

Critical concentration of Glucose changes human serum albumin conformation: Circular Dichroism (CD) and UV Spectroscopy approaches

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ABSTRACT

HSA plays an important role in transporting metabolites and drugs throughout the vascular system. In as much as its performance is very vital in the presence of different kinds of ligands at the specific body temperatures, its examination is crucial. This molecule can undergo increased glycation in diabetes. Therefore, glucose as the one of the most fundamental ligands dealing with albumin in human body is examined in this study at 100 mg/dl concentration in correspond to normal condition on human body, 175 mg/dl as a kidney glucose tolerance point and also 400 mg/dl as the critical point at the two most important temperatures in diabetic patients. Thermal conformational changes of (HSA) are important. These conformational alterations are accompanied by a mild alteration of secondary structures. For this reason, possible secondary structural changes of HSA in presence of glucose has been investigated by circular dichroism (CD) using Hepes buffer at the normal temperature 37°C and 42°C as a high fever condition. UV spectroscopy studies confirmed CD findings and indicate that critical concentration of glucose lead to generation of new structural feature of albumin similar to 42°C. However, as the temperature increases from 37°C to 42°C this process is no more capable of responding to glucose concentration changes. These results indicate that the native form of HSA is changed in the severe diabetic condition; likewise, same consequences can be achieved as the temperature arises from 37°C to 42°C.

Keywords: Human serum albumin (HSA); Temperature; Structural changes; Glucose; Circular dichroism (CD); UV spectroscopy

INTRODUCTION

Human serum albumin (HSA) as a monomeric multidomain globular protein is the most common circulating protein in blood. This molecule consists of 585 amino acids with an average molecular weight of 66500 Da [1]. This molecule represents the major controlling element of fluid dispersal among body compartments, and the most vital factor in regulating plasma osmotic pressure. Furthermore, ligand binding capability enabled this vital molecule for carrying many endogenous and exogenous compounds. In fact, albumin influences pharmacokinetics of many drugs, signifies the chief carrier for fatty acids, control the metabolic change of some ligands, transforms possible toxins mild, makes up for most of the anti-oxidant capacity of human plasma, and shows (pseudo-)enzymatic activities [2-4]. Additionally, albumin is a biological marker in a number of clinical disorders, adaptation mechanisms may be involved [5].

Remarkably, a number of factors are identified to influence HSA structure and dynamics, like pH, temperature, and binding of different ligands [6]. It has been marked that not only protein activity, unfolding, and degradation but also cell functioning is highly related to protein glycation. This globular heart-shaped molecule contains three homologous domains typically specified as I (1-195), II (196-383), and III (384-585). These domains are comparable both in the amino acid sequence and in the secondary and tertiary structures. Amazingly, even in the presence of a wide variety of ligands albumin conformation is completely preserved [7-9]. From recent studies, glycated albumin has biological impact on cell physiology and functioning; it is known as a marker of diabetes [3, 10, 11]. For instance, a risk factor for diabetes mellitus (DM) is urinary albumin excretion, which is independent of primary metabolic profile and increase of insulin resistance [12, 13]. Moreover,

glycation has key implications for albumin performance and influence on cell. The incubation of HSA with glucose results in its non-enzymatic glycoxidation in a concentration, incubation time, and temperature dependent manner. Non-enzymatic glycation of HSA alters its conformation and function[14]. In this study, it is proposed that glucose binds to HSA and alters the structural aspects. Circular dichroism (CD) and UV spectroscopic methods, consequently, have been applied to evaluate the structural modifications of human serum albumin in the temperature of 37-42°C in the presence and absence of glucose.

MATERIALS AND METHODS

Materials

Human serum albumin was purchased from Sigma chemical Co., USA. The other substances of reagent grade were obtained from Merck chemical Co., Germany; also, the buffer used all through the study was HEPES 100 mM, pH 7.

Methods

CD spectra were recorded by a Jasco J-715 spectropolarimeter (Japan). Results are expressed as ellipticity, $[\theta]$ (degree $\text{cm}^2 \text{dmol}^{-1}$), based on a mean amino acid residue weight (MRW). This value was assumed to be 113 Da for HSA. The molar ellipticity was determined as

$$[\theta]_{\lambda} = (\theta \times 100 \text{ MRW}/c)$$

Where c is the protein concentration in mM, l is the length of light path in cm and θ is the measured

ellipticity in degree at a given wavelength. The data were smoothed using the Jasco J-715 software, which includes a fast Fourier-transform noise reduction routine. All experiments were repeated three times. The concentration of the protein solution was 0.5 mg/ml. Percentage of secondary structures was calculated with the method of Chen et al.[15]. The UV-V absorption spectra of HSA were obtained by Unicop spectrophotometer[16]. HSA incubated 5 minutes in HEPES buffer 100 mM, pH 7 under the applied conditions and then the spectra were taken. The results were presented as mean \pm standard deviation for continuous variables.

RESULT

For examining albumin (HSA) exposure to glucose, CD technique was applied for detection of structural changes of this protein. For understanding the role of glucose on albumin structure at fever condition the mentioned experiment was repeated at 42°C (see figure 2). Absorption UV spectroscopy is a method that is applied widely in protein studies [17-20]. Based on this method it has been reported that Acetaminophen as a pain killer induces conformational change in human serum albumin [20, 21] (see figure 3). For more investigation UV spectra of albumin in the presence of two concentrations of glucose in the range of temperatures are provided, and represented in figures 4 and 5.

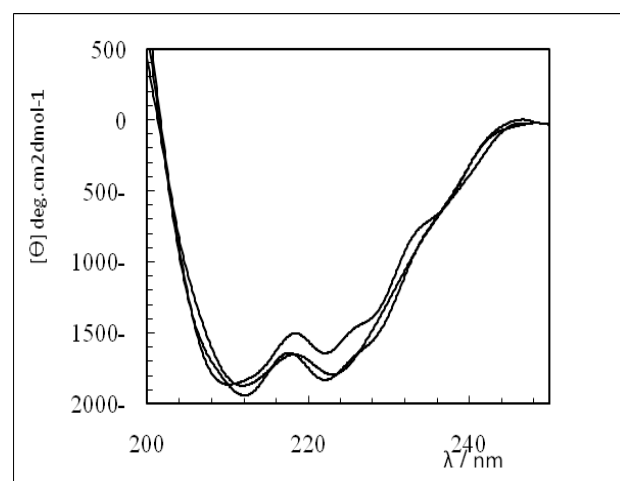


Figure 1. CD spectra of HSA in the presence of 100 mg/dl, 175 mg/dl, and also 400 mg/dl concentrations of glucose in HEPES buffer 100 mM, pH 7 at 37°C. Glucose concentration decreases from up to down.

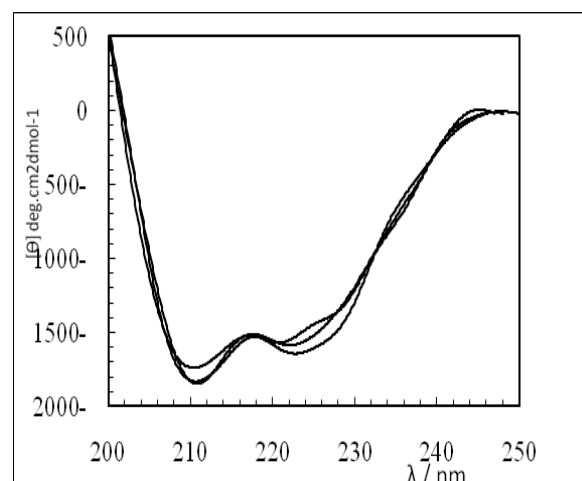


Figure 2. CD spectra of HSA in the presence of 100 mg/dl, 175 mg/dl, and also 400 mg/dl concentrations of glucose in HEPES buffer 100 mM, pH 7 at 42°C. Glucose concentration decreases from up to down.

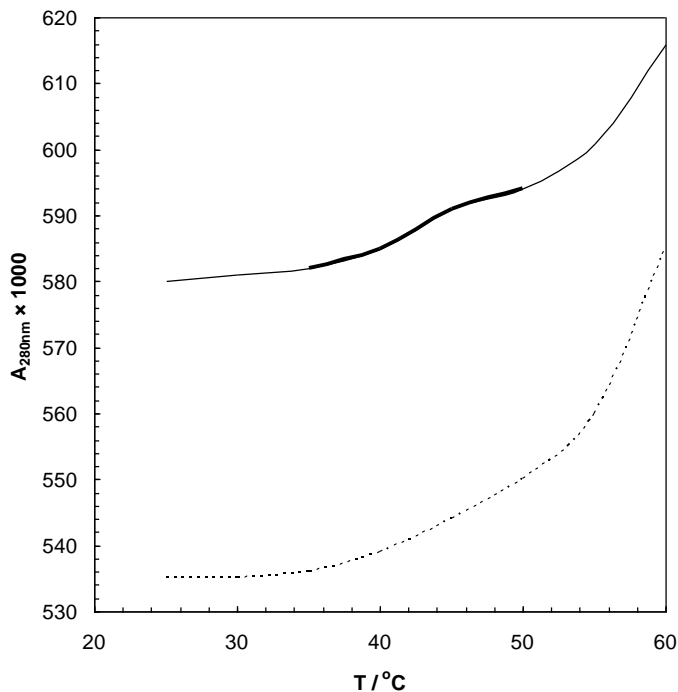


Figure 3. Acetaminophen induces conformational change in albumin, so the solid part of albumin absorption curve (up curve) is not seen in the presence of drug (low curve) [20].

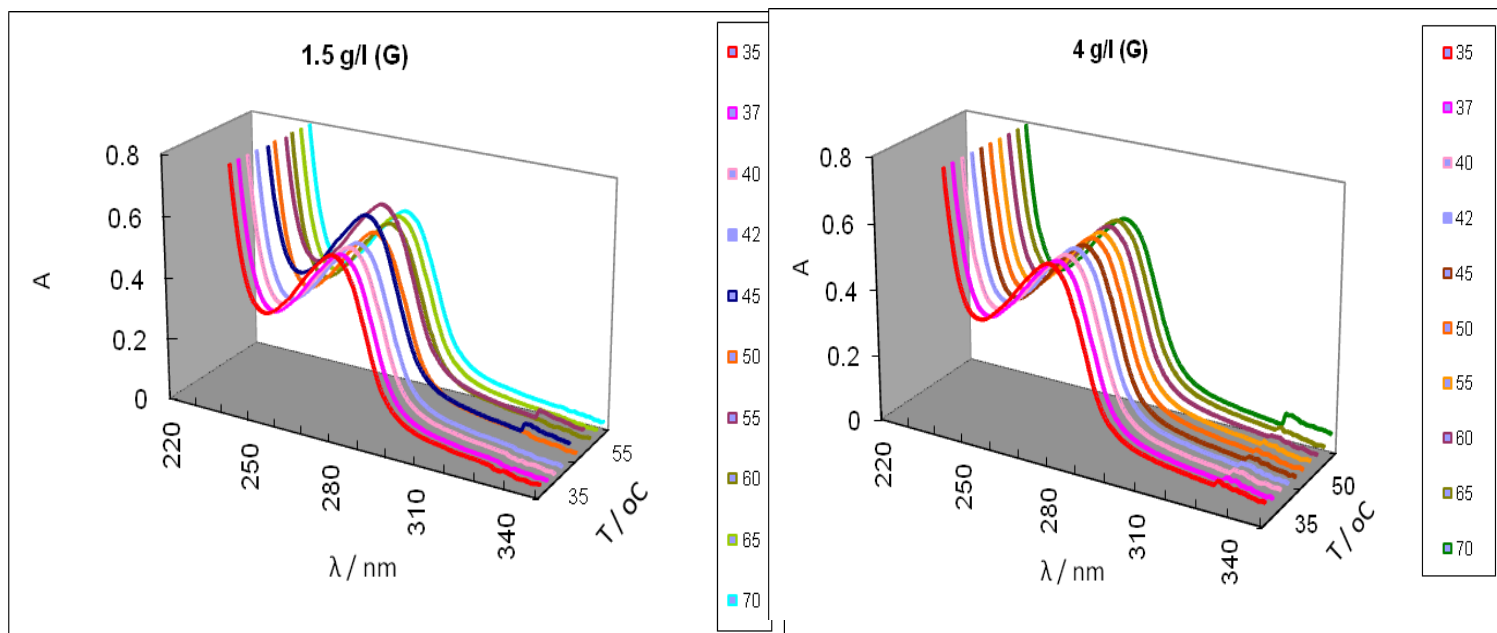


Figure 4. UV spectra of human serum albumin in the presence of 150 mg/dl concentration of glucose in the range of temperatures, Hepes buffer 100 mM and pH 7.

Figure 5. UV spectra of human serum albumin in the presence of 400 mg/dl concentration of glucose in the range of temperatures, Hepes buffer 100 mM and pH 7.

DISCUSSION

HSA as high abundance protein in our blood has particular tasks [22-24]. HSA has remarkable ligand binding despite being monomeric; in fact, endogenous and exogenous HSA ligand binding modulation is included by ligand-ligand competition for the same site as well as by intermolecular communication(s) within multiple clefts. HSA, furthermore, signifies a prominent biomarker for many diseases such as cancer, ischemia, severe acute graft-versus-host disease, and sicknesses that require monitoring glycemic control. Here secondary structural changes associated with incubated HSA with glucose in the 100, 175 and 400 mg/dl range in both 37°C and 42°C has been studied. Near UV-CD is applied widely for the study of tertiary structural changes of protein while far UV-CD is one of the best ways for studying the secondary structural changes of proteins during phase transition [25, 26]. Accordingly, far UV-CD technique was employed for the determination of the effect of glucose on the secondary structure of HSA [27]. It can be suggested that, glucose at high concentration which is the exact dosage that diabetic patients dealing with, shows a highly modification in secondary structure of albumin which is basically related to β sheets and α helix of this molecule. Our calculation shows that albumin loses about 4% α -helix component in the presence of 400 mg/dl concentration of glucose at 37°C. It is reported that secondary structural change may be accompanied by alteration of molecular function [1, 28], so it is an important process that glucose induces in albumin structure. The 100 mg/dl as normal dose of glucose in human body and control, 175 mg/dl concentration of glucose as a glucose tolerance point for kidney and also 400 mg/dl as the critical point for

REFERENCES

1. Rezaei-Tavirani M, Moghaddamnia SH, Ranjbar B, Zolfaghari SN. The Effects of Acetaminophen on Human Serum Albumin (HSA). *Iranian Journal of Pharmaceutical Research*. 2005;4:239-44.
2. Gabriella Fanali AdM, Viviana Trezza, Maria Marino, Mauro Fasano, Paolo Ascenzi. Human

glucose in diabetic condition are selected. As it is depicted in the figure 2 this structural transition is not seen at 42°C temperature. The 42°C temperature corresponds to severe condition of fever in human body. Previous study [6] showed that 42°C changes conformational structure of albumin in a reversible manner. In both cases conformational changes are accompanied by mild alteration of α -helix component of albumin.

As it is shown in figure 3 [20], UV spectroscopy method is a suitable technique for analysis of protein structural change in some cases, on the other hand CD findings confirmed relationship between glucose concentrations, temperature and Albumin conformational change; here UV spectra (see figures 4 and 5) are taken and compared with CD results. UV findings indicate that albumin structure in the presence of sub threshold dosage of glucose has a flexible unit and responses to temperature alteration in pre-denaturation range, but arising glucose concentration to 400 mg/dl diminishes this ability of albumin. This finding verifies CD findings. To sum up, it is suggested that thermal conformational changes may be alike to conformational changes of albumin in the presence of glucose at 37°C.

CONCLUSION

It can be concluded that, high doses of glucose such as diabetic condition can possibly effect on albumin structure and function. It seems that this effect of glucose is similar to the effect of fever on structure and function of albumin.

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serum albumin: From bench to bedside *Molecular Aspects of Medicine*. 2012;33:209-90.

3. Rondeau P, Bourdon E. The glycation of albumin: Structural and functional impacts. *Biochimie*. 2011;93:645-58.

4. Shahani M, Daneshi-Mehr F, Tadayon R, Hoseinzade Salavati B, Akbar Zadeh-Baghban AR, Zamanian A, et al. Glucose and Flouretin

- induce fine structural change in human serum albumin. *Iranian Journal of Pharmaceutical Research*. 2012;12: 185-191.
5. Prinsen BHCMT, de Sain-van der Velden MGM. Albumin turnover: experimental approach and its application in health and renal diseases. *Clinica Chimica Acta*. 2004;347:1-14.
 6. Rezaei Tavirani M, Moghaddamnia SH, Ranjbar B, Amani M, Marashi SA. Conformational study of human serum albumin in pre-denaturation temperatures by differential scanning calorimetry, circular dichroism and UV spectroscopy. *J Biochem Mol Biol*. 2006;39:530-6.
 7. Curry S. Lessons from the crystallographic analysis of small molecule binding to human serum albumin. *Drug. Metab. Pharmacokinet*. 2009;24:342-57.
 8. Fasano M, Curry, S., Terreno, E., Galliano, M., Fanali, G., Narciso, P., Notari, S., . The extraordinary ligand binding properties of human serum albumin. *IUBMB Life Ascenzi*. 2005; 57:787-96.
 9. Ascenzi P, Fasano, M. Allostery in a monomeric protein: the case of human serum albumin. *BiophysChem*. 2010;148:16-22.
 10. Kisugi R, Kouzuma T, Yamamoto T, Akizuki S, Miyamoto H, Someya Y, et al. Structural and glycation site changes of albumin in diabetic patient with very high glycated albumin. *Clinica Chimica Acta*. 2007;382:59-64.
 11. Guerin-Dubourg A, Catan A, Bourdon E, Rondeau P. Structural modifications of human albumin in diabetes. *Diabetes & Metabolism*. 2012;38:171-8.
 12. Uchida J, Iwai T, Kuwabara N, Machida Y, Iguchi T, Naganuma T, et al. Glucose intolerance in renal transplant recipients is associated with increased urinary albumin excretion. *Transplant Immunology*. 2011;24:241-5.
 13. Viberti GC, Pickup JC, Jarrett RJ, Keen H. Effect of control of blood glucose on urinary excretion of albumin and beta2 microglobulin in insulin-dependent diabetes. *The New England Journal of Medicine*. 1979;300:638-641.
 14. Mohamadi-Nejad A, Moosavi-Movahedi AA, Hakimelahi GH, Sheibani N. Thermodynamic analysis of human serum albumin interactions with glucose: insights into the diabetic range of glucose concentration. *The International Journal of Biochemistry & Cell Biology*. 2002;34:1115-24.
 15. Chen YH, Yang, J. T. and Martinez, H. M. . Determination of the secondary structures of proteins by circular dichroism and optical rotatory dispersion. *Biochemistry*. 1972;11:4120-31.
 16. Biglar M, Soltani K, Nabati F, Bazl R, Mojab F, Amanlou M. A Preliminary Investigation of the Jack-bean Urease Inhibition by Randomly Selected Traditional used Herbal Medicine. *Iranian Journal of Pharmaceutical Research*. 2012;11:831-7.
 17. Mergny JL, Phan AT, Lacroix L. Following G-quartet formation by UV-spectroscopy. *FEBS letters*. 1998;435:74-8.
 18. Korshin GV, Li CW, Benjamin MM. Monitoring the properties of natural organic matter through UV spectroscopy: a consistent theory. *Water Research*. 1997;31:1787-95.
 19. Calandra P, Goffredi M, Liveri VT. Study of the growth of ZnS nanoparticles in water/AOT/n-heptane microemulsions by UV-absorption spectroscopy. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 1999;160:9-13.
 20. Rezaei-Tavirani M, Moghaddamnia SH, Ranjbar B, Namaki S, Zolfaghari P. The Effects of acetaminophen on human serum albumin (HSA). *Iranian Journal of Pharmaceutical Research*. 2010;4:239-44.
 21. Hesami Takallu S, Rezaei Tavirani M, Kalantari S, Amir Bakhtiarvand M, Mahdavi SM. Co-amoxiclav Effects on the Structural and Binding Properties of Human Serum Albumin. *Iranian Journal of Pharmaceutical Research*. 2010;9:251-7.
 22. Guevara M, Terra C, Nazar A, Solà E, Fernández J, Pavesi M, et al. Albumin for bacterial infections other than spontaneous bacterial peritonitis in cirrhosis. A randomized, controlled study. *Journal of Hepatology*. 2012;57:759-65.
 23. Safari MR, Sheikh N, Mani Kashani K. Study on the Effect of Vitamin C on the In Vitro Albumin Glycation Reaction. *Iranian Journal of Pharmaceutical Research*. 2010;5:275-9.
 24. Jahangard-Rafsanjan Z, Javadi MR, Torkamandi H, Alahyari S, Hajhossein Talasaz A, Gholami K. The Evaluation of Albumin Utilization in a Teaching University Hospital in

Iran. Iranian Journal of Pharmaceutical Research. 2011;10:385-90.

25. Rezaei-Tavirani M, Moosavi-Movahedi AA, Moosavi-Nejad SZ, Chamani J, Ajloo D. Domain analysis of human apotransferrin upon interaction with sodium n-dodecyl sulphate: differential scanning calorimetry and circular dichroism approaches. *Thermochimica Acta*. 2003;408:9-16.

26. Rezaei-Tavirani M, Moosavi-Movahedi AA, Saboury AA, Hakimelahi GH, Ranjbar B, Housaindokht MR. Thermodynamic domain analysis of fresh and incubated human

apotransferrin. *Thermochimica Acta*. 2002;383:103-8.

27. Ramasamy T, Khandasamy US, Shanmugamb S, Ruttalad H. Formulation and Evaluation of Chondroitin Sulphate Tablets of Aceclofenac for Colon Targeted Drug Delivery. *Iranian Journal of Pharmaceutical Research*. 2012;11:465-79.

28. Streicher E, Bergval I, Dheda K, Böttger E, van Pittius NCG, Bosman M, et al. Mycobacterium tuberculosis population structure determines the outcome of genetics-based second-line drug resistance testing. *Antimicrobial agents and chemotherapy*. 2012;56:2420-7.