



Full length article

Surface modification of P_{DL}LGA microspheres with gelatine methacrylate: Evaluation of adsorption, entrapment, and oxygen plasma treatment approaches



Abdulrahman Baki^a, Cheryl V. Rahman^a, Lisa J. White^a, David J. Scurr^b, Omar Qutachi^a, Kevin M. Shakesheff^{a,*}

^a Division of Drug Delivery and Tissue Engineering, School of Pharmacy, University of Nottingham, University Park, Nottingham NG7 2RD, UK

^b Laboratory of Biophysics and Surface Analysis, School of Pharmacy, University of Nottingham, University Park, Nottingham NG7 2RD, UK

ARTICLE INFO

Article history:

Received 6 July 2016

Received in revised form 11 January 2017

Accepted 12 January 2017

Available online 16 January 2017

Keywords:

Surface modification
P_{DL}LGA microspheres
Gelatine methacrylate
Drug delivery
Tissue engineering

ABSTRACT

Injectable poly (dl-lactic-co-glycolic acid) (P_{DL}LGA) microspheres are promising candidates as biodegradable controlled release carriers for drug and cell delivery applications; however, they have limited functional groups on the surface to enable dense grafting of tissue specific biocompatible molecules. In this study we have evaluated surface adsorption, entrapment and oxygen plasma treatment as three approaches to modify the surfaces of P_{DL}LGA microspheres with gelatine methacrylate (gel-MA) as a biocompatible and photo cross-linkable macromolecule. Time of flight secondary ion mass spectroscopy (TOF SIMS) and X-ray photoelectron spectroscopy (XPS) were used to detect and quantify gel-MA on the surfaces. Fluorescent and scanning electron microscopies (SEM) were used to image the topographical changes. Human mesenchymal stem cells (hMSCs) of immortalised cell line were cultured on the surface of gel-MA modified P_{DL}LGA microspheres and Presto-Blue assay was used to study the effect of different surface modifications on cell proliferation. Data analysis showed that the oxygen plasma treatment approach resulted in the highest density of gel-MA deposition. This study supports oxygen plasma treatment as a facile approach to modify the surface of injectable P_{DL}LGA microspheres with macromolecules such as gel-MA to enhance proliferation rate of injected cells and potentially enable further grafting of tissue specific molecules.

Statement of Significance

Poly (dl lactic-co-glycolic acid) (P_{DL}LGA) microspheres offer limited functional groups on their surface to enable proper grafting of tissue specific bioactive molecules. To overcome this limitation, previous approaches have suggested using alkaline solutions to introduce active groups to the surface; however, they may compromise surface topography and lose any potential surface patterns. Plasma polymerisation of bioactive monomers has been suggested to enhance surface biocompatibility; however, it is not applicable on low vapour pressure macromolecules such as most extracellular matrix (ECM) proteins and growth factors. This study aims to evaluate three different approaches to modify the surface of P_{DL}LGA microspheres with gelatine-methacrylate (gel-MA) to enable further grafting of cross-linkable biomolecules without compromising the surface topography or the biocompatibility of the system.

© 2017 Acta Materialia Inc. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Injectable P_{DL}LGA microspheres have been proposed as cell delivery systems as they can form porous scaffolds [1–3] and con-

trol the release of signalling molecules into the cell niche [4–7]. However, P_{DL}LGA microspheres have limited reactive groups, such as carboxyl groups, available on the surface; this may limit the potential for further grafting of molecules required to support cell attachment and proliferation [8,9]. To overcome this limitation, different approaches have been proposed to functionalise the surface and to enable further grafting [10,11].

* Corresponding author.

E-mail address: kevin.shakesheff@nottingham.ac.uk (K.M. Shakesheff).

Some approaches have been reported to modify the surface by incorporating or copolymerising specific biomolecules such as collagen or biotinylated poly ethylene glycol (PEG) with the bulk polymer or monomer units conforming polymer chains [12,13]. However, as these approaches may compromise the physicochemical properties of the bulk polymer, subsequent approaches focused on the surface modification instead [14]. Both surface adsorption and layer by layer deposition (LBL) of bioactive molecules used the electrostatic charges on the polymer surface to introduce oppositely charged molecules without changing polymer structure; however, these methods require a prior surface activation step to increase the surface charge [15–18].

To increase the available surface charge, wet chemistry approaches have been suggested to functionalise the polymer surface by breaking polymer chains using alkaline solutions [19–21]; however, these approaches can also degrade the polymer surface and change surface topography [22]. Surface entrapment of molecules can functionalise inert polymer surfaces without altering their bulk chemistry [23]. This approach involves inducing polymer swelling with a modifying mixture made of a swelling agent [24] or a diluted solvent [25] with a dissolved modifying molecule. On the collapse of the swollen layer, the modifying molecule is trapped as a penetrating network at the surface of the polymer structure [26–28]. Electron beam irradiation and plasma induced polymerisation were reported to functionalise the surface with a minor effect on surface topography [29–37].

Gelatine has been used as cell delivery carrier and as a collagen derived molecule to enhance cell adhesion and proliferation [38–40]. Grafting methacrylate groups to the chains of macromolecules such as gelatine and gellan gum produces methacrylated chains such as gelatine methacrylate (gel-MA) and gellan gum methacrylate (GG-MA) which can be chemically crosslinked to obtain stiffer hydrogels with controllable degradation rate and mechanical properties [41–44]. Gel-MA has also multiple cross-linkable sites which can be exploited for chemical grafting of other molecules for different tissue engineering applications [45].

Functionalising the polymer surface with cross-linkable small molecules such as vinyl based monomers was also reported to enable grafting of cross-linkable macromolecules. However, this approach may involve multistep reactions with corrosive chemicals to enable grafting which may compromise the activity of the grafted macromolecules and affect the topography of polymer surface [46,47].

In this study, surface adsorption, surface entrapment, and oxygen plasma treatment approaches were investigated as single step approaches to modify the surface of P_{DL}LGA microspheres with gel-MA macromolecules. The proposed surface modification approaches were later evaluated using different surface imaging and analytical techniques to determine the highest density of gel-MA molecules on the modified surface. Gel-MA modified P_{DL}LGA microspheres were proposed to enhance stem cell interaction and to enable further grafting of further gel-MA molecules for different stem cell therapy applications.

2. Materials and methods

2.1. Material Preparation and fabrication

Poly (DL lactic-co-glycolic acid) (uncapped P_{DL}LGA 8515, MW 52 kDa, Evonik – USA) microspheres were prepared with a preparation method reported earlier [7]. Briefly, 20% (w/v) P_{DL}LGA solution in dichloromethane (DCM, Thermofisher Scientific, UK) was poured into 0.3% (w/v) poly(vinyl alcohol) (PVA, 86–89% hydrolysed Alfa Aesar-USA) solution in demineralised water and homogenised at 3000 RPM for 2 min using a propeller homogeniser (Silverson

L5 M, UK). The resultant (o/w) emulsion was left stirring overnight to allow DCM solvent evaporation. P_{DL}LGA microspheres were then centrifuged (MSE Mistral 1000, UK) and washed twice with distilled water, freeze-dried for 48 h (ModulyoD, Thermofisher Scientific, USA) and stored in a vacuum packed containers at (–20 °C).

Gelatine methacrylate (Gel-MA) was prepared following a published method [41]. Briefly, 10% gelatine (Type B gelatin from Bovine skin, Sigma-UK) solution in phosphate buffered saline (PBS Gibco, Thermofisher Scientific-UK) was prepared at 60 °C and kept under vigorous stirring (~600 rpm, Thermofisher Scientific-UK) while 20 mL of methacrylic anhydride (MAA, Sigma-Germany) was injected dropwise over 20 min. The reaction mixture stirred at 60 °C for a further two hours until a white viscous liquid formed. The reaction was then stopped using 500 mL of warm PBS and the mixture was moved into dialysis tubes (Biodesign Dialysis Tubing D118, cut-off at 8000 MWCO, 30.5 cm length * 49.5 cm wet diameter, UK) and dialysed against daily changed distilled water for one week. After dialysis, the mixture became a clear to pale yellow solution. Gel-MA solution was later lyophilised to obtain white porous foam of gel-MA and stored at (–80 °C).

Fluorescein isothiocyanate (Fit-C isomer 1, Sigma-USA) was used to prepare Fit-C conjugated gel-MA as per manufacturer protocol to allow fluorescent detection of gel-MA layer on surface modified P_{DL}LGA microspheres. Briefly, 80 mg of sodium hydroxide (NaOH, Sigma-UK) and 168 mg of sodium bicarbonate (NaHCO₃, Sigma-UK) were dissolved in 20 mL of distilled water to obtain 1 N carbonate buffer solution with pH adjusted at 9.6 (Mettler Toledo – UK). Labelling solution was prepared by dissolving 10 mg of Fit-C in 1 mL of dimethyl sulfoxide (DMSO, Fischer Scientific-USA) in the dark. A small amount of gel-MA (500 mg) was dissolved in the freshly prepared carbonate buffer solution and the conjugation reaction was started by adding Fit-C solution dropwise in the dark with continuous stirring (~300 rpm) to prevent Fit-C precipitation. Once all the fit-C solution was added, the mixture was incubated at 4 °C for 24 h in the dark with gentle stirring (~200 rpm) to allow Fit-C conjugation to gel-MA free amine groups. The conjugation reaction was later neutralised by adding 20 mL of ammonium chloride (NH₄Cl 1 N – Sigma, USA) to obtain a solution with (pH = 7). The final mixture was later dialysed (8000 MW cut-off dialysis bags) against distilled water to filter out unconjugated fit-C, freeze-dried, and stored at –80 °C (Supplementary Data 1).

2.2. Surface modification of P_{DL}LGA microspheres

2.2.1. Surface adsorption

P_{DL}LGA microspheres (500 mg) were dispersed in 2 mL of 10% (w/v) gel-MA /phosphate buffered saline (PBS) solution. The mixture was stirred at 300 rpm for 24 h before washing twice with water for collection as shown in provided schematic (Fig. 1).

2.2.2. Surface entrapment

A modifying mixture was freshly prepared by mixing 0.75 mL of 2,2,2-trifluoroethanol (TFE, Sigma-UK) with 4.25 mL of 10% (w/v) gel-MA/PBS solution and P_{DL}LGA microspheres (500 mg) were dispersed in 2 mL of the modifying mixture and kept under continuous stirring (~300 rpm) to induce swelling and allow sufficient time for gel-MA to infiltrate the P_{DL}LGA polymer matrix. After 24 h, the mixture was quenched with 20 mL of distilled water and kept under continuous stirring for further 30 min to induce the collapse of the swollen polymer surface and entrap gel-MA molecules. Treated microspheres were washed twice with water, freeze-dried, and stored at –20 °C for later analysis (Fig. 1).

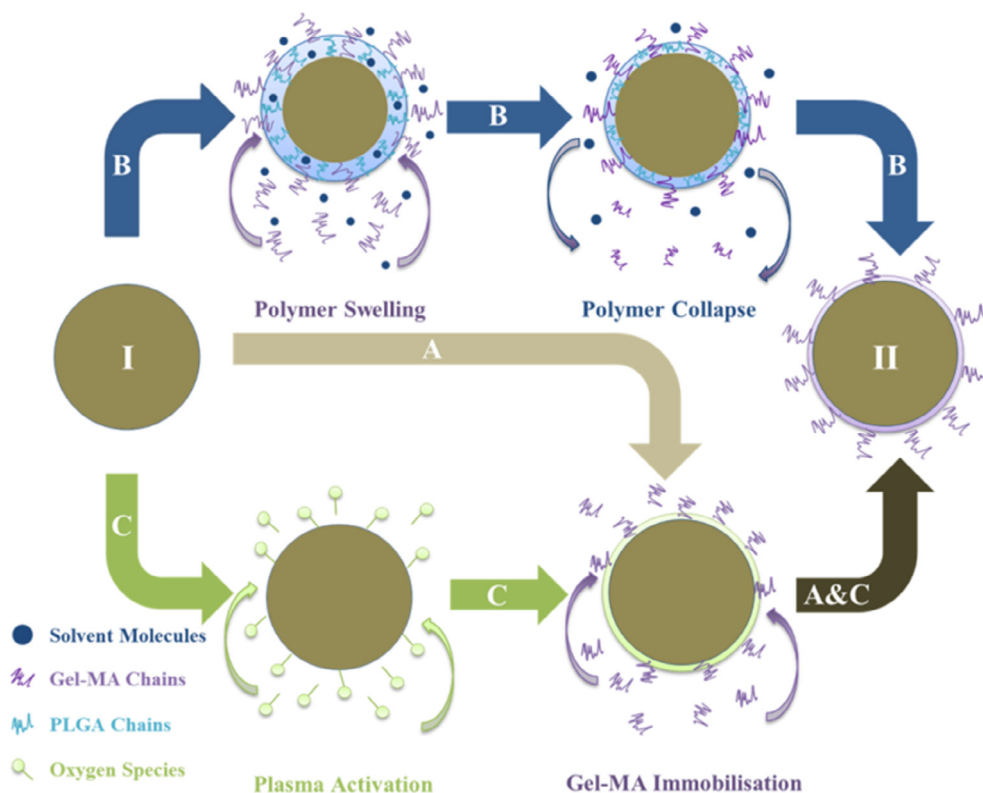


Fig. 1. Schematic figure showing the different surface treatment approaches used to modify the surface of PDLGA microspheres (I) with gel-MA to obtain gel-MA modified microspheres (II). Adsorption (A) is used to utilise the potential charge on the polymer chains to enable direct deposition of gel-MA molecules on the surface. Surface entrapment (B) is proposed to entrap gel-MA molecules on the surface of PDLGA microspheres without compromising polymer chain composition. Finally, plasma treatment is proposed to immobilise gel-MA molecules on the surface of PDLGA microspheres by introducing reactive oxygen species to PDLGA polymer chains on the surface (C).

2.2.3. Plasma treatment

Low pressure oxygen plasma treatment was performed to introduce oxygen species to the surface. Briefly, PDLGA microspheres (500 mg) were put in 25 mL glass vials in a plasma machine chamber (Diener Nano Type F, Diener Electronic GMBH – Germany) and kept rotating at 60 rpm using a rotary holder. Chamber pressure was pumped down to 20 mbar then oxygen gas was pumped into the chamber for 2 min to obtain a working pressure of 50 mbar. As working pressure was maintained at 50 mbar with continuous oxygen gas supply, plasma activation was initiated at 150 W (low frequency generator – 40 kHz) for 5 min. Following plasma activation, the chamber was purged with nitrogen gas for 2 min to dispose reactive oxygen ions and then air vented to atmospheric pressure to collect the samples. Following plasma treatment, PDLGA microspheres were dispersed in 2 mL of 10% (w/v) gel-MA/PBS solution and kept stirring for 24 h to allow enough time for surface modification. Treated microspheres were later washed twice with water, freeze-dried, and stored at -20°C for later analysis (Fig. 1).

2.3. Surface analysis of modified PDLGA microspheres

2.3.1. X-ray photoelectron spectroscopy (XPS)

XPS (Kratos Axis ULTRA DLD) with a monochromatic X-ray source of aluminium (Al K α) with emission at (1486.6 eV) was used to detect and quantify gel-MA concentration on the surface of modified PDLGA microspheres. Microspheres were mounted on carbon discs and both wide and high resolution scans were performed on three different areas of 0.25×0.25 mm size at room temperature while obtained peaks were analysed to quantify gel-

MA related (N1s) peaks on the surface of modified microspheres using Casa XPS program (version 2.3.16 PR 1.6, Casa Software Ltd.).

2.3.2. Time of flight secondary ion mass spectroscopy (ToF SIMS)

The ToF-SIMS instrument (ION-TOF, ION-TOF GmbH) was used to analyse treated PDLGA microspheres using a bismuth (Bi⁺) ion source at room temperature. Three different areas of 0.25×0.25 mm size were scanned in each sample to analyse chemical composition of modified microsphere surface. Negative spectrums were collected and processed using a program provided by ION-TOF GmbH. ToF-SIMS spectra were analysed to create images of gel-MA distribution on the surface of modified PDLGA microspheres.

2.4. Preparation of cross-sectioned PDLGA microspheres

PDLGA microspheres were mixed with 1 mL of O.C.T compound (Mounting Medium for Cryotomy, VWR international – Belgium) and moulded into aluminium sheet cylinders of $10 \times 20 \times 20$ mm. The moulds were later immersed in N-Isopentane bath (Sigma-UK) frozen in liquid nitrogen until the O.C.T became solid white. Frozen moulds were mounted and cross-sectioned at -20°C using a micro-cryotome (Leica CM1100 – Germany) to obtain $20 \mu\text{m}$ thickness slices of the O.C.T/microsphere mixture. Slices containing cross-sectioned microspheres were washed twice with distilled water to remove residual O.C.T, dispersed on a glass slide, and dried using a blast of N₂ gas to obtain final microsphere cross-sections.

2.5. Imaging of surface modified P_DLGA microspheres

2.5.1. Fluorescent microscopy imaging

Cross-sectioned P_DLGA microspheres modified with Fit-C labelled gel-MA were visualised on a Leica inverted fluorescent microscope (Leica DM IRB, Germany) using Fit-C light filter cube set at 495 nm. Images were taken using a microscope mounted camera (Qicam imaging, Canada) at a gain of 1 and a camera exposure time of 500 ms.

2.5.2. Focused ion beam scanning electron microscopy (FIB-SEM)

Surface modified P_DLGA microspheres were mounted on carbon discs and coated with platinum for 5 min in a chamber attached to the FIB SEM (FEI Quanta 200 3D Dual Beam FIB-SEM). A focused ion beam was emitted on one of the P_DLGA microspheres to mill through the surface and to obtain a square hole exposing the inner structure of the microsphere. The FIB-SEM stage was later tilted to (30°) degree to enable imaging of the inner structure of P_DLGA microspheres using an electron beam acceleration of 15 kV.

To image cross-sectioned P_DLGA microspheres, samples were dispersed on carbon discs mounted on aluminium stubs (Agar, UK) and gold coated with benchtop plasma coater for 5 min at 25 mA (Leica EM SCD005 Sputter Coater) and imaged with SEM (JEOL 6060LV variable pressure SEM, UK) at electron beam acceleration of 14 kV and a working distance of 25 mm between electron beam source and sample surface.

2.6. Mesenchymal stem cell proliferation study

2.6.1. Bicinchoninic acid (BCA) assay

Bicinchoninic acid assay (Micro BCA Protein Assay Kit, ThermoFisher Scientific-UK) was used as per manufacturer's protocol to quantify the total content of gel-MA on the surface of P_DLGA microspheres. Briefly, P_DLGA microspheres (100 mg) were dispersed in 1 mL of PBS and mixed with 1 mL of BCA mixture then incubated at 37 °C for 2 h. The mixture was injected through 0.2 µm mesh size filter to obtain clear P_DLGA microsphere-free solution. The solution was pipetted into 5 wells of a 96 well plate (Transparent 96 well plate, Falcon-USA) with 300 µL pipetted into each well. A plate reader (Tecan 200, Switzerland) was used to read the absorbance value of UV light at 562 nm to determine colour intensity. Intensity values were normalised against a standard curve of known gel-MA concentrations.

2.6.2. Mesenchymal stem cell seeding and proliferation

A cell culture medium was prepared using Dulbecco's Modified Eagle Medium (Gibco DMEM, ThermoFisher Scientific, UK) supplemented with (10% v/v) of foetal calf serum (ThermoFisher Scientific, UK), (1% v/v) of penicillin/streptomycin solution (anti-biotic/anti-mycotic 1x Gibco, ThermoFisher Scientific, UK), (1%v/v) of L-Glutamine (2 mM Gibco, ThermoFisher Scientific, UK), and (1% v/v) of MEM non-essential amino acids solution (Sigma-Aldrich, UK). Immortalised human mesenchymal stem cells obtained from the bone marrow of a healthy female donor (commercial cell, JCRB cell bank, Japan) were cultured and passaged 4 times, and detached using trypsin EDTA (Sigma Aldrich, UK) [48]. Cells were counted and centrifuged at 180g for 5 min, suspended in a freshly prepared cell culture medium at a cell density of 100,000 cells/mL and seeded onto 10 mg of P_DLGA microspheres at a seeding density of 10,000 cells/mg microspheres per well (non-tissue culture plate, Falcon, USA). Plates were then gently shaken to disperse cells, and incubated in a humidified tissue-culture incubator at 37 °C with 5% CO₂.

The Presto-Blue cell viability assay (Invitrogen Life Sciences, UK) was used as per manufacturer protocol to assess cell viability

and proliferation for different time points over 3 days post seeding on P_DLGA microspheres. Cultured cells were aspirated 4 h post seeding to wash away any non-attached cells on the microspheres (day 0), and Presto-Blue assay was performed on day 0, 1, and 3 to evaluate cell adhesion and proliferation on each sample. Briefly, Presto-Blue mixture was prepared as 1:9 parts of Presto-Blue solution: cell culture medium and each sample was incubated in 1 mL of the previous mixture for 45 min at 37 °C. After 45 min, five 100 µL aliquots of each sample were pipetted in 5 wells (R = 5) of 96 well plates (Flat Bottom Black, Coaster-US) and read on a Tecan plate reader at excitation/emission wavelengths of 535 nm and 590 nm respectively. Samples were later aspirated to remove excess Presto Blue mixture and incubated again with 1 mL of cell culture medium at 37 °C with 5% CO₂ for the next reading time point.

2.7. Study design and statistics

Statistical analysis has been performed using GraphPad Prism (Version 6) analysis software. All values recorded in the graphs are averages of 3 different samples ± standard deviation (SD) and differences among groups were determined by ANOVA Tukey-Kramer multiple comparisons test and were considered to be significantly different if p value is less than 0.05.

3. Results

3.1. Detection of gel-MA on the modified microspheres

Wide XPS scan spectra included distinct peaks at C1s (~285–289 eV) and O1s (~531–533 eV) regions that are attributed to P_DLGA. A peak at N1s region (~398–402 eV) records the presence of a molecule with nitrogen containing bonds detected on gel-MA modified microspheres (Fig. 2). The different peak intensity of N1s regions on wide XPS scans between different surface modification approaches may indicate different densities of gel-MA on the surfaces (Supplementary Data 2). Quantification of the gel-MA related N1s peak showed an increase in the surface area of the N1s peak on the surface of plasma modified microspheres compared to adsorption and entrapment modified microspheres. As gel-MA is the only molecule containing nitrogen bonds, the increase in the surface area of N1s peaks indicate a respective increase in the density of gel-MA molecules on the surface (Table 1).

High resolution scan analysis of the C1s region was performed on non-modified P_DLGA microspheres and gel-MA films to obtain the control peaks. Data obtained from non-modified P_DLGA microspheres were assigned to three P_DLGA peaks (C–C, C–O, and O=C–O) and one (C–O) poly vinyl alcohol (PVA) peak (1, 2, 3, and 4 peaks respectively – Fig. 2) as previously reported [49]. Control data from gel-MA films have shown three peaks at (~286.3 eV), (~288.1 eV), and (~289.1 eV) assigned to (C–O), (N–C=O), and (O–C=O) (peak 6, 5, and 7 respectively) with allowed variability of peak position of ±0.1 eV and full width at half

Table 1

Data analysis shows peak area distribution of different elements and their chemical states as obtained using XPS. Reported values represent averages of peak area percentage ± standard deviation obtained from wide XPS scans of three different scan areas.

Peak Area%	C1s	O1s	N1s
P _D LGA	70.13 ± 0.41	29.38 ± 0.65	–
Adsorption	70.3 ± 0.17	27.66 ± 0.47	0.23 ± 0.31
Entrapment	67.37 ± 0.96	31.44 ± 0.54	1.17 ± 0.36
Plasma	67.99 ± 0.83	22.39 ± 0.22	6.27 ± 0.9
Gel-MA	65.32 ± 1.11	20.55 ± 0.5	9.51 ± 0.11

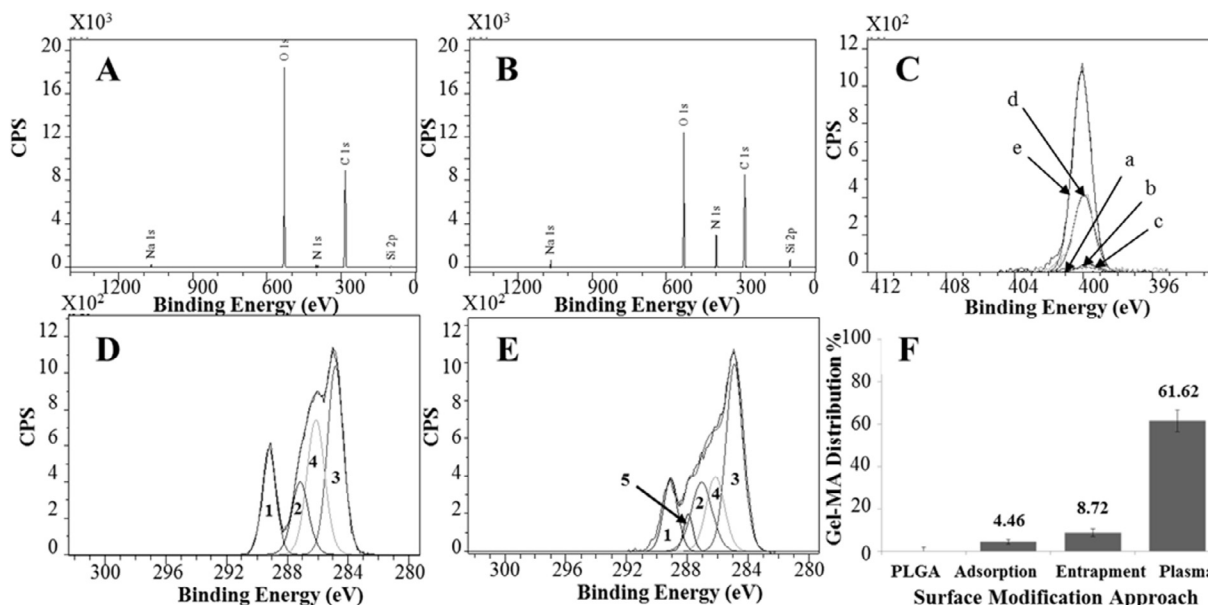


Fig. 2. X-ray Spectroscopy (XPS) wide scan shows peak spectra of PDLGA microspheres before (A) and after surface modification with gel-MA using plasma modification approach (B) where (N1s) peak from gel-MA peptide bonds can be seen at binding energy of ~ 400 eV. A comparison between N1s peaks obtained from PDLGA microspheres (C) shows the relative change in the intensity of N1s before (a), and after modification with gel-MA using adsorption (b), entrapment (c), and plasma approaches (d) compared to pure gel-MA (e). High resolution scan of C1s region from PDLGA microspheres (D) shows three distinct peaks of PDLGA (1,2, and 3) and a fourth peak of PVA (4), while a further gel-MA related peak (5) can be seen on plasma modified microspheres (E). Relative (N1s) peak area ratio of samples was normalised against (N1s) peak area of pure gel-MA films and used to quantify the concentration of gel-MA on the surface of gel-MA modified PDLGA microspheres where a significant increase in the (N1s) area ratio can be seen on plasma and entrapment modified microspheres compared to adsorption and non-modified PDLGA microspheres (F).

maximum values (FWHM) of ± 0.2 eV (Supplementary Data 3). As gel-MA peaks (3) and (6) overlap PDLGA and PVA peaks (3) and (4) respectively, the presence of the gel-MA peak (5-arrow) on C1s curves may indicate the presence of gel-MA on the surface of plasma modified microspheres (Fig. 2).

High resolution scan of the N1s region showed the increase in the intensity of the peak at ~ 398 nm on gel-MA modified PDLGA microspheres compared to non-modified PDLGA microspheres (Supplementary Data 4). Different C1s peaks were assigned to relevant chemical states of carbon atoms found on the different amino acids found in gel-MA molecules (Supplementary Data 5). However, as most of gel-MA assigned C1s peaks overlap those of PDLGA, the surface area of N1s peaks was used instead of that of C1s peaks to quantify gel-MA density on the surface.

Negative ion spectra of gel-MA modified PDLGA microspheres have shown specific ion peaks at an ion mass of (26 m/z) and (42 m/z) which have been assigned to gel-MA peptide bond ions (CN^-) and (CNO^-), while specific PDLGA ion peaks have been obtained from non-modified PDLGA microspheres at (71 m/z) and assigned to ($\text{C}_3\text{H}_3\text{O}_2^-$) (Fig. 3-I). While non-modified PDLGA microspheres have not shown a detectable number of gel-MA ions, an increase in the number of gel-MA specific ions has been observed on the surface of PDLGA microspheres following gel-MA modification using plasma compared to less increase with entrapment and adsorption approaches. Images of gel-MA specific ion distribution on the surface of PDLGA microspheres have shown a homogenous distribution of gel-MA ions on the surface of PDLGA microspheres modified using plasma approach. On the other hand, PDLGA microspheres modified using entrapment or adsorption approaches have shown a heterogeneous -or island like-distribution of gel-MA specific ions with limited surface coverage (Table 2) compared to no observable distribution on the surface of non-modified PDLGA microspheres (Fig. 3-II). Other figures obtained from the optimisation process of both plasma and entrapment approaches can be found in the appendix (Supplementary Data 6, 7, and 8).

Table 2

Average surface coverage of gel-MA molecules on the surface of PDLGA microspheres following modification with different surface modification approaches. Values are obtained from ToF SIMs image data analysis and correspond to the average coverage with gel-MA on the surface of 9 different microspheres \pm standard deviation.

Surface Modification Approach	PLGA	Adsorption	Entrapment	Plasma
Gel-MA Surface Coverage%	0	15.7 \pm 6.1	2.3 \pm 0.8	89.3 \pm 21.3

3.2. Imaging of modified PDLGA microspheres

Surface structure of PDLGA microspheres before and after modification with gel-MA was investigated using SEM. Microspheres modified using both adsorption and plasma modification approaches were spherical with smooth surfaces similar to the non-modified PDLGA microspheres, while microspheres modified using the entrapment approach had a rougher surface with some evidence of porosity (Fig. 4-top row). Further probing of the structural changes using (FIB-SEM) showed no clear changes to the internal structures of PDLGA microspheres before and after surface modification using both adsorption and plasma approaches (Fig. 4-magnified top). On the other hand, a porous internal structure was observed inside microspheres modified using the surface entrapment approach (Fig. 4-magnified top).

Further investigation of the internal structures using cross-sectioned PDLGA microspheres revealed a highly porous internal structure underneath the surface following modification with the entrapment approach. The microspheres modified using adsorption and plasma approaches did not show any observable changes to internal structure compared to non-modified PDLGA microspheres (Fig. 4-middle row).

Fluorescent microscope images showed a fluorescent layer on the outer surface of the cross-sectioned PDLGA microspheres modified using both plasma and entrapment approaches, while a neg-

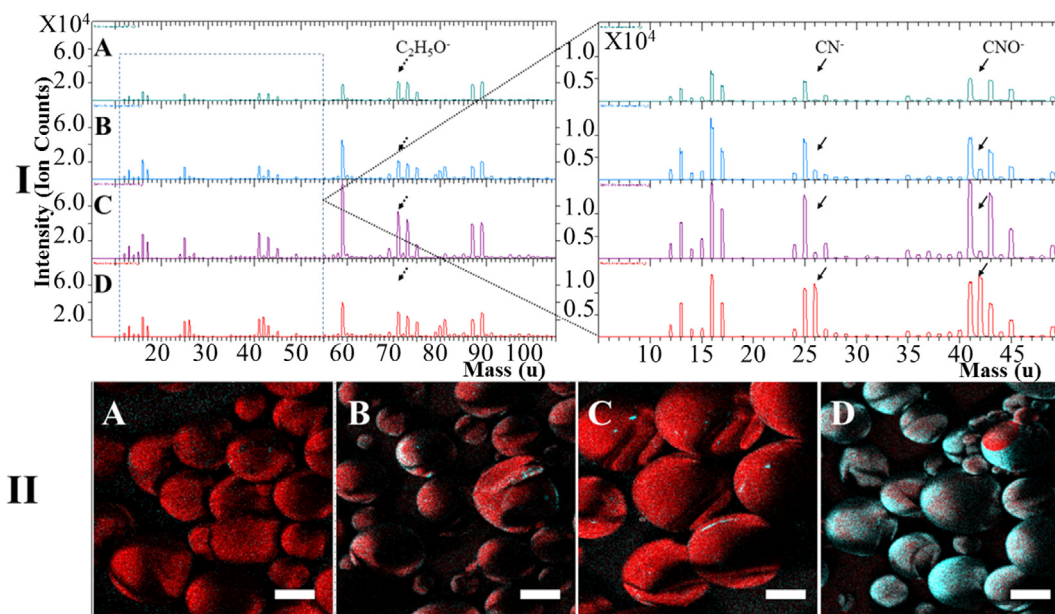


Fig. 3. Time of Flight Secondary Ion Mass Spectrometry (ToF SIMS) ion peak spectra of PDLGA microspheres before (A) and after modification with gel-MA using adsorption (B), entrapment (C), and plasma treatment approaches (D). Increasing ion counts of peptide bond related peaks (CN⁻) and (CNO⁻) can be seen on gel-MA modified microspheres (solid arrows) using adsorption, entrapment and plasma treatment approaches compared to (C₂H₅O⁻) PDLGA ions (dotted arrow) respectively (I). Ion mapping images of (C₂H₅O⁻) and (CNO⁻) ions related to PDLGA (red) and gel-MA (cyan) respectively, show homogenous distribution of gel-MA ions on the surface of plasma modified microspheres (D) compared to heterogeneous – or spotty – distribution on the surface of entrapment (C) and adsorption (B) modified microspheres (II). On the other hand, no clear sign of gel-MA specific ions can be seen on non-modified PDLGA microspheres (A) (Scale bar = 50 μm).

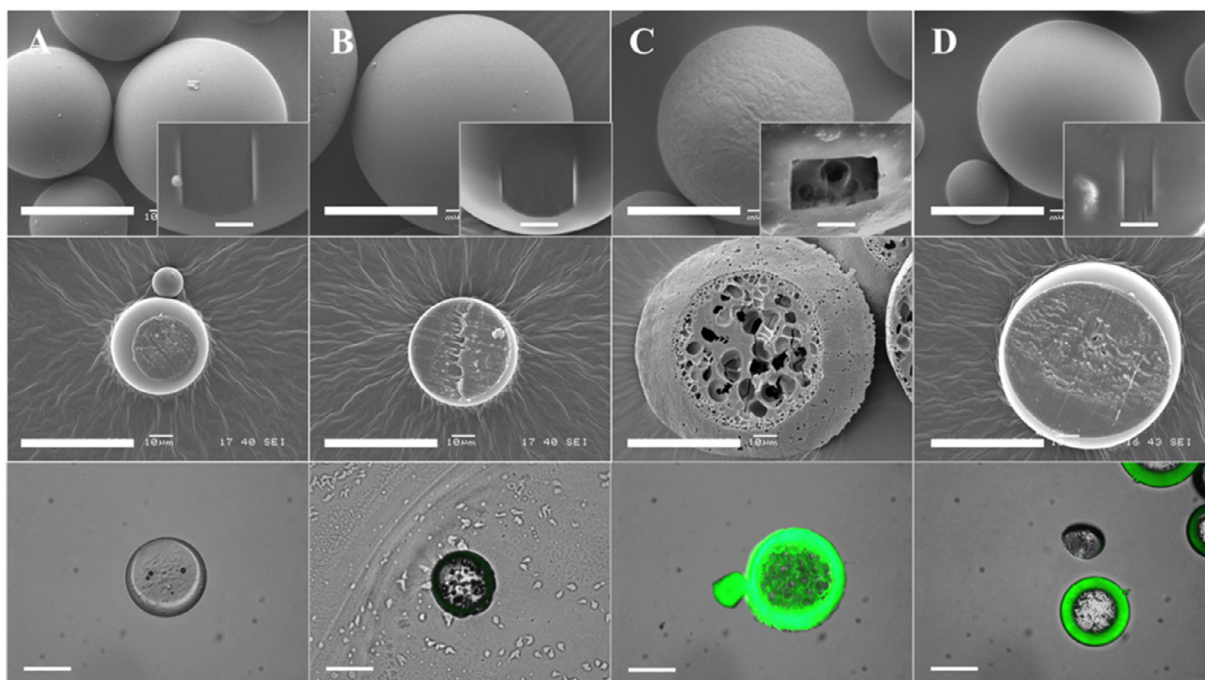


Fig. 4. Scanning electron microscopy (SEM) images of PDLGA microspheres before (A) and after modification with gel-MA using surface adsorption (B), surface entrapment (C), and oxygen plasma treatment approaches (D). Morphological changes can be observed on the surface of entrapment modified microspheres compared to a smooth round surface of all of the other conditions (Top row). Further inspection of the morphological changes using a focused ion beam (FIB SEM) has revealed a porous structure underneath the surface of entrapment modified microspheres compared to solid fill inside all of the other conditions (magnified figures). Images of cross-sectioned PDLGA microspheres (middle row) obtained with SEM show a porous structure with a decreasing pore size closer to the surface of entrapment modified microspheres while no observable changes have been noticed using other approaches. Fluorescent images of cross-sectioned microspheres (bottom row) show the depth of surface modification effect using Fit-C labelled gel-MA. While no clear presence of gel-MA on the non-modified (A) and adsorption modified PDLGA microspheres (B), a considerable amount of Fit-C labelled gel-MA can be observed inside entrapment modified microspheres (C) compared to more surface specific localisation of fit-C labelled gel-MA on plasma modified microspheres (D) (Scale bar: all figures = 50 μm, magnified figures = 5 μm).

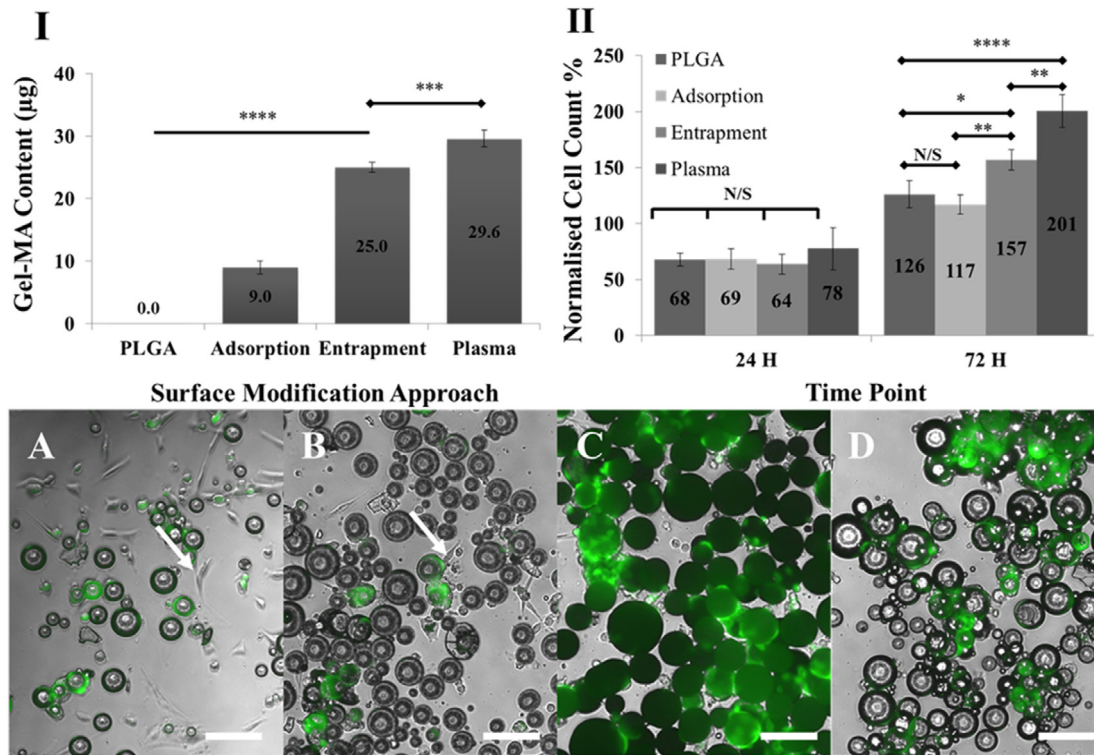


Fig. 5. Quantification of gel-MA content on the surface of 100 mg of PDLGA microspheres using BCA assay shows significant increase in gel-MA amount following modification with adsorption, entrapment, and plasma modification approaches respectively (I). Quantification of metabolic activity of immortalised human mesenchymal stem cells cultured on non-modified and gel-MA modified PDLGA microspheres using Presto Blue assay shows significant increase in cell proliferation after 72 h of cell culture. All measurements are normalised to the activity measurements of cells cultured on a tissue culture treated plates over the same period of time. While no significant changes in cell metabolic activity can be noticed after 24 h; a significant increase in cell metabolic activity can be observed on plasma and entrapment modified PDLGA microspheres compared to adsorption modified PDLGA microspheres. Images taken after 72 h of cells cultured on microspheres show weak attachment of cells on both non-modified (A) and adsorption modified (B) PDLGA microspheres where cells tend to grow on the substrate as well (arrows) while good cell attachment can be seen on both entrapment (C) and plasma modified PDLGA microspheres (D). (n = 3, P Value is (****) <math>P < 0.0001</math>, (***) <math>P < 0.001</math>, (**) <math>P < 0.01</math>, (*) <math>P < 0.05</math>, and (ns) $P > 0.05$) (Scale Bar = 200 µm).

ligible fluorescent layer not detectable against the background fluorescence could be seen on the adsorption modified compared to non-modified PDLGA microspheres where no observable fluorescent layer could be seen on the surface (Fig. 4-bottom row). Moreover, a fluorescent deposition could also be seen inside the entrapment modified microspheres compared to no fluorescent deposition inside plasma or adsorption modified microspheres (Supplementary Data 9).

3.3. Cell viability and proliferation

BCA assay data normalised against readings from PDLGA microspheres have shown an increase in the amount of gel-MA on the surface of plasma and entrapment modified PDLGA microspheres compared to adsorption modified and non-modified PDLGA microspheres respectively ($P < 0.001$) (Fig. 5-I). Immortalised hMSCs metabolic activity was measured using a Presto-Blue assay to study the effect of different surface modification approaches on cell adhesion and proliferation on the surfaces of PDLGA microspheres over time. All Presto-Blue assay readings were normalised against readings from cells cultured on tissue culture plates. A cell survival study using a Presto-Blue assay over 72 h has shown that the immortalised hMSCs proliferation on gel-MA modified PDLGA microspheres has not been compromised compared to non-modified PDLGA microspheres. Presto-Blue assay readings after 24 h of cell culture did not show any significant change between different samples, while readings after 72 h showed a significant increase in both plasma and entrapment modified microspheres compared to adsorption modified and non-modified microspheres

(Fig. 5-II). Microscope images showed a relatively higher number of cells growing on the surface of entrapment and plasma modified microspheres compared to both adsorption and non-modified microspheres. Cells were exclusively localised on the surface of plasma and entrapment modified microspheres, while they seemed to be non-specifically presented onto the surface of adsorption modified and non-modified PDLGA microspheres as well as on the cell culture substrate (Fig. 5-A, B, C, and D).

4. Discussion

PDLGA microspheres offer a potential injectable cell delivery system with controllable growth factor release for different tissue engineering applications [4–7]. However, as with other polyester polymers, PDLGA have limited functional groups on the surface which limit grafting of tissue specific bioactive molecules to control cell behaviour. This study has proposed modifying the surface of PDLGA microspheres with gel-MA to enable grafting of further tissue specific biomolecules through the methacrylate group without compromising surface biocompatibility. Gel-MA molecules have the potential to form hydrogels with tunable elastic properties upon photo-crosslinking. Unlike gelatine, gel-MA molecules contain the methacrylate vinyl groups which can be used to further graft gel-MA or other bioactive molecules containing vinyl groups [43,50]. Moreover, non-crosslinked gelatine hydrogels dissolve over 24 h of incubation at 37 °C while gel-MA hydrogels prepared using UV photo-crosslinking offer controllable elastic properties and do not dissolve over 24 h of incubation at 37 °C (Supplementary Data 11).

Terminally uncapped P_DLGA was used in this study as it was reported to have higher surface energy due to the presence of carboxyl end groups compared to the lower surface energy of terminally capped P_DLGA which has a methylated carboxyl end groups [51–53]. Surface entrapment was proposed in this study to evaluate the potential incorporation of gel-MA molecules within P_DLGA polymer network on the surface as previously suggested [54]. Likewise, oxygen plasma treatment was proposed in this study to improve the available oxygen species on the surface of P_DLGA microspheres and immobilise gel-MA molecules.

This study has demonstrated that P_DLGA microspheres modified using oxygen plasma treatment had higher density of gel-MA molecules on the surface compared to those modified using entrapment or adsorption approaches respectively. Since adsorption of gel-MA molecules would mainly depend on the available surface energy, the low density of deposited gel-MA molecules on the surface of adsorption modified microspheres indicate a low surface energy on the surface of uncapped P_DLGA microspheres.

On the other hand, as plasma induced oxygen species have been suggested to include peroxide, hydroxyl, and carboxylic groups [55–57]; the increase in gel-MA content following oxygen plasma modification could be attributed to these oxygen species as well. Peroxide groups have been reported to be chemically unstable and could potentially form reactive radicals which may attack electron rich vinyl groups and immobilise gel-MA molecules through covalent bonding. Unstable peroxide groups may also transform into hydroxyl or carboxyl groups which could increase surface energy and further enhance the deposition of gel-MA molecules on the surface [58].

The structural changes seen on the surface as well as inside the entrapment modified microspheres could be attributed to the partial restructuring of the P_DLGA network during the entrapment process. The prolonged exposure to solvent molecules enables deeper penetration and higher densities of modifying molecules to entrap on the polymer surface as previously suggested in literature [59]. This study has shown that prolonged exposure to the solvent can also induce porous structures underneath the surface.

Fluorescent microscope images have shown a fluorescent layer on the surface of entrapment and plasma modified microspheres only which indicates successful modification with fit-C gel-MA. No detectable fluorescent layer was observed on the adsorption modified microspheres which indicates little deposition of fit-C gel-MA on the surface. Moreover, the detection of fit-C gel-MA on the surface along with the porous structures inside the entrapment modified microspheres may indicate a modification not just on the surface, but deep inside the microspheres as well.

The density of gel-MA on the surface of entrapment modified microspheres was comparable to that on the adsorption modified microspheres as shown with XPS and ToF SIMs data. However, the overall content of gel-MA quantified with BCA assay on the entrapment modified microspheres was higher than that detected on the surface of adsorption modified microspheres. This may indicate that entrapment approach has increased the available surface area. As the BCA assay measures the total gel-MA content of microspheres in contact with the BCA mixture, the high surface area of the porous entrapment modified microspheres may potentially contain higher amounts of gel-MA. Therefore, the BCA mixture would react with a higher number of gel-MA molecules on the surface of entrapment modified microspheres compared to the relatively less gel-MA molecules found on the lower surface area of non-porous adsorption modified microspheres.

The low density of gel-MA molecules detected by XPS and ToF SIMs on the surface of entrapment modified microspheres may be attributed to the fact that both techniques are surface specific. Both ToF SIMs and XPS can probe few nanometres into the

analysed surface [60] while fluorescent microscopy and BCA can detect the whole surface including the internal surfaces of the porous microspheres. This may explain the relatively higher gel-MA density detected with BCA and fluorescent microscopy on the surface of entrapment modified microspheres compared to the non-porous adsorption modified microspheres.

The significant increase in the metabolic activity of immortalised hMSCs cultured on the plasma and entrapment modified microspheres compared to the non-modified and adsorption modified microspheres can be attributed to the high density of gel-MA on the surface. On the other hand, the significant increase in the metabolic activity with the entrapment modified compared to the adsorption modified microspheres can be attributed to the high surface area of the porous entrapment modified microspheres which seems to introduce higher gel-MA domains for cell adhesion. This may also explain the localised growth of the immortalised hMSCs on the surface of plasma and entrapment modified microspheres compared to the non-specific growth on the cell culture substrates as well as on the surface of adsorption modified and non-modified microspheres where gel-MA content is negligible.

Finally, fit-C labelled gel-MA has been proposed to evaluate the potential grafting of crosslinkable macromolecules on the surface of gel-MA modified P_DLGA microspheres. ToF SIMs images of gelatine modified microspheres prepared using the plasma treatment approach showed homogenous distribution of gelatine molecules on the surface of P_DLGA microspheres (Supplementary Data 12). Grafting fit-C labelled gel-MA molecules on the surface of non-modified, gelatine, and gel-MA modified P_DLGA microspheres showed a fluorescent layer related to the fit-C gel-MA molecules on the surface of gel-MA modified microspheres only. In comparison, no detectable fluorescent layer could be seen on the surface of non-modified and gelatine modified P_DLGA microspheres indicating a negligible deposition of fit-C gel-MA. This can be explained with the presence of the methacrylate vinyl groups on the surface of gel-MA modified P_DLGA microspheres which can form covalent bonds with fit-C gel-MA molecules upon photo-grafting. In comparison, as both gelatine molecules and P_DLGA chains do not offer the crosslinkable vinyl groups, they would not enable grafting of fit-C gel-MA molecules (Supplementary Data 13).

5. Conclusion

This study has evaluated three different approaches to modify the surface of P_DLGA microspheres with gel-MA: surface adsorption, surface entrapment, and oxygen plasma treatment. Different surface analytical and quantitative assays have shown that the oxygen plasma treatment approach produced the highest density of gel-MA on the surface of modified P_DLGA microspheres followed by entrapment and adsorption approaches respectively. Moreover, different microscopy images have shown porous structures inside P_DLGA microspheres modified using surface entrapment approach. This may suggest a deep penetration of gel-MA molecules underneath the surface of microspheres and a possible restructuring of the polymer microsphere during the process. On the other hand, the significant increase in cell metabolic activity of immortalised hMSCs cultured for 72 h on the surface of plasma and entrapment modified microspheres compared to adsorption and non-modified microspheres can be related to the gel-MA content on the surface. This in turn would suggest no adverse effects of the proposed surface modification approaches on the biocompatibility of the system.

The modified microspheres could be utilised to induce further grafting of cross-linkable biomolecules to obtain tissue specific functionalities. Moreover, as gel-MA has been reported to produce hydrogels with tissue specific mechanical properties, gel-MA

modified PLGA microspheres could potentially be further grafted with gel-MA molecules to produce hydrogel coated microspheres with tissue specific elastic properties.

Acknowledgments

Authors would gratefully acknowledge regenerative medicine platform hub “acellular approaches for therapeutic delivery” (MR/K026682/1) for support. Authors would also like to thank Dr Christopher Parmenter and Dr Emily Smith for their help on, FIB SEM and XPS, and Dr Ahmed Aied for helpful discussions. L J White was funded by a Marie Curie International Outgoing Fellowship under REA grant agreement no. 624841.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.actbio.2017.01.042>.

References

- [1] M. Sokolsky-Papkov, K. Agashi, A. Olaye, K. Shakesheff, A.J. Domb, Polymer carriers for drug delivery in tissue engineering, *Adv. Drug Deliv. Rev.* 59 (4–5) (2007) 187–206.
- [2] J.M. Anderson, M.S. Shive, Biodegradation and biocompatibility of PLA and PLGA microspheres, *Adv. Drug Deliv. Rev.* 64 (Supplement(0)) (2012) 72–82.
- [3] O. Qutachi, J.R. Vetsch, D. Gill, H. Cox, D.J. Scurr, S. Hofmann, R. Müller, R.A. Quirk, K.M. Shakesheff, C.V. Rahman, Injectable and porous PLGA microspheres that form highly porous scaffolds at body temperature, *Acta Biomater.* 10 (12) (2014) 5090–5098.
- [4] H.K. Makadia, S.J. Siegel, Poly lactic-co-glycolic acid (PLGA) as biodegradable controlled drug delivery carrier, *Polymers* 3 (3) (2011) 1377–1397.
- [5] V. Mouriño, A.R. Boccacini, Bone tissue engineering therapeutics: controlled drug delivery in three-dimensional scaffolds, *J. R. Soc. Interface* 7 (43) (2010) 209–227.
- [6] C.V. Rahman, D. Ben-David, A. Dhillon, G. Kuhn, T.W. Gould, R. Müller, F.R. Rose, K.M. Shakesheff, E. Livne, Controlled release of BMP-2 from a sintered polymer scaffold enhances bone repair in a mouse calvarial defect model, *J. Tissue Eng. Regen. Med.* (2012).
- [7] L.J. White, G.T.S. Kirby, H.C. Cox, R. Qodratnama, O. Qutachi, F.R.A.J. Rose, K.M. Shakesheff, Accelerating protein release from microparticles for regenerative medicine applications, *Mater. Sci. Eng. C* 33 (5) (2013) 2578–2583.
- [8] C.V. Rahman, A. Saeed, L.J. White, T.W.A. Gould, G.T.S. Kirby, M.J. Sawkins, C. Alexander, F.R.A.J. Rose, K.M. Shakesheff, Chemistry of polymer and ceramic-based injectable scaffolds and their applications in regenerative medicine, *Chem. Mater.* 24 (5) (2011) 781–795.
- [9] Y.-P. Jiao, F.-Z. Cui, Surface modification of polyester biomaterials for tissue engineering, *Biomed. Mater.* 2 (4) (2007) R24.
- [10] X. Liu, J.M. Holzwarth, P.X. Ma, Functionalized synthetic biodegradable polymer scaffolds for tissue engineering, *Macromol. Biosci.* 12 (7) (2012) 911–919.
- [11] M. Tallawi, E. Rosellini, N. Barbani, M.G. Cascone, R. Rai, G. Saint-Pierre, A.R. Boccacini, Strategies for the chemical and biological functionalization of scaffolds for cardiac tissue engineering: a review, *J. R. Soc. Interface* 12 (108) (2015).
- [12] G. Chen, T. Ushida, T. Tateishi, Hybrid biomaterials for tissue engineering: a preparative method for PLA or PLGA–collagen hybrid sponges, *Adv. Mater.* 12 (6) (2000) 455–457.
- [13] S.M. Cannizzaro, R.F. Padera, R. Langer, R.A. Rogers, F.E. Black, M.C. Davies, S.J.B. Tendler, K.M. Shakesheff, A novel biotinylated degradable polymer for cell-interactive applications, *Biotechnol. Bioeng.* 58 (5) (1998) 529–535.
- [14] J.M. Goddard, J.H. Hotchkiss, Polymer surface modification for the attachment of bioactive compounds, *Prog. Polym. Sci.* 32 (7) (2007) 698–725.
- [15] H. Zhu, J. Ji, J. Shen, Biomacromolecules electrostatic self-assembly on 3-dimensional tissue engineering scaffold, *Biomacromolecules* 5 (5) (2004) 1933–1939.
- [16] T.I. Croll, A.J. O'Connor, G.W. Stevens, J.J. Cooper-White, A blank slate? Layer-by-layer deposition of hyaluronic acid and chitosan onto various surfaces, *Biomacromolecules* 7 (5) (2006) 1610–1622.
- [17] E. Kharlampieva, V. Kozlovskaya, S.A. Sukhishvili, Layer-by-layer hydrogen-bonded polymer films: from fundamentals to applications, *Adv. Mater.* 21 (30) (2009) 3053–3065.
- [18] Y. Zhu, C. Gao, T. He, X. Liu, J. Shen, Layer-by-layer assembly to modify poly(L-lactic acid) surface toward improving its cytocompatibility to human endothelial cells, *Biomacromolecules* 4 (2) (2003) 446–452.
- [19] J. Gao, L. Niklason, R. Langer, Surface hydrolysis of poly(glycolic acid) meshes increases the seeding density of vascular smooth muscle cells, *J. Biomed. Mater. Res.* 42 (3) (1998) 417–424.
- [20] G. Khang, J.-H. Choe, J.M. Rhee, H.B. Lee, Interaction of different types of cells on physicochemically treated poly(L-lactide-co-glycolide) surfaces, *J. Appl. Polym. Sci.* 85 (6) (2002) 1253–1262.
- [21] T.I. Croll, A.J. O'Connor, G.W. Stevens, J.J. Cooper-White, Controllable surface modification of poly(lactic-co-glycolic acid) (PLGA) by hydrolysis or aminolysis I: physical, chemical, and theoretical aspects, *Biomacromolecules* 5 (2) (2004) 463–473.
- [22] H. Li, Y. Xia, J. Wu, Q. He, X. Zhou, G. Lu, L. Shang, F. Boey, S.S. Venkatraman, H. Zhang, Surface modification of smooth poly(L-lactide) films for gelatin immobilization, *ACS Appl. Mater. Interfaces* 4 (2) (2012) 687–693.
- [23] R.A. Quirk, M.C. Davies, S.J.B. Tendler, K.M. Shakesheff, Surface engineering of poly(lactic acid) by entrapment of modifying species, *Macromolecules* 33 (2) (2000) 258–260.
- [24] Q. Hou, R. Freeman, L.D.K. Buttery, K.M. Shakesheff, Novel surface entrapment process for the incorporation of bioactive molecules within preformed alginate fibers, *Biomacromolecules* 6 (2) (2005) 734–740.
- [25] H. Guo, M. Ulbricht, The effects of (macro)molecular structure on hydrophilic surface modification of polypropylene membranes via entrapment, *J. Colloid Interface Sci.* 350 (1) (2010) 99–109.
- [26] H. Guo, Polymer Surface Modification via Noncovalent Binding of Functional (Macro) Molecules: Investigation of Entrapment Strategy and Mechanism, Universität Duisburg-Essen, Fakultät für Chemie Technische Chemie, 2010.
- [27] E. Berndt, S. Behnke, M. Ulbricht, Influence of alkyl chain length and molecular weight on the surface functionalization via adsorption/entrapment with biocidal cationic block copolymers, *Eur. Polymer J.* 47 (12) (2011) 2379–2390.
- [28] N.P. Desai, J.A. Hubbell, Surface physical interpenetrating networks of poly(ethylene terephthalate) and poly(ethylene oxide) with biomedical applications, *Macromolecules* 25 (1) (1992) 226–232.
- [29] Z.X. Meng, Q.T. Zeng, Z.Z. Sun, X.X. Xu, Y.S. Wang, W. Zheng, Y.F. Zheng, Immobilizing natural macromolecule on PLGA electrospun nanofiber with surface entrapment and entrapment-graft techniques, *Colloids Surf. B* 94 (2012) 44–50.
- [30] Z. Jin, W. Feng, S. Zhu, H. Sheardown, J.L. Brash, Protein-resistant polyurethane via surface-initiated atom transfer radical polymerization of oligo(ethylene glycol) methacrylate, *J. Biomed. Mater. Res. A* 91A (4) (2009) 1189–1201.
- [31] M. Källrot, U. Edlund, A.C. Albertsson, Surface functionalization of degradable polymers by covalent grafting, *Biomaterials* 27 (9) (2006) 1788–1796.
- [32] Z. Ma, C. Gao, J. Yuan, J. Ji, Y. Gong, J. Shen, Surface modification of poly-L-lactide by photografting of hydrophilic polymers towards improving its hydrophilicity, *J. Appl. Polym. Sci.* 85 (10) (2002) 2163–2171.
- [33] J.-P. Deng, W.-T. Yang, B. Rånby, Auto-initiating performance of styrene on surface photografting polymerization, *Macromol. Rapid Commun.* 22 (7) (2001) 535–538.
- [34] D. Leonhardt, C. Muratore, S.G. Walton, D.D. Blackwell, R.F. Fernsler, R.A. Meger, Generation of electron-beam produced plasmas and applications to surface modification, *Surf. Coat. Technol.* 177–178 (2004) 682–687.
- [35] T. Desmet, R. Morent, N.D. Geyter, C. Leys, E. Schacht, P. Dubruel, Nonthermal plasma technology as a versatile strategy for polymeric biomaterials surface modification: a review, *Biomacromolecules* 10 (9) (2009) 2351–2378.
- [36] M. Zelzer, R. Majani, J.W. Bradley, F.R.A.J. Rose, M.C. Davies, M.R. Alexander, Investigation of cell–surface interactions using chemical gradients formed from plasma polymers, *Biomaterials* 29 (2) (2008) 172–184.
- [37] P.M. López-Pérez, R.M.P. da Silva, R.A. Sousa, I. Pashkuleva, R.L. Reis, Plasma-induced polymerization as a tool for surface functionalization of polymer scaffolds for bone tissue engineering: An in vitro study, *Acta Biomater.* 6 (9) (2010) 3704–3712.
- [38] M. Santoro, A.M. Tatara, A.G. Mikos, Gelatin carriers for drug and cell delivery in tissue engineering, *J. Control. Release* 190 (2014) 210–218.
- [39] M.H. Zaman, Understanding the molecular basis for differential binding of integrins to collagen and gelatin, *Biophys. J.* 92 (2) (2007) L17–L19.
- [40] M.C. Gómez-Guillén, B. Giménez, M.E. López-Caballero, M.P. Montero, Functional and bioactive properties of collagen and gelatin from alternative sources: a review, *Food Hydrocolloids* 25 (8) (2011) 1813–1827.
- [41] J.W. Nichol, S.T. Koshy, H. Bae, C.M. Hwang, S. Yamanlar, A. Khademhosseini, Cell-laden microengineered gelatin methacrylate hydrogels, *Biomaterials* 31 (21) (2010) 5536–5544.
- [42] K. Yue, G. Trujillo-de Santiago, M.M. Alvarez, A. Tamayo, N. Annabi, A. Khademhosseini, Synthesis, properties, and biomedical applications of gelatin methacryloyl (GelMA) hydrogels, *Biomaterials* 73 (2015) 254–271.
- [43] J. Silva-Correia, A. Gloria, M.B. Oliveira, J.F. Mano, J.M. Oliveira, L. Ambrosio, R.L. Reis, Rheological and mechanical properties of acellular and cell-laden methacrylated gellan gum hydrogels, *J. Biomed. Mater. Res. A* 101 (12) (2013) 3438–3446.
- [44] J. Silva-Correia, J.M. Oliveira, S.G. Caridade, J.T. Oliveira, R.A. Sousa, J.F. Mano, R. L. Reis, Gellan gum-based hydrogels for intervertebral disc tissue-engineering applications, *J. Tissue Eng. Regen. Med.* 5 (6) (2011) e97–e107.
- [45] A. Barbeta, M. Dentini, E.M. Zannoni, M.E. De Stefano, Tailoring the porosity and morphology of gelatin-methacrylate polyHIPE scaffolds for tissue engineering applications, *Langmuir* 21 (26) (2005) 12333–12341.
- [46] S. Van Vlierbergh, E. Vanderleyden, V. Boterberg, P. Dubruel, Gelatin functionalization of biomaterial surfaces: strategies for immobilization and visualization, *Polymers* 3 (1) (2011) 114.
- [47] Y. Zhu, C. Gao, J. Guan, J. Shen, Promoting the cytocompatibility of polyurethane scaffolds via surface photo-grafting polymerization of acrylamide, *J. Mater. Sci. – Mater. Med.* 15 (3) (2004) 283–289.

- [48] Y. Kosaka, N. Kobayashi, T. Okitsu, N. Tanaka, T. Nakanishi, Immortalized human mesenchymal stem cell lines for tissue engineering, *ASAIO J.* 49 (2) (2003) 220.
- [49] K.M. Shakesheff, C. Evora, I. Soriano, R. Langer, The adsorption of poly(vinyl alcohol) to biodegradable microparticles studied by X-ray photoelectron spectroscopy (XPS), *J. Colloid Interface Sci.* 185 (2) (1997) 538–547.
- [50] M.G. Ondeck, A.J. Engler, Mechanical characterization of a dynamic and tunable methacrylated hyaluronic acid hydrogel, *J. Biomech. Eng.* 138 (2) (2016). 021003–021003.
- [51] M. Müller, J. Vörös, G. Csúcs, E. Walter, G. Danuser, H.P. Merkle, N.D. Spencer, M. Textor, Surface modification of PLGA microspheres, *J. Biomed. Mater. Res. A* 66A (1) (2003) 55–61.
- [52] D.P. Go, J.A. Palmer, S.L. Gras, A.J. O'Connor, Coating and release of an anti-inflammatory hormone from PLGA microspheres for tissue engineering, *J. Biomed. Mater. Res. A* 100A (2) (2012) 507–517.
- [53] K.W. Chun, H.S. Yoo, J.J. Yoon, T.G. Park, Biodegradable PLGA microcarriers for injectable delivery of chondrocytes: effect of surface modification on cell attachment and function, *Biotechnol. Prog.* 20 (6) (2004) 1797–1801.
- [54] H. Guo, M. Ulbricht, Preparation of thermo-responsive polypropylene membranes via surface entrapment of poly(N-isopropylacrylamide)-containing macromolecules, *J. Membr. Sci.* 372 (1–2) (2011) 331–339.
- [55] Y. Wan, X. Qu, J. Lu, C. Zhu, L. Wan, J. Yang, J. Bei, S. Wang, Characterization of surface property of poly(lactide-co-glycolide) after oxygen plasma treatment, *Biomaterials* 25 (19) (2004) 4777–4783.
- [56] H. Shen, X. Hu, F. Yang, J. Bei, S. Wang, Combining oxygen plasma treatment with anchorage of cationized gelatin for enhancing cell affinity of poly(lactide-co-glycolide), *Biomaterials* 28 (29) (2007) 4219–4230.
- [57] G.H. Ryu, W.-S. Yang, H.-W. Roh, I.-S. Lee, J.K. Kim, G.H. Lee, D.H. Lee, B.J. Park, M.S. Lee, J.-C. Park, Plasma surface modification of poly (D, L-lactic-co-glycolic acid) (65/35) film for tissue engineering, *Surf. Coat. Technol.* 193 (1–3) (2005) 60–64.
- [58] N. Inagaki, *Plasma Surface Modification and Plasma Polymerization*, CRC Press, 1996.
- [59] R.A. Quirk, D. Briggs, M.C. Davies, S.J.B. Tendler, K.M. Shakesheff, Characterization of the spatial distributions of entrapped polymers following the surface engineering of poly(lactic acid), *Surf. Interface Anal.* 31 (1) (2001) 46.
- [60] J.B. Gilbert, M.F. Rubner, R.E. Cohen, Depth-profiling X-ray photoelectron spectroscopy (XPS) analysis of interlayer diffusion in polyelectrolyte multilayers, *Proc. Natl. Acad. Sci. U.S.A.* 110 (17) (2013) 6651–6656.