

Antimicrobial effect of the aqueous and ethanolic *Teucrium polium* L. extracts on gram positive and gram negative bacteria “*in vitro*”

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ABSTRACT

The genus *Teucrium* (germander) belongs to the family Lamiaceae, within the subfamily Ajugoideae. *Teucrium polium* L. (Labiatae) has long been recognized in folk medicine in the treatment of many pathophysiological implications, such as gastrointestinal disorders, inflammations, diabetes and rheumatism. In this study antimicrobial activity of two crude extracts obtained from *Teucrium polium* L. was tested against bacterial species. The Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) for both species determined by using “dilution method”. The antimicrobial effects of extracts were evaluated on *Streptococcus pyogenes* PTCC 1447, *Pseudomonas aeruginosa* PTCC 1310 and *Staphylococcus epidermidis* PTCC 1435 by “using the method of Collins” and “disk agar diffusion method”. The results showed that aqueous and ethanolic extracts were quite effective in 2000 µg/ml concentration on *Streptococcus pyogenes* and *Staphylococcus epidermidis* and were prevented from growth them on medium, while both extracts have no certain antimicrobial effect on *Pseudomonas aeruginosa*. In “disk agar diffusion method”, 10, 20, 30 and 40 mg/ml aqueous and ethanolic extracts concentrations, was inhibition effect on *Streptococcus pyogenes* and *Staphylococcus epidermidis*, and 30 and 40 mg/ml aqueous and ethanolic extracts concentrations, has inhibition effect on *Pseudomonas aeruginosa* prevent them growing, but at 10 and 20 mg/ml concentrations, no inhibitory effect on *Pseudomonas aeruginosa* was observed. The results indicate that ethanolic and aqueous extracts of *Teucrium polium* L. have the greatest effect on gram-positive bacterium *Streptococcus pyogenes* ($p < 0.05$). Results showed, aqueous and ethanolic extracts of *Teucrium polium* L., have been strong antimicrobial activity against many food pathogen bacteria.

Keywords: *Teucrium polium* L.; Extract; Antimicrobial; Pathogen bacteria.

INTRODUCTION

The genus *Teucrium* (germander) belongs to the family Lamiaceae, within the subfamily Ajugoideae. In the flora of Europe the genus *Teucrium* has been divided into six sections with 49 species. The section *Stachyobotrys* is represented by the species *Teucrium arduini* L., the section *Scorodonia* by *Teucrium scordium* L., the section *Chamaedrys* by *Teucrium chamaedrys* L. and the section *polium* by *Teucrium polium* L. and *Teucrium montanum* L. These are mostly perennial herbs, shrubs or sub shrubs, while *Teucrium botrys* is an herbaceous annual plant. The species of this genus are widespread in all continents of the world but a very large number of species are present in the Mediterranean [1].

Teucrium polium L. are rich in a wide variety of secondary metabolites such

as tannins, alkaloids and flavonoids, which have been found “*in vitro*” to have antimicrobial properties [2]. Finding healing power in plant is an ancient. Numerous studies have been carried out in different parts of the globe to extract plant products for screening antimicrobial activity [3]. Herbal medicines are one of the important cultural and traditional part of the people. Today, most of the world population depends on herbal medicines for caring their health [4]. Antibiotics provide the main basis for the therapy of microbial (bacterial and fungal) infections. Since the discovery of these antibiotics and their use as chemotherapeutic agents there was a belief in the medical fraternity that this would lead to the eventual eradication of infectious diseases. However, overuse of antibiotics has become the major factor in the emergence and

dissemination of multi-drug resistant strains of several groups of microorganisms [5].

Darabpour *et al.*, (2010) demonstrated that the methanolic extract of Iranian *Teucrium polium* L. inhibited the growth of several bacteria with different MIC. This extract inhibited the growth of *Staphylococcus aureus* and *Salmonella typhi* with a MIC of 40 mg/ml. In addition, 10 mg/ml represents also the MBC against *Bacillus anthracis* [6]. *Streptococcus pyogenes*, and *Staphylococcus epidermidis*, organisms that cause respiratory and cutaneous infections, and members of the *Pseudomonas* families, organisms that cause diarrhea, urinary infection, and sepsis, are now resistant to virtually all of the older antibiotics. *Pseudomonas aeruginosa* is opportunistic pathogens. *Pseudomonas aeruginosa* are frequently associated with infection of the urinary tract. Also, *Pseudomonas aeruginosa* is commonly found in hospital environment and can easily infect immunosuppressed patients.

The aim of this study was evaluated of antimicrobial effects of aqueous and ethanolic extracts *Teucrium polium* L. against some of the important food pathogens bacteria.

MATERIALS AND METHODS

I. Plant Preparation

Teucrium polium L. was collected from Birjand (southeast of Khouzestan state, Iran). Taxonomic identification was performed by the Faculty of Science Herbarium, Ferdowsi University of Mashhad, Iran.

II. Preparation of aqueous and ethanolic extracts from *Teucrium polium* L.

The amount 100 gram of *Teucrium polium* L. powder was added to 500 ml ethanol 96 degree or distilled water. The ethanolic extract mixture was preserved at laboratory temperature for 24 hours and was stirred every few hours with a glass rod. The aqueous mixture was boiled for 20 minutes with low flame until the cream colored liquid was obtained. The collecting supernatant was centrifuged by 6000 rpm for 5 min. The resulting extract (supernatant) volume has reached to the original with ethanol or distilled water, and then samples were stored into the dark container at refrigerator temperature after filtering by 0.45 μ Whatman filter paper [7, 8].

III. Source of microorganisms

Three bacterial species, Gram-positive (*Streptococcus pyogenes* PTCC 1447,

Staphylococcus epidermidis PTCC 1435) and Gram-negative (*Pseudomonas aeruginosa* PTCC 1310) the microorganisms used in this study are significant. *Pseudomonas aeruginosa* are frequently associated with infection of the urinary tract. Also, *Pseudomonas aeruginosa* is commonly found in hospital environment and can easily infect immunosuppressed patients. *Staphylococcus epidermidis* can infect skin and wounds causing acne, boils, and pimples.

IV. Preparation of Microbial suspension

Some of the bacterial suspension was poured in sterile tubes containing Ringer solution and its turbidity, was measured by spectrophotometer at 530 nm wavelength. It was diluted by Ringer solution until the solution turbidity equalizes with 0.5 McFarland standard solutions. Suspension should have contains 1.5×10^8 CFU / ml [9].

V. Evaluation of antimicrobial activity

0.2 g of the extract was reconstituted in 5ml sterile distilled water and vortexes for homogeneity. Then 1 ml of this solution was added to sterile plates. The final concentration of the extract was 2000 μ g/ml [10]. In the next step, Mueller Hinton agar (Merck-Germany) medium were sterile and used for bacteria, were added to the plates, and placed at room temperature, so the medium was prepared. One loop of each standard strain culture media was cultured inoculums on these medium. The plates were incubated for 48 hours at 37 °C. The culture with extract and without bacteria was used as control [10]. In the "disk agar diffusion" method, a loop of each standard strain culture media was cultured on the plates, and then paper discs (from Whatman filter with 6 mm diameter) plates were saturated with 100 μ l of the test compound allowed to dry and was introduced on the upper layer of the seeded agar plate with 10, 20, 30 and 40 mg/ml extract concentrations, were prepared in distilled water and was treated with *Teucrium polium* L. extract and placed in culture medium by Sterile loop. Then it was fixed on the media with a light little pressure. After incubation time, the diameter of free zone was measured exactly by using a ruler in millimeters. All experiments were performed with 3 replicates [11].

VI. Determination of Minimum Inhibitory Concentration (MIC)

MIC was determined according to agar dilution method [12]. Various concentrations of extract were prepared in 10 cm experimental tubes

containing Mueller Hinton broth for bacteria. Each tube contains 9 ml of Mueller Hinton for bacteria were sterilized by autoclaving. On cooling, 1 ml of each extract (watery & ethanoli) concentration were added to each tube, to make the final concentrations of (2, 4, 8, 16, 32 and 64mg/ml).

The mixture of Mueller Hinton and extract was poured into plates aseptically in a laminar flow cabinet. On solidification of the agar medium, 2 µl of adjusted spore suspension were added to each plate by micropipette and incubated 37 °C for bacteria [13]. The Mueller Hinton without any herbal extract served as control. The MIC was regarded as the lowest concentration of the extract that did not show any visible growth after 12 days of incubation (compared with control).

VII. Minimum bactericidal concentration (MBC)

MBC was determined according to agar dilution method [14] with slight modifications. The MBC were determined by incorporating various concentrations of extracts (2, 4, 8, 16, 32, 64, 128 and 256 mg/ml) in Mueller Hinton broth for bacteria. The tubes which showed no visible growth after 2 days incubation were subculture on extract free Mueller Hinton plates and incubated at 37 °C for 2 days [13]. The MBC was regarded as the lowest concentration of the extract that prevented the growth of any bacteria colony on the solid medium.

VIII. Statistical analysis

All the assays were carried out in triplicates. The experimental results were expressed as mean ± standard deviation. The data were analyzed using one way analysis of variance (ANOVA) using SPSS version 17. One-way analysis of variance (ANOVA) was used to analyze each treatment over storage. In the case of significant differences, Tukey test ($p < 0.05$) was used.

RESULTS

The results of the antimicrobial effects of aqueous and ethanolic extracts, by “using the method of Collins [15]” were showed in Tables 1 and 2. The results showed 2000 µg/ml concentration of both aqueous and ethanolic *Teucrium polium* L. extracts, were quite effective on reduce of growth *Streptococcus pyogenes* and *Staphylococcus epidermidis* were had prevent growth over the medium ($p < 0.05$). However, 2000 µg/ml concentration aqueous and ethanolic extracts, have no significant antibacterial effect on *Pseudomonas aeruginosa* and it is not able to prevent the growth of bacteria on culture ($p < 0.05$). The results of the antimicrobial effects of aqueous and ethanolic *Teucrium polium* L. extracts, by “the agar diffusion method” are presented in Tables 3 and 4. Minimum Inhibitory Concentration (MIC) results of the aqueous and ethanolic extracts of *Teucrium polium* L. are given in Table 5 and 6. Minimum Bacterial Concentration (MBC) results of the aqueous and ethanolic extracts of *Teucrium polium* L. are given in Table 7 and 8.

Table 1. Antimicrobial effects of 2000 µg/ml ethanolic *Teucrium polium* L. extract concentrations, on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis* (using the method of Collins [15])

Microorganism	<i>Teucrium polium</i> L.
<i>P. aeruginosa</i>	-
<i>S. pyogenes</i>	++
<i>S. epidermidis</i>	+

- (-) in Table showed the growth of bacteria on culture and the lack of antibacterial activity of ethanolic *Teucrium polium* L. extract
- (+) in Table showed no bacterial growth on culture and antibacterial activity of ethanolic *Teucrium polium* L. extract
- (++) in Table showed no bacterial growth on culture and strong antibacterial activity of ethanolic *Teucrium polium* L. extract

Table 2. Antimicrobial effects of 2000µg/ml aqueous *Teucrium polium* L. extract concentrations, on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis* (using the method of Collins [15])

Microorganism	<i>Teucrium polium</i> L.
<i>P. aeruginosa</i>	-
<i>S. pyogenes</i>	+
<i>S. epidermidis</i>	+

- (-) in Table showed the growth of bacteria on culture and the lack of antibacterial activity of aqueous *Teucrium polium* L. extract
- (+) in Table showed no bacterial growth on culture and antibacterial activity of aqueous *Teucrium polium* L. extract

Table 3. Average diameter (mm) of microbial free zone area of aqueous *Teucrium polium* L. leaves extract, on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis* (disk agar diffusion method)

Microorganism		<i>P. aeruginosa</i>		
<i>Teucrium polium</i> L. concentration	10	20	30	40
Average diameter (mm) of microbial free zone area	-	-	6.30±0/50	6.90 ±0/50
Microorganism		<i>S. pyogenes</i>		
<i>Teucrium polium</i> L. concentration	10	20	30	40
Average diameter (mm) of microbial free zone area	12.30±0/57	14.30±0/50	16 ±0/28	19.10±0/28
Microorganism		<i>S. epidermidis</i>		
<i>Teucrium polium</i> L. concentration	10	20	30	40
Average diameter (mm) of microbial free zone area	11.20±0/57	12.40±0/57	14.10±0/50	16.90±0/28

▪ (-) in Table showed no inhibitory effects was shown

Table 4. Average diameter (mm) of microbial free zone area of ethanolic *Teucrium polium* L. leaves extract, on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis* (disk agar diffusion method)

Microorganism		<i>P. aeruginosa</i>		
<i>Teucrium polium</i> L. concentration	10	20	30	40
Average diameter (mm) of microbial free zone area	-	-	6.70 ±0/57	8.80±0/28
Microorganism		<i>S. pyogenes</i>		
<i>Teucrium polium</i> L. concentration	10	20	30	40
Average diameter (mm) of microbial free zone area	14.90±0/76	16.90±0/50	18.80 ±0/50	22.90±0/57
Microorganism		<i>S. epidermidis</i>		
<i>Teucrium polium</i> L. concentration	10	20	30	40
Average diameter (mm) of microbial free zone area	13.10±0/28	14.90±0/50	16.90 ±0/76	18.60±0/57

▪ (-) in Table showed no inhibitory effects was shown

Table 5. Minimum Inhibitory Concentration (MIC) of *Teucrium polium* L. extract (aqueous) on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis*

Microorganism species	Concentration (mg/ml)						
	Control	2	4	8	16	32	64
<i>P. aeruginosa</i>	-	-	-	-	-	-	+
<i>S. pyogenes</i>	-	-	-	-	+	+	+
<i>S. epidermidis</i>	-	-	-	-	-	+	+

+: Positive inhibition

- : Negative inhibition

Table 6. Minimum Inhibitory Concentration (MIC) of *Teucrium polium* L. extract (ethanolic) on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis*

Microorganism species	Concentration (mg/ml)						
	Control	2	4	8	16	32	64
<i>P. aeruginosa</i>	-	-	-	-	-	+	+
<i>S. pyogenes</i>	-	-	-	+	+	+	+
<i>S. epidermidis</i>	-	-	-	-	+	+	+

+: Positive inhibition

- : Negative inhibition

Table 7. Minimum Bacterial Concentration (MBC) of *Teucrium polium* L. extract (aqueous) on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis*

Microorganism species	Concentration (mg/ml)								
	Control	2	4	8	16	32	64	128	256
<i>P. aeruginosa</i>	-	-	-	-	-	-	-	-	+
<i>S. pyogenes</i>	-	-	-	-	-	-	+	+	+
<i>S. epidermidis</i>	-	-	-	-	-	-	-	+	+

+: Positive inhibition

- : Negative inhibition

Table 8. Minimum Bacterial Concentration (MBC) of *Teucrium polium* L. extract (ethanolic) on *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus epidermidis*

Microorganism species	Concentration (mg/ml)									
	Control	2	4	8	16	32	64	128	256	
<i>P. aeruginosa</i>	-	-	-	-	-	-	-	+	+	
<i>S. pyogenes</i>	-	-	-	-	-	+	+	+	+	
<i>S. epidermidis</i>	-	-	-	-	-	-	+	+	+	

+: Positive inhibition

- : Negative inhibition

DISCUSSION

The antimicrobial compounds from plants may inhibit bacterial growth by different mechanisms than those presently used. Antimicrobials therefore, may have a significant clinical value in treatment of resistant microbial strains. In particular, the antimicrobial activities of plant oils and extracts have formed the basis of many applications including raw and processed food preservation, pharmaceuticals, alternative medicine, and natural therapies [16]. The results showed 2000 µg/ml concentration of both aqueous and alcoholic extracts, were quite effective on reduce of growth *Streptococcus pyogenes* and *Staphylococcus epidermidis* and were had prevent growth over the medium. However, 2000 µg/ml concentration aqueous and alcoholic extracts, have no significant antibacterial effect on *Pseudomonas aeruginosa* and it is not able to prevent the growth of bacteria on culture. Based on the results ethanolic extract of *Teucrium polium* L. in this study have significant antimicrobial activity on the studied microorganisms.

The results show that *Teucrium polium* L. ethanolic and aqueous extracted at all concentrations (10, 20, 30 and 40 mg/ml) had the inhibitory effect on *Streptococcus pyogenes* and *Staphylococcus epidermidis*. The results show that *Teucrium polium* L. extracted at concentrations (30 and 40 mg/ml) had the inhibitory effect on *Pseudomonas aeruginosa*. However, 10 and 20 mg/ml concentration extracts, have no significant antimicrobial effect on *Pseudomonas aeruginosa* and it is not able to prevent the growth of bacteria on culture. Gram-positive bacteria are more sensitive than gram-negative bacteria to *Teucrium polium* L. extract, due to differences in cell structure of gram-negative and gram-positive bacteria, because gram-positive bacteria have more mucopeptide in their cell wall composition while gram-negative bacteria have only a thin layer of mucopeptide and most of their cell structure is

lipoprotein and lipo-polysaccharides. Thus, gram-negative bacteria are more resistant [17, 18]. These points were consistent with the results obtained in this study.

Alizadeh Behbahani *et al.*, 2012 reports that, ethanol extract compared to the aqueous extract was more effective and have a greater deterrent. The reason of these phenomena may be extracting more effective materials extracted by ethanol from *Avicennia marina*. These points were consistent with the results obtained in this study [19]. Antimicrobial activity of *Teucrium polium* L. extracts is attributed to the presence of numerous bioactive secondary metabolites. Phenolic compounds, terpenoids and alkaloids are very important components in antimicrobial activity. Kizil *et al.*, 2003 expressed in their study where they used the disc diffusion method that *Teucrium spicata* var. *spicata* essential oil had a strong antibacterial impact on *Staphylococcus aureus*, *Escherichia coli* K 12, *Escherichia coli* pMK 3, *Escherichia coli* pBR 322 and *Escherichia coli* pUC [20].

In a study where antibacterial activity of *Teucrium spicata* var. *spicata* essential oil was examined through the method of disc diffusion, it was indicated that the essential *Staphylococcus aureus* ATCC 25923 was effective on *Bacillus cereus* ATCC 11778 and *Escherichia coli* ATCC 29998 and created the largest inhibition zone against *Bacillus cereus* ATCC 11778 [21].

In another study aiming at studying the effects of the essential oil of *Teucrium polium* plant obtained by hydro distillation on various bacteria and yeasts carried out by its absorption into empty antibiotic discs at different amounts through the method of disc diffusion, it was found that the 2µl and 4µl applications of the essential oil had no effect on *E. coli* DM, which is one of the bacteria of the study, it had also no effect on *Staphylococcus aureus* Cowan 1 in the application of 2µl while its application of 4µl created an inhibition zone [22].

The results shows that MIC of aqueous extract of *Teucrium polium* L. for *Streptococcus pyogenes* was 16mg/ml, for *Pseudomonas aeruginosa* was 64 mg/ml and for the *S. epidermidis* was 32 mg/ml. The results shows that MIC of ethanolic extract of *Teucrium polium* L. for *Streptococcus pyogenes* was 8 mg/ml, for *Pseudomonas aeruginosa* was 32 mg/ml and for the *Staphylococcus epidermidis* was 16 mg/ml. The results indicate that ethanolic extract of *Teucrium polium* L. mostly had been effective on *Streptococcus pyogenes* and has the least impact on *Pseudomonas aeruginosa*. The result shows that MBC of aqueous extract of *Teucrium polium* L. for *Streptococcus pyogenes* was 64 mg/ml, for *Pseudomonas aeruginosa* was 256 mg/ml and *Staphylococcus epidermidis* was 128 mg/ml. The result shows that MBC of ethanolic extract of *Teucrium polium* L. for *Streptococcus pyogenes* was 32 mg/ml, for *Pseudomonas aeruginosa* was 128 mg/ml and *Staphylococcus epidermidis* was 64 mg/ml. The obtained results were similar to the results in other studies, the ethanolic extract of *Teucrium polium* L.

REFERENCES

1. Ricci D, Fraternali D, Giamperi L. Chemical composition, antimicrobial and antioxidant activity of the essential oil of *Teucrium marum* (Lamiaceae). *J of ethnopharmacology*.2005; 98: 195-200.
2. Bajpai, M., A. Pande, S.K. Tewari and D. Prakash. Phenolic contents and antioxidant activity of some food and medicinal plants. *Inter J Food Sci Nutr*.2005; 56(4): 287-291.
3. Barbour EK, Al Sharif M, Sagherian VK, Habre AN, Talhouk RS, Talhouk SN. Screening of selected indigenous plants of Lebanon for antimicrobial activity. *J of ethnopharmacology*. 2004(93):1-7.
4. Mahmoud M, Al-Ameri SA, Abbas S. Extraction, Identification and Antimicrobial Activity of Some Phenolic Acids as Antioxidants in *Teucrium polium* Plant. *Kerbala J Pharmaceutical S*. 2011 (2) 170-188.
5. Khan R, Islam B, Akram M. Antimicrobial activity of five herbal extracts against multi drug resistant (MDR) strains of bacteria and fungus of clinical origin. *Molecules*.2009 (14) 586-597.
6. Darabpour, E., H. Motamedi and S. M. Seyyed Nejad. Antimicrobial properties of *Teucrium polium* against some clinical pathogens. *Asian Paci J Tropic Med*.2010; 3(2): 124-127.
7. Alizadeh Behbahani, B., Tabatabaei Yazdi, F., Shahidi, F., & Mohebbi, M. Antifungal effect of aqueous and ethanolic mangrove plant extract on pathogenic fungus "in vitro". *Inte J Agronomy and Plant Produc*.2013; 4 (7), 1652-1658.
8. Ahmad I. Beg AZ. Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogen. *J Ethnopharmacol*. 2001(74)113-123.
9. Valero M, Salmeron M. Antimicrobial activity of 11 essential oils against *Bacillus cereus* in Tyndallized carrot broth. *Int J Food Microbiology*. 2003(8) 73-81.
10. Babayi, H., Kolo, I., Okogun, J., & Ijah, U. The antimicrobial activities of methanolic extracts of *Eucalyptus camaldulensis* and *Terminaliacatappa* against some pathogenic microorganisms. *Biokemistri*. 2004;16(2); 106-111.
11. He F, Zhou J. A new antimicrobial susceptibility testing method of *Escherichia coli* against ampicillin by MSPQC. *J microbiologi methods*. 2007; 68(3): 563-7.

12. Benger S, Townsend P, Ashford RL, Lambert P. An in vitro study to determine the minimum inhibitory concentration of *Melaleuca alternifolia* against the dermatophyte *Trichophyton rubrum*. *The Foot*. 2004(14) 86 - 91.
13. Yazdani D RS, Amin Gh, Zainal Abidin MA, Shahnazi S, Jamalifar H. Antifungal Activity of Dried Extracts of Anise (*Pimpinella anisum* L.) and Star anise (*Illicium verum* Hook. f.) Against Dermatophyte and Saprophyte Fungi. *J Medicinal Plants*. 2009;8(5):24-9.
14. Espinel-Ingroff, A., Fothergill, A., Peter, J., Rinaldi, M., & Walsh, T. Testing conditions for determination of minimum fungicidal concentrations of new and established antifungal agents for *Aspergillus* spp.: NCCLS collaborative study. *J clinical microbiology*. 2001;40(9): 3204-3208.
15. Collins, C. H., Lynes, P. M. and Grange, J.M. *Microbiological Methods*. (7thEdn.) Butterworth-Heinemann Ltd., Britain, 1995;pp.175-190.
16. Sarac, N. and A. Ugur. Antimicrobial activities and usage in folkloric medicine of some Lamiaceae species growing in Mugla, Turkey. *EurAsia J BioSci*. 2007(4) 28-37.
17. Tassou CC, Nychas GJ. Antimicrobial activity of the essential oil of Mastic fume on gram positive and gram negative bacteria in broth and model food systems. *Inter Biodeterio. biodegrad*. 1998(36)411- 20.
18. Ghalem, B. R., & Mohamed, B. Antibacterial activity of leaf essential oils of *Eucalyptus globulus* and *Eucalyptus camaldulensis*. *African J Pharmacy and Pharmacology*. 2008;2(10), 211-215.
19. Alizadeh Behbahani, B., Tabatabaei Yazdi, F., Shahidi, F., & Mohebbi, M. Antimicrobial activity of *Avicennia marina* extracts ethanol, methanol & glycerin against *Penicillium digitatum* (citrus green mold). *Sci J Microbio*. 2012;1(7): 147-151.
20. S. Kizil, T. Sogut. Investigation of Antibacterial Effects of Some Spices. *Crop Research*. 2003;25(1): 86-90.
21. Akin, M., Oguz, D., & Saracoglu, H. Antibacterial Activity of Essential oil from *Thymbra spicata* var. *spicata* L. and *Teucrium polium* (Stapf Brig.). *Interventions*. 2010;8 (9): 53-58.
22. Pauli, A., & Knobloch, K. Inhibitory effects of essential oil components on growth of food-contaminating fungi. *Zeitschrift für Lebensmitteluntersuchung und-Forschung A*. 1987;185(1): 10-13.