Original Article

Seroprevalence of *Toxocara* infection among healthy individuals referred to the medical center laboratories in Tehran City, Capital of Iran

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Abstract

Background: Toxocarosis is a zoonotic disease with worldwide distribution. Humans' infection occurred by incidental ingestion of eggs shed in feces of dogs or cats. Studies on general population are rare in Iran. In this cross-sectional study, we investigated seroprevalence, and risk factors associated with toxocariasis among the healthy individuals in Tehran, capital if Iran.

Materials and Methods: In total, 374 sera samples were investigated for the presence of anti-Toxocara IgG. We applied ELISA as screening test using available commercial kit. In addition, demographic data were obtained from participant's questionnaires. Data analysis was performed using $SPSS_{16}$.

Results: The overall seroprevalence of toxocariasis was found 5.6% (21/374). Regarding the sociodemographic variables, age (P<0.001) and eating unwashed vegetables (P=0.049) were significantly associated to toxocariasis in univariate analysis. In the logistic regression analysis, only age (P<0.001) was identified as potential risk factor associated with Toxocara infection.

Conclusion: This study revealed that seroprevalence of toxocariasis is relatively low in the healthy individuals in Tehran. We suggest carrying out further studies in the different part of Iran and investigate on the prevalence of toxocariasis in high-risk groups such as asthma, hyper-eosinophilic, epilepsy, rheumatism and schizophrenia patients.

Keywords: Toxocariasis, Seroprevalence, Tehran, Iran

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Introduction

Toxocara cati and *Toxocara canis* parasites cause a zoonotic disease known as toxocariasis with a worldwide distribution, particularly in communities with low levels of sanitation and hygiene¹. Both of the causative agents belong to nematodes and inhabit in the intestine of their definitive hosts, including

dogs and cats². It was estimated that 2 billion people are at risk of the infection, and because of the global with human animals' warming, along and immigration, the geographical distribution of toxocariasis is expanding³. Humans are the accidental host of the Toxocara spp., and ingestion of ova-contaminated embryonated soil/water and vegetables or raw infected larvae-contaminated animal tissue are main routes of infection⁴⁻⁶. The risk of infection is higher in the first decade of life, due to more geophagic behaviors in children⁷.

Most of the infected humans are asymptomatic and probably its morbidity depends on parasite burden and immune response of the host⁸. Symptomatic toxocariasis is classified into three main forms, including visceral larva migrans (VLM), ocular larva migrans (OLM), and neurological toxocariasis (NT). The occurrence of any form depends on organs affected. It should be mentioned that some other helminth larvae can migrate in the human body and result in VLM⁹⁻¹¹. The common clinical manifestations of toxocariasis may include dyspnea, cough, chest discomfort asthma, skin itching, or gastrointestinal disorders¹².

The adult stage of the *Toxocara* spp. is not present in the human body, so microscopic examination of stool for ova is not possible. Biopsy is the gold standard for diagnosis of human toxocariasis, although it is an extremely difficult method, so serological methods such as enzyme-linked immunosorbent assay (ELISA) and indirect fluorescent antibody test (IFAT) are the most common¹³.

A number of seroprevalence studies on toxocariasis were conducted in Iran, mostly focused on high-risk groups like children¹⁴. Tehran is the biggest and most populous city in Iran. To the best of our knowledge, none of the previous studies have been carried out to evaluate the epidemiology of toxocariasis in Tehran city. In addition, since many stray dogs and cats are widely distributed in around of city and public parks, Tehran can be a neglected endemic area for toxocariasis. The objectives of the present study were a better understanding of prevalence of *Toxocara* spp. infection and determination of toxocariasis associated potential risk factors in this area.

Methods

Study area and study population: This crosssectional study was conducted at the Department of Medical Parasitology, Shahid Beheshti University of Medical sciences (SBMU), Tehran, Iran. The Ethics Committee of the SBMU (grant No 1394-263), approved this research. The Study involved subjects were healthy individuals referred to medical laboratories for routine health checkup in the Tehran city. This area $(35^{\circ}41'46''N \text{ and } 51^{\circ}25'23''E)$ is located meanly 1700 meters above the sea level and has a cold semi-arid climate (with an average annual temperature of 16.4°C and about 429 mm rain receives annually).

Sampling strategy: The subjects were obtained from six hospitals between January 2014 and September 2015. A previously designed questionnaire was used to record potential risk factors associated with toxocariasis include: sex, age, educational level (illiterate, primary education, high school, and college graduated education and above), occupation, eating unwashed vegetables, and contact with dog, cat and contaminated soils. Objectives and protocol of the study were explained to all the enrolled participants. The written consents were signed by participants or their parents. Medical laboratory specialist collected blood samples after receiving consent forms and questionnaires.

Laboratory tests: A total of 374 participants were enrolled in this study. About 3-5 mL of whole blood samples were taken from each participant using venipuncture. The samples were allowed to clot and centrifuged at 1000 g for 3 minutes in order to the separation of sera. The collected sera were transported in ice to the Helminthology Laboratory of Shahid Beheshti University of Medical Science, where the sera were stored at -20°C until the examination. Sera samples were screened for anti-*Toxocara* IgG antibodies using ELISA kit (Nova Tec, Germany) as its instruction. The kit has sensitivity and specificity of >95%.

Statistical analysis: The SPSS statics software, version 21 (IBM, NY, USA), was used for analysis of results. Frequency was used for the description of characteristics of participants and prevalence of the parasite, and risk factors. Associations between seropositivity for toxocariasis and the potential risk factors were evaluated by Pearson's chi-square test.

Results

Population sample comprised 192 (51.4 %) females and 182 (48.6%) males, aged from 1 to 81 years with a mean age (standard deviation; SD) of 28.6 (18.2) years. Most of the participants were aged \geq 61 years (23.8%). The majority of women (105/192; 54.7%) were housewife. Concerning education level, only 16 (4.2%) were unable to read and write. One hundred

Characteristic	No. Persons (%)	% seropositive	P value in	P value in	
		-	χ^2 test	Logistic	
				regression	
Age (yr)			< 0.001	0.001	
≤9	64 (17.1)	0 (0.0)			
10-19	72 (19.3)	1 (1.38)			
20-29	75 (20)	3 (4)			
30-39	74 (19.8)	2 (2.7)			
≥61	89 (23.8)	15 (16.8)			
Sex			>0.05	0.1	
Male	182 (48.6)	13 (7.1)			
Female	192 (51.4)	8 (4.1)			
Education			0.16	0.2	
College and above	97 (26)	4 (4.1)			
High school	200 (53.5)	12 (6)			
Primary school	61 (16.3)	4 (6.5)			
Illiterate	16 (4.2)	1 (6.2)			
Occupation			0.09	0.1	
Gov't employer & Other	194 (51.9)	13 (6.7)			
Student	73 (19.5)	0 (0.0)			
Housewife	105 (28.1)	8 (7.6)			
Farmer & shepherd	2 (0.5)	0 (0.0)			
Eating not sterile raw vegetables			0.046	0.06	
Yes	177 (47.3)	15 (8.5)			
No	197 (52.7)	6 (3.04)			
Contact with dog and cat			0.08	0.1	
Yes	118 (31.5)	11 (9.3)			
No	256 (68.5)	10 (3.9)			
Contact to soil			0.23	0.3	
Yes	67 (17.9)	2 (2.98)			
No	307 (82.1)	19 (6.1)			
Eosinophilia			0.47	0.6	
Yes	11 (2.9)	1			
No	363 (97.1)	20			

Table 1: Seroprevalence of *Toxocara* infection among healthy individuals referred to the medical centers Tehran city, according to sociodemographic characteristics (n=374).

and eighteen (31.5%) and 67 (17.9%) participants reported a frequent contact with domestic animals (dog and cat) and soil, respectively. More sociodemographic characteristics are presented in Table 1. Out of 374 participants, the overall prevalence of anti-*Toxocara* antibodies was 5.6% (21/374). The seropositivity rate among males (13/182 cases; 7.1%) was slightly lower than in females (8/192; 4.1%). There was no significant difference in the prevalence among males and females (P>0.05) in chi-square test. Results by the chi-square test showed that Toxocara seropositivity was associated with age (P<0.001) and eating unwashed vegetables (P=0.046). In logistic regression only age was as potential risk factor (P<0.001). Further data are presented in Table 1.

Discussion

In spite of the importance of toxocariasis for the human health, the infection is classified as a neglected disease by World Health Organization²², and is taken little attention as an important health problem, in the developing countries like Iran²³. However, few studies have been conducted to determine the seroprevalence of toxocariasis among potentially at-risk groups in Iran¹⁴⁻¹⁶.

Seroprevalence of *Toxocara* spp. infection was reported 3-86% in deferent studies from different parts of the world, and from 5 to 29% in different parts of Iran^{12,24}. In our study, the overall seroprevalence of 5.6% was found to *Toxocara* spp. infection in the Tehran city that is significantly lower than previously reported mean (15.8%) seroprevalence of the infection in the Iran¹⁴. To compare of our results with other

Seroprevalence of *Toxocara* infection among healthy individuals referred to the ...

First author/Ref	Study province	Study population	Sample size	Infected (%)
Sharif/ ³³	Mazandaran/Sari	School-children	1210	297 (25)
Nourian/ ³⁴	Zanjan	School-children	810	22 (2.7)
		Hypereosinophilic	100	19 (19)
Maraghi/ ³⁵	Khuzestan/Ahwaz	patients		
		Healthy controls	100	1 (1)
Sajjadi/ ¹⁵	Fars/Shiraz	School-children	519	133 (25.6)
Fallah/ ¹⁶	Hamadan	School-children	544	29 (5.3)
Hosseini-Safa/ ³⁶	Isfahan	Children	427	6 (1.39)
Shahraki/ ³⁷	Sistan-Baluchestan	Children	364	14 (3.8)
Alavi/ ³⁸	Khuzestan/Ahwaz	Children with chronic	115	16 (13.9)
		cough		

Table	2: Seroi	prevalence o	of <i>Toxocara</i> spp	reported by	previous	studies i	in the	different	parts of Ir	an.
I unic .	- • DUIU	prevalence o	I IONOCUIU SPP	· reported by	previous	bluares 1		uniterent	puito or m	un.

reports in different part of Iran, we have summarized previously performed studies in Table 2. Moreover, the seroprevalence of toxocariasis in the present study is lower than the 44.92% of seroprevalence reported Peru¹⁷, 51.6% in Brazil¹⁸, 13.9% in United States¹⁹, 51.2% in South Korea²⁰, 23.5% in Serbia²¹ and 22.1% in Roma population of Slovakia²². Furthermore, seroprevalence rate in our study is lower than their reported from Turkey (16.97%) in neighboring Iran²³. On the other hand, the seroprevalence in this study higher than results of the study from Denmark (2.4%)²⁴ and approximately in consistent with results from studies in Egypt (7.7%) and Austria (6.3%)^{25,26}.

Considering to risk factors for Toxocara infection results from this study demonstrated that age and unwashed vegetable were potential risk factors to the acquisition of infection. In our study rate of infection was elevated with increase in age. These results are in agreement with those reported by Won et al. in the USA^{19} and Lee *et al.* in South Korea²⁰ and other studies in the different part of world^{21,22}. The possible explanation for this would be an enhanced exposure to Toxocara eggs by means contaminated soil and raw vegetables or undercooked meat. Moreover, it should be noticed that anti-Toxocara antibodies remain for a long time, and the increment of the seroprevalence with age is associated with lifetime exposure. Given that unwashed vegetables, it should be mentioned that many previous studies in Iran have reported moderate contamination of vegetables with *Toxocara* eggs^{5,6,27}. Therefore, raw vegetables could be a potential risk factor, especially if they are washed inappropriately.

In our study we observed not statistically significant

association of Toxocara infection seropositivity with the gender, education, occupation, contact with dog or cat and contact with soil. In consistent with our results. Gabrielli et al. in Serbia, Lötsch et al. in Gabon and Espinoza et al. in Peru have reported that contact with dog or cat could not be a risk factor for Toxocara infection^{21,28,29}. Interestingly, Rubinsky-Elefant et al. have found cat as protective factor in their study 30 . However, unlike to our study in some studies, contacts with dogs have described as important risk factors for the infection 19,31,32 . In contrast with our study, Won *et* al. have found that low level of education is significantly associated *Toxocara* seropositivity¹⁹. They mentioned that lower education levels are often associated with lower socioeconomic status, employing in occupations involving more soil exposure, live in areas with high environmental contamination, in which all of these could be associated with increased rate of Toxocara infection in the overall population¹⁹. Although our results showed that male were more infected than women, but statistically significant association was not observed. The different findings may be due to difference in sample size, time the study was done, type of studies population in different studies.

Our study had some limitations. First, financial issue and time limits did not allow us to include wider area and more participants in our study. Another limitation of our study is the use of only ELISA method. It would have been ideal to use of western blot to confirmation of seropositive individuals.

Conclusion

The relatively low seroprevalence of *Toxocara* infection in our study can be caused by environmental

and socio-cultural conditions, moderate or high standards of hygiene, in Tehran city. We suggest carrying out further studies on the prevalence of toxocariasis in at-risk groups (such as asthma, hypereosinophilic, epilepsy, rheumatism and schizophrenia patients). In addition, to decrease *Toxocara* spp. infections, knowledge of people regarding toxocariasis and its risk factors must be increased.

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References

1. Smith H, Holland C, Taylor M, Magnaval J, Schantz P, Maizels R. How common is human toxocariasis? Towards standardizing our knowledge. Trend Parasitol. 2009;25(4):182-8.

2. Overgaauw PA, van Knapen F. Veterinary and public health aspects of *Toxocara* spp. Vet Parasitol. 2013;193(4):398-403.

3. Jenkins EJ, Castrodale LJ, de Rosemond S, Dixon BR, Elmore SA, Gesy KM, et al. Tradition and transition: parasitic zoonoses of people and animals in Alaska, northern Canada, and Greenland. Adv Parasitol. 2013;82:33-204.

4. Choi D, Lim JH, Choi D-C, Lee KS, Paik SW, Kim SH, et al. Transmission of *Toxocara canis* via ingestion of raw cow liver: a cross-sectional study in healthy adults. Korean J Parasitol. 2012;50(1):23-7.

5. Rostami A, Ebrahimi M, Mehravar S, Omrani VF, Fallahi S, Behniafar H. Contamination of commonly consumed raw vegetables with soil transmitted helminth eggs in Mazandaran province, northern Iran. Int J Food Microbiol. 2016;225:54-8.

6. Siyadatpanah A, Tabatabaei F, Zeydi AE, Spotin A, Omrani VF, Assadi M, et al. Parasitic contamination of raw vegetables in Amol, North of Iran. Arch Clin Infect Dis. 2013;8(2): e15983.

7. Despommier D. Toxocariasis: clinical aspects, epidemiology, medical ecology, and molecular aspects. Clin Microbiol Rev. 2003;16(2):265-72.

8. Macpherson CN. The epidemiology and public health importance of toxocariasis: a zoonosis of global importance. Int J Parasitol. 2013;43(12):999-1008.

9. Ma G, Holland CV, Wang T, Hofmann A, Fan C-K, Maizels RM, et al. Human toxocariasis. Lancet Infect Dis. 2017.

http://dx.doi.org/10.1016/ S1473-3099(17)30331-6

10. Fragoso RP, Monteiro MBM, Lemos EM, Pereira FEL. Anti-*Toxocara* antibodies detected in children attending elementary school in Vitoria, State of Espírito Santo, Brazil: prevalence and associated factors. Rev Soc Bras Med Trop. 2011;44(4):461-6.

11. Gavignet B, Piarroux R, Aubin F, Millon L, Humbert P. Cutaneous manifestations of human toxocariasis. J Am Acad Dermatol 2008;59 (6):1031-42.

12. Kwon N-H, Oh M-J, Lee S-P, Lee B-J, Choi D-C. The prevalence and diagnostic value of toxocariasis in unknown eosinophilia. Ann Hematol. 2006;85(4):233-8.

13. Fillaux J, Magnaval J-F. Laboratory diagnosis of human toxocariasis. Vet Parasitol. 2013; 193(4):327-36.

14. Abdi J, Darabi M, Sayehmiri K. Epidemiological situation of toxocariasis in Iran: meta-analysis and systematic review. Pak J Biol Sci. 2012;15:15(22):1052-5.

15. Sadjjadi S, Khosravi M, Mehrabani D, Oryan A. Seroprevalence of *Toxocara* infection in school children in Shiraz, Southern Iran. J Trop Pediatr. 2000;46(6):327-30.

16. Fallah M, Azimi A, Taherkhani H. Seroprevalence of toxocariasis in children aged 1-9 years in western Islamic Republic of Iran, 2003. East Mediterr Health J. 2007;13(5):1073-7.

17. Roldán WH, Espinoza YA, Huapaya PE, Huiza AF, Sevilla CR, Jiménez S. Frequency of human toxocariasis in a rural population from Cajamarca, Peru determined by DOT-ELISA test. Rev Inst Med Trop Sao Paulo. 2009;51(2):67-71.

18. Colli CM, Rubinsky-Elefant G, Paludo ML, Falavigna DL, Guilherme EV, Mattia S, et al. Serological, clinical and epidemiological evaluation of toxocariasis in urban areas of south Brazil. Rev Inst Med Trop Sao Paulo 2010;52(2):69-74.

19. Won KY, Kruszon-Moran D, Schantz PM, Jones JL. National seroprevalence and risk factors for zoonotic *Toxocara* spp. infection. Am J Trop Med Hyg. 2008;79(4):552-7.

20. Lee J-Y, Yang MH, Hwang J-H, Kang M, Paeng JW, Yune S, et al. The prevalence of toxocariasis and diagnostic value of serologic tests in asymptomatic Korean adults. Allergy Asthma Immunol Res. 2015;7(5):467-75.

21. Gabrielli S, Tasić-Otašević S, Ignjatović A, Fraulo M, Trenkić-Božinović M, Momčilović S, et al. Seroprevalence and Risk Factors for *Toxocara canis* Infection in Serbia During 2015. Foodborne Pathog Dis. 2017;14(1):43-9.

22. Antolová D, Jarčuška P, Janičko M, Madarasová-Gecková A, Halánová M, Čisláková L, et al. Seroprevalence of human *Toxocara* infections in the Roma and non-Roma populations of Eastern Slovakia: a cross-sectional study. Epidemiol Infect. 2015;143(10):2249-58.

23. Doğan N, Dinleyici EÇ, Bor Ö, Töz SÖ, Özbel Y. Seroepidemiological survey for Toxocara canis infection in the northwestern part of Turkey. Turkiye Parazitol Derg. 2007;31(4):288-91.

24. Stensvold CR, Skov J, Møller LN, Jensen PM, Kapel CM, Petersen E, et al. Seroprevalence of human toxocariasis in Denmark. Clin Vaccine Immunol. 2009;16(9):1372-3.

25. El-Shazly A, Abdel Baset S, Kamal A, Mohammed KA, Sakrs T, Hammad S. Seroprevalence of human toxocariasis (visceral larva migrans). J Egypt Soc Parasitol. 2009;39 (3):731-44.

26. Poeppl W, Herkner H, Tobudic S, Faas A, Mooseder G,

Burgmann H, et al. Exposure to Echinococcus multilocularis, *Toxocara canis*, and *Toxocara cati* in Austria: a nationwide cross-sectional seroprevalence study. Vector Borne Zoonotic Dis. 2013;13(11):798-803.

27. Fallah AA, Pirali-Kheirabadi K, Shirvani F, Saei-Dehkordi SS. Prevalence of parasitic contamination in vegetables used for raw consumption in Shahrekord, Iran: influence of season and washing procedure. Food Control. 2012;25:617-20.

28. Lötsch F, Obermüller M, Mischlinger J, Mombo-Ngoma G, Groger M, Adegnika AA, et al. Seroprevalence of *Toxocara* spp. in a rural population in Central African Gabon. Parasitol Int. 2016;65(6):632-634.

29. Espinoza YA, Huapaya PE, Roldán WH, Jiménez S, Abanto EP, Rojas CA, et al. Seroprevalence of human toxocariasis in Andean communities from the Northeast of Lima, Peru. Rev Inst Med Trop Sao Paulo. 2010;52(1):31-6.

30. Rubinsky-Elefant G, da Silva-Nunes M, Malafronte RS, Muniz PT, Ferreira MU. Human toxocariasis in rural Brazilian Amazonia: seroprevalence, risk factors, and spatial distribution. Am J Trop Med Hyg. 2008;79(1):93-8.

31. Loukas A, Hintz M, Linder D, Mullin NP, Parkinson J, Tetteh KK, et al. A family of secreted mucins from the parasitic nematode *Toxocara canis* bears diverse mucin domains but shares similar flanking six-cysteine repeat motifs. J Biol Chem. 2000:15;275(50):39600-7.

32. Silva MB, Amor AL, Santos LN, Galvão AA, Oviedo Vera AV, Silva ES, et al. Risk factors for *Toxocara* spp. seroprevalence and its association with atopy and asthma phenotypes in school-age children in a small town and semi-rural areas of Northeast Brazil. Acta Trop. 2017;174:158-64.

33. Sharif M, Daryani A, Barzegar G, Nasrolahei M, Khalilian A. Seroprevalence of toxocariasis in schoolchildren in Northern Iran. Pak J Biol Sci. 2010;13(4):180-4.

34. Nourian A, Amiri M, Ataeian A, Haniloo A, Mosavinasab S, Badali H. Seroepidemiological study for toxocariasis among children in Zanjan-northwest of Iran. Pak J Biol Sci. 2008;11(14):1844-7.

35. Maraghi S, Rafiei A, Hajihossein R, Sadjjadi S. Seroprevalence of toxocariasis in hypereosinophilic individuals in Ahwaz, south-western Iran. J Helminthol. 2012;86(2):241-4.

36. Hosseini-Safa A, Mousavi SM, Badorani MBB, Samani MG, Mostafaei S, Darani HY. Seroepidemiology of Toxocariasis in Children (5–15 yr Old) Referred to the Pediatric Clinic of Imam Hossein Hospital, Isfahan, Iran. Iran J Parasitol. 2015;10(4):632-7.

37. Shahraki MK, Dabirzadeh M, Afshari M, Maroufi Y. Epidemiological Study of Toxocar canis in Children under 14-Years-Old and Dogs in Zabol and Chabahar Districts, Southeast of Iran. Iran J Parasitol. 2017;12(1):101-7.

38. Alavi SM, Sefidgaran G. Frequency of anti Toxocara antibodies in school children with chronic cough and eosinophilia in Ahwaz, Iran. Pak J Med SCI. 2008;24(3):360-3.