

Original Article

Evaluation of Matrix Metalloproteinase 2 and 9 Activity in Patients with Prostate Cancer and Benign Prostate Hyperplasia Compared with Healthy Individuals

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Abstract

Background and Aim: Prostate cancer (PC) is one of the most prevalent cancers with high mortality and morbidity in men, which can be treated in different ways before the progression and metastasis to distant organs. Destruction of extracellular matrix by matrix metalloproteinase (MMP), particularly by the 2 and 9 subtypes, has an important role in the metastasis of PC. We aimed to assess the activity of MMP 2 and 9 and some related metalloproteinases in PC and with benign prostate hyperplasia (BPH) patients in comparison to healthy individuals.

Methods: In this case-control study, 72 individuals referred to Imam Khomeini hospital (Tehran, Iran), have been divided into 3 groups, including PC, BPH, and healthy control. Age and body mass index (BMI) for all groups have been matched. Venous blood samples were used to assess the enzyme activity by the zymography technique.

Results: The activity of MMP-2 and 9 was significantly higher in PC than BPH and control groups. But there was no difference in the activity of enzymes in patients with PC according to the Gleason score.

Conclusion: The results suggested that MMPs activity can be considered a diagnostic marker for PC. However, further studies are required to establish this concept.

Keywords: Benign prostate hyperplasia; Metalloproteinase; Prostate Cancer.

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Introduction

Prostate cancer (PC) is one of the most prevalent malignancies in men worldwide. In 2018, 164690 new cases of the disease have been reported in the United States. Among them, approximately 29430 patients passed away. PC contains 19.23% of all cancer morbidity and 9% of all cancer-related death in the men population worldwide (1).

This cancer can be treated by current strategies if it is limited to just prostate tissue. But, the treatment almost is not completely effective when it metastases to other tissues. The metastasis is a complex cascade process leading to tumor cell migration, attachment, and invasion. Cellular invasion contains the tumor cell translocation across the extracellular matrix barrier as a known important biological event needed for tumor metastasis (2).

The MMPs are a family of proteolytic enzymes that degrade extracellular matrix components such as collagen, fibronectin, and laminin (3). The most important component of the basal membrane is collagen IV, which is degraded by MMP-9. In this regard, the evaluation of MMP-9 proteolytic activity is essential for an understanding of basal membrane change and repair mechanisms and abnormal collagen destruction in pathologic conditions such as atherosclerosis, cancer, and rheumatoid arthritis (4-7).

Among all types of MMPs, MMP-2, 9 can degrade abnormal collagen and the types of IV, V, VII, IX. Recently their role in apoptosis, differentiation, angiogenesis, immune responses, and tumor cell growth has been revealed (8). The MMP-9 expression was associated with a higher rate of metastasis; it was confirmed that the enzyme inhibitors reduced the metastasis rate of PC (9). The MMP-9 expression was higher in the serum and tissue of PC compared to BPH (10, 11). The increasing MMP-2 expression is associated with a decreased survival rate in patients with PC (12, 13). MMP-2 also was known as an activator of MMP-9. (14). Therefore, it aimed to assess the activity of MMP-2 and 9 in PC and BPH patients in comparison to healthy men.

Methods

Among the patients who were referred to the urology center in the Imam Khomeini hospital in Tehran City during 2018-2019, Forty-eight patients were selected and after the biopsy and pathological tests were divided into two groups: PC group (n=24) and BPH (n= 24). The Third group included 24 healthy people. Exclusion criteria in the PC group include the patients who were diagnosed with more than one year of diseases and who received anti-cancer drugs, chemotherapy, hormone therapy, and radiotherapy. Inclusion criteria in the BPH group include the BPH detection and PC ruling out according to histological survey following open prostatectomy.

Exclusion criteria in the BPH group include the patients with a history of cancer, received finasteride more than one month and anti-cancer

drugs, and whose prostate histological evaluation showed a section suspected to prostate intraepithelial neoplasia (PIN).

All the individuals who entered the study signed the testimonial consciously and with desire.

Blood sampling

From all individuals, about 2.5 mL of blood samples were collected into tubes without any coagulant agents from the forearm, and serum was extracted and used for zymography tests.

Zymography

Serum samples were electrophoresed on polyacrylamide gel 10% with sodium dodecyl sulfate (SDS-PAGE) and gelatin 1% (gelatinase substrate). After electrophoresis, the gel was incubated in 2% Triton X-100 solution in Phosphate-buffered saline (PBS) for one hour at room temperature and then in Tris-HCL (pH 7.4, containing 10 mmol calcium-chloride) for 16 hours at 37°C. Following washing, the gel was stained by 0.05% coomassie brilliant blue G-250. Then, the destaining was done with a solution of water: methanol: acetic acid with a ratio of 60%, 30%, and 10%, respectively. The destained bands produced by MMP-9, MMP-2, MMP-9/NGAL, and dimmer MMP-9 activity appeared in a purple background. Protein weight markers (Color Burst, Sigma Aldrich; USA) were used to confirm the identity of the gelatinase band. After the complete destaining, the gels have been filled between two transparent films and scanned by a Canon scanner (LiDE110, Japan).

Quantification of bands produced by gelatinase activation

Colorless bands of zymography gels produced by the activity of MMP-9, MMP-2, MMP-9/NGAL, and dimmer MMP-9 were quantified by measurement of the bands' area with ImageJ software.

Statistical analysis

To compare the mean activity of MMP-9, MMP-2, MMP-9/NGAL, and Dimmer MMP-9 enzymes in patients groups with the healthy group, the SPSS-20 software was used for statistical analysis and T-test, ANOVA and Tukey tests were used and the results were reported in Mean \pm standard deviation (SD). Bivariate correlation test and Spearman's rho

statistical method were also used to investigate the association between marker activity and cancer stage. All results reported in 95% confidence interval (CI) and p-value <0.05 were considered significant.

Results

The demographic results were compared between three PC, BPH, and cancer (Table 1). There were no statistical differences between the three groups in age and BMI, so we considered that the groups are matched. The groups were also examined for smoking and family history. There were no statistical differences between groups according to familial history and smoking.

Table 1. Comparison of the demographic result between the three groups

Characteristics		N	Mean± SD	P-value	
Age (year)	PC	24	64.00± 6.11	0.597	
	BPH	24	66.63± 6.40		
	control	24	65.85±10.69		
BMI (Kg/m ²)	PC	24	23.21± 3.21	0.297	
	BPH	24	22.16± 3.20		
	control	24	23.61± 3.08		
Familial history	PC	N		0.357	
		percent			
	Yes	2	8.34		
	No	22	91.66		
	BPH	Yes	1		4.17
		No	23		95.83
	control	Yes	0		0
		No	24		100
	Smoking	PC	Yes		7
No			17	70.83	
BPH		Yes	4	16.67	
		No	20	83.33	
control		Yes	10	41.67	
		No	14	58.33	

PC: Prostate cancer; MMP: matrix metalloproteinase; BPH: benign prostate hyperplasia

It can be seen from the data in Table 2 that there is a significant difference in the quantitative activity of MMPs except for MMP-9/NGAL between the three groups. Tukey test was used to determine the difference of quantitative activity

of MMPs between each group. The data shown in Table 3 demonstrated that the Dimmer MMP-9 between cancer and control groups, the MMP-9 and MMP-2 between the cancer group with the control and BPH groups differed significantly.

Table 2. Comparison of the quantified activity level of MMPs between three groups

Variables	Mean±SD	p-value
Dimmer MMP-9 (IU)	PC	163.35± 40.85
	BPH	125.86±52.77
	control	94.31±72.98
MMP-9/NGAL (IU)	PC	20.56±18.98
	BPH	34.42±33.59
	control	18.13±7.70
MMP-9 (IU)	PC	291.98±141.81
	BPH	144.90±90.77
	control	193.96±50.25

MMP-2 (IU)	PC	113.18±65.38	0.019
	BPH	70.59±44.23	
	control	71.24±26.33	

PC: Prostate cancer; MMP: matrix metalloproteinase; BPH: benign prostate hyperplasia

Table 3. Multiple Comparison of the quantified activity level of MMPs between each group

Enzyme	Groups		p-value
Dimmer MMP-9 (IU)	PC	control	0.006
		BPH	0.000
MMP-9 (IU)	PC	control	0.041
		BPH	0.026
MMP-2 (IU)	PC	control	0.041
		BPH	0.041

PC: Prostate cancer; MMP: matrix metalloproteinase; BPH: benign prostate hyperplasia

The prognosis of men with prostate cancer evaluated by the Gleason grading system (Table 4). The prostate tissues that they have prepared by the biopsy, are evaluated by the pathologists and as cancer progresses, they give it a score of 2 to 9. The higher numbers indicate greater risks and higher mortality. The results demonstrate that when patients with PC are divided into two

groups according to the Gleason score of >7 and ≤7, no differences are seen in the activity of the enzymes. The correlation analysis between different factors demonstrated that MMP-9 positively correlated with MMP-2 (R: 0.708; P < 0.001). Other factors showed no significant correlations.

Table 4. Comparison of the quantified activity level of MMPs in patients with PC based on the Gleason score

Variables	GS divided	Mean	Std. Deviation	p-value
Age (year)	>7	64.50	7.06	0.810
	≤7	63.70	5.85	
BMI (kg/m ²)	>7	22.24	1.75	0.285
	≤7	23.79	3.80	
Dimmer MMP-9 (IU)	>7	161.41	30.67	0.899
	≤7	164.57	48.16	
MMP-9/NGAL (IU)	>7	22.09	17.87	0.806
	≤7	19.64	20.50	
MMP-9 (IU)	>7	293.31	180.96	0.977
	≤7	290.99	118.13	
MMP-2 (IU)	>7	143.97	63.78	0.282

Discussion

The important processes in cancer development are angiogenesis, metastasis, and distribution of tumor cells far from the original location. The process of angiogenesis includes the growth and branching of blood vessels in tumor tissue (15, 16). Several studies illustrated that MMPs such as MMP-2 and MMP-9 have a key role in angiogenesis (17-21).

Trudel D et al. demonstrated MMP-9 is over-expressed in high-grade prostate tumor cells and also in benign stromal and epithelial cells in the early stages (22). These findings were in line with

our results. In the present study, MMP-9 activity was 291.98 in patients with PC, while in the BPH and control groups, it was 144.9 and 193.96, respectively. Moreover, these results showed that MMP-9 activity significantly increased in prostate cancer. These results were not consistent with Rodríguez G *et al.* results that they didn't observe any association between the plasma expression of MMP-9 and prostate syndromes, so it can't be considered as a marker for PC diagnosis (23). According to previous studies, it can be concluded that MMP-9 isn't a proper diagnostic marker but can be considered as a target molecule for inhibiting

tumor progression. Based on current results, both MMP-2 and MMP-9 enzymes can be used as diagnostic markers because their expression is not significantly different in healthy men and benign prostatic hyperplasia, but in the group of prostate cancer with both healthy and BPH is significantly different.

Morgia G et al. investigated the MMP-13 as a diagnostic marker and MMP-2, 9 as prognostic markers in PC. They concluded that the plasma level and activity of MMPs concomitant with PSA determination can play a key role in the diagnosis, treatment, and screening of prostate cancer (11).

Wilson et al. reported that produced protease-activated receptors 1 and 2 (PAR-1 and PAR-2) can increase the activity of MMP-2, 9 in PC cells that confirm their role in PC metastasis (24). Our result consistent with Wilson's research demonstrated MMP-2 activity in patients with PC is remarkably higher than the BPH and the control group. Hamdy F.C et al. and Festuccia C et al. showed the elevation of MMP-9 levels in patients with PC compared to BPH (25, 26). But several studies reported opposite results. Lokeshwar et al. (27) revealed that BPH caused a higher level of MMP-9 in comparison to patients with PC, while the level of MMP-2 in PC samples was more than BPH (28-31). Similar studies proved an increased level of MMP-2 in PC compare to BPH, however, Upadhyay et al. (32) didn't observe any significant difference in MMP-2 expression among patients with PC and healthy control. No definitive specificity of these enzymes for cancer was found in this study, but it is confirmed that they are significantly higher in cancer patients. On the other hand, in people who have recently been diagnosed with prostate cancer, they can be used as specific therapeutic targets in the cancer site.

Conclusion

The results suggested that MMPs activity can be considered a diagnostic marker for PC. However, further studies are required to establish this concept.

Conflict of Interest

The authors declared that they have no conflict of interest.

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Ethics

The protocol of experiments was permitted by the ethical committee of MAZUMS (code: IR.MAZUMS.REC.1398.1065).

Author contributions

MM, HJ, HB, and SP performed sampling and experiments and assembled input data. AK-T and HM-S designed the study, analyzed and interpreted the data, and wrote the paper.

References

1. Zamanian Z, Dehghani M, Mohammady H, Rezaeiani M, Daneshmandi H. Investigation of shift work disorders among security personnel. *International Journal of Occupational Hygiene*. 2012;4:39-42.
2. Bogenrieder T, Herlyn M. Axis of evil: Molecular mechanisms of cancer metastasis. *Oncogene*. 2003;22:6524.
3. Nabeshima K, Inoue T, Shimao Y, Sameshima T. Matrix metalloproteinases in tumor invasion: Role for cell migration. *Pathology international*. 2002;52:255-264.
4. Busti C, Falcinelli E, Momi S, Gresele P. Matrix metalloproteinases and peripheral arterial disease. *Internal and emergency medicine*. 2010;5:13-25.
5. Murphy G, Nagase H. Progress in matrix metalloproteinase research. *Molecular aspects of medicine*. 2008;29:290-308.
6. Egeblad M, Werb Z. New functions for the matrix metalloproteinases in cancer progression. *Nature Reviews Cancer*. 2002;2:161.
7. Björklund M, Koivunen E. Gelatinase-mediated migration and invasion of cancer cells. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*. 2005;1755:37-69.
8. Mook OR, Frederiks WM, Van Noorden CJ. The role of gelatinases in colorectal cancer progression and metastasis. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*. 2004;1705:69-89.

9. Sehgal G, Hua J, Bernhard EJ, Sehgal I, Thompson TC, Muschel RJ. Requirement for matrix metalloproteinase-9 (gelatinase b) expression in metastasis by murine prostate carcinoma. *The American journal of pathology*. 1998;152:591.
10. Sauer CG, Kappeler A, Späth M, Kaden JJ, Michel MS, Mayer D, et al. Expression and activity of matrix metalloproteinases-2 and-9 in serum, core needle biopsies and tissue specimens of prostate cancer patients. *Virchows Archiv*. 2004;444:518-26.
11. Morgia G, Falsaperla M, Malaponte G, Madonia M, Indelicato M, Travali S, et al. Matrix metalloproteinases as diagnostic (mmp-13) and prognostic (mmp-2, mmp-9) markers of prostate cancer. *Urological research*. 2005;33:44-50.
12. Trudel D, Fradet Y, Meyer F, Harel F, Têtu B. Significance of mmp-2 expression in prostate cancer: An immunohistochemical study. *Cancer research*. 2003;63:8511-5.
13. Trudel D, Fradet Y, Meyer F, Harel F, Têtu B. Membrane-type-1 matrix metalloproteinase, matrix metalloproteinase 2, and tissue inhibitor of matrix proteinase 2 in prostate cancer: Identification of patients with poor prognosis by immunohistochemistry. *Human pathology*. 2008;39:731-9.
14. Fridman R, Toth M, Peña D, Mobashery S. Activation of progelatinase b (mmp-9) by gelatinase a (mmp-2). *Cancer research*. 1995;55:2548-55.
15. Ferrara N, Kerbel RS. Angiogenesis as a therapeutic target. *Nature*. 2005;438:967
16. Michael A, Relph K, Pandha H. Emergence of potential biomarkers of response to anti-angiogenic anti-tumour agents. *International journal of cancer*. 2010;127:1251-8.
17. Cross MJ, Claesson-Welsh L. Fgf and vegf function in angiogenesis: Signalling pathways, biological responses and therapeutic inhibition. *Trends in pharmacological sciences*. 2001;22:201-7.
18. Cooke JP, Losordo DW. Nitric oxide and angiogenesis. 2002;105:2133-5.
19. Nissen LJ, Cao R, Hedlund E-M, Wang Z, Zhao X, Wetterskog D, et al. Angiogenic factors fgf2 and pdgf-bb synergistically promote murine tumor neovascularization and metastasis. *The Journal of clinical investigation*. 2007;117:2766-77.
20. Stetler-Stevenson WG. Matrix metalloproteinases in angiogenesis: A moving target for therapeutic intervention. *The Journal of clinical investigation*. 1999;103:1237-41.
21. Vanderslice P, Munsch CL, Rachal E, Erichsen D, Sughrue KM, Truong AN, et al. Angiogenesis induced by tumor necrosis factor- α ; is mediated by α 4 integrins. *Angiogenesis*. 1998;2:265-5.
22. Trudel D, Fradet Y, Meyer F, Têtu B. Matrix metalloproteinase 9 is associated with gleason score in prostate cancer but not with prognosis. *Human pathology*. 2010;41:1694-701.
23. González RI, Gil UR, Del Fresno MR, Fernández GI, Tardón A. 537 the value of plasmatic determination of matrix metalloproteinase 9 as a diagnostic marker in prostate cancer. Closing the controversies. *European Urology Supplements*. 2012;11:e537.
24. Wilson SR, Gallagher S, Warpeha K, Hawthorne SJ. Amplification of mmp-2 and mmp-9 production by prostate cancer cell lines via activation of protease-activated receptors. *The Prostate*. 2004;60:168-74.
25. Hamdy F, Fadlon E, Cottam D, Lawry J, Thurrell W, Silcocks P, et al. Matrix metalloproteinase 9 expression in primary human prostatic adenocarcinoma and benign prostatic hyperplasia. *British journal of cancer*. 1994;69:177.
26. Festuccia C, Bologna M, Vicentini C, Tacconelli A, Miano R, Violini S, et al. Increased matrix metalloproteinase-9 secretion in short-term tissue cultures of prostatic tumor cells. *International journal of cancer*. 1996;69:386-93.
27. Lokeshwar BL, Selzer MG, Block NL, Gunja-Smith Z. Secretion of matrix metalloproteinases and their inhibitors (tissue inhibitor of metalloproteinases) by human prostate in explant cultures: Reduced tissue inhibitor of metalloproteinase secretion by malignant tissues. *Cancer research*. 1993;53:4493-8.
28. Stearns ME, Wang M. Type iv collagenase (mr 72,000) expression in human prostate: Benign and malignant tissue. *Cancer research*. 1993;53:878-83.
29. Monfironi R, Fabris G, Lucarini G, Biagini G. Location of 72-kd metalloproteinase (type iv collagenase) in untreated prostatic adenocarcinoma. *Pathology-Research and Practice*. 1995;191:1140-6.
30. Montironi R, Lucarini G, Castaldini C, Galluzzi C, Biagini G, Fabris G. Immunohistochemical evaluation of type iv collagenase (72-kd metalloproteinase) in prostatic intraepithelial neoplasia. *Anticancer research*. 1996;16:2057-62.
31. Still K, Robson CN, Autzen P, Robinson MC, Hamdy FC. Localization and quantification of mrna for matrix metalloproteinase-2 (mmp-2) and tissue inhibitor of matrix metalloproteinase-2 (timp-2) in human benign and malignant prostatic tissue. *The Prostate*. 2000;42:18-25.
32. Upadhyay J, Shekarriz B, Nemeth JA, Dong Z, Cummings GD, Fridman R, et al. Membrane type 1-matrix metalloproteinase (mt1-mmp) and mmp-2 immunolocalization in human prostate: Change in cellular localization associated with high-grade prostatic intraepithelial neoplasia. *Clinical Cancer Research*. 1999;5:4105-10.