Review Article

A Review on Relationship Between Human Endogenous Retrovirus Groups and Human Diseases

Reza Aramideh khouy¹, Mehrnaz Hosseini Tehrani¹, Hamed Nosrati², Maryam Esghaei^{1*}

¹ Department of Virology, Iran University of Medical Sciences, Tehran, Iran ² Department of Tissue Engineering, Faculty of Advanced Technologies, Shahrekord University of Medical Sciences, Shahrekord, Iran

Received: 23 June, 2018, Accepted: 24 November, 2018

Abstract

Various factors are involved in the incidence of some diseases like autoimmune, psychiatric and cancerous ones. One of these probable factors is considered as the endogenous retroviruses, for example, proviruses that have been introduced in previous generations in some organisms' genome, and make up over 8% of the human genome. Recent studies have indicated that these factors and their related products (including RNA, cytosolic DNA, and proteins) may affect and also change the host cell function and immune system. This review summarizes the detailed information about the structure, classification, and pathogenesis mechanism of human endogenous retroviruses and their relationship with the autoimmune diseases and some kinds of cancers.

Keywords: Endogenous Retroviruses, Autoimmune disease, Mental Disorders, Neoplasms.

***Corresponding Author:** Maryam Esghaei, Department of Virology, Iran University of Medical Sciences, Tehran, Iran. Email: maryam.esghaei@gmail.com, esghaei.m@iums.ac.ir.

Please cite this article as: Aramideh khouy R, Hosseini Tehrani M, Nosrati H, Esghaei M. A Review on Relationship Between Human Endogenous Retrovirus Groups and Human Diseases. Arch Med Lab Sci. 2018;4(4): 1-9.

Introduction

About 45% of the human genome contains the intergenic regions. The genome of all studied eukaryotes (from yeast to humans) has these intergenic regions variety and it has aspects of hereditary into the next generation. Retrotransposons are an important part of these areas, which were divided into elements containing long terminal repeat (LTR) and LTR-free. The most common retrotransposons are endogenous retroviruses (ERVs) containing LTR in humans, and can express viral proteins similar to the exogenous retroviruses (1, 2). HERV products in most cases can increase antigenic sensitivity and reciprocal reactivity against host antigens. Current studies use retroviruses as an indication of autoimmune disease and cancer pathology (3). Here, we reviewed endogenous retroviruses and discussed about their classification,

pathogenesis and possible association of various human endogenous retrovirus (HERV) groups with autoimmune diseases, nervous system diseases and cancer. Identification of endogenous viruses association with various diseases not only helps to diagnose the pathology, but also provides an appropriate therapeutically approach for related disorders.

Human endogenous retrovirus (HERV)

Endogenous viruses have been derived from their exogenous counterparts, due to the germline infection and their proliferation amongst their host genomes (4). These viruses are a big part of a larger family of retroviral elements, which is composed of about 5-8% of the human genome (5). The HERVs structure includes gag, pol and env that are surrounded

by LTR regions on both sides, as same as exogenous retroviruses. Briefly, gag encodes the matrix, the capsid and the nucleocapsid proteins, and pol encodes protease (PR), reverse transcriptase (RT), Ribonuclease H (RNase H), integrase (IN), and env encodes virus coat proteins. HERVs also contains the primer binding site (PBS) sequence, between 5'LTR and gag, and polypurine tract (PPT), between 3'LTR and env. Specifically, the first is the binding site of the tRNA primer for the negative strand synthesis, and the second one is the binding site of the primer for the positive strand synthesis (6-8). Moreover, the HERV-K group (HML2) encodes two accessory proteins named Np9 and Rec, which are derived from alternative splicing of the env transcript. The expression of these two proteins has been demonstrated in cancerous cells (9). The LTR sequence that contains U3-R-U5 plays an important role in the regulating viral gene expression. These regions, which can act as alternate promoters and enhancers, often contribute to the transcriptomes production in different cells(6, 8).

Classification of HERVs

Because of the lack of appropriate nomenclature, HERVs classification has been remained uncompleted (8). Based on their homology with animal retroviruses, HERVs are categorized into three main classes: class I is similar to the Gammaretrovirus and Epsilonretrovirus; class II is similar to Betaretrovirus and finally class III is similar to the Spumaretrovirus. Individual nomenclature of HERVs is accomplished based on various characteristics in which there is no specific pattern. for example, some HERVs were named based on the amino acids along with tRNA-Primer, which were bonded to PBS (like HERV-E for glutamic acid and HERV-W for tryptophan). Some others were named

based on their proximity to a specific gene (like HERV-ADP) or a specific motif (like HERV-FRD). In terms of the common structural features of HERVs, they have been categorized into 39 "canonical" groups and 31 "noncanonical" clades using the Retro Tector software (10-15)(Table1).

Pathogenicity mechanism

HERVs and their products (including RNA, cytosolic DNA and proteins) could affect and alter the host's immune system, for example it is recommended that these factors may play an important role in the immune system's evolution and its function(16). They affect the host with different ways:

- Integration of the DNA of provirus into the • genome: HERVs integration at any location on the host genome could affect their adjacent genes activity even if they were not translated or replicated. Also, LTRs can have an inhibitory or activating effect on the promoter / gene enhancer in proximity to the HERV integration region (17). HERVs integration in the introns, can modify the starting and ending of the transcription region, and would lead to an mRNA with abnormal activities. They can also create different splicing locations, which ultimately affect the produced mRNA functions in negative way(18).
- *HERVs effect on the innate immune system:* These agents may play an important role in forming and expanding of the interferon signaling network and the host innate immune system. Consequently, their replication leads to inflammatory and autoimmune disorders and control the immune system over-activation throughout inhibitory properties (16, 19-21).

Class	nss Type species		
Class I (gamma-like, epsilon-like)	Murine leukemia virus (MLV) Feline leukemia virus (FeLV)	Canonical 27 Noncanonical 25	
	Walleye dermal sarcoma virus (WDSV)	23	
Class II (beta-like)	Mouse mammary tumor virus (MMTV)	Canonical 10	
	Mason-Pfizer monkey virus (MPMV)	Caliblical 10	
	Jaagsiekte sheep retrovirus (JSRV)		
Class III (spuma-like)	Simian foamy virus (SFV)	Canonical 2 Noncanonical 5	
Uncertain_Errantilike	Gypsy retrovirus	Noncanonical 1	

Table1. Classification of HERVs (adapted from Vargiu et al., 2016)

• The HERV proteins effect on the immune system: Evidences from HERV protein expression indicated that these products affect their hosts immune system. The analogy of these proteins with antigenic epitopes and their identification by immune cells causes some responses by the host's immune system and that can lead to higher pathogens tolerance in the host immune system in the long run.

In one animal model study, it was shown that those encoded antigen by ERV can be detected by T cells, and an immune response can be created for counteraction (22). Another study showed that HERV-K18 env stimulates the V β 7-expressing T-cells in humans and transgenic mice (21, 23). Other products including env in HERV-E and HERV-H or Syncytin in HERV-W can be identified by the host immune system and moreover they can stimulate the immune system. (21).

HERV in autoimmune diseases and psychiatric disorders

Studies have suggested the direct and indirect relationship between the HERVs and various kinds of diseases. For example, HERV-H/F and HERV-W/MSRV have been demonstrated to be activated in patients with multiple sclerosis (MS). In addition, the level of HERV RNA and associated antigens increase in the mononuclear brain and peripheral blood cells (PBMCs) (24, 25).

Due to uncertainty about the pathology of schizophrenia, different speculations have been deduced, however, the HERV-W and HERV-K presence in clinical samples of these patients can be considered as important keys for understanding of the disease pathology (26, 27). An increase in the HERV-related products such as reverse transcriptase has been observed in the patients with amyotrophic lateral sclerosis (ALS) (28).

The relationship between endogenous retroviral agents with other autoimmune diseases like RA, SLE, T1D, EAE, etc. has been also investigated, and indicated the role of these agents in immune-related diseases and psychiatric disorders (29-31) (Table 2).

HERVs and cancer

Cancer refers to a set of diseases resulting from the uncontrolled cell proliferation. Genetic and environmental risk factors are considered as the most important factors in the cancers incidence. Studies have shown a correlation between HERVs and many types of tumors like melanoma, breast cancer, germ cell tumors, liver cancer and also ovarian cancer (32, 33). The unusual HERVs expression in some of the body cells, like the HERV-K elements in the germ cell tumor and melanoma indicates the association between these factors with cancer cells (34). Some investigations have indicated a significant increase in antibody titer against the provirus in patients with ovarian cancer (35). The HERV-K-related proteins expression has been also observed in patients with breast cancer (36). Studies have indicated the association between different HERV groups with different cancer types (Table3).

Conclusion

This study provides a review of the relation between different human endogenous viruses and the autoimmune, neurological and cancerous disorders. The endogenous proviruses presence amongst human genes and their compliance with Mendelian inheritance laws have raised questions, which researchers have been seeking for answering to it over the past few years. Existence of many questions and few answers indicated our little knowledge about this provirus's role. The definite relationship between these proviruses and MS is established by accomplishing some clinical studies on MS patients. However, the mechanism of this relationship is still unclear. Other researches in this field also recommend that numbers of autoimmune and psychiatric diseases and cancers are associated with endogenous proviruses. The presence of these factors and the possibility of interference in the immune and cellular systems normal process would result in the best and worst fate for the cells and cellular factors. The former one led us to have a deeper view into the usage of antiviral drugs in the endogenous retrovirus-related diseases treatment. However, the latter one led to use these factors in the treatment and control of further diseases.

Disease	Group	Туре	rovirus groups related Molecular technique	Sample	Ref		K119 HERV- K106				
		syncy tin-1	RT-PCR	PBMC	(37)		HERV- K18	LTR	PCR	Blood	(70)
		syncy tin-1	ELISA	PBMC	(38)		HERV- K10s	Env	PCR	Plasma	(71)
		syncy	Immunohistological	Brain	(39)		HERV-K HERV-	LTR syncy	PCR	PBMC	(72)
		tin syncy	PCR	PBMC	(40)		Env59	tin	RT-PCR	PBMC	(73)
		tin syncy	RT-PCR	Serum Brain	(40)		HRES-I	LTR	PCR , Southern blot hybridization	PBL	(74)
	HERV-W	tin syncy	qRT-PCR	PBMC	(41)		HRES-1	Gag	Western blot analysis ELISA	Serum	(75)
		tin Env	RT-PCR	Brain Blood	(43)	SLE	HERV-E		in silico		
		Env	ELISA	Serum	(44)		HERV-K	LTR	PCR	PBMC	(61)
		Synci tin	Immunohistology	Brain	(45)		HERV- K113	Gag	PCR	Genomic DNA	(63)
		Pol	FISH PCR	РВ	(46)		HERV	P30 GAG	Western blot RT-PCR	Serum	(76)
MS		Syncy tin-1	Immunohistochemist ry qPCR	Brain	(47)	-	ERV-9 HERV-H	Env	ELISA	Blood	(77)
-	HERV- K113	Gag Pol Env	PCR and mass- spectrometry	Blood PBMC	(48)		HERV-K	Env	Immunoblotting- ELISA	Serum	(78)
	HERV-E HERV-	Gag	qPCR	Brain	(49)		HRES-1	LTR	PCR Southern blot	Blood	(79)
_	K10 HERV-	Env	qPCR	Brain				syncy	RT-PCR, cell culture ELISA, immunohistology Immunocytofluoresc	Blood	
	K18 HERV-	Env	PCR	Blood	(50)	CIDP	HERV-W	tin		Brain	(80)
	K10	Env	RNA-PCR RT-PCR	PBMC	(51)	EAE	HERV-W	Env	- ence	Mice	(81)
	HERV-H	Gag	South-Western blots	Blood	(52)				Immunohistochemist ry	Salivary gland biopsies	(82)
	HERV-E HERV-	Gag Env	qPCR qPCR	Brain Brain	(53)	SS Psoriasis Addison	HRES-I	Gag			
	K10 HERV-H	Env	serology	Blood	(54)		HERV	P30	Western blot	Serum	(76)
	HERV-Fc1	Env	PCR	PBMC	(55)		HRES-1	Gag Gag	RT-PCR ELISA	Serum	(67)
	HERV-H	Env	Western Blot flow cytometric	PBMC	(56)		ERV-9 /HERV-W	Pol	RT-PCR	Tissue	(83)
1	HERV-Fc1	Env	PCR and mass- spectrometry	Blood PBMC	(56)		HERV-K HERV-E			Blood	(85)
	HERV-K	Gag	RT-PCR ELISA	Blood	(57)		HERV-K	dUTP ase	ELISPOT ELISA	Serum PBMC	(84)
	HERV-K	LTR Pol	PCR-SSP	Blood	(58)		HERV	LTR1	PCR	Blood	(85)
	HERV-K	Env	NASBA technology	Plasma Synovial	(59)	-	HERV-W	3 syncy	RT-PCR	PBMC	(86)
	HERV- K10 HERV-K HERV-L	RT Pol	RT-PCR RT-PCR	fluid Cell PBMC Synovial	(60)		HERV-W	tin syncy tin	Western blotting Immunohistochemist ry Recombinant	Brain	(87)
	ERV-9				(61)		HERV-K	LTR	proteins qRT-PCR	Tissue	(88)
RA	HERV-K HERV-	Env	Indirect ELISA	Serum Genomic	(62)		HERV-W	Gag Env	ELISA	Serum	(89)
	K113	Gag	PCR	DNA Synovial	(63)	schizophrenia	HERV- K115	LTR	PCR	Leukocyte	(90)
	HERV-W Pol	Pol	PCR	fluid	(64)	-	HERV- K	-	COBRA	PBL	(91)
				Plasma samples			HERV-W	Gag	Haplotype analysis PCR	Plasma	(92)
	HERV-K HERV-	Gag	RT-PCR	Blood	(57)		HERV-W	Env (U25	Nested RT-PCR	Blood	(49)
	K10	Gag	RT-PCR	PBMC	(65)			1)			
	ERV3	Env	RFLP	Synovial tissues PBMC	(66)		HERV-W HERV K-	Pol UTR	nested RT-PCR PCR	Blood Lymphocy	(93)
	HRES-1	Gag	ELISA	Serum	(67)]	18	1 2.11		te	()4)
	ERV-9 HERV-K HERV-L	Pol	RT-PCR	Synovial	(68)	MS, Multiple Sclerosis; RA, Rheumatoid arthritis; T1DM, Type 1 diabetes me SLE, Systemic lupus erythematosus; CIDP, Chronic Inflammatory Demyelir					
	HERV-K	Env	Immunoblotting ELISA	Serum	(69)	Polyneuropat		perimental	autoimmune encephalor		
T1DM	HERV-K HERV-H HERV-	Gag Pol Env	PCR and mass- spectrometry	Blood PBMC	(48)		, _r piterar				

-

Cancer	Group	Туре	Molecular technique	Sample	Ref
	HERV-K	Gag	SEREX analysis Western blotting	Serum	(95)
Prostate Cancer	HERV-K	LTR	RT–PCR	Cell lines	(96)
	HERV-K	LTR	qRT-PCR	Tissue and Cell lines	(97)
	HERV-E	Env	RT-PCR	Tissue	(98)
	HERV-K	Env	Real-TimePCR cytometry analysis	Cell lines and culture conditions	(99)
	HERV-K	Env	immunohistochemistry	Cell lines	(100)
	HERV-K	Gag Env	PCR	Serum	(101)
Melanoma	HERV-K	Gag Env Pol	qRT-PCR Indirect immunofluorescence	Tissue	(102)
	HERV-K	Gag	Immunohistochemistry Immunoblotting assay	Tissue	(103)
	ERV	Gag Env Pol	qRT–PCR	Tissue	(104)
	HERV-K	Pol Env	Real-Time PCR	Tissue	(105)
	HERV-K	Gag Env	qRT-PCR	Melanoma Cell lines and Human tissue	(34)
	HERV-K	Gag Env Pol	qRT-PCR	Tissue	(106)
	HERV-K	LTR	qRT-PCR	Cell lines	(107)
Ovarian Cancer	HERV-K	Env	RT–PCR	Blood Tissue	(108)
Ovarian Cancer	HERV-K	LTR	PCR COBRA	Tissue	(109)
Lung Cancer	HERV-K113	LTR	Nested inverse PCR genotyping PCR	Tissue	(110)
Pancreatic cancer	HERV-K	Env	RT-PCR	Tissue	(111)
Colon Cancer	HERV-H	LTR	RT-PCR	Tissue	(112)
Hepatocellular Carcinoma	HERV-K	Env	qRT-PCR	Tissue	(113)
Seminoma	HERV-W	syncytin-1	DdPCR qRT-PCR	Tissue	(114)
Urothelial Carcinoma	HERV-E	Gag	qRT-PCR	Tissue	(115)

 Table 3. List of endogenous retrovirus groups related with Cancers

Conflict of Interests

The authors report no conflicts of interest.

Funding & Acknowledgments

We need to thank Virology Department of Iran University of Medical Sciences for providing opportunities for retroviral studies.

References

1. Hancks DC, Kazazian HH. Roles for retrotransposon insertions in human disease. Mobile DNA. 2016;7(1):9.

2. Hurst T, Magiorkinis G. Epigenetic control of human endogenous retrovirus expression: focus on regulation of long-terminal repeats (LTRs). Viruses. 2017;9(6):130.

3. Singh SK. Endogenous retroviruses: suspects in the disease world. 2007.

4. Belshaw R, Pereira V, Katzourakis A, Talbot G, Pačes J, Burt A, et

al. Long-term reinfection of the human genome by endogenous retroviruses. Proceedings of the National Academy of Sciences. 2004;101(14):4894-9.

5. Nelson PN, Hooley P, Roden D, Davari Ejtehadi H, Rylance P, Warren P, et al. Human endogenous retroviruses: transposable elements with potential? Clinical & Experimental Immunology. 2004;138(1):1-9.

6. Antony JM, DesLauriers AM, Bhat RK, Ellestad KK, Power C. Human endogenous retroviruses and multiple sclerosis: innocent bystanders or disease determinants? Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease. 2011;1812(2):162-76.

7. Armbruester V, Sauter M, Krautkraemer E, Meese E, Kleiman A, Best B, et al. A novel gene from the human endogenous retrovirus K expressed in transformed cells. Clinical Cancer Research. 2002;8(6):1800-7.

8. Grandi N, Tramontano E. HERV envelope proteins: physiological role and pathogenic potential in cancer and autoimmunity. Frontiers in microbiology. 2018;9:462.

9. Büscher K, Hahn S, Hofmann M, Trefzer U, Özel M, Sterry W, et al. Expression of the human endogenous retrovirus-K transmembrane envelope, Rec and Np9 proteins in melanomas and melanoma cell

lines. Melanoma research. 2006;16(3):223-34.

10. Urnovitz HB, Murphy WH. Human endogenous retroviruses: nature, occurrence, and clinical implications in human disease. Clinical microbiology reviews. 1996;9(1):72-99.

11. Medstrand P, Blomberg J. Characterization of novel reverse transcriptase encoding human endogenous retroviral sequences similar to type A and type B retroviruses: differential transcription in normal human tissues. Journal of virology. 1993;67(11):6778-87.

12. Vargiu L, Rodriguez-Tomé P, Sperber GO, Cadeddu M, Grandi N, Blikstad V, et al. Classification and characterization of human endogenous retroviruses; mosaic forms are common. Retrovirology. 2016;13(1):7.

13. Nelson PN, Carnegie P, Martin J, Ejtehadi HD, Hooley P, Roden D, et al. Demystified... Human endogenous retroviruses. Molecular Pathology. 2003;56(1):11.

14. Jern P, Sperber GO, Blomberg J. Use of endogenous retroviral sequences (ERVs) and structural markers for retroviral phylogenetic inference and taxonomy. Retrovirology. 2005;2(1):50.

15. Grandi N, Cadeddu M, Blomberg J, Tramontano E. Contribution of type W human endogenous retroviruses to the human genome: characterization of HERV-W proviral insertions and processed pseudogenes. Retrovirology. 2016;13(1):67.

16. Grandi N, Tramontano E. Human Endogenous Retroviruses Are Ancient Acquired Elements Still Shaping Innate Immune Responses. Frontiers in immunology. 2018;9:2039.

17. Jern P, Coffin JM. Effects of retroviruses on host genome function. Annual review of genetics. 2008;42:709-32.

18. Cohen CJ, Lock WM, Mager DL. Endogenous retroviral LTRs as promoters for human genes: a critical assessment. Gene. 2009;448(2):105-14.

19. Christensen T. Human endogenous retroviruses in neurologic disease. APMIS : acta pathologica, microbiologica, et immunologica Scandinavica. 2016;124(1-2):116-26.

20. Weiss RA. Human endogenous retroviruses: friend or foe? APMIS : acta pathologica, microbiologica, et immunologica Scandinavica. 2016;124(1-2):4-10.

21. Young GR, Stoye JP, Kassiotis G. Are human endogenous retroviruses pathogenic? An approach to testing the hypothesis. BioEssays : news and reviews in molecular, cellular and developmental biology. 2013;35(9):794-803.

22. Young GR, Ploquin MJ, Eksmond U, Wadwa M, Stoye JP, Kassiotis G. Negative selection by an endogenous retrovirus promotes a higher-avidity CD4+ T cell response to retroviral infection. PLoS pathogens. 2012;8(5):e1002709.

23. Conrad B, Weissmahr RN, Boni J, Arcari R, Schupbach J, Mach B. A human endogenous retroviral superantigen as candidate autoimmune gene in type I diabetes. Cell. 1997;90(2):303-13.

24. Johnston JB, Silva C, Holden J, Warren KG, Clark AW, Power C. Monocyte activation and differentiation augment human endogenous retrovirus expression: Implications for inflammatory brain diseases. Annals of Neurology. 2001;50(4):434-42.

25. Christensen T. Human endogenous retroviruses in the aetiology of MS. Acta neurologica Scandinavica. 2017;136 Suppl 201:18-21.

26. Lewis DA. Retroviruses and the pathogenesis of schizophrenia. Proceedings of the National Academy of Sciences of the United States of America. 2001;98(8):4293-4.

27. Frank O, Giehl M, Zheng C, Hehlmann R, Leib-Mosch C,

Seifarth W. Human endogenous retrovirus expression profiles in samples from brains of patients with schizophrenia and bipolar disorders. J Virol. 2005;79(17):10890-901.

28. McCormick AL, Brown RH, Cudkowicz ME, Al-Chalabi A, Garson JA. Quantification of reverse transcriptase in ALS and elimination of a novel retroviral candidate. Neurology. 2008;70(4):278.

29. Volkman HE, Stetson DB. The enemy within: endogenous retroelements and autoimmune disease. Nature immunology. 2014;15(5):415-22.

30. Br, #xfc, tting C, Emmer A, Kornhuber ME, Staege MS. Cooccurrences of Putative Endogenous Retrovirus-Associated Diseases. BioMed Research International. 2017;2017:11.

31. Suntsova M, Garazha A, Ivanova A, Kaminsky D, Zhavoronkov A, Buzdin A. Molecular functions of human endogenous retroviruses in health and disease. Cellular and molecular life sciences : CMLS. 2015;72(19):3653-75.

32. Krishnamurthy J, Rabinovich BA, Mi T, Switzer KC, Olivares S, Maiti SN, et al. Genetic Engineering of T Cells to Target HERV-K, an Ancient Retrovirus on Melanoma. Clinical cancer research : an official journal of the American Association for Cancer Research. 2015;21(14):3241-51.

 Grabski DF, Hu Y, Sharma M, Rasmussen SK. Close to the Bedside: A Systematic Review of Endogenous Retroviruses and Their Impact in Oncology. Journal of Surgical Research. 2019;240:145-55.
 Schmitt K, Reichrath J, Roesch A, Meese E, Mayer J. Transcriptional profiling of human endogenous retrovirus group HERV-K(HML-2) loci in melanoma. Genome biology and evolution. 2013;5(2):307-28.

 Boller K, Janssen O, Schuldes H, Tonjes RR, Kurth R. Characterization of the antibody response specific for the human endogenous retrovirus HTDV/HERV-K. J Virol. 1997;71(6):4581-8.
 Wang-Johanning F, Li M, Esteva FJ, Hess KR, Yin B, Rycaj K, et al. Human endogenous retrovirus type K antibodies and mRNA as serum biomarkers of early-stage breast cancer. International journal of cancer. 2014;134(3):587-95.

37. Arru G, Leoni S, Pugliatti M, Mei A, Serra C, Delogu LG, et al. Natalizumab inhibits the expression of human endogenous retroviruses of the W family in multiple sclerosis patients: a longitudinal cohort study. Multiple sclerosis (Houndmills, Basingstoke, England). 2014;20(2):174-82.

38. Madeira A, Burgelin I, Perron H, Curtin F, Lang AB, Faucard R. MSRV envelope protein is a potent, endogenous and pathogenic agonist of human toll-like receptor 4: Relevance of GNbAC1 in multiple sclerosis treatment. Journal of neuroimmunology. 2016;291:29-38.

39. van Horssen J, van der Pol S, Nijland P, Amor S, Perron H. Human endogenous retrovirus W in brain lesions: Rationale for targeted therapy in multiple sclerosis. Multiple sclerosis and related disorders. 2016;8:11-8.

40. Perron H, Germi R, Bernard C, Garcia-Montojo M, Deluen C, Farinelli L, et al. Human endogenous retrovirus type W envelope expression in blood and brain cells provides new insights into multiple sclerosis disease. Multiple sclerosis (Houndmills, Basingstoke, England). 2012;18(12):1721-36.

41. Antony JM, van Marle G, Opii W, Butterfield DA, Mallet F, Yong VW, et al. Human endogenous retrovirus glycoproteinmediated induction of redox reactants causes oligodendrocyte death and demyelination. Nature neuroscience. 2004;7(10):1088-95.

42. Mameli G, Astone V, Arru G, Marconi S, Lovato L, Serra C, et al. Brains and peripheral blood mononuclear cells of multiple sclerosis (MS) patients hyperexpress MS-associated retrovirus/HERV-W endogenous retrovirus, but not Human herpesvirus 6. The Journal of general virology. 2007;88(Pt 1):264-74.

43. Morandi E, Tanasescu R, Tarlinton RE, Constantin-Teodosiu D, Gran B. Do Antiretroviral Drugs Protect From Multiple Sclerosis by Inhibiting Expression of MS-Associated Retrovirus? Frontiers in immunology. 2019;9:3092-.

44. Arru G, Sechi E, Mariotto S, Farinazzo A, Mancinelli C, Alberti D, et al. Antibody response against HERV-W env surface peptides differentiates multiple sclerosis and neuromyelitis optica spectrum disorder. Multiple sclerosis journal - experimental, translational and clinical. 2017;3(4):2055217317742425-.

45. Perron H, Lazarini F, Ruprecht K, Pechoux-Longin C, Seilhean D, Sazdovitch V, et al. Human endogenous retrovirus (HERV)-W ENV and GAG proteins: physiological expression in human brain and pathophysiological modulation in multiple sclerosis lesions. Journal of neurovirology. 2005;11(1):23-33.

46. Zawada M, Liwien I, Pernak M, Januszkiewicz-Lewandowska D, Nowicka-Kujawska K, Rembowska J, et al. MSRV pol sequence copy number as a potential marker of multiple sclerosis. Polish journal of pharmacology. 2003;55(5):869-75.

47. Antony JM, Ellestad KK, Hammond R, Imaizumi K, Mallet F, Warren KG, et al. The human endogenous retrovirus envelope glycoprotein, syncytin-1, regulates neuroinflammation and its receptor expression in multiple sclerosis: a role for endoplasmic reticulum chaperones in astrocytes. Journal of immunology (Baltimore, Md : 1950). 2007;179(2):1210-24.

48. Nexø BA, Villesen P, Nissen KK, Lindegaard HM, Rossing P, Petersen T, et al. Are human endogenous retroviruses triggers of autoimmune diseases? Unveiling associations of three diseases and viral loci. Immunologic research. 2016;64(1):55-63.

49. Huang W, Li S, Hu Y, Yu H, Luo F, Zhang Q, et al. Implication of the env gene of the human endogenous retrovirus W family in the expression of BDNF and DRD3 and development of recent-onset schizophrenia. Schizophrenia bulletin. 2011;37(5):988-1000.

50. Tai AK, O'Reilly EJ, Alroy KA, Simon KC, Munger KL, Huber BT, et al. Human endogenous retrovirus-K18 Env as a risk factor in multiple sclerosis. Multiple sclerosis (Houndmills, Basingstoke, England). 2008;14(9):1175-80.

51. Rasmussen HB, Geny C, Deforges L, Perron H, Tourtelotte W, Heltberg A, et al. Expression of endogenous retroviruses in blood mononuclear cells and brain tissue from multiple sclerosis patients. Multiple Sclerosis Journal. 1995;1(2):82-7.

52. Christensen T, Dissing Sorensen P, Riemann H, Hansen HJ, Munch M, Haahr S, et al. Molecular characterization of HERV-H variants associated with multiple sclerosis. Acta neurologica Scandinavica. 2000;101(4):229-38.

53. Bhetariya PJ, Kriesel JD, Fischer KF. Analysis of Human Endogenous Retrovirus Expression in Multiple Sclerosis Plaques. Journal of emerging diseases and virology. 2017;3(2).

54. Christensen T, Petersen T, Thiel S, Brudek T, Ellermann-Eriksen S, Moller-Larsen A. Gene-environment interactions in multiple sclerosis: innate and adaptive immune responses to human

endogenous retrovirus and herpesvirus antigens and the lectin complement activation pathway. Journal of neuroimmunology. 2007;183(1-2):175-88.

55. Nissen KK, Laska MJ, Hansen B, Pedersen FS, Nexo BA. No additional copies of HERV-Fc1 in the germ line of multiple sclerosis patients. Virology journal. 2012;9:188.

56. Brudek T, Christensen T, Aagaard L, Petersen T, Hansen HJ, Moller-Larsen A. B cells and monocytes from patients with active multiple sclerosis exhibit increased surface expression of both HERV-H Env and HERV-W Env, accompanied by increased seroreactivity. Retrovirology. 2009;6:104.

57. Freimanis G, Hooley P, Ejtehadi HD, Ali HA, Veitch A, Rylance PB, et al. A role for human endogenous retrovirus-K (HML-2) in rheumatoid arthritis: investigating mechanisms of pathogenesis. Clinical and experimental immunology. 2010;160(3):340-7.

58. Seidl C, Donner H, Petershofen E, Usadel KH, Seifried E, Kaltwasser JP, et al. An endogenous retroviral long terminal repeat at the HLA-DQB1 gene locus confers susceptibility to rheumatoid arthritis. Human Immunology. 1999;60(1):63-8.

59. Reynier F, Verjat T, Turrel F, Imbert PE, Marotte H, Mougin B, et al. Increase in human endogenous retrovirus HERV-K (HML-2) viral load in active rheumatoid arthritis. Scandinavian journal of immunology. 2009;70(3):295-9.

60. Nelson PN, Lever AM, Smith S, Pitman R, Murray P, Perera SA, et al. Molecular investigations implicate human endogenous retroviruses as mediators of anti-retroviral antibodies in autoimmune rheumatic disease. Immunological investigations. 1999;28(4):277-89.
61. Nakkuntod J, Sukkapan P, Avihingsanon Y, Mutirangura A, Hirankarn N. DNA methylation of human endogenous retrovirus in systemic lupus erythematosus. Journal of human genetics. 2013;58(5):241-9.

62. Mameli G, Erre GL, Caggiu E, Mura S, Cossu D, Bo M, et al. Identification of a HERV-K env surface peptide highly recognized in Rheumatoid Arthritis (RA) patients: a cross-sectional case-control study. Clin Exp Immunol. 2017;189(1):127-31.

63. Krzysztalowska-Wawrzyniak M, Ostanek M, Clark J, Binczak-Kuleta A, Ostanek L, Kaczmarczyk M, et al. The distribution of human endogenous retrovirus K-113 in health and autoimmune diseases in Poland. Rheumatology (Oxford, England). 2011;50(7):1310-4.

64. Gaudin P, Ijaz S, Tuke PW, Marcel F, Paraz A, Seigneurin JM, et al. Infrequency of detection of particle-associated MSRV/HERV-W RNA in the synovial fluid of patients with rheumatoid arthritis. Rheumatology (Oxford, England). 2000;39(9):950-4.

65. Ejtehadi HD, Freimanis GL, Ali HA, Bowman S, Alavi A, Axford J, et al. The potential role of human endogenous retrovirus K10 in the pathogenesis of rheumatoid arthritis: a preliminary study. Annals of the rheumatic diseases. 2006;65(5):612-6.

66. Takeuchi K, Katsumata K, Ikeda H, Minami M, Wakisaka A, Yoshiki T. Expression of endogenous retroviruses, ERV3 and lambda 4-1, in synovial tissues from patients with rheumatoid arthritis. Clin Exp Immunol. 1995;99(3):338-44.

67. Brookes SM, Pandolfino YA, Mitchell TJ, Venables PJ, Shattles WG, Clark DA, et al. The immune response to and expression of cross-reactive retroviral gag sequences in autoimmune disease. British journal of rheumatology. 1992;31(11):735-42.

68. Nakagawa K, Brusic V, McColl G, Harrison LC. Direct evidence

for the expression of multiple endogenous retroviruses in the synovial compartment in rheumatoid arthritis. Arthritis and rheumatism. 1997;40(4):627-38.

69. Herve CA, Lugli EB, Brand A, Griffiths DJ, Venables PJ. Autoantibodies to human endogenous retrovirus-K are frequently detected in health and disease and react with multiple epitopes. Clin Exp Immunol. 2002;128(1):75-82.

70. Ramos-Lopez E, Ghebru S, Van Autreve J, Aminkeng F, Herwig J, Seifried E, et al. Neither an intronic CA repeat within the CD48 gene nor the HERV-K18 polymorphisms are associated with type 1 diabetes. Tissue antigens. 2006;68(2):147-52.

71. Lan M, Mason A, Coutant R, Chen Q-Y, Vargas A, Rao J, et al. HERV-K10s and immune-mediated (type 1) diabetes1998. 14-6; discussion 6 p.

72. Pani MA, Wood JP, Bieda K, Toenjes RR, Usadel KH, Badenhoop K. The variable endogenous retroviral insertion in the human complement C4 gene: a transmission study in type I diabetes mellitus. Hum Immunol. 2002;63(6):481-4.

73. Laska MJ, Troldborg A, Hauge EM, Bahrami S, Stengaard-Pedersen K. Human Endogenous Retroviral Genetic Element With Immunosuppressive Activity in Both Human Autoimmune Diseases and Experimental Arthritis. Arthritis & rheumatology (Hoboken, NJ). 2017;69(2):398-409.

74. Pullmann R, Jr., Bonilla E, Phillips PE, Middleton FA, Perl A. Haplotypes of the HRES-1 endogenous retrovirus are associated with development and disease manifestations of systemic lupus erythematosus. Arthritis and rheumatism. 2008;58(2):532-40.

75. Perl A, Colombo E, Dai H, Agarwal R, Mark KA, Banki K, et al. Antibody reactivity to the HRES-1 endogenous retroviral element identifies a subset of patients with systemic lupus erythematosus and overlap syndromes. Correlation with antinuclear antibodies and HLA class II alleles. Arthritis and rheumatism. 1995;38(11):1660-71.

76. Hishikawa T, Ogasawara H, Kaneko H, Shirasawa T, Matsuura Y, Sekigawa I, et al. Detection of antibodies to a recombinant gag protein derived from human endogenous retrovirus clone 4-1 in autoimmune diseases. Viral immunology. 1997;10(3):137-47.

77. Bengtsson A, Blomberg J, Nived O, Pipkorn R, Toth L, Sturfel G. Selective antibody reactivity with peptides from human endogenous retroviruses and nonviral poly(amino acids) in patients with systemic lupus erythematosus. Arthritis & Rheumatism. 1996;39(10):1654-63. 78. Herve CA, Lugli EB, Brand A, Griffiths DJ, Venables PJW. Autoantibodies to human endogenous retrovirus-K are frequently detected in health and disease and react with multiple epitopes. Clinical and experimental immunology. 2002;128(1):75-82.

79. Magistrelli C, Samoilova E, K. Agarwal R, Banki K, Ferrante P, Vladutiu A, et al. Polymorphic genotypes of the HRES-1 human endogenous retrovirus locus correlate with systemic lupus erythematosus and autoreactivity1999. 829-34 p.

80. Faucard R, Madeira A, Gehin N, Authier F-J, Panaite P-A, Lesage C, et al. Human Endogenous Retrovirus and Neuroinflammation in Chronic Inflammatory Demyelinating Polyradiculoneuropathy. EBioMedicine. 2016;6:190-8.

81. Perron H, Dougier-Reynaud H-L, Lomparski C, Popa I, Firouzi R, Bertrand J-B, et al. Human endogenous retrovirus protein activates innate immunity and promotes experimental allergic encephalomyelitis in mice. PloS one. 2013;8(12):e80128-e.

82. Shattles WG, Brookes SM, Venables PJ, Clark DA, Maini RN.

Expression of antigen reactive with a monoclonal antibody to HTLV-1 P19 in salivary glands in Sjögren's syndrome. Clinical and experimental immunology. 1992;89(1):46-51.

83. Moles JP, Tesniere A, Guilhou JJ. A new endogenous retroviral sequence is expressed in skin of patients with psoriasis. The British journal of dermatology. 2005;153(1):83-9.

84. Lai OY, Chen H, Michaud H-A, Hayashi G, Kuebler PJ, Hultman GK, et al. Protective effect of human endogenous retrovirus K dUTPase variants on psoriasis susceptibility. The Journal of investigative dermatology. 2012;132(7):1833-40.

85. Pani MA, Seidl C, Bieda K, Seissler J, Krause M, Seifried E, et al. Preliminary evidence that an endogenous retroviral long-terminal repeat (LTR13) at the HLA-DQB1 gene locus confers susceptibility to Addison's disease. Clinical Endocrinology. 2002;56(6):773-7.

86. Yao Y, Schroder J, Nellaker C, Bottmer C, Bachmann S, Yolken RH, et al. Elevated levels of human endogenous retrovirus-W transcripts in blood cells from patients with first episode schizophrenia. Genes, brain, and behavior. 2008;7(1):103-12.

87. Weis S, Llenos IC, Sabunciyan S, Dulay JR, Isler L, Yolken R, et al. Reduced expression of human endogenous retrovirus (HERV)-W GAG protein in the cingulate gyrus and hippocampus in schizophrenia, bipolar disorder, and depression. Journal of neural transmission (Vienna, Austria : 1996). 2007;114(5):645-55.

88. Suntsova M, Gogvadze EV, Salozhin S, Gaifullin N, Eroshkin F, Dmitriev SE, et al. Human-specific endogenous retroviral insert serves as an enhancer for the schizophrenia-linked gene PRODH. Proceedings of the National Academy of Sciences of the United States of America. 2013;110(48):19472-7.

89. Perron H, Mekaoui L, Bernard C, Veas F, Stefas I, Leboyer M. Endogenous Retrovirus Type W GAG and Envelope Protein Antigenemia in Serum of Schizophrenic Patients. Biological Psychiatry. 2008;64(12):1019-23.

90. Otowa T, Tochigi M, Rogers M, Umekage T, Kato N, Sasaki T. Insertional polymorphism of endogenous retrovirus HERV-K115 in schizophrenia. Neuroscience Letters. 2006;408(3):226-9.

91. Mak M, Samochowiec J, Frydecka D, Pelka-Wysiecka J, Szmida E, Karpinski P, et al. First-episode schizophrenia is associated with a reduction of HERV-K methylation in peripheral blood. Psychiatry research. 2019;271:459-63.

92. Karlsson H, Schroder J, Bachmann S, Bottmer C, Yolken RH. HERV-W-related RNA detected in plasma from individuals with recent-onset schizophrenia or schizoaffective disorder. Molecular psychiatry. 2004;9(1):12-3.

93. Huang W, Li S, Hu Y, Yu H, Luo F, Zhang Q, et al. Implication of the env gene of the human endogenous retrovirus W family in the expression of BDNF and DRD3 and development of recent-onset schizophrenia. Schizophrenia bulletin. 2011;37(5):988-1000.

94. Dickerson F, Rubalcaba E, Viscidi R, Yang S, Stallings C, Sullens A, et al. Polymorphisms in human endogenous retrovirus K-18 and risk of type 2 diabetes in individuals with schizophrenia. Schizophrenia research. 2008;104(1-3):121-6.

95. Ishida T, Obata Y, Ohara N, Matsushita H, Sato S, Uenaka A, et al. Identification of the HERV-K gag antigen in prostate cancer by SEREX using autologous patient serum and its immunogenicity. Cancer immunity. 2008;8:15-.

96. Agoni L, Guha C, Lenz J. Detection of Human Endogenous Retrovirus K (HERV-K) Transcripts in Human Prostate Cancer Cell

Lines. Frontiers in oncology. 2013;3:180-.

97. Goering W, Schmitt K, Dostert M, Schaal H, Deenen R, Mayer J, et al. Human endogenous retrovirus HERV-K(HML-2) activity in prostate cancer is dominated by a few loci. The Prostate. 2015;75(16):1958-71.

98. Wang-Johanning F, Frost AR, Jian B, Azerou R, Lu DW, Chen DT, et al. Detecting the expression of human endogenous retrovirus E envelope transcripts in human prostate adenocarcinoma. Cancer. 2003;98(1):187-97.

99. Argaw-Denboba A, Balestrieri E, Serafino A, Cipriani C, Bucci I, Sorrentino R, et al. HERV-K activation is strictly required to sustain CD133+ melanoma cells with stemness features. Journal of experimental & clinical cancer research : CR. 2017;36(1):20.

100. Huang G, Li Z, Wan X, Wang Y, Dong J. Human endogenous retroviral K element encodes fusogenic activity in melanoma cells. Journal of carcinogenesis. 2013;12:5-.

101. Hahn S, Ugurel S, Hanschmann KM, Strobel H, Tondera C, Schadendorf D, et al. Serological response to human endogenous retrovirus K in melanoma patients correlates with survival probability. AIDS research and human retroviruses. 2008;24(5):717-23.

102. Oricchio E, Sciamanna I, Beraldi R, Tolstonog GV, Schumann GG, Spadafora C. Distinct roles for LINE-1 and HERV-K retroelements in cell proliferation, differentiation and tumor progression. Oncogene. 2007;26(29):4226-33.

103. Li Z, Sheng T, Wan X, Liu T, Wu H, Dong J. Expression of HERV-K correlates with status of MEK-ERK and p16INK4A-CDK4 pathways in melanoma cells. Cancer investigation. 2010;28(10):1031-7.

104. Pothlichet J, Mangeney M, Heidmann T. Mobility and integration sites of a murine C57BL/6 melanoma endogenous retrovirus involved in tumor progression in vivo. Int J Cancer. 2006;119(8):1869-77.

105. Schanab O, Humer J, Gleiss A, Mikula M, Sturlan S, Grunt S, et al. Expression of human endogenous retrovirus K is stimulated by ultraviolet radiation in melanoma. Pigment cell & melanoma research. 2011;24(4):656-65.

106. Serafino A, Balestrieri E, Pierimarchi P, Matteucci C, Moroni G, Oricchio E, et al. The activation of human endogenous retrovirus K (HERV-K) is implicated in melanoma cell malignant

transformation. Experimental cell research. 2009;315(5):849-62.

107. Stengel S, Fiebig U, Kurth R, Denner J. Regulation of human endogenous retrovirus-K expression in melanomas by CpG methylation. Genes, chromosomes & cancer. 2010;49(5):401-11.

108. Rycaj K, Plummer JB, Yin B, Li M, Garza J, Radvanyi L, et al. Cytotoxicity of human endogenous retrovirus K-specific T cells toward autologous ovarian cancer cells. Clin Cancer Res. 2015;21(2):471-83.

109. Iramaneerat K, Rattanatunyong P, Khemapech N, Triratanachat S, Mutirangura A. HERV-K hypomethylation in ovarian clear cell carcinoma is associated with a poor prognosis and platinum resistance. International journal of gynecological cancer : official journal of the International Gynecological Cancer Society. 2011;21(1):51-7.

110. Kahyo T, Tao H, Shinmura K, Yamada H, Mori H, Funai K, et al. Identification and association study with lung cancer for novel insertion polymorphisms of human endogenous retrovirus. Carcinogenesis. 2013;34(11):2531-8.

111. Li M, Radvanyi L, Yin B, Li J, Chivukula R, Lin K, et al. Downregulation of Human Endogenous Retrovirus Type K (HERV-K) Viral env RNA in Pancreatic Cancer Cells Decreases Cell Proliferation and Tumor Growth. Clinical cancer research : an official journal of the American Association for Cancer Research. 2017;23(19):5892-911.

112. Liang Q, Xu Z, Xu R, Wu L, Zheng S. Expression patterns of non-coding spliced transcripts from human endogenous retrovirus HERV-H elements in colon cancer. PloS one. 2012;7(1):e29950-e.

113. Ma W, Hong Z, Liu H, Chen X, Ding L, Liu Z, et al. Human Endogenous Retroviruses-K (HML-2) Expression Is Correlated with Prognosis and Progress of Hepatocellular Carcinoma. BioMed Research International. 2016;2016:9.

114. Benešová M, Trejbalová K, Kovářová D, Vernerová Z, Hron T, Kučerová D, et al. DNA hypomethylation and aberrant expression of the human endogenous retrovirus ERVWE1/syncytin-1 in seminomas. Retrovirology. 2017;14(1):20-.

115. Gosenca D, Gabriel U, Steidler A, Mayer J, Diem O, Erben P, et al. HERV-E-Mediated Modulation of PLA2G4A Transcription in Urothelial Carcinoma. PLOS ONE. 2012;7(11):e49341.