Detachment of hard ticks (Acari: Ixodidae) from hunted sika deer (Cervus nippon)

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Present address: Department of Vector Ecology and Environment, Institute of Tropical Medicine, Nagasaki University, 1-12-4 Sakamoto, Nagasaki 852-8523, Japan E-mail: tsunoda@nagasaki-u.ac.jp Telephone number: +81 95 819 7809 Fax number: +81 95 819 7812 Ixodid ticks attach to their host to suck blood meal, with attachment often lasting 1-2 weeks (Balashov 1972). However, when a host dies, ticks gradually detach and leave. The distribution of tick species and genders on a host may change as a function of time after the host's death. Since hundreds or thousands of ticks often detach from a dead host, it is difficult to count the number of ticks that detach from a host after its death. Little is known about detachment of ixodid ticks from dead hosts.

The hypostome of an attached ixodid tick penetratees its host for a blood meal (Arthur 1962; Moorhouse 1967). A cement-like substance from the salivary glands glues the mouthparts firmly into the host to ensure continuous attachment during the long feeding times (Kemp et al. 1982). It is assumed that the fraction of ixodid ticks on a host with their mouthparts glued to the host, relative to the fraction that have detached and left, gradually decreases after the death of the host.

Sixteen species of ixodid ticks have been reported to be ectoparasites of sika deer in Japan (Ohdachi et al. 2009). In the Boso Peninsula of Japan, the bush tick *Haemaphysalis longicornis* is the dominant parasite of sika deer in the summer and *Haemaphysalis megaspinosa* is dominant in the winter (Mori et al. 1995), with adults of both species abundant in March (Tsunoda 2007).

Ticks were obtained from 13 sika deer, *Cervus n. nippon*, shot in Amatsu, the eastern part of Kamogawa City in the Boso Peninsula in central Japan, from late February to early March 1999. Hunters put each deer in a large polyvinyl plastic bag immediately after they shot it. Cadavers were kept in the bags until the hunters dissected them 1 to 3 hr after culling. Fur from the head, legs, and body were put in another polyvinyl plastic bag, brought to a laboratory 3 hr later, and kept in a -20°C freezer to kill ticks.

Fur was kept at 25°C for at least for 12 hr before collection of ticks. For collection from a deer body surface, ticks were collected from either the left or right side of the head and body and from the legs. Ticks were also collected from the polyvinyl plastic bag. A tick with its hypostome inserted into the host skin (designated attached) was distinguished from a tick that was detached and on the body surface. In addition, female ticks were classified in three groups based on their blood feeding stage: engorged (i.e. swollen into a globular shape and with a black color), partially-engorged (i.e. swollen, but not into a globular shape, and with a brown color), and unfed (i.e. unfed or little feeding, although these were difficult to discriminate).

A total of 3,954 adult, 4,495 nymph and 83 larva *Haemaphysalis* ticks were collected from 13 sika deer. The most abundant adult ticks collected were *H. megaspinosa*, and smaller numbers of *H. longicornis*, *H. flava*, *H. kitaokai*, and *H.*

cornigera (Table 1). *H. ias* is a synonym of *H. cornigera* (Takada 1990) However, 92 *Haemaphysalis* females, most of them were partially-engorged, could not be identified because they had lost their capitulum when they were pulled out of their hosts with forceps. In addition, 35 *Ixodes ovatus* adults and 1 *Amblyomma testudinarium* nymph were also collected.

Males were more abundant than females for the *H. longicornis*, *H. megaspinosa*, and *H. flava* ticks collected (Fig. 1A-C) but not for *H.kitaokai* (Fig. 1D) (Wilcoxson signed-rank test, *H. longicornis*: Z=91, P<0.01; *H. megaspinosa*: Z=91, P<0.001; *H. flava*: Z=90, P<0.01; *H.kitaokai*: Z=62.5, P=0.07). There was a significant difference in the sex ratio of *H. megaspinosa*, *H. longicornis*, and *H. kitaokai* ticks that were attached or detached (Table 2a). A greater fraction of the male than female *H. megaspinosa*, *H. longicornis*, and *H. kitaokai* (Fig. 2).

A generalized linear model (GLM) with a binomial distribution and a log link function was used to test whether detached or attached ticks were significantly different in terms of sex ratio and blood feeding stage. For feeding stage analysis, engorged ticks were grouped with partially-engorged ticks because the number of engorged ticks was small. Overdispersion was resolved by use of a quasi-binomial distribution. Model comparisons were analyzed using the F-test. There was a significant relationship between whether *H. megaspinosa* and *H. longicornis* ticks were attached or detached and their feeding stage (Table 2b). More detached *H. longicornis* and *H. megaspinosa* unfed female ticks were collected than engorged and partially-engorged females (Fig. 3). However, for *H. kitaokai*, there was no significant relationship between whether female ticks were attached or detached and their feeding stage.

More *H. megaspinosa*, *H. longicornis*, *H. flava*, and *H. kitaokai* male than female ticks were collected on the sika deer in this study. It has been reported that there were more *H. megaspinosa* and *H. flava* male than female ticks on wild boar, and more *H. longicornis* male than female ticks on Reeve's muntjac (Tsunoda 2012). Metastriata ticks mate on their host and prostriata ticks mate both on and off their host (Sonenshine 1991). Although some prostriata male ticks of the metastriata species *Amblyomma americanum* can inseminate up to 37 females (Gladney and Drummond 1970). The sex ratio of metastriata ticks, with more males than females on hosts, suggests that the male's reproductive strategy may be to remain longer on their hosts to enable them to encounter and mate with more females.

Although more *H. longicornis* and *H. kitaokai* females than males were attached, more *H. megaspinosa*, *H. longicornis*, *H. flava*, and *H. kitaokai* males than females

detached after their host died. These results indicated that *H. megaspinosa*, *H. longicornis*, *H. flava*, and *H. kitaokai* male ticks detached sooner than female ticks after their host died.

Ixodes dammin (the same species as *I.scapularis*) eggs are fertilized by sperm from males a second copulation when a female copulates with two males (Yuval and Spielman 1990). Although there is no information on sperm displacement in metastriata, it should be advantageous for males to guard females after insemination until females detach from the host. In general, female ticks that have larger blood meals lay more eggs (Sonenshine 1991). In *Rhipicephalus appendiculatus*, a male helps its female mate feed by modulating the host immune response during co-feeding (Wang et al. 1998). However, if a host is dead, the male does not have to stay attached long because it is difficult for females to take a blood meal. Therefore, it may be beneficial for males to detach from a dead host and wait for a new host.

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Figure legends

Figure 1. Number of male and female ticks per deer. (A) *Haemaphysalis* longicornis, (B) *H. megaspinosa*, (C) *H. flava*, (D) *H. kitaokai*. The mean \pm SE of each population is shown. Asterisks show significant difference: **p<0.01, ***p<0.001; NS indicates lack of significance at p>0.05.

Figure 2. Percent of male and female ticks that were detached or attached. (A) *Haemaphysalis longicornis*, (B) *H. megaspinosa*, (C) *H. flava*, (D) *H. kitaokai*. The numbers in parenthesis denote the total numbers of ticks.

Figure 3. Percent of fed (engorged + partially-engorged) and unfed ticks that were detached or attached. (A) *Haemaphysalis longicornis*, (B) *H. megaspinosa*, (C) *H. kitaokai*. The numbers in parenthesis denote the total numbers of ticks.

| Species | Male | | Female | | | |
|---------------|------|----------|--------------------|-------|--|--|
| | | Engorged | Partially-engorged | Unfed | | |
| H.longicornis | 244 | 1 | 63 | 70 | | |
| H.megaspinosa | 1982 | 30 | 649 | 388 | | |
| H.flava | 91 | 0 | 6 | 13 | | |
| H.kitaokai | 176 | 9 | 48 | 53 | | |
| H.cornigera | 1 | 0 | 1 | 0 | | |
| H.sp. | 0 | 4 | 84 | 4 | | |
| I.ovatus | 16 | 5 | 11 | 3 | | |
| I.sp. | 0 | 0 | 2 | 0 | | |

 Table 1. Adult Haemaphysalis and Ixodes ticks collected from 13 sika deer in the

 Boso Peninsula

| (A) | | | | | |
|-------------------|----------------------|----------|------|-------|---------|
| Response variable | Explanatory variable | Estimate | SE | Т | Р |
| H. megaspinosa | Intercept | 0.69 | 0.06 | 11.61 | < 0.001 |
| | Condition* | -0.76 | 0.20 | -3.85 | < 0.001 |
| H.longicornis | Intercept | 0.78 | 0.13 | 5.85 | < 0.001 |
| | Condition* | -1.63 | 0.42 | -3.92 | < 0.001 |
| H. flava | Intercept | 1.49 | 0.25 | 5.88 | < 0.001 |
| | Condition* | 0.91 | 1.05 | 0.87 | 0.40 |
| H. kitaokai | Intercept | 1.07 | 0.22 | 4.83 | < 0.001 |
| | Condition* | -2.15 | 0.43 | -4.97 | < 0.001 |

Table 2. Results of GLM for (A) sex ratio of four species of ticks and (B) bloodfeeding level of three species

*Condition: detached or attached.

| (| R) |
|----|----|
| L) | D) |
| | |

| Response variable | Explanatory variable | Estimate | SE | Т | Р |
|-------------------|----------------------|----------|------|-------|---------|
| H. megaspinosa | Intercept | 1.63 | 0.38 | 4.28 | < 0.001 |
| | Condition* | -1.33 | 0.40 | -3.32 | < 0.005 |
| H.longicornis | Intercept | 1.05 | 0.58 | 1.82 | 0.09 |
| | Condition* | -1.57 | 0.65 | -2.41 | 0.03 |
| H. kitaokai | Intercept | 0.14 | 0.35 | 0.41 | 0.69 |
| | Condition* | 0.25 | 0.48 | 0.53 | 0.61 |

*Condition: detached or attached.

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Figure 1.



Figure 2.



Figure 3.

