital map in shape file format (Kenya-Boundaries, FAO Africover, http://www.glc.n.org/activities/africover_en.jsp) was used for mapping of the collection sites. The geographical positions of the collection sites were plotted on the map using ArcGIS 10.1 (ESRI Japan Corp, Tokyo, Japan).

The daily average number of mosquitoes collected per house in the 8 time zones was calculated. The generalized linear mixed model using the poisson distribution was used to examine whether the collection times explained the activity patterns using the package lme4 in R. The house and the date of collection were used as random effects. The multiple comparison of the number of collected mosquitoes at different time zones was performed by means of Tukey honestly significant difference test using the package multcomp in R. Comparison of the number of human blood-fed and unfed mosquitoes



during bedtime (10:00 PM–6:00 AM) and active time (4:00–10:00 PM and 6:00–8:00 AM) was performed with χ^2 test using JMP 7.0 J (SAS Institute Japan Inc., Tokyo, Japan).

Ethics statement

The protocol for the study (case no. 1775) was approved by the Scientific Steering Committee and the National Ethics Review Committee of the Kenya Medical Research Institute. All necessary permits were obtained for the described field studies. No mosquito collection was done

without the approval of the head of the village and the owner and occupants of the collection house.

Results

The indoor activity patterns of the 3 major malaria vectors in western Kenya are shown in Figure 3. The number of trapped female *An. gambiae* s.s. (fed and unfed) significantly rose at night (10:00 PM to 2:00 AM), falling before and after this period (ANOVA, $df = 7$, $\chi^2 = 84.9$, $p < 0.0001$). The number of human blood–fed mosquitoes

Table 1 Information on the houses used for all-night trapping of *An. gambiae* s.s. by CDC traps equipped with a collection bottle rotator in May to June 2011

| House no. ^a | No. of olyset® nets | No. of persons sleeping | | No. of livestock around house | | |
|------------------------|---------------------|-------------------------|-------------|-------------------------------|-------|------|
| | | Bedroom ^b | Living room | Cow | Sheep | Goat |
| MFA 1 | 3 | 2A | 3C | 13 | 0 | 7 |
| MFA 7 | 2 | 2A | 3C | 7 | 30 | 10 |
| MFA 9 | 2 | 1A + 1C | 0 | 5 | 0 | 0 |
| MFA 11 | 1 | 1A + 1C | 0 | 7 | 30 | 10 |
| MFA 12 | 2 | 1A | 2A | 3 | 0 | 0 |

^aMFA, Mfangano; A, adults (>15 years old); C, children (≤15 years old).

^bPersons sleeping in a bedroom usually used one bed under a single bed net.

Table 2 Information on the houses used for all-night trapping of *An. arabiensis* and *An. funestus* s.s. by CDC traps equipped with a collection bottle rotator

| House no. ^a | Date of collection | No. of olyset [®] nets | No. of persons sleeping ^b | | No. of livestock around house | | |
|------------------------|--------------------|---------------------------------|--------------------------------------|-------------|-------------------------------|-------|------|
| | | | Bedroom ^c | Living room | Cow | Sheep | Goat |
| NYAR 11 | Oct 2011 Jul 2012 | 2 ^d | 2A | 1C | 18 | 2 | 26 |
| NYAR 14 | Oct 2011 Jul 2012 | 1 | 1A | 0 | 22 | 8 | 9 |
| NYAR 15 | Oct 2011 Jul 2012 | 2 ^d | 2A | 0 | 0 | 0 | 2 |
| NYAR 19 | Oct 2011 Jul 2012 | 2 | 2A | 0 | 2 | 0 | 0 |
| NYAR 20 | Oct 2011 Jul 2012 | 2 | 2A + 3C | 0 | 22 | 8 | 9 |
| NYAR 21 | Oct 2011 | 1 | 1A | 0 | 2 | 0 | 0 |
| NGO 3 | Feb–Mar 2012 | 2 | 2A | 1C | 8 | 0 | 10 |
| NGO 4 | Feb–Mar 2012 | 1 | 2A | 0 | 6 | 0 | 4 |
| NGO 5 | Feb–Mar 2012 | 2 | 1A | 2C | 10 | 0 | 4 |
| NGO 6 | Feb–Mar 2012 | 2 | 2A | 1C | 10 | 0 | 4 |
| NGO 7 | Feb–Mar 2012 | 3 | 2A | 4C | 0 | 0 | 0 |
| NGO 8 | Feb–Mar 2012 | 2 | 1A + 2C | 0 | 10 | 0 | 4 |

^aNYAR, Nyaroya; NGO, Ungoe.

^bA, adults (>15 years old); C, children (≤15 years old).

^cPersons sleeping in a bedroom usually used one bed under a single bed net.

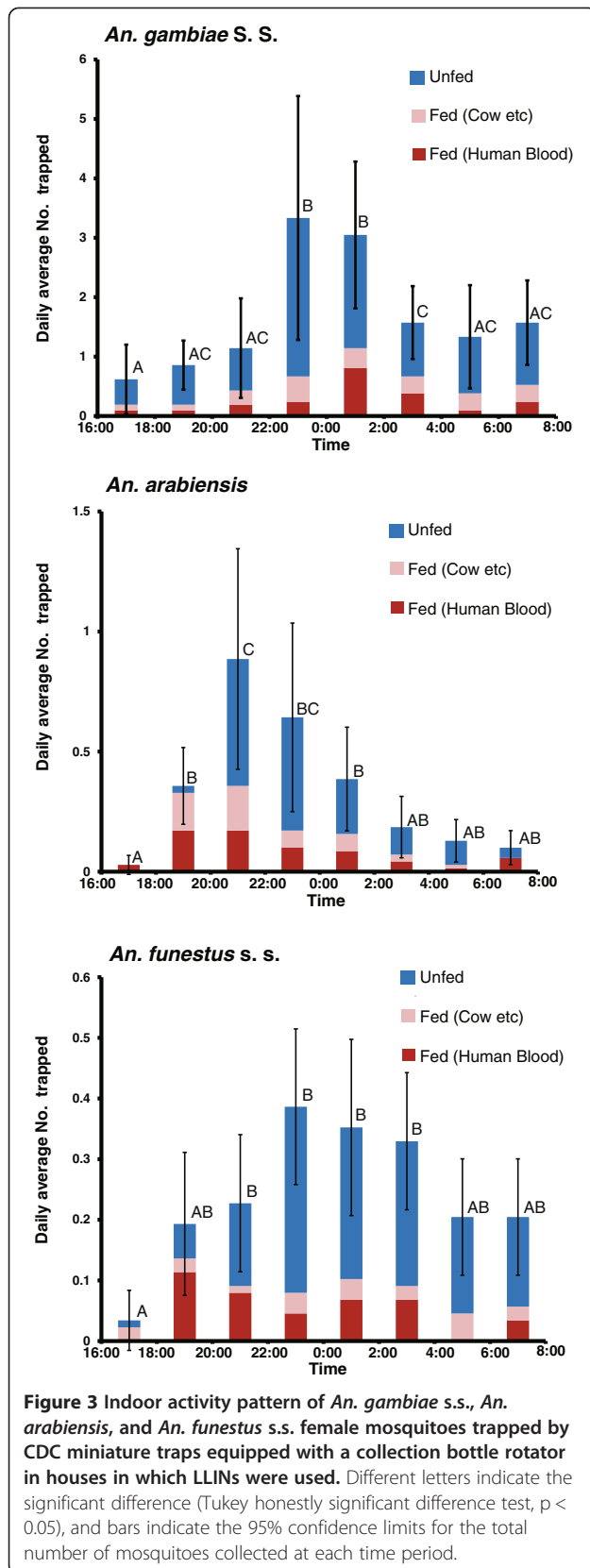
^dOne PermaNet[®] 2.0 was used occasionally instead of an Olyset[®] Net in the living room.

also peaked during this period (ANOVA, $df = 7$, $\chi^2 = 26.7$, $p < 0.001$), but there was no significant peak in the number of animal blood-fed mosquitoes (ANOVA, $df = 7$, $\chi^2 = 7.20$, $p = 0.41$). The allelic frequency of the point mutation in the voltage-gated sodium channel (L1014S) in the collected mosquitoes was 98.4% ($n = 96$, Accession AB776705, AB776706). Among the 283 female *An. gambiae* s.s. collected (13.4 ± 0.70 /house per a night), 195 were unfed (68.9%, 9.3 ± 0.81 /house per a night), 45 contained blood from a human source (15.9%, 2.0 ± 0.26 /house per a night), and 43 had fed from an animal source (15.2%, 2.0 ± 0.26 /house per a night). Among the 45 female mosquitoes engorged with human blood, 22 (48.9%) were trapped from 10:00 PM to 2:00 AM. There was no significant difference in the ratio of human blood-fed mosquitoes to unfed plus animal blood-fed mosquitoes between bedtime (10:00 PM to 6:00 AM), when most residents were assumed to sleep under bed nets, and active time (4:00–10:00 PM and 6:00–8:00 AM), when most residents were active outside bed nets ($\chi^2 = 0.03$, $p = 0.73$, Figure 4).

The number of trapped female *An. arabiensis* (fed and unfed) significantly rose from early night (8:00–10:00 PM) to midnight (10:00 PM to 12:00 AM), falling before and after this period (ANOVA, $df = 7$, $\chi^2 = 122.6$, $p < 0.0001$) (Figure 3). The number of human blood-fed (ANOVA, $df = 7$, $\chi^2 = 22.0$, $p = 0.0025$) and animal blood-fed mosquitoes (ANOVA, $df = 7$, $\chi^2 = 37.5$, $p < 0.0001$) peaked earlier than the above period (6:00–10:00 PM). Among the 191 female *An. arabiensis* collected (2.7 ± 0.32 /house per a night), 106 were unfed (55.5%, 1.5 ± 0.32 /house per a night), 47 contained blood from a human source (24.6%,

0.67 ± 0.19 /house per a night), and 37 had fed from an animal source (19.4%, 0.53 ± 0.13 /house per a night). Among the 47 female mosquitoes engorged with human blood, 24 (51.1%) were trapped from 6:00 to 10:00 PM. A significant difference in the ratio of human blood-fed mosquitoes to unfed plus animal blood-fed mosquitoes was observed between bedtime (10:00 PM to 6:00 AM) and active time (4:00–10:00 PM and 6:00–8:00 AM) ($\chi^2 = 3.58$, $p = 0.039$, Figure 4).

The number of trapped female *An. funestus* s.s. (fed and unfed) showed broader peaks as compared to those of *An. gambiae* s.s. and *An. arabiensis*, with significant peaks through almost the whole night (8:00 PM to 4:00 AM) (ANOVA, $df = 7$, $\chi^2 = 39.7$, $p < 0.0001$) (Figure 3). The number of human blood-fed mosquitoes also showed broad peaks that peaked earlier at night (6:00–10:00 PM) (ANOVA, $df = 7$, $\chi^2 = 25.7$, $p < 0.0001$), whereas there was no significant peak in the number of animal blood-fed mosquitoes (ANOVA, $df = 7$, $\chi^2 = 2.49$, $p = 0.93$). Among the 170 female *An. funestus* s.s. collected (1.9 ± 0.12 /house per a night), 115 were unfed (67.6%, 1.3 ± 0.14 /house per a night), 36 contained blood from a human source (21.2%, 0.41 ± 0.038 /house per a night), and 19 had fed from an animal source (11.2%, 0.22 ± 0.12 /house per a night). Among the 36 female mosquitoes engorged with human blood, 17 (47.2%) were trapped from 6:00 to 10:00 PM. A significant difference in the ratio of human blood-fed mosquitoes to unfed plus animal blood-fed mosquitoes was observed between bedtime (10:00 PM to 6:00 AM) and active time (4:00–10:00 PM and 6:00–8:00 AM) ($\chi^2 = 8.17$, $p = 0.0022$, Figure 4).

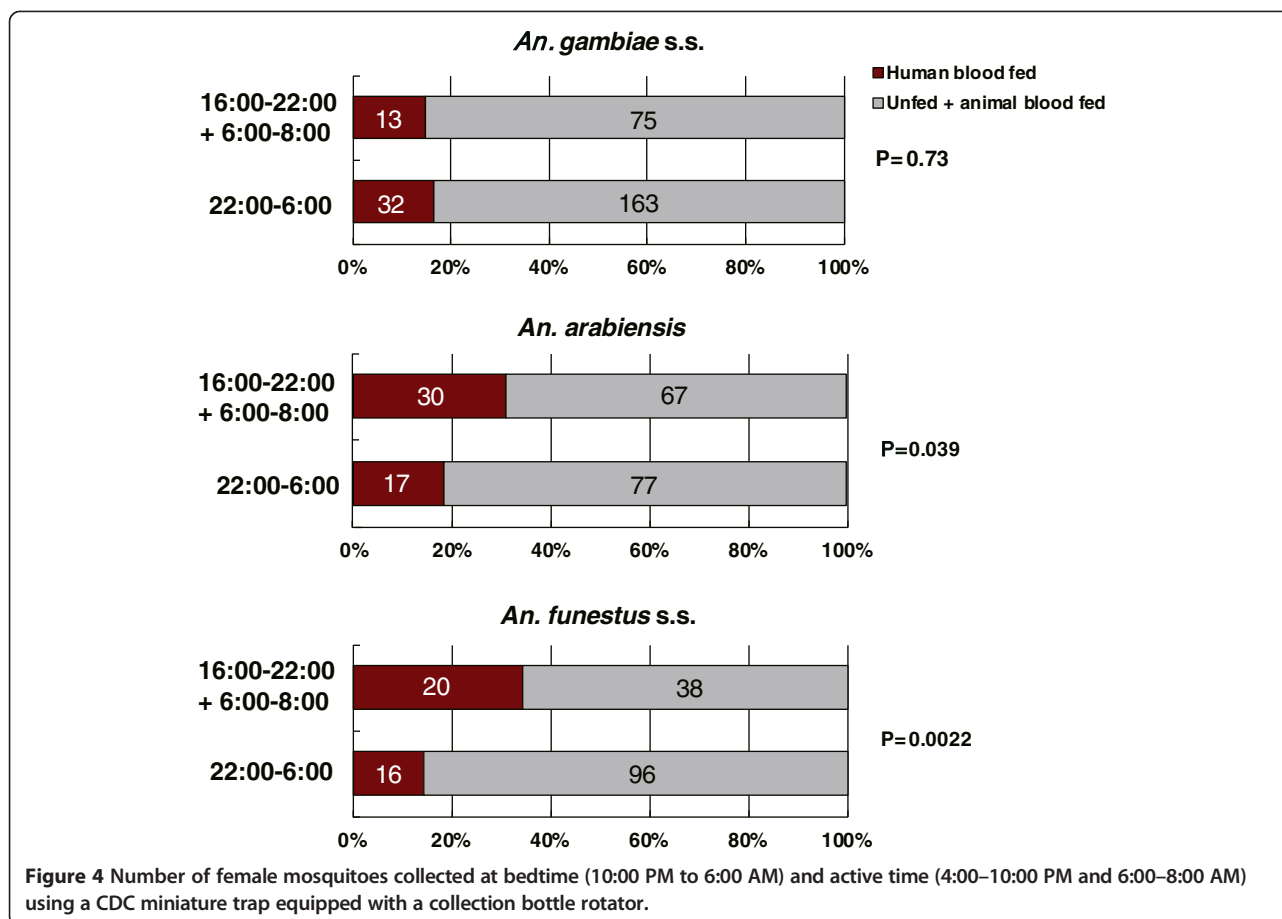


Discussion

In Nyanza province, dieldrin was reported to be administered mainly through aerial spraying especially for tsetse fly control through 1968–1971 [27], while the organized intensive spray of dichloro-diphenyl-trichloroethane (DDT) for mosquito control was not performed in the 1970s and 1980s, and no indoor residual spraying (IRS) has been administered since then. Therefore, the extensive use of ITNs and LLINs in the study area is thought to be a major factor causing high pyrethroid resistance [21,28].

In the present report we hypothesized that activity patterns of mosquitoes as indicated by the number trapped in the light trap corresponded to the host seeking or blood feeding activity patterns. The hypothesis is strongly supported by the experimental data of Kawada and Takagi [29] who showed that the activity patterns recorded by the photoelectric sensing devices in the laboratory well explained the host seeking activity patterns in the field as indicated by the number of mosquitoes by human bait collection. The activity of *An. gambiae* s.s. peaked at midnight (10:00 PM–2:00 AM), and human biting corresponded to this activity pattern. There was no significant difference in the ratio of human blood-fed female mosquitoes to unfed + animal blood-fed female mosquitoes between bedtime (10:00 PM–6:00 AM) and active time (4:00–10:00 PM and 6:00–8:00 AM), suggesting that LLINs did not affect human blood feeding activity of *An. gambiae* s.s., although the LLINs reduced the feeding success in this species at low level (15.9%). The similar consistency of anthropophagy in *An. gambiae* s.s. under the existence of permethrin-impregnated bed nets was reported by Bøgh et al. [30]. The activity peak in *An. arabiensis* was early at night (10:00 PM–12:00 AM), followed by human and animal biting peaks (6:00–10:00 PM), and human biting at bedtime was significantly reduced as compared with active time in this species. The small human biting peak was also noted in the morning (6:00–8:00 AM), when most people were assumed to be active outside of bed nets. In comparison to the 2 previously described species, the activity peaks in *An. funestus* s.s. had a broader range throughout the night. However, the same discrepancy in active time and human blood feeding time as shown in *An. arabiensis* was observed in this species. Additionally, the small human biting peaks were noted in the morning (6:00–8:00 AM) in this species.

In conclusion, LLINs were effective against 3 major malaria vectors during bedtime, since the percentages of human feeding success were found to be reduced to a low level during bedtime (15.9–24.6%). Although we could not arrange the untreated control houses in the present study simply because of ethical reason, feeding successes of *An. gambiae* in the houses without LLINs or with insecticide-untreated and badly torn bed nets were reported to be more than 80% [31,32]. The overall



activities of the 3 major malaria vectors in this study area have not changed from those reported before the extensive use of ITNs and LLINs started in Africa [33–35]. However, a notable human blood feeding activity in evening and a slight activity in early morning was shown in both *An. arabiensis* and *An. funestus s.s.*, while there was no such events in *An. gambiae s.s.* The same human biting patterns were recently reported in *An. gambiae s.l.* (most of which were thought to be *An. arabiensis*) [36], *An. arabiensis* [37], and *An. funestus s.s.* [36,38]. Though simple explanation for such species dependent difference in the host feeding activities under LLIN use observed in the present report is difficult, the difference in physiological factors such as the intrinsic host preference, insecticide resistance mechanisms, and repellency to pyrethroids, etc. might be involved as the plausible answers. Gatton *et al.* [39] recently reviewed the behavioral changes in mosquitoes by the impact of indoor insecticides. Most interventions against malaria vectors have involved indoor treatment of insecticides such as IRS and ITNs and they are thought to have selected exophagy or exophily and time shift of human feeding. Bayoh *et al.* [40] reported the historical population decline in *An. gambiae s.s.*, but it was not easy to

explain this population decline simply because of the changes in climate, the variations in abundance of cattle, etc. The authors concluded that the high coverage of LLINs during the decades was one of the most plausible answer to the above decline. However, the killing effect of LLINs against *An. gambiae s.s.* population, which has developed *kdr* mutations at high frequency (>90%), seemed to be not enough to cause such population decline. The historical population decline of this species might also be explained by the behavioral characteristics in *An. gambiae s.s.* population observed in the present report. *Anopheles gambiae s.s.* might have not changed their characteristics as “midnight feeder” by some physiological reasons notwithstanding existence of LLINs and has to rely on the limited human blood sources, most of which are protected by LLINs, resulting in the decline in their population size.

Bed nets are effective only when the vector mosquitoes are endophagous and their feeding time corresponds to the bedtime. The most important limitation for the effective use of bed nets is that they are only effective when people are sleeping inside. LLINs might not be effective in cases where most blood feeding takes place when people are active outside of bed nets. The

importance of the human host feeding in malaria vectors during when peoples are active outside of LLINs as reported in the present study will need to be intensively addressed in African countries in the near future. Changes in house design, such as screening or closing of eaves, might be effective in reducing human exposure to malaria vectors in such conditions [41,42]. The use of excito-repellency of pyrethroids as a spatial repellent, which acts as a spatial barrier to invasion of mosquitoes or reduces feeding motivation of mosquitoes might be another countermeasure [43].

Conclusion

LLINs provided effective protection against three pyrethroid-resistant malaria vector populations (*An. gambiae* s.s., *An. arabiensis*, and *An. funestus* s.s.) during bedtime hours. However, protection of LLINs was insufficient during the hours when people were active outside of the bed nets in *An. arabiensis* and *An. funestus* s.s. Screening or closing of eaves and the use of excito-repellency of pyrethroids as a spatial repellent might be effective in reducing human exposure to malaria vectors in such conditions.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

HK and KO designed the study, carried out the experiments, and drafted the manuscript. GOD and GS arranged the fields and informed consents of the participants for the study and organized the staffs for the experiments. SMN, CM, and NM critically revised the protocol for the study. All authors read and approved the final version of the manuscript.

Acknowledgments

This study was funded by a joint research effort between Nagasaki University and Sumitomo Chemical Co Ltd to create field and semi-field bioassay systems for delivering new insecticide technology. The protocol for the study (case no. 1775) was reviewed and approved by the Scientific Steering Committee and the National Ethics Review Committee of the Kenya Medical Research Institute. We are grateful to the staff at the Thomas Odhiambo campus of the International Center of Insect Physiology and Ecology for providing facilities and experimental insects. We also thank Fredrick Sonye, Anwal Orenge' Obilo, Godfrey Juma, Martin Shikuku, and Jairo Orenge' (Springs of Hope, Mbita, Kenya) and Emiko Kawashima, Kogomi Minagawa, and Chiaki Kimura (Institute of Tropical Medicine, Nagasaki University, Nagasaki, Japan) for providing technical support and assisting with this study.

Author details

¹Department of Vector Ecology & Environment, Institute of Tropical Medicine, Nagasaki University, Nagasaki, Japan. ²Health and Crop Sciences Research Laboratory, Sumitomo Chemical Co Ltd, Hyogo, Japan. ³School of Public Health, Maseno University, Kisumu, Kenya. ⁴Springs of Hope, Mbita, Kenya. ⁵Eastern and Southern Africa Centre of International Parasite Control, Nairobi, Kenya. ⁶Kenya Medical Research Institute, Nairobi, Kenya. ⁷The Global Center of Excellence Program, Nagasaki University, Nagasaki, Japan.

Received: 24 February 2014 Accepted: 9 August 2014

Published: 20 August 2014

References

1. Zaim M, Jambulingam P: *Global insecticide use for vector-borne disease control*. Geneva: World Health Organization; 2007.
2. Nevill CG, Some ES, Mung'ala VO, Mutemi W, New L, Marsh K, Lengeler C, Snow RW: *Insecticide-treated bed nets reduce mortality and severe*

3. morbidity from among children on the Kenyan Coast. *Trop Med Int Health* 1996, **1**:139–146.
3. Binka FN, Kubaje A, Adjuik M, Williams LA, Lengeler C, Maude GH, Armah GE, Kajihara B, Adiamah JH, Smith PG: *Impact of permethrin treated bed nets in child mortality in Kassa-Nankana district, Ghana: a randomized controlled trial*. *Trop Med Int Health* 1996, **1**:147–154.
4. Lines J: *Mosquito nets and insecticides for nets treatment: a discussion of existing and potential distribution systems in Africa*. *Trop Med Int Health* 1996, **1**:616–632.
5. Guillet P, Alnwick D, Cham MK, Neira M, Zaim M, Heymann D, Mukelabai K: *Long-lasting treated mosquito nets: a breakthrough in malaria prevention*. *Bull World Health Organ* 2001, **79**:998.
6. RBM: *The Global Partnership to Roll Back Malaria*, Initial period covered July 1998–December 2001 (preparatory phase: July 1998–December 1999). Proposed strategy and work plan. Draft 3.1b/12 Geneva: World Health Organization; 1999.
7. Curtis CF, Myamba J, Wilkes TJ: *Various pyrethroids on bednets and curtains*. *Mem Inst Oswaldo Cruz* 1992, **87**:363–370.
8. Zaim M, Aitio A, Nakashima N: *Safety of pyrethroid-treated mosquito nets*. *Med Vet Entomol* 2000, **14**:1–5.
9. Lindsay SW, Gibson ME: *Bednets revised - old idea, new angle*. *Parasitol Today* 1988, **4**:270–272.
10. Bhatt RM, Sharma SN, Urugayala S, Dash AP, Kamaraju R: *Effectiveness and durability of Interceptor® long-lasting insecticidal nets in a malaria endemic area of central India*. *Malar J* 2012, **11**:189.
11. Russell TL, Lwetoijera DW, Maliti D, Chipwaza B, Kihonda J, Charlwood JD, Smith TA, Lengeler C, Mwanyangala MA, Nathan R, Knols BG, Takken W, Killeen GF: *Impact of promoting longer-lasting insecticide treatment of bed nets upon malaria transmission in a rural Tanzanian setting with pre-existing high coverage of untreated nets*. *Malar J* 2010, **9**:187.
12. Sharma SK, Upadhyay AK, Haque MA, Tyagi PK, Mohanty SS, Raghavendra K, Dash AP: *Field evaluation of Olyset nets: a long-lasting insecticidal net against malaria vectors Anopheles culicifacies and Anopheles fluviatilis in a hyperendemic tribal area of Orissa, India*. *J Med Entomol* 2009, **46**:342–350.
13. Takken W: *Do insecticide-treated bednets have an effect on malaria vectors?* *Trop Med Int Health* 2002, **7**:1022–1030.
14. Ranson H, N'guessan R, Lines J, Moiroux N, Nkuni Z, Corbel V: *Pyrethroid resistance in African anopheline mosquitoes: what are the implications for malaria control?* *Trends Parasitol* 2011, **27**:91–98.
15. Mathias D, Ochomo EO, Atieli F, Ombok M, Bayoh MN, Olang G, Muhia D, Kamau L, Vulule JM, Hamel MJ, Hawley WA, Walker ED, Gimnig JE: *Spatial and temporal variation in the kdr allele L1014S in Anopheles gambiae s.s. and phenotypic variability in susceptibility to insecticides in Western Kenya*. *Malar J* 2011, **10**:10.
16. Dabiré RK, Diabaté A, Baldet T, Paré-Toé L, Guiguemdé RT, Ouedraogo JB, Skovmand O: *Personal protection of long lasting insecticide-treated nets in areas of Anopheles gambiae s.s. resistance to pyrethroids*. *Malar J* 2006, **5**:12.
17. Van Bortel W, Chinh VD, Berkvens D, Speybroeck N, Trung HD, Coosemans M: *Impact of insecticide-treated nets on wild pyrethroid resistant Anopheles epiroticus population from southern Vietnam tested in experimental huts*. *Malar J* 2009, **8**:248.
18. Gouagna LC, Okech BA, Kabiru EW, Killeen GF, Obare P, Ombonya S, Bier JC, Knols BG, Githure JI, Yan G: *Infectivity of Plasmodium falciparum gametocytes in patients attending rural health centers in western Kenya*. *East Afr Med J* 2003, **80**:627–634.
19. Noor AM, Gething PW, Alegana VA, Patil AP, Hay SI, Muchiri E, Juma E, Snow RW: *The risks of malaria infection in Kenya in 2009*. *BMC Infect Dis* 2009, **9**:180.
20. Kawada H, Futami K, Komagata O, Kasai S, Tomita T, Sonye G, Mwatele C, Njenga SM, Mwandawiro C, Minakawa N, Takagi M: *Distribution of a knockdown resistance mutation (L1014S) in Anopheles gambiae s.s. and Anopheles arabiensis in Western and Southern Kenya*. *PLoS One* 2011, **6**:e24323.
21. Kawada H, Dida GO, Ohashi K, Komagata O, Kasai S, Tomita T, Sonye G, Maekawa Y, Mwatele C, Njenga SM, Mwandawiro C, Minakawa N, Takagi M: *Multimodal pyrethroid resistance in malaria vectors, Anopheles gambiae s.s., Anopheles arabiensis, and Anopheles funestus s.s. in western Kenya*. *PLoS One* 2011, **6**:e22574.
22. Kawada H, Ohashi K, Dida GO, Sonye G, Njenga SM, Mwandawiro C, Minakawa N: *Insecticidal and repellent activities of pyrethroids to the*

- three major pyrethroid-resistant malaria vectors in western Kenya. *Parasit Vectors* 2014, **7**:208.
23. Gillies MT, Coetzee M: A supplement to the Anophelinae of Africa south of the Sahara (Afrotropical region). *S Afr Inst Med Res* 1987, **55**:1–143.
 24. Scott JA, Brogdon WG, Collins FH: Identification of single specimens of the *Anopheles gambiae* complex by the polymerase chain reaction. *Am J Trop Med Hyg* 1993, **49**:520–529.
 25. Koekemoer LL, Kamau L, Hunt RH, Coetzee M: A cocktail polymerase chain reaction (PCR) assay to identify members of the *Anopheles funestus* (Diptera: Culicidae) group. *Am J Trop Med Hyg* 2002, **66**:804–811.
 26. Kent BJ, Norris DE: Identification of mammalian blood meals in mosquitoes by a multiplexed polymerase chain reaction targeting cytochrome B. *Am J Trop Med Hyg* 2005, **73**:336–342.
 27. Bertram DS: Tsetse and trypanosomiasis control in Nyanza Province, Kenya. *Trans R Soc Trop Med Hyg* 1969, **63**:125.
 28. Kawada H, Dida GO, Sonye G, Njenga SM, Mwandawiro C, Minakawa N: Reconsideration of *Anopheles rivulorum* as a Vector of *Plasmodium falciparum* in Western Kenya: some evidence from biting time, blood preference, sporozoite positive rate, and pyrethroid resistance. *Parasit Vectors* 2012, **5**:230.
 29. Kawada H, Takagi M: Photoelectric sensing device for recording mosquito host-seeking behavior in the laboratory. *J Med Entomol* 2004, **41**:873–881.
 30. Bøgh C, Pedersen EM, Mukoko DA, Ouma JH: Permethrin-impregnated bednet effects on resting and feeding behaviour of lymphatic filariasis vector mosquitoes in Kenya. *Med Vet Entomol* 1998, **12**:52–59.
 31. N'Guessan R, Corbel V, Akogbeto M, Rowland M: Reduced efficacy of insecticide-treated nets and indoor residual spraying for malaria control in pyrethroid resistance area, Benin. *Emerg Infect Dis* 2007, **13**:199–206.
 32. Chandre F, Dabire RK, Hougard JM, Djogbenou LS, Irish SR, Rowland M, N'Guessan R: Field efficacy of pyrethroid treated plastic sheeting (durable lining) in combination with long lasting insecticidal nets against malaria vectors. *Parasit Vectors* 2010, **3**:65.
 33. Mathenge EM, Gimnig JE, Kolczak M, Ombok M, Irungu LW, Hawley WA: Effect of permethrin-impregnated nets on exiting behavior, blood feeding success, and time of feeding of malaria mosquitoes (Diptera: Culicidae) in western Kenya. *J Med Entomol* 2001, **38**:531–536.
 34. Shillu J, Ghebremeskel T, Seulu F, Mengistu S, Fekadu H, Zerom M, Asmelash GE, Sintasath D, Mbogo C, Githure J, Brantly E, Beier JC, Novak RJ: Seasonal abundance, vector behavior, and malaria parasite transmission in Eritrea. *J Am Mosq Control Assoc* 2004, **20**:155–164.
 35. Amenesheba B, Service MW: Blood-feeding behaviour of *Anopheles arabiensis* Patton (Diptera: Culicidae) in central Ethiopia. *J Afr Zool* 1997, **11**:235–245.
 36. Russell TL, Govella NJ, Azizi S, Drakeley CJ, Kachur SP, Killeen GF: Increased proportions of outdoor feeding among residual malaria vector populations following increased use of insecticide-treated nets in rural Tanzania. *Malar J* 2011, **10**:80.
 37. Yohannes M, Boelee E: Early biting rhythm in the Afro-tropical vector of malaria, *Anopheles arabiensis*, and challenges for its control in Ethiopia. *Med Vet Entomol* 2012, **26**:103–105.
 38. Moiroux N, Gomez MB, Pennetier C, Elanga E, Djèntontin A, Chandre F, Djègbé I, Guis H, Corbel V: Changes in *Anopheles funestus* biting behavior following universal coverage of long-lasting insecticidal nets in Benin. *J Infect Dis* 2012, **206**:1622–1659.
 39. Gattton ML, Chitnis N, Churcher T, Donnelly MJ, Ghani AC, Godfray HC, Gould F, Hastings I, Marshall J, Ranson H, Rowland M, Shaman J, Lindsay SW: The Importance of mosquito behavioral adaptations to malaria control in Africa. *Evolution* 2013, **67**:1218–1230.
 40. Bayoh MN, Mathias DK, Odiero MR, Mutuku FM, Kamau L, Gimnig JE, Vulule JM, Hawley WA, Hamel MJ, Walker ED: *Anopheles gambiae*: historical population decline associated with regional distribution of insecticide-treated bed nets in western Nyanza Province. *Kenya Malar J* 2010, **9**:62.
 41. Lindsay SW, Jawara M, Paine K, Pinder M, Walraven GE, Emerson PM: Changes in house design reduce exposure to malaria mosquitoes. *Trop Med Int Health* 2003, **8**:512–517.
 42. Kawada H, Dida GO, Ohashi K, Sonye G, Njenga SM, Mwandawiro C, Minakawa N, Takagi M: Preliminary evaluation of insecticide-impregnated ceiling nets with coarse mesh size as a barrier against the invasion of malaria vectors. *Jpn J Infect Dis* 2012, **65**:243–246.
 43. Kawada H: New mosquito control techniques as countermeasures against insecticide resistance. In *Insecticides - Advances in Integrated Pest Management*. Edited by Perveen F, Rijeka, Croatia: InTech; 2012:657–682.

doi:10.1186/1756-3305-7-383

Cite this article as: Kawada et al.: Preventive effect of permethrin-impregnated long-lasting insecticidal nets on the blood feeding of three major pyrethroid-resistant malaria vectors in western Kenya. *Parasites & Vectors* 2014 **7**:383.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

