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Evaluation of Aquaporins 1 and 5 Expression in Rat Parotid Glands After Volumetric Modulated Arc Radiotherapy and Use of Low-Level Laser Therapy at Different Times

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Abstract

Introduction: This experimental study investigated the mRNA expression of aquaporins (AQPs) 1 and 5 in the parotid glands of rats irradiated with volumetric modulated arc therapy (VMAT) and subjected to low-level laser therapy (LLLT) at different time points.

Methods: The sample consisted of 30 Wistar rats (Rattus norvegicus) divided into the following groups: control, LLLT alone (LG), radiotherapy alone (RG), and experimental groups that received LLLT at 24 hours (early experimental group [EEG], n = 12) and 120 hours (late experimental group [LEG], n = 12) after radiotherapy. VMAT was delivered at a single dose (12 Gy) and LLLT was performed with an aluminium-gallium-arsenide diode laser (660 nm, 100 mW), spot area of 0.0028 cm2, energy of 2 J/cm² applied to 3 spots in the region corresponding to the right parotid gland, for 10 consecutive days. The right parotid gland was resected and prepared for RNA extraction. The gene expression of AQPs was evaluated by quantitative polymerase chain reaction (qPCR) using specific TaqMan probes, with the HPRT gene as an internal control.

Results: The lowest AQP1 gene expression was 0.83 (0.27) with the use of LLLT 24 hours after radiotherapy (EEG), and the highest was 1.56 (0.80) with the use of LLLT alone (LG). Likewise, the lowest AQP5 gene expression was found in the EEG (mean = 0.88; SD = 0.49) and the highest in the LG (mean = 1.29; SD = 0.33).

Conclusion: The use of LLLT after radiotherapy may contribute to the maintenance and an increase of these proteins, even when used at a later time point after radiotherapy.

Keywords: Aquaporins; Salivary glands; Intensity-modulated radiotherapy; Low-level light therapy; Rats.



Introduction

Xerostomia due to radiotherapy-induced hyposalivation, one of the main clinical complications of cancer therapy, is caused by the action of ionising radiation on cellular structures of the salivary glands.^{1,2} Because the salivary glands are in the radiation field in most cases of treatment of head and neck cancer, radiation causes atrophy and apoptosis of fluid-secreting acinar cells.³⁻⁵ Volumetric modulated arc therapy (VMAT) has shown more effective results and less damage to the normal tissue than other radiotherapy modalities, with the dose and frequency delivered in fractions and directed to the target tissue.^{6,7} Among the treatment modalities used to improve the sequelae of radiotherapy to the salivary glands, lowlevel laser therapy (LLLT) has produced favourable results in terms of tissue repair and biomodulation of the inflammatory process,⁸⁻¹³ especially in clinical and histological studies evaluating the effects of LLLT on the salivary glands in the ionising radiation field.¹⁴⁻¹⁷

Damage caused by radiotherapy to the stroma and structures involved in the process of salivary secretion has a direct impact on salivary production and flow. Water transport by the acinar cells for primary production of saliva occurs through specific proteins, namely aquaporins (AQPs). AQPs form a family of proteins with 13 different molecular structures, each of which is present in a particular tissue type and performs specific functions associated with water and solute transport.¹⁸⁻²⁰ In addition

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to being involved in the processes of cell proliferation, migration, and apoptosis, these transmembrane proteins consist of water channels present in the cell membrane of several tissues, contributing to transepithelial water movement.²¹⁻²³

In the major salivary glands, AQP1 and AQP5 are strongly associated with water transport and fluid secretion; therefore, the functional and structural integrity of these proteins must be preserved.^{9,24} AQPs present in the membrane of the acinar cells and duct cells may be affected by radiotherapy, thus modifying the dynamics of the salivary secretion process.²⁵

Previous studies in animal models investigating the mechanisms of action and participation of AQPs in the process of salivary secretion in relation to the physiology^{18,26} and maintenance of these proteins in the salivary glands after cobalt 60 (Co⁶⁰) radiotherapy²⁵ have demonstrated the down-regulation of AQP expression. However, the effects of using VMAT radiotherapy and LLLT on the expression of AQPs in the major salivary glands have not yet been reported, especially regarding the potential action of LLLT on the synthesis of nucleic acids and proteins, which may lead to increased expression of AQPs in the plasma membrane, thereby facilitating water permeability during the process of salivary flow.

Therefore, this study aimed to investigate AQP1 and AQP5 mRNA expression in the parotid glands of rats irradiated with VMAT and subjected to LLLT at early (24 hours) and late (120 hours) time points after radiotherapy.

Methods

This experimental study in an animal model is part of a project approved by the Animal Care and Use Committee of Universidade Luterana do Brasil (approval number 2014-10P). Animal handling and experimentation followed the Brazilian ethical principles of animal experimentation and international standards and guidelines for the care and use of laboratory animals. All efforts were made to minimise pain and discomfort, as well as to use only the minimum number of animals required to produce reliable scientific data.

The sample consisted of the right parotid glands of 30 adult (7- to 8-week-old) male Wistar rats (*Rattus norvegicus*) weighing 240 to 300 g each. The rats were randomly divided into five groups as shown in Figure 1.

The groups were divided into one control group that received no intervention (CG), one that received radiotherapy alone (RG), one that received LLLT alone



Figure 1. Flow Diagram of Group Distribution and Interventions.

Legend: LLLT (low-level laser therapy); CG (control group); LG (laser group); RG (radiotherapy group); EEG (early experimental group); LEG (late experimental group); CG (control group); LG (laser group); RG (radiotherapy group); EEG (early experimental group); LEG (late experime

(LG), and two experimental groups that received LLLT at 24 hours (early experimental group [EEG]) and 120 hours (late experimental group [LEG]) after radiotherapy. One animal in the EEG was lost 48 hours after radiotherapy, resulting in a total sample of 29 parotid glands for analysis.

Animals in the groups receiving radiotherapy (RG, EEG, and LEG) were subjected to VMAT delivered with a Trilogy linear accelerator (Varian Medical Systems, Palo Alto, CA, USA) at a single dose of 12 Gy with 6 MV photons and a radiation field of 9×40 cm², at 1 cm in diameter from the head region (Figure 2a).

Animals in the groups receiving LLLT (LG, EEG, and LEG) were subjected to the same laser irradiation protocol. At a distance of 1 cm from the external area corresponding to the right parotid gland (Figure 2b), the laser was applied to three spots with an aluminium-gallium-arsenide diode laser (Photon Lase III DMC Ltda) at a wavelength of 660 nm,²⁷ 100 mW output power, spot area of 0.0028 cm², and energy density of 2 J/cm² over an irradiation time of 20 seconds per spot, for a total of 60 seconds and 70 J/cm²/d for 10 consecutive days.

Rats were euthanised by intraperitoneal administration of barbiturate and local anaesthetic (thiopental and lidocaine) at a concentration of 80 mg/kg at different time points, according to the distribution of the experimental groups (Figure 1), always 24 hours after the last LLLT application. In the CG, LG, and RG, euthanasia was performed at the same time as in the EEG.

Subsequently, the right parotid gland was resected and stored in a tube containing RNA stabilisation solution (RNAlater; Ambion, Applied Biosystems, Carlsbad, CA, USA) for later analysis by quantitative polymerase chain reaction (qPCR) in a thermal cycler (StepOnePlus[™] Real-Time PCR System; Life Technologies, Carlsbad, CA, USA). DNA was extracted from the cells according to the conventional silica-based process after individually grinding each sample.²⁸

The DNA fragments were amplified by qPCR using probes specific to AQPs (Integrated DNA Technologies Inc., Coralville, IA, USA) under the following conditions: 95°C for 3 minutes, then 40 cycles of 95°C for 15 seconds and 60°C for 1 minute. The hypoxanthine



Figure 2. (a) Rat positioning for radiotherapy protocol with VMAT using a Trilogy linear accelerator. (b) Low-level laser irradiation protocol with the laser applied to three spots at a distance of 1 cm from the area corresponding to the right parotid gland.

phosphoribosyltransferase (HPRT) gene was used as an internal control.

The expression of mRNA was analysed by using kits with RNA-specific primers. The samples were rotated 40 times to obtain a higher number of cycles and to improve data accuracy.⁵ All qPCR procedures were performed by a single examiner with experience in the method, who was blinded to group assignment (the groups were coded for analysis).

The 2^{- $\Delta\Delta$ Ct} algorithm was used to analyse the data obtained from AQP1 and AQP5 expression assays in all groups for quantification of the mean relative expression levels per group, which were then compared to the control group (CG). The Mann-Whitney U test was used to analyse the results, and a *P* value < 0.05 was considered significant.

Results

The analysis of the mRNA expression of AQP1 and AQP5 by qPCR showed a mean value of 1.0 for both types in the CG, which was used as a reference value for comparisons with the other groups (Table 1).

The lowest AQP1 gene expression was 0.83 (0.27) with the use of LLLT 24 hours after radiotherapy (EEG), and the highest was 1.56 (0.80) with the use of LLLT alone (LG). The highest expression of AQP1 was in the groups using LLLT alone (LG) and late use after radiotherapy (LEG).

Likewise, the lowest AQP5 gene expression was found in the EEG (mean = 0.88; SD = 0.49), and the highest in the LG (mean = 1.29; SD = 0.33) (Table 1). In the comparison between the groups, the average expression of AQP5 was higher in the LG group which used only LLLT. In summary, the mean values obtained for the mRNA expression of AQP1 and AQP5 showed little variation among the treatments, with no statistically significant difference between the groups.

Discussion

AQP1 and AQP5 were expressed in all samples of parotid glands, regardless of group assignment. However, AQP1 showed the highest mean expression levels, especially in the samples from the groups that received LLLT, whether with or without previous radiotherapy. Mean AQP5 expression was highest in the group that received LLLT alone, however, without statistical significance. Studies evaluating gene expression by qPCR after the use of LLLT have shown conflicting results regarding its effect.²⁹

These results may be related to the heterogeneity of sample composition, given that the samples included the entire anatomical structure of the salivary glands rather than only the sites where the proteins were located, and also to the preparation of samples for qPCR, which were ground without separating the acini from the ducts. This may have contributed to the different mean expression

Groups	AQP1 Expression Mean (SD)	Р	AQP5 Expression Mean (SD)	Р
RG	0.98 (0.27)	0.922	1.16 (0.54)	0.715
LG	1.56 (0.80)	0.363	1.29 (0.33)	0.102
EEG	0.83 (0.27)	0.423	0.88 (0.49)	0.553
LEG	1.11 (0.31)	0.608	0.90 (0.37)	0.667

Table 1. Distribution of the Mean (SD) mRNA Expression of AQP1 and AQP5 in the Different Groups Compared to the Control Group (CG)

RG, Radiotherapy group; LG, LLLT group; EEG, Early experimental group; LEG, Late experimental group.

values observed among the groups and represented by their respective standard deviations. However, studies have shown that AQP1 expression in the salivary glands is higher in ductal cells than in myoepithelial cells, while AQP5 expression is higher in the basal cells of the acini at different sites.^{25,26,30-33}

The groups receiving radiotherapy did not differ in AQP expression from the control group in contrast to previous reports that these proteins are radiosensitive, especially AQP5.²⁵ However, the radiotherapy modality used should be taken into consideration. For instance, conventional Co⁶⁰ radiotherapy is often associated with greater tissue damage and less selectivity than VMAT, the modality used in the present study. Also, in the study conducted by Li et al,²⁵ qPCR performed on days 3 and 30 after radiotherapy showed a progressive decrease in AQP5 expression levels. This is different from our results, which showed that mean AQP5 expression levels remained superior in the group that received LLLT at a later time point compared to the control group.

The group receiving LLLT alone had the highest mean AQP expression values. This result may be associated with the different actions of laser irradiation on the salivary glands. Simões et al,³³ evaluating the effects of the infrared diode laser, found an increase in protein synthesis in the parotid and submandibular glands of healthy rats compared to the control group. This may be attributed to photobiological reactions induced by LLLT on glandular tissues, which may occur in the presence or absence of light. In the latter, this may occur days after laser irradiation, leading to biochemical changes in the cells that modify the mitochondrial respiratory chain and activate intracellular signalling, which increases DNA and RNA synthesis, thereby increasing cell proliferation and protein synthesis.³⁴⁻³⁶

The group receiving LLLT 24 hours after radiotherapy had lower mean AQP expression values than the group receiving LLLT at a later time point. This may be related to a cellular stress adaptive response caused by the immediate use of the laser, without sufficient time for adequate tissue repair. Conversely, if a longer time is allowed for tissue repair, the subsequent use of LLLT can lead to the modulation of the inflammatory process and activation of transcription factors, which promote the growth of fibroblasts, pro-inflammatory cytokines, and chemokines involved in tissue repair.^{14,33,36-38}

This study has some limitations that need to be

addressed. A healthy animal model was used, without simulation or presence of malignancy, which appears to have promoted a selective effect, or even a protective effect, of VMAT. This radiotherapy modality is able to minimise frequency, dose, and toxicity to the normal tissue by being directed to the neoplastic cells.^{6,7} Also, we were unable to perform a qualitative analysis of the cells and gland parenchyma, thus precluding the determination of the exact location of AQPs in the parotid glands. The method used for obtaining the samples required grinding of the tissues to perform qPCR, which is a quantitative method for mRNA analysis. Therefore, further studies using other types of analysis are still needed to determine the location and structure of AQPs in order to clarify how LLLT acts on the salivary glands after radiotherapy. The use of immunohistochemistry seems to be an effective option for evaluating the location and structure of AQPs in the salivary glands.^{25,39,40-42}

To date, no studies have been published reporting the expression of AQP1 and AQP5 in salivary glands (without separating the acini from the ducts) irradiated with VMAT and subjected to LLLT at different time points. Therefore, the present results are expected to provide a basis for future studies analysing AQPs in the salivary glands that are in the radiation field using different radiotherapy modalities, doses and LLLT protocols, in an attempt to establish an effective therapy for the management of xerostomia due to radiotherapyinduced hyposalivation.

Conclusion

The use of LLLT seems to contribute even to an increase of AQP1 and AQP5 in the rat's parotid gland even when used at a later time point after radiotherapy for VMAT. The results suggest that the use of LLLT contributes to the maintenance of the salivary secretion process.

Ethical Considerations

This study has been approved by the Animal Care and Use Committee of Universidade Luterana do Brasil – RS/ Brazil (CEUA/ULBRA; approval number 2014-10P).

Conflict of Interests

The authors declare that they have no conflict of interest.

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