Abstract

# **Signaling Pathways Involved in Auditory Hair Cells Development**

Shahrokh Khoshsirat<sup>1</sup>, Navid Ahmady Roozbahany<sup>1</sup>, Somayeh Niknazar<sup>1\*</sup>

1. Hearing Disorders Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

#### Article Info

Received: Jun 2017 Accepted: July 2017 Publish: 20 Aug 2017

**Corresponding Author:** Somayeh Niknazar

**Email:** niknazar@sbmu.ac.ir

#### **Keywords:**

Signaling pathways, Auditory Hair Cells, Regeneration. Auditory hair cells (HCs) cannot be spontaneously regenerated or replaced in mammalian damaged cochlea which leads to permanent deafness. On the other hand, regenerative ability of HCs in lower vertebrates such as birds and amphibians causes that researchers investigate underlying mechanisms and pathways which can possibly induce mammalian cochlear HCs regeneration and hearing recovery. Signaling cascades of HCs regeneration in lower vertebrate can be considered as the potential therapeutic option for the hearing loss in human. This paper reviews current knowledge about the main signaling pathways involved in HCs development in the mammalian cochlea.

**Cite this article that:** Khoshsirat Sh, Ahmady Roozbahany N, Niknazar S. Signaling Pathways Involved in Auditory Hair Cells Development. Journal of Otorhinolaryngology & Facial Plastic Surgery. 2017; 2017; e3.

#### Introduction

Spiral organ of the cochlea, the organ of Corti is composed of sensory HCs and accessory supporting cells (SCs) (1). Auditory HCs are highly specialized cells that transduce sound vibrations into neural signals (2). Unlike nonmammalian vertebrates, mammalian HCs have not regenerative capacity which leads to irreversible hearing loss (3, 4). Studies of birds' auditory system show that damage to HCs can be regenerated and hearing function restored (4). During regeneration, SCs can be differentiated or transdifferentiated into HCs through a combination of mitotic and nonmitotic mechanisms. Proliferation and transdifferentiation process of SCs is started by the signals of dying HCs (5). In mammals, this mechanism only occurs during inner ear development in fetal period (6). Although it is recently reported that auditory HCs regeneration is happened in the mice cochlea during a short time of postnatal period (7), but definite mechanism of HCs regeneration is not clear yet. Studies have shown that various cascades including Atoh1, Wnt, Shh, Fgf and Notch signaling pathways are critical during HCs development and regeneration (8-12). In this review we provide an overview of the main mechanisms and signaling cascades that lead to auditory HCs regeneration.

#### Atoh1

basic Atoh1 (atonal helix-loop-helix transcription factor 1) or Math1 is the first gene that was discovered during inner ear development and involved in HCs differentiation (13). Atoh1 expression was observed in the basal region of the cochlea at embryonic day 14.5 (E14.5) which responsible for HCs differentiation (one row of inner HCs and three rows of outer HCs) at E15.5. Previous studies provide evidence indicating that Atoh1 positive cells can generate SCs in addition to HCs. In neonatal cochlea, SCs can convert to HCs in response to Atoh1 overexpression (14-16). However, Atoh1 ability in HCs generation from surrounding SCs is extremely decreased in adult or uninjured cochlea (17, 18). It has been reported that adenoviral-mediated overexpression of Atoh1 in kanamycin-induced damaged cochlea leads to HCs regeneration and improve deafness in guinea pigs (19). Although Atoh1 gene expression plays a critical role in development of cochlear HCs, it is insufficient to produce functional auditory HCs. Other signaling mechanisms (e.g. Wnt, Shh, Fgf and Notch) also contribute in regulation of HCs development and regeneration.

#### Wnt

Wnt signaling pathway has a many key roles in animal development and includes intracellular signaling pathways (canonical Wnt pathway, the non-canonical planar cell polarity (PCP) and Wnt/calcium pathways) that transmit signals into a cell via cell surface receptors. This pathway is an evolutionarily conserved and highly complex signaling cascade that involves in regulation of important events such as cell proliferation and migration, neural patterning and cell polarity during development (20). Wnt/β-catenin and PCP pathways have been observed in the development of the mammalian cochlea. Wnt/β-catenin signaling is required for regulation of cell proliferation, cell fate and differentiation of HCs during early cochlear development (21, 22). It has been demonstrated that inhibition of canonical Wnt/β-catenin signaling blocks prosensory cell early stages of cochlea evolution at development (21). Furthermore, Wnt/β-catenin up regulation can induce Leucine-rich repeat G-protein-coupled receptor 5 (Lgr5) which acts as HCs progenitors in neonatal mammalian cochlea (23, 24). Planar polarity of HCs is established by the non-canonical Wnt/PCP pathway during cochlear development (25).

# Shh

The Hedgehog (HH) signaling pathway is essential for vertebrate embryonic development. Sonic hedgehog (SHH) is the best known ligand of the hedgehog signaling cascade. SHH is critical for patterning of the central nervous system (CNS) like induction of neural tube, tooth and lung development (26-28). SHH modulates cochlear HCs through regeneration regulating the retinoblastoma (Rb) proteins activity in rat. Rb proteins repress cell growth by preventing cell from entering the cell cycle. Inhibition of Rb proteins activity causes cells to divide. Previous study has shown that SHH treatment can lead to inhibition of Rb protein in neonatal cochlea explant culture, which induces HCs regeneration following neomycin damage (29). However, SHH role in the auditory HCs regeneration is largely stays unknown.

## Fgf

Fibroblast growth factor (FGF) signaling pathway is initiated through the binding of FGF ligands to FGF receptors (FGFR1, FGFR2, FGFR3 and FGFR4) (30). FGF signaling pathway has important roles during CNS development (31). During the development of the inner ear, it is responsible for otic placode induction and initial otocyst formation (32). FGF also regulates cochlear HCs formation at later stages (33, 34). Interruption of FGF signaling by FGF receptor inhibitor significantly decreases HCs and SCs development (11). In addition, FGF signaling cascade is linked to Atoh1 expression during inner ear evolution (35).

# Notch

The Notch signaling pathway is important in cell differentiation, proliferation and cell death. This pathway is critically required during the development of the inner ear (36), and acts through two mechanisms including lateral induction and lateral inhibition. Interaction between the Notch ligand and Notch receptor influences on neighboring cells and promote prosensory cell formation signal-mediated through Notch lateral induction in early stage of cell development (37, 38). Lateral inhibition of Notch signaling is critical for establishing the size and prosensory regions in later stage of the inner ear evolution. HCs and surrounding SCs are precisely arranged in mosaic pattern in the mammalian organ of Corti by Notch lateral inhibition effect (39). It was reported that the blockade of Notch signaling by  $\gamma$ -secretase inhibitors can lead to excessive regeneration of HCs in the zebrafish lateral line (40). In mammals, inhibition of Notch signaling by  $\gamma$ secretase inhibitor causes adjacent SCs can be converted into HCs in the postnatal period (41).

## Conclusion

Understanding the mechanisms and signaling cascades involved in auditory HCs development and regeneration can lead to designing effective therapeutic strategies for hearing loss recovery in human.

## **Funding/ Support**

None to declare.

## **Conflict of Interest**

The authors declare no conflict of interest.

## **References:**

1. Ekdale, Eric G. "Form and function of the mammalian inner ear." Journal of anatomy 228, no. 2 (2016): 324-337.

This open-access article distributed under the terms of the Creative Commons Attribution Non Commercial 3.0 License (CC BY-NC 3.0). Copyright © 2017 Shahid Beheshti University of Medical Sciences. All rights reserved. www.journals.sbmu.ac.ir/otolaryngology

- 2. Raphael, Yehoash, and Richard A. Altschuler. "Structure and innervation of the cochlea." Brain research bulletin 60, no. 5 (2003): 397-422.
- 3. Peyvandi, A., & Roozbahany, N. A. (2013). Hearing loss in chronic renal failure patient undergoing hemodialysis. Indian Journal of Otolaryngology and Head & Neck Surgery, 65(3), 537-540.
- 4. Lu, Xiaoling, Yilai Shu, Mingliang Tang, and Huawei Li. "Mammalian Cochlear Hair Cell Regeneration and Ribbon Synapse Reformation." Neural plasticity 2016 (2016).
- White, Patricia M., Angelika Doetzlhofer, Yun Shain Lee, Andrew K. Groves, and Neil Segil. "Mammalian cochlear supporting cells can divide and trans-differentiate into hair cells." Nature 441, no. 7096 (2006): 984.
- Malgrange, Brigitte, Marc Thiry, Thomas R. Van De Water, Laurent Nguyen, Gustave Moonen, and P. P. Lefebvre. "Epithelial supporting cells can differentiate into outer hair cells and Deiters' cells in the cultured organ of Corti." Cellular and molecular life sciences 59, no. 10 (2002): 1744-1757.
- Cox, Brandon C., Renjie Chai, Anne Lenoir, Zhiyong Liu, LingLi Zhang, Duc-Huy Nguyen, Kavita Chalasani et al. "Spontaneous hair cell regeneration in the neonatal mouse cochlea in vivo." Development 141, no. 4 (2014): 816-829.
- Chonko, Kurt T., Israt Jahan, Jennifer Stone, Margaret C. Wright, Tomoyuki Fujiyama, Mikio Hoshino, Bernd Fritzsch, and Stephen M. Maricich. "Atoh1 directs hair cell differentiation and survival in the late embryonic mouse inner ear." Developmental biology 381, no. 2 (2013): 401-410.
- Chai, Renjie, Bryan Kuo, Tian Wang, Eric J. Liaw, Anping Xia, Taha A. Jan, Zhiyong Liu et al. "Wnt signaling induces proliferation of sensory precursors in the postnatal mouse cochlea." Proceedings of the National Academy of Sciences 109, no. 21 (2012): 8167-8172.
- 10. Li, Wenyan, Jingfang Wu, Jianming Yang, Shan Sun, Renjie Chai, Zheng-Yi Chen, and Huawei Li. "Notch inhibition induces mitotically generated hair cells in mammalian cochleae via activating the Wnt pathway." Proceedings of the National Academy of Sciences 112, no. 1 (2015): 166-171.
- 11. Pickles, J. O. "The expression of fibroblast growth factors and their receptors in the

embryonic and neonatal mouse inner ear." Hearing research 155, no. 1 (2001): 54-62.

- 12. Żak, Magdalena, Sjaak FL Klis, and Wilko Grolman. "The Wnt and Notch signalling pathways in the developing cochlea: formation of hair cells and induction of regenerative potential." International journal of developmental neuroscience 47 (2015): 247-258.
- Chen, Ping, Jane E. Johnson, Huda Y. Zoghbi, and Neil Segil. "The role of Math1 in inner ear development: Uncoupling the establishment of the sensory primordium from hair cell fate determination." Development 129, no. 10 (2002): 2495-2505.
- 14. Kelly, Michael C., Qing Chang, Alex Pan, Xi Lin, and Ping Chen. "Atoh1 directs the formation of sensory mosaics and induces cell proliferation in the postnatal mammalian cochlea in vivo." Journal of Neuroscience 32, no. 19 (2012): 6699-6710.
- 15. Shou, Jianyong, J. Lisa Zheng, and Wei-Qiang Gao. "Robust generation of new hair cells in the mature mammalian inner ear by adenoviral expression of Hath1." Molecular and Cellular Neuroscience 23, no. 2 (2003): 169-179.
- 16. Yang, Juanmei, Ning Cong, Zhao Han, Yibo Huang, and Fanglu Chi. "Ectopic hair cell-like cell induction by Math1 mainly involves direct transdifferentiation in neonatal mammalian cochlea." Neuroscience letters 549 (2013): 7-11.
- 17. Kawamoto, Kohei, Shin-Ichi Ishimoto, Ryosei Minoda, Douglas E. Brough, and Yehoash Raphael. "Math1 gene transfer generates new cochlear hair cells in mature guinea pigs in vivo." Journal of Neuroscience 23, no. 11 (2003): 4395-4400.
- 18. Liu, Zhiyong, et al. "Age-dependent in vivo conversion of mouse cochlear pillar and Deiters' cells to immature hair cells by Atoh1 ectopic expression." Journal of Neuroscience 32.19 (2012): 6600-6610.
- 19. Atkinson, Patrick J., Andrew K. Wise, Brianna O. Flynn, Bryony A. Nayagam, and Rachael T. Richardson. "Hair cell regeneration after ATOH1 gene therapy in the cochlea of profoundly deaf adult guinea pigs." PloS one 9, no. 7 (2014): e102077.
- Logan, Catriona Y., and Roel Nusse. "The Wnt signaling pathway in development and disease." Annu. Rev. Cell Dev. Biol. 20 (2004): 781-810.

This open-access article distributed under the terms of the Creative Commons Attribution Non Commercial 3.0 License (CC BY-NC 3.0). Copyright © 2017 Shahid Beheshti University of Medical Sciences. All rights reserved. www.journals.sbmu.ac.ir/otolaryngology

- 21. Jacques, Bonnie E., Chandrakala Puligilla, Rachel M. Weichert, Anna Ferrer-Vaquer, Anna-Katerina Hadjantonakis, Matthew W. Kelley, and Alain Dabdoub. "A dual function for canonical Wnt/ $\beta$ -catenin signaling in the developing mammalian cochlea." Development 139, no. 23 (2012): 4395-4404.
- 22. Shi, Fuxin, Lingxiang Hu, Bonnie E. Jacques, Joanna F. Mulvaney, Alain Dabdoub, and Albert SB Edge. "β-Catenin is required for haircell differentiation in the cochlea." Journal of Neuroscience 34, no. 19 (2014): 6470-6479.
- 23. Shi, Fuxin, Lingxiang Hu, and Albert SB Edge. "Generation of hair cells in neonatal mice by βcatenin overexpression in Lgr5-positive cochlear progenitors." Proceedings of the National Academy of Sciences 110, no. 34 (2013): 13851-13856.
- 24. Chai, Renjie, Bryan Kuo, Tian Wang, Eric J. Liaw, Anping Xia, Taha A. Jan, Zhiyong Liu et al. "Wnt signaling induces proliferation of sensory precursors in the postnatal mouse cochlea." Proceedings of the National Academy of Sciences 109, no. 21 (2012): 8167-8172.
- 25. Waqas, Muhammad, Shasha Zhang, Zuhong He, Mingliang Tang, and Renjie Chai. "Role of Wnt and Notch signaling in regulating hair cell regeneration in the cochlea." Frontiers of medicine 10, no. 3 (2016): 237-249.
- 26. Litingtung, Ying, and Chin Chiang. "Control of Shh activity and signaling in the neural tube." Developmental Dynamics 219, no. 2 (2000): 143-154.
- 27. Hardcastle, Zoë, Rong Mo, C. C. Hui, and Paul T. Sharpe. "The Shh signalling pathway in tooth development: defects in Gli2 and Gli3 mutants." Development 125, no. 15 (1998): 2803-2811.
- 28. Bellusci, Savério, Yasuhide Furuta, Margaret G. Rush, Randall Henderson, Glenn Winnier, and B. L. Hogan. "Involvement of Sonic hedgehog (Shh) in mouse embryonic lung growth and morphogenesis." Development 124, no. 1 (1997): 53-63.
- 29. Lu, Na, Yan Chen, Zhengmin Wang, Guoling Chen, Qin Lin, Zheng-Yi Chen, and Huawei Li. "Sonic hedgehog initiates cochlear hair cell regeneration through downregulation of retinoblastoma protein." Biochemical and biophysical research communications 430, no. 2 (2013): 700-705.

- 30. Eswarakumar, V. P., I. Lax, and J. Schlessinger. "Cellular signaling by fibroblast growth factor receptors." Cytokine & growth factor reviews 16, no. 2 (2005): 139-149.
- 31. Hébert, Jean M. "FGFs: neurodevelopment's Jack-of-all-trades-how do they do it?." Frontiers in neuroscience 5 (2011).
- 32. Schimmang, Thomas. "Expression and functions of FGF ligands during early otic development." International Journal of Developmental Biology 51, no. 6-7 (2007): 473-481.
- 33. Hayashi, Toshinori, Catherine A. Ray, and Olivia Bermingham-McDonogh. "Fgf20 is required for sensory epithelial specification in the developing cochlea." Journal of Neuroscience 28, no. 23 (2008): 5991-5999.
- 34. Pirvola, Ulla, Jukka Ylikoski, Ras Trokovic, Jean M. Hébert, Susan K. McConnell, and Juha Partanen. "FGFR1 is required for the development of the auditory sensory epithelium." Neuron 35, no. 4 (2002): 671-680.
- 35. Su, Yi-Xun, Cong-Cong Hou, and Wan-Xi Yang. "Control of hair cell development by molecular pathways involving Atoh1, Hes1 and Hes5." Gene 558, no. 1 (2015): 6-24.
- 36. Yamamoto, Norio, Kenji Tanigaki, Masayuki Tsuji, Daisuke Yabe, Juichi Ito, and Tasuku Honjo. "Inhibition of Notch/RBP-J signaling induces hair cell formation in neonate mouse cochleas." Journal of molecular medicine 84, no. 1 (2006): 37-45.
- 37. Lewis, Julian. "Notch signalling and the control of cell fate choices in vertebrates." In Seminars in cell & developmental biology, vol. 9, no. 6, pp. 583-589. Academic Press, 1998.
- 38. Daudet, Nicolas, and Julian Lewis. "Two contrasting roles for Notch activity in chick inner ear development: specification of prosensory patches and lateral inhibition of hair-cell differentiation." Development 132, no. 3 (2005): 541-551.
- 39. Lanford, Pamela J., Yu Lan, Rulang Jiang, Claire Lindsell, Gerry Weinmaster, Thomas Gridley, and Matthew W. Kelley. "Notch signalling pathway mediates hair cell development in mammalian cochlea." Nature genetics 21, no. 3 (1999).
- 40. Ma, Eva Y., Edwin W. Rubel, and David W. Raible. "Notch signaling regulates the extent of hair cell regeneration in the zebrafish lateral

This open-access article distributed under the terms of the Creative Commons Attribution Non Commercial 3.0 License (CC BY-NC 3.0). Copyright © 2017 Shahid Beheshti University of Medical Sciences. All rights reserved. www.journals.sbmu.ac.ir/otolaryngology

line." Journal of Neuroscience 28, no. 9 (2008): 2261-2273.

41. Pan, Wei, Ying Jin, Jing Chen, Robbert J. Rottier, Karen P. Steel, and Amy E. Kiernan. "Ectopic expression of activated notch or SOX2 reveals similar and unique roles in the development of the sensory cell progenitors in the mammalian inner ear." Journal of Neuroscience 33, no. 41 (2013): 16146-16157.

This open-access article distributed under the terms of the Creative Commons Attribution Non Commercial 3.0 License (CC BY-NC 3.0). Copyright © 2017 Shahid Beheshti University of Medical Sciences. All rights reserved. www.journals.sbmu.ac.ir/otolaryngology