

Medicinal uses and chemistry of flavonoid contents of some common edible tropical plants

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ABSTRACT

Flavonoids are wide range of phytochemical with various pharmacological effects including antioxidant, anti-inflammation, anti-platelet, anti-allergic, cytotoxicity, reduce risk for heart disease or cancer etc. These flavonoids inhibited various enzymes such as prostaglandin synthase, lipoxygenase, cyclooxygenase, and induced detoxifying enzyme systems such as glutathione S-transferase. Flavonoids have significant vitamin C sparing activity and powerful antioxidants than the traditional vitamins such as quercetin, myricetin, rutin and apigenin. In vegetables, quercetin glycosides predominate, but glycosides of kaempferol, luteolin, and apigenin are also present. Fruits exclusively contain quercetin glycosides, whereas kaempferol and myricetin glycosides are found only in trace quantities. In this article we mainly discussed on three major flavonols (kaempferol, quercetin, and myricetin) and two major flavones (luteolin and apigenin) of edible tropical plants.

Keywords: Flavonoids; myricetin; quercetin; kaempferol; luteolin; apigenin

INTRODUCTION

Today we appears to be increasingly interested in the health benefits of foods and look beyond the basic nutritional benefits of food stuffs to disease prevention and health enhancing ingredients. Traditional systems of medicine be indebted their significance to the bioactive components from natural sources and most of them were associated with food habits. Plants have aromatic substances, most of which are phenols or their oxygen substituted derivatives [1]. Most are secondary metabolites, of which at least 12000 have been isolated, a number estimated to be less than 10% of the total [2]. In many cases these substances serve as plant defense mechanism against microorganisms, insects and herbivores. Polyphenols are widely distributed plant derived dietary constituents and implicated as the active components in a number of medicines [3]. More than 5000 plant polyphenols have been identified and several of them are known to possess a wide spectrum of biological activities [4] such as antiinflammatory, anti-microbial, anti-

carcinogenic, anti-HIV, cardioprotective and neuroprotective influence etc. In view of their wide range of pharmacological activities they seem to have a great therapeutic potential.

Chemical Structure

Flavonoids are the major polyphenols present in wide variety of plants. The basic structure of flavonoids contains a heterocyclic skeleton of flavan (2-phenylbenzopyrane). The structure is represented by a benzene ring (A), condensed with a heterocyclic six membered pyran or pyrone ring (C), which in the 2 or 3 position carries a phenyl ring (B) as a substituent (Figure 1). The constituent polyphenolic units are derived from the secondary plant metabolism of the shikimate pathway [5]. Flavonoids are often hydroxylated at positions 3,5,7,2',3',4',5'. These flavonoids exist in conjugated forms, the most common being the glycosides. When glycosides are formed, the glycosidic linkage is normally located at position 3 or 7 and the carbohydrate moiety can be L rhamnose, D-glucose, gluco-rhamnose, galactose or arabinose [6].

Sources and Bioavailability

For any chemical moiety to exert a biological effect, it should be bioavailable and have the potential to exert effects *in vivo*. Most of the polyphenols are known to be readily absorbed [7,8] but these compounds are prone to be modified into other forms inside biological systems, one such common chemical modification being conjugation [9]. Curcumin undergoes metabolic O-conjugation to curcumin glucuronide and curcumin sulfate and bioreduction to tetrahydrocurcumin, hexahydrocurcumin, and hexahydrocurcuminol [10]. Certain curcumin metabolites, such as tetrahydrocurcumin, possess anti-inflammatory [11] and antioxidant activities [12] similar to those of their metabolic progenitor. Dietary resveratrol is rapidly absorbed and predominantly present in plasma as glucuronide and sulphate conjugates. When administered in food, such as wine or grape juice, resveratrol metabolism is significantly inhibited by other polyphenols due to competitive reactions with metabolizing phase-II enzymes resulting in an increased concentration of the free form [13]. Isoflavones such as genistein are undergo conjugation with glycosides and is metabolized in human intestine to dihydrogenistein and 6'-hydroxy-O-desmethylangolensin. Concentration of genistein has been shown to be higher in individuals consuming soy rich diet [14] and as a result genistein and its metabolites have been detected in plasma, breast aspirate and prostatic fluid [15]. Similarly, other polyphenols are also known to be absorbed and metabolized into various end products which may or may not possess the biological effects of the parent compound.

Antioxidant compounds in food play an important role as a health-protecting factor. Antioxidants reduce the risk for chronic diseases including cancer and heart disease. Primary sources of naturally occurring antioxidants are whole grains, fruits and vegetables. Plant sourced food antioxidants like vitamin C, vitamin E, carotenes, phenolic acids, phytate and phytoestrogens having the potential to reduce disease risk. Most of the antioxidant compounds in a typical diet are derived from plant sources and belong to various classes of compounds with a wide variety of physical and chemical properties. Some compounds, such as gallates, have strong antioxidant activity, while

others, such as the mono-phenols are weak antioxidants. The main characteristic of an antioxidant is its ability to trap free radicals. Highly reactive free radicals and oxygen species are present in biological systems from a wide variety of sources. These free radicals may oxidize nucleic acids, proteins, lipids or DNA and can initiate degenerative disease. Antioxidant compounds like phenolic acids, polyphenols and flavonoids scavenge free radicals such as peroxide, hydroperoxide or lipid peroxy and thus inhibit the oxidative mechanisms that lead to degenerative diseases. The number of studies suggesting that the antioxidants in fruits, vegetables, tea and red wine are the main factors for the observed efficacy of these foods in reducing the incidence of chronic diseases including heart disease and some cancers. Flavonoids possess wide range of biochemical and pharmacological effects including anti-oxidation, anti-inflammation, anti-platelet, anti-thrombotic action, and anti-allergic effects [16,17]. They can inhibit enzymes such as prostaglandin synthase, lipoxygenase, and cyclooxygenase, closely related to tumorigenesis [18], and induce detoxifying enzyme systems such as glutathione S-transferase [19]. Quercetin inhibited oxidation and cytotoxicity of low-density lipoprotein (LDL) [20], and can reduce risk for heart disease or cancer [21]. Quercetin, myricetin, and rutin are being more powerful antioxidants than the traditional vitamins [22]. Flavonols and flavones possess antioxidant and free radical scavenging activity in foods [23], and have significant vitamin C sparing activity [16], with myricetin being one of the most active. In vegetables, quercetin glycosides predominate, but glycosides of kaempferol, luteolin, and apigenin are also present. Fruits almost exclusively contain quercetin glycosides, whereas kaempferol and myricetin glycosides are found only in trace quantities. Flavonoids are polyphenols with diphenylpropane skeletons. The four major classes are the 4-oxoflavonoids (flavones, flavonols, etc.), anthocyanins, isoflavones, and the flavan-3-ol derivatives (catechin and tannins) [24]. Flavonoids are a large family of over 4000 secondary plant metabolites, many being present as sugar conjugates. Epidemiological studies have indicated a relationship between a diet rich in flavonols and a reduced incidence of heart disease [25]. Three major flavonols (kaempferol,

quercetin, and myricetin) and two major flavones (luteolin and apigenin) of some edible tropical plants are studied. These tropical plants were studied such as guava (*Psidium guajava*), Chinese cabbage (*Brassica oleracea*), cabbage (*Brassica oleracea*), broccoli (*Brassica oleracea*), cauliflower (*Brassica oleracea*), winged bean (*Psophocarpus tetragonolobus*), French beans (*Phaseolus vulgaris*), French peas (*Pisum sativum*), lady fingers (*Hibiscus esculentus*), string beans (*Vigna sinensis*), bell pepper (*Capsicum annum*), bird chili (*Capsicum frutescens*), green chili (*Capsicum annum*), red chili (*Capsicum annum*), carrot (*Daucus carota*), white radish (*Raphanus sativus*), garlic (*Allium sativum*), lemongrass (*Cymbopogon citratus*), betel (*Piper betle*), Chinese chives (*Allium odorum*), onion (*Allium fistulosum*), drumstick leaves (*Moringa oleifera*), wolfberry (*Lycium chinense*), limau purut (*Citrus hystrix*), red spinach (*Amaranthus gangeticus*), kangkung (*Ipomoea aquatica*), sweet potato (*Ipomoea batatas*), tapioca (*Manihot utilissima*), cashew (*Anacardium occidentale*), fern (*Diplazium esculentum*), papaya (*Carica papaya*), yam (*Colocasia esculentum*), pegaga (*Hydrocotyle asiatica*), selom (*Oenanthe javanica*), brinjal (*Solanum melongena*), angular loofah (*Luffa acutangula*), pumpkin (*Cucurbita maxima*), sengkuang (*Pachyrrhizus erosus*), snake gourd (*Trichosanthes anguina*), mung bean (*Phaseolus aureus*), soy bean (*Glycine max*), mint (*Mentha arvensis*), oyster mushrooms (*Pleurotus sajor-caju*), peria (*Momordica charantia*), petai (*Parkia speciosa*), asam gelugur (*Garcinia atroviridis*), black tea (*Camellia chinensis*), turmeric (*Curcuma longa*), plaintain (*Musa sapientum*), bunga kantan (*Phaeomeria speciosa*), perilla (*Perilla frutescences*). In these above edible plants, five major food flavonoids, viz quercetin, kaempferol, myricetin, apigenin, and luteolin, were determined in dried plant samples of the flavonoid glycosides.

Vegetables

The total phenol contents in the vegetable subgroup were very low as compared to berries. The major phenolics were chlorogenic and *p*-coumaric acids [26]. The difference in antioxidant activity between potato varieties may result partly from the presence of anthocyanins, although pelargonin has been reported to show poor antioxidant activity [27]. Methanolic extracts of

purple potatoes as well as other anthocyanin-rich plant products, such as blueberries, sweet cherries, purple sunflower hulls, and red onion scales, showed strong antioxidative activities in a \hat{a} -carotene bleaching method [28]. In the present study, the antioxidant activity order was approximately similar at both tested concentration levels, which indicates that the yield of the vegetable extract was good. However, while the total phenol content of many vegetable extracts was extremely small. Sugar removal could have increased the antioxidant activities of some of the vegetable extracts.

Cereals

The concentrations of total phenolic compounds in cereals were even lower than the concentrations in vegetables. Cereal extracts did not show remarkable antioxidant activity. Oat bran exerted a moderate effect. In oat flour, *p*-hydroxybenzoic, protocatechuic, vanillic, *trans-p*-coumaric, (*p*-hydroxyphenyl)-acetic, syringic, *trans*-sinapic, caffeic, and ferulic acids were found, with ferulic acid being the most abundant [29]. Ferulic acid is also the dominant form of phenolic acid in rye, wheat, and barley. Ferulic acid showed moderate, caffeic acid showed strong, and *p*-coumaric acid showed negligible antioxidant activity [30]. In addition, oats contain a series of anionic substituted cinnamic acid conjugates, avenanthramides, which also are potential antioxidants [31]. The processing seems to be important when evaluating the antioxidant activity of cereals as brans were more active than other products from the same cereal species. This is obviously due to the localization of the phenolics in the grain: the outer layers of the grain (husk, pericarp, testa, and aleurone cells) contain the greatest concentrations of total phenolic, whereas their concentrations are considerably lower in the endosperm layers. About 80% of the *trans*-ferulic acid of both rye and wheat grain was found in the bran [29].

Polyphenolics

Flavonoids are polyphenols, the most widespread class of metabolites in nature, and their distribution is almost ubiquitous. It is estimated that 100,000 to 200,000 secondary metabolites exist. They generate the majority of the natural phenolics, such as flavonoids. Although, monophenols, such as *p*-coumaric acid, are not polyphenols, they have been same properties and characteristics as functional

polyphenols. Although a large variety of plant phenols exists, most of these compounds arise from a common origin: the amino acids phenylalanine or tyrosine. This well established antioxidant activity of flavonoids is also responsible for other biological activities. For example, the anticancer activity of some compounds is due to their ability to scavenge free radicals, thus avoiding the early stages of cancer promotion [32]. Besides this mechanism, flavonoids have also been reported to act as anticancer agents, modulation of enzyme activity related to detoxification, oxidation and reduction, stimulation of the immune system and DNA repair, and regulation of hormone metabolism. Some other flavonoid classes as potent molecules for the treatment of other pathologies that do not involve in the antioxidant activity. The some isoflavones, whose estrogen-like capacity is now well established, is related with their similarity to estradiol estrogen for the treatment of conditions in which the agonist effect in estrogen receptors is beneficial, such as menopause conditions. The venoprotective properties of flavonoids are also explored for the enhancement of micro-circulation in pathological conditions in which this function is compromised [32].

Flavonoids

Flavonoids are characterized by a phenylbenzopyran chemical structure. The general structure includes a C15 (C6-C3-C6) skeleton joined to a chroman ring (benzopyran moiety). The heterocyclic benzopyran ring is known as the C ring, the fused aromatic ring as the A ring, and the phenyl constituent as the B ring. The A ring can be of two types: a phloroglucinol type that is *meta*-trihydroxylated, or a resorcinol type that is *meta*-dihydroxylated. The B ring can be monohydroxylated, *ortho*-dihydroxylated or vicinal-trihydroxylated. The center heterocycle most commonly exists in one of three forms: pyran, pyrilium, or γ -pyrone. According to the position of the aromatic ring to the benzopyrane moiety, flavonoids can be grouped in four classes: major flavonoids (2-phenylbenzopyrans), isoflavonoids (3-benzopyrans), neoflavonoids (4-benzopyranes) and minor flavonoids. In plants, these compounds occur in nearly all species. Increasingly, flavonoids have been reported to possess many useful properties, including anti-

inflammatory, oestrogenic, enzyme inhibition, antimicrobial, antiallergic, vascular and cytotoxic antitumour activity, but the antioxidant activity is, without a doubt, the most studied one attributed to flavonoids.

Berries and Fruits

Overall, berries exhibited high total phenolic contents and high antioxidant activity. Berries with a strong purple color, such as crowberry, aronia, bilberry, and whortleberry, had clearly higher phenolic contents than the yellowish rowanberries and cloudbberries. The strongly colored berries are rich in anthocyanins. The bilberry, cranberry, and cowberry are rich in flavonols, mainly quercetin, while strawberry, red raspberry, and cloudberry have an especially high content of ellagic acid, and rowanberry contains high amounts of ferulic acid [33]. In addition, bilberries are also especially rich in hydroxycinnamic acid derivatives, which have been shown to exert antioxidant activity [34,35]. Extracts of black currants along with red currants, blackberries, blueberries, and black and red raspberries possessed also a remarkably high scavenging activity toward chemically generated superoxide radicals [36]. The total phenol contents of the two apple varieties studied were almost similar but lower than the contents in berries. However, apples exerted strong antioxidant activities.

Flavonoid Contents

The contents of five types of flavonoids, namely myricetin, quercetin, luteolin, kaempferol, and apigenin, in methanol extracts of common edible plants. All the species showed significant amounts of flavonoids except sweet potato shoots, winged bean, string bean, petai, peria, bayam duri, betel leaves, plaintain flower, and oyster mushroom. The results clearly indicated that some of the tested plants were rich in these natural antioxidant flavonoids, and the qualities and quantities of the flavonoids in these plants seemed to be very different among the kinds of samples used.

Flavonoids in Some Consumed Plants

Some of the locally consumed plants such as papaya shoot, tapioca shoot, cashew shoot, pegaga, maman, kesom leaves, bunga kantan, and daun turi are found to be rich in flavonoid content. *Piper betel* (Piperaceae) leaves are chewed alone or with other plant materials including the areca nut, *Areca catechu* L. Seven phenols were identified in *P.*

betel flowers. Safrole was the major phenol, followed by hydroxychavicol, eugenol, methyl eugenol, isoeugenol, flavone, and quercetin [37]. Flavonoids such as isorhamnetin, hyperoside, and persicarin were isolated from *Oenanthе javanica* [38]. The fruits and leaves of box thorn or Wolfberry leaves (*Lycium chinense*) have been used as foods, tea, and medicine. Box thorn leaves are known to be capable of reducing the risk of certain diseases such as arteriosclerosis, essential arterial hypertension, diabetes, and nightblindness [39] contained the anti-aging ascorbic acid and tocopherols [40]. The box thorn leaves contained rutin (1.1-2.7% dry weight basis), a preventive phytochemical for hypertension and stroke. Rhizomes of *Curcuma spp.*, such as *C. longa* are used in traditional medicine. The rhizomes of *C. longa* are also used as a yellow coloring additive for food because it contains curcuminoids [41]. Flavonoid extracted from the fruits of *Solanum melongena* showed hypolipidemic action.

Flavonols in the Plants Tested

The quercetin is a potent anticancer agent in man. Myricetin, with its three adjacent hydroxyl groups was one of the most active antioxidants. Myricetin is not only a good antioxidant, but also been shown to be a potent anticarcinogen and antimutagen [42]. Quercetin is also a strong antioxidant that can contribute to the prevention of atherosclerosis [22,23]. Quercetin is a suppressing chemopreventive and chemotherapeutic agent that can relieve local pain caused by inflammation, headache, oral surgery, and stomach ulcer. Recently, quercetin has been shown to reduce the carcinogenic activity of several cooked food mutagens, enhance the antiproliferative activity of anticancer agents, and inhibit the growth of transformed tumorigenic cells [43]. Currently, kaempferol is in interest because of its antioxidant [22], antitumor, antiinflammatory, and antiulcer activity [44], and its inhibitory activity of HIV protease [45]. The flavones had the poorest antioxidant activity because of lack of *o*-dihydroxy groups. Kaempferol and its derivative have been identified in various vegetables, fruits, and beverages such as French beans [46], onions [47], teas [48], and honey [49-51]. there was variation in the levels of quercetin in cherry tomatoes purchased at different times. The concentrations of flavones and flavonols, like those of all secondary

plant metabolites, vary within certain limits and are dependent on a number of factors: for example, growing condition, degree of ripeness, size of the fruit, and variety [52]. The consumption of controlled diets high in fruits and vegetables increased significantly the antioxidant capacity of plasma, and the increase could not be explained by the increase in the plasma R-tocopherol or carotenoid concentration [53]. On a milligram-per-day basis, the intake of the antioxidant flavonoids still exceeds that of the antioxidant \hat{a} -carotene and vitamin E. Thus, flavonoids represent an important source of antioxidant activity in the human diet [54]. Supplementation of these natural antioxidants through a balanced diet could be much more effective and economical than supplementation of an individual antioxidant, such as ascorbic acid or R-tocopherol, in protecting the body against various oxidative stresses. Polyphenols are effective hydrogen donors, particularly flavonols such as quercetin [55]. Studies on the natural antioxidants flavonoids and phenolics compounds in temperate edible plants are quite established [56]. These foods include tea, alliums, tomatoes, lettuce, and celery [24,52], apples, vegetables, fruits, citrus fruit juices such as fresh orange, grapefruit, and lemon juices [54]; beer, coffee, chocolate milk, white wine, tea infusion, and red wine [57,58]; apple juice, tomato juice, grape juice, orange juice, grapefruit juice and lemon juice, cauliflower, radish, pea, broccoli, Chinese cabbage, and carrot [50]; sixteen leafy vegetables and fruits such as *Colocasia*, cabbage, and *Hibiscus sabdariffa*; lettuce, kale, chive, garlic chive, leek, horseradish, red radish, and red cabbage tissues; soybean sprouts, Japanese radish, grapefruit, and burdock root [51]; citrus fruits [59]; juices of orange, apple, pineapple, peach, apricot, pear, grape [58,60] and citrus fruits contain almost exclusively flavanones [59]. Quercetin was also found in some of the citrus fruit juices such as fresh orange, grapefruit, and lemon juices [48]. Flavanones, flavones, and flavonols are the flavonoids present in citrus. However, flavones and flavonols were in low concentration in citrus tissues in relationship to flavanones. These types of compounds have been shown to be powerful antioxidants and free radical scavengers [61].

Effect of Drying Temperature on Flavonoid Content

Phenolic compounds are usually susceptible to different factors (eg., acidic solution and high temperature) during the extraction process. Drying at temperatures below 50 °C yields the highest amount of total phenolics. Drying at room temperature may enhance the enzymatic degradation and thus lower the amount of phenolics in the samples. Increasing the temperature above 60 °C lowered the phenolic amount considerably. At high temperatures, certain phenolics may decompose/combine with the other plant components. Cooking lowered the quercetin content of both tomatoes and onions, with greater reduction being detected following microwaving and boiling than after frying [52]. This could be due to flavonoid breakdown during cooking and/or conjugated quercetin being extracted from the tomato and onion tissues by hot water more effectively than with hot oil used in the study. There are only little gross changes in either the overall level or the composition of quercetin glucosides during normal commercial storage [62]. Boiling and frying do not result in gross changes in glucosides composition, although an overall loss of up to 25% is found for both processes, in the former by leaching into the cooking water and in the latter by thermal degradation into products.

Therapeutic Potential

Exploring healing powers in plants is an ancient phenomenon. Traditional healers have long used plants to prevent or cure various diseased conditions. An insight into the investigations, both in vitro and in vivo, reveals the properties of plant polyphenols that could form the basis of their use in the prevention and cure of several disorders. Some of the important therapeutic properties of plant-derived polyphenols with strong evidences from the existing literature have been discussed below.

Anti-HIV Properties

Human immunodeficiency virus (HIV), the etiologic agent for acquired immunodeficiency syndrome (AIDS) has been the most successful pathogen to challenge the humans in the last three decades. Globally, about 39.5 million adults are living with the syndrome. In the past few years, several therapies have been tried but as of now, there is no conclusive treatment to eliminate this virus from the body once the infection has taken place. The efforts to develop vaccines against HIV

have not been successful so far due to their everchanging variants [63]. Flavonoids and their derivatives have been reported to inhibit the growth and development of HIV by interrupting at several stages of its life cycle. Derivatives of hesperidin, particularly sulphonated and phosphorylated forms, have been studied by various scientists as hyaluronidase inhibitors and antimicrobial agent [64]. Acute HIV-1 infection has been shown to be suppressed by certain flavonoids and evidence for inhibition of HIV-1 protease, integrase and reverse transcriptase by flavonoids also exists [65]. Anti-HIV activity of scutellarin has been reported against three strains of human deficiency virus including laboratory-derived virus (HIV-1 IIB), drug resistant virus (HIV-1 74V) and low passage clinically isolated virus (HIV-1 KM018) [66]. Scutellarin was found to inhibit several stages of HIV-1 replication with different potencies. It appeared to inhibit HIV-1 reverse transcriptase activity, HIV-1 particle attachment and cell fusion. In cells harboring proviral HIV-1 DNA, viral transcription represents a potential therapeutic target, if selective inhibitors can be developed [67]. Chrysin, a flavonoid has been characterized as a potent inhibitor of HIV-1 transcription in chronically infected cells [65]. The flavonoid halts the transcription by inhibiting casein kinase-II (CK-II) activity [68]. CK-II may regulate HIV-1 transcription by phosphorylating cellular proteins involved in HIV-1 transactivation. Isoflavones have also been shown to inhibit transcription by repressing HIV-1 promotor activity [69]. The multiple steps of the HIV-1 life cycle each lend themselves to potential therapeutic intervention. The interaction between the viral products and the host factors are critical to develop the host-pathogen relationship. HIV-1 cellular entry via binding to CD4 and chemokine receptors well defines the principle of HIV-1 and host factor interaction [70]. Epigallocatechin-3-gallate (EGCG), the major polyphenol in tea has been reported to bind with CD4 receptor of TH cells, thus interfering with its ability to interact with gp120, an envelope protein of HIV-1 [71]. Inhibition of viral adsorption by flavonoids such as epicatechin has been attributed to an irreversible interaction with gp120 [72]. This protective effect against HIV infection is mediated by inhibiting virions from binding to the target cell surface.

Recent studies documented that the beta-chemokine receptors, CCR2b, CCR3 and CCR5, and the alpha-chemokine receptors, CXCR1, CXCR2, and CXCR4 serve as entry coreceptors for HIV-1 [73]. Grape seed polyphenols, proanthocyanidins, have been shown to down regulate HIV-1 entry coreceptors CCR2b, CCR3 and CCR5 gene expression by normal peripheral blood mononuclear cells [74]. These studies have clinical significance since the ability of polyphenols to interfere at multiple target sites of HIV might determine their successful use against ever changing variants.

Antimicrobial Properties

Microbiologists and natural product chemists are exploring the Earth for phytochemicals, which could be developed for the treatment of infectious diseases [75]. Polyphenols particularly, flavonoids are found to be effective antimicrobial agents against a wide array of microorganism. This is probably due to their ability to complex with extracellular and soluble proteins and also with the bacterial cell wall [76]. Phenolics present in plants are known to be toxic to microorganisms [77]. Many plant extracts derived from different parts of the plant have been analyzed for their active constituents possessing antibacterial activities. Antibacterial activity of leaf and stem bark of *Pterocarpus santalinus* was investigated for both gram-positive and gram-negative bacteria [78]. The stem bark and leaf extracts showed inhibitory activity against a number of infectious microbial strains including *Enterobacter aerogenes* and *Staphylococcus aureus*. The broadspectrum antibacterial activity exhibited by *Pterocarpus santalinus* may be attributed to its richness in isoflavone glucosides [79]. Flavonoids are known to be synthesized by plants in response to microbial infections [80] and therefore, very obviously they have been found *In vitro* to be effective antimicrobial substance against a wide range of microorganisms. Catechins, an important group of flavonoids, have been extensively investigated due to their occurrence in oolong green teas. It has been reported in the past, that teas possess antimicrobial activity [81] and that they contain a mixture of catechin compounds. These compounds inhibited *Vibrio cholerae* as products of fermentation [82], *Streptococcus mutans* [83], *Shigella* [84] and other bacterial strains *in vitro*. The catechins have been

found to inactivate the cholera toxin from *V. cholerae* [76] and inhibit isolated bacterial glucosyl transferase in *S. mutans* [85]. Many of the flavonoids are known to be hydroxylated and the site(s) and number of hydroxyl groups associated with the rings are thought to be related to their relative toxicity to microorganisms, with evidence that increased hydroxylation results in increased toxicity [1]. Catechol and pyrogallol both of which are hydroxylated phenols, have been shown to be toxic to microorganisms. Inhibitory interaction between the polyphenols and protein or DNA has also been observed. Quinones are known to complex irreversibly with nucleophilic amino acids in proteins [86] often leading to inactivation of the protein via loss of function. This has been attributed for the wide range of quinone antimicrobial effects [87]. The above references have well established the antimicrobial properties of plant polyphenols supporting their use to combat infective diseases. While 25 to 50% of current pharmaceuticals are derived from plants, none are used as antimicrobials, since our choice had been restricted to bacterial and fungal sources for these activities. However, with the increasing complexities of antibiotic resistance, the use of antibiotic needs to be checked and antimicrobial agents from plant origin be given a favourable insight for their therapeutic use.

Cardioprotective properties

A longstanding tenet of nutrition holds that people with diets rich in fruits and vegetables enjoy better health than those eating few. Consequently, research has sought the components or compounds responsible for this apparent health benefit. Much of current research shows that free radicals are the connecting link between otherwise physiologically distinct diseases. As a result dietary antioxidants hold promise in at least delaying the onset/progression of these diseases. The "French Paradox"—the observation that mortality from coronary heart disease is relatively low in France despite relatively high levels of dietary saturated fat led to the idea that regular consumption of red wine (rich source of polyphenols) might provide additional protection from cardiovascular disease [88]. In the prevention of cardiovascular disease, many of the observed effects of polyphenols can therefore be attributed to their recognized antioxidant and radical scavenging properties,

which may delay the onset of atherogenesis by reducing chemically and enzymatically mediated peroxidative reaction [89]. Regular, moderate consumption of red wine is linked to a reduced risk of coronary heart disease [90]. Resveratrol, a component of red wine has been linked to a number of potentially cardioprotective effects [91]. Anthocyanidins have also been found to have antioxidant potential [92]. Studies suggest that EGCG can suppress reactive oxygen species and thereby prevent the development of cardiac hypertrophy [90]. Increase in LDL is taken as a parameter for the occurrence and susceptibility of cardiovascular diseases. Polyphenols such as dicvertin have been reported to produce a 12% decrease in LDL along with a 14% increase in HDL in coronary heart disease patients [93]. Lipid-lowering activity has also been reported in tea flavonoids [90]. Endothelial dysfunction is the pathophysiologic principle involved in the initiation and progression of arteriosclerosis. Some polyphenols have been shown to relax endothelium-denuded arteries. There have been several reports that extracts from grape and wine induce endothelium-dependent relaxation via enhanced and/ or increased biological activity of nitric oxide (NO) which leads to the elevation of cGMP levels [94]. Resveratrol has been found to promote vasodilation by enhancing the production of NO [95]. Genistein, one of the major isoflavones in soy protein, binds to estrogen receptor b with much higher affinity than to ERa [96] and can elicit endothelium dependant vasorelaxation in vitro [97] and in vivo [98]. Other isoflavones such as dihydrodaidzeins have also been reported to enhance endothelial function [99]. Flavonoids have also been found to be good hypochlorite scavenger in vitro and could have favorable effects in diseases such as atherosclerosis in which hypochlorite is known to play a significant role [100]. As documented above, it is evident that natural polyphenolic compounds possess antioxidant, vasorelaxant and antihypertensive properties that are beneficial to cardio-vascular health.

Neuroprotective properties:

Neurodegenerative disorders are a heterogeneous group of diseases of the nervous system, including the brain, spinal cord and peripheral nerves, which have different aetiologies. The multifactorial

etiology of these diseases suggests that interventions having multiple targets such as polyphenols could have therapeutic potential for them. Moreover, epidemiological studies indicate that dietary habits and antioxidants from diet can influence the incidence of neurodegenerative disorders such as Alzheimer and Parkinson's diseases [101]. The nervous system is rich in fatty acids and iron. High levels of iron can lead to oxidative stress via the iron-catalyzed formation of ROS [102]. In addition brain regions that are rich in catecholamines are vulnerable to free radical generation. One such region of the brain is the substantia nigra, where a connection has been established between antioxidant depletion and tissue degeneration [103]. There is substantial evidence that oxidative stress is a causative or at least an ancillary factor in the pathogenesis of many neurodegenerative diseases, including Alzheimer's disease (AD), Parkinson's disease (PD), Amyotrophic lateral sclerosis (ALS) [104], Huntington's disease (HD) and Schizophrenia [105]. Flavonoids exhibit biological effects such as anti-inflammatory, antioxidant and metal chelating properties, which augment their role in neuroprotection. Reports also suggest that red wine that contains high levels of antioxidant polyphenols reduces the incidence of AD [106]. Polyphenols such as EGCG, curcumin, extracts of blue berries and Scutellaria are also known to help in AD [107]. In vitro studies show that green tea extract rich in catechins could protect neurons from the amyloid beta-induced damages in AD [108]. EGCG is also found to be of use in ALS [109,110] and PD [111]. Extract of Scutellaria stem and polyphenols such as curcumin and naringenin also exhibit neuroprotection in PD [112]. Alzheimer's disease is characterized by chronic inflammation and oxidative damages in the brain. Curcumin possesses antioxidative and anti-inflammatory properties and has thus been shown to exert a protective effect against oxidative damages initiated by divalent metals or suppress inflammatory damage and also inhibits amyloid beta fibril formation [113]. Dietary polyphenols have potential as protective agents against neuronal apoptosis, through selective actions within stress-activated cellular responses including protein kinase signaling cascade [114]. Several dietary supplements with blueberries extracts have been

reported to reduce some neurological deficits in aged animal models. Blueberries are a rich source of polyphenols such as catechins, epicatechins and anthocyanins. Recent studies investigating the effect of polyphenols in cognitive performance have demonstrated that dietary supplementation with blueberries extracts reversed cognitive deficits in Morris water maze performance test and Y-maze test in aged mice models [115,116]. The studies therefore present a promising class of compound for possible application as health supplements and nutraceuticals for neuroprotection.

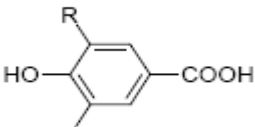
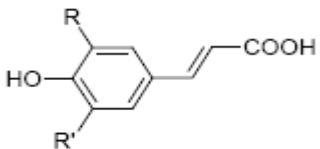
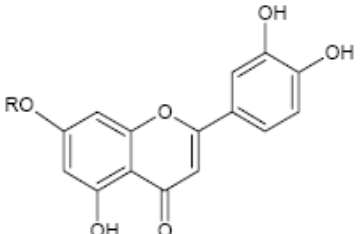
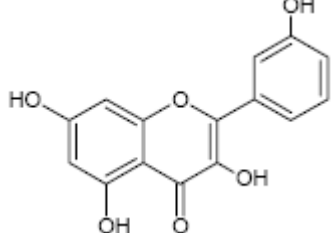
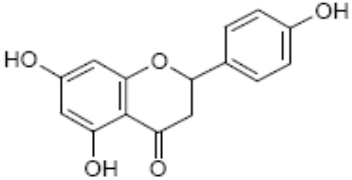
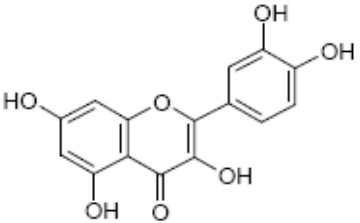
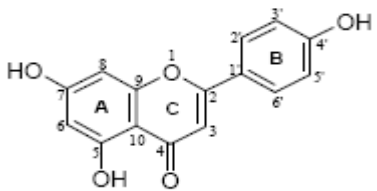
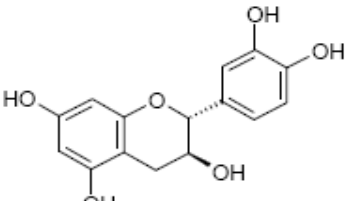
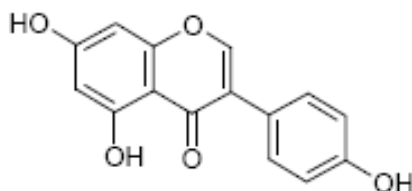
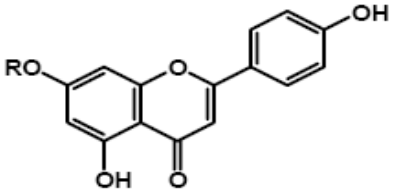
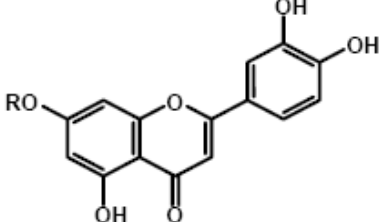
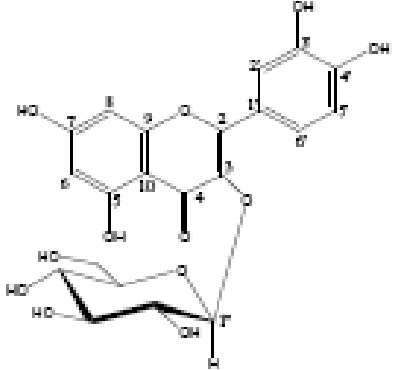
Anti-Carcinogenic Properties

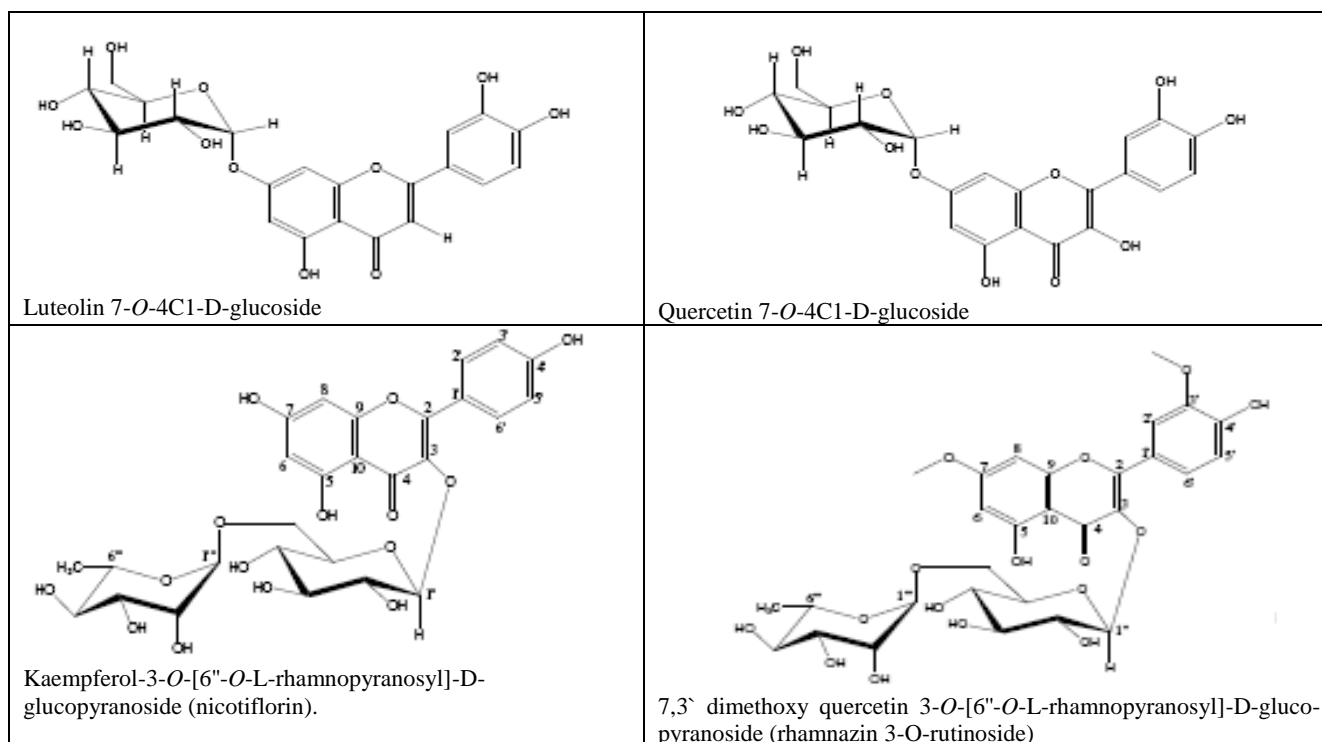
Evidence in the literature suggests that the differential regulation (progression and prevention) of several disorders may be characterized through the food habits prevalent in different geopolitical regions of the world. Soybean food comprises a significant portion of Asian diet, providing 10% of the total per capita protein intake in Japan and China where the incidence of breast and prostate cancer is much less than that in the United States [117]. Various classes of dietary polyphenols are under investigation for their anticancer properties in order to design novel strategies for chemoprevention [118]. Studies have found that genistein, an isoflavone from soy can inhibit the growth of various cancer cell lines including leukemia, lymphoma, prostate, breast, lung and head and neck cancer cells [119]. Pretreatment with genistein, potentiated cell killing induced by radiation in human PC-3 prostate cancer cells in vitro [120] and prostate tumor growth in vivo [121]. Another isoflavone present in soy, biochanin A has been found to have cytotoxic effect on cell growth in the mammary carcinoma cell line MCF-7 [122], myeloid leukemia [123] and pancreatic tumor cells [124]. Biochanin A also induces a dose dependent inhibition of proliferation in LNCaP-cell and [3H] thymidine incorporation that is correlated with increased DNA fragmentation, indicative of apoptosis [125]. Protein tyrosine kinases (PTKs) are known to play key roles in carcinogenesis, cell growth and apoptosis [126]. Genistein has been identified as a PTK inhibitor [127]. Transcription of genes is critical to cell growth and proliferation. Transcription factors interact with enhancer and promoter regions of target genes, which allow the binding of RNA polymerase and initiation of gene

transcription process [128]. Investigations have revealed that genistein treatment could inhibit DNA binding activity of a major transcription factor NF- κ B in PC3 and LNCaP prostate cancer cells [129]. Curcumin, a natural phenolic compound found in turmeric have been shown to have antiproliferative action against colon cancer, breast cancer and myeloid leukemia [130,131]. Antitumor activity of curcumin is believed to be in part due to its ability to block the NF kappa B pathway [132]. Other studies have shown that curcumin inhibits cell growth and induces apoptosis in MCF-7, a human breast carcinoma cell line through modulation of insulin-like growth factor-1 (IGF-1) system, including IGFs (IGF-1 and IGF-2), IGF-1R (IGF-1 receptor) and IGF-BPs (IGF binding proteins), which have been implicated to play a critical role in the development of breast cancer [133]. Resveratrol, the phenol antioxidant found in berries and grapes has been reported to possess anticancer properties (Aggarwal et al., 2004) and is able to inhibit the formation of prostate tumors by acting on the regulatory genes such as p53 [134]. Androgen independent DU145 human prostate cancer cells manifest resistance to radiation-induced apoptotic death [135]. Scarlatti et al have reported that pre-treatment with resveratrol significantly enhances radiation induced cell death in DU145 cells [136]. Some polyphenols are known to inhibit the cancerous growth by arresting the cell cycle progression rather than inducing apoptosis. Genistein has been demonstrated to induce a G2/M cell cycle arrest in breast cancer cells, gastric adenocarcinoma cells and human melanoma cells [137]. Citrus fruit flavonoids, tangeretin and nobiletin have also been shown to inhibit human breast cancer cell lines MDAMB-435 and MCF-7 and human colon cancer cell line HT-29 by blocking cell cycle progression at G1 stage of the cell cycle [138]. Such flavonoids which inhibit the growth and proliferation of cancer cell lines by arresting cell cycle are cytostatic and significantly block the proliferation without apoptosis. Inhibition of proliferation of human cancers without inducing cell death may be advantageous in treating tumors as it would restrict proliferation in a manner less likely to induce cytotoxicity and death in normal, non tumor tissues. Inhibition of tumor invasion and angiogenesis by popular flavonoids such as

luteolin may also account for the antiproliferative properties of plant polyphenols [139].

Figure 1. Chemical Structures of major Polyphenolic Compounds

Chemical classes of polyphenolic compounds.		
<p>PHENOLIC ACIDS <i>Benzoic acid derivatives</i></p>  <p>R = R' = H; <i>p</i>-hydroxybenzoic acid R = OH, R' = H; protocatechuic acid R = OCH₃, R' = H; vanillic acid R = R' = OH; gallic acid R = R' = OCH₃; syringic acid</p>	<p><i>Cinnamic acid derivatives</i></p>  <p>R = R' = H; <i>p</i>-coumaric acid R = OH, R' = H; caffeic acid R = OCH₃, R' = H; ferulic acid R = R' = OCH₃; sinapic acid</p>	 <p>R = H, Luteolin R = Glc, Luteolin-7-O-glucoside</p>
 <p>Kaempferol</p>	<p><i>Flavanones</i></p>  <p>Naringenin</p>	<p><i>Flavonols</i></p>  <p>Quercetin</p>
<p>FLAVONOIDS: Flavones</p>  <p>Apigenin</p>	<p><i>Flavan-3-ols</i></p>  <p>(+)-catechin</p>	<p><i>Isoflavones</i></p>  <p>Genistein</p>
 <p>R = H Apigenin R = Glu-Caf Apigenin 7-O-caffeoylglucoside R = GlcU-GlcU Apigenin 7-O-diglucuronide</p>	 <p>R = H Luteolin R = GlcU Luteolin 7-O-glucuronide R = GlcU-GlcU Luteolin 7-O-diglucuronide</p>	 <p>Quercetin 3-O-4C1-D-glucopyranoside</p>



Another convincing antiproliferative mechanism includes the oxidative DNA breakage by the prooxidant action of plant polyphenols in the presence of transition metals especially copper [140-142]. Flavonoids are recognized as naturally occurring antioxidants and this property has been implicated for their anticancer activity [143]. However, evidence in the literature suggests that antioxidant properties of plant polyphenols may not fully account for their anticancer effects [144]. In the context of copper being an essential constituent of chromatin (Bryan, 1979) and that the copper levels in tissues [145] and serum [146] are considerably elevated in various malignancies, the mechanism of oxidative DNA breakage holds significance. In addition to their potential as anticancer agents, an important role of plant polyphenols as natural modulators of cancer multidrug resistance (MDR) has been realized recently [147]. Resistance of recurrent disease to cytotoxic drugs is the principal factor limiting long-term treatment success against cancer. Flavonoids, a major class of plant polyphenol has been found to inhibit breast cancer resistance protein (BCRP), an ABC transporter, which plays an important role in drug disposition leading to chemoresistance in breast cancer [148].

Isoflavones such as biochanin A, daidzein [149] and green tea polyphenol EGCG [150] have also been shown to exhibit anti MDR activities in various drug resistant cancer cell lines such as doxorubicin resistant KB-A1 cells through the inhibition of P glycoprotein transporters. Curcumin has been reported to induce apoptosis in chemoresistant ovarian cancer cell lines SKOV₃ and ES-2 [151]. The above findings suggest that the plant polyphenols have indeed emerged as an area of great promise for delineating innovative strategies in cancer chemoprevention.

DISCUSSION

The importance of dietary supplements for their therapeutic and preventive bioactive components due to their elevated margin of safety and desired range of efficacy. The above observation made centuries ago has now gained scientific verifications with epidemiological studies showing that the incidence of cancer and cardiovascular diseases are least in countries like India and China where vegetables, fruits and spices form an essential part of human diet. With regard to the extensive consumption of polyphenols in the diet, the biological activity of these compounds is an important area of scientific

investigation. Given the potential therapeutic tendencies of these compounds, one would expect to observe their favourable effects in human population. The investigations, both in vitro and in vivo provide a definite link between the dietary intake of Plant polyphenols mobilize endogenous copper in human peripheral polyphenols and their associated health benefits. However, the issue of bioavailability has to be addressed before any targeted therapy could be designed effectively [152-160]. On the basis of what is known about the bio availabilities, it seems likely that the organ sites that are most accessible to dietary polyphenols experience the protective effects of these compounds. Moreover, another way is to explore the biofactors that in combination with dietary polyphenols could stabilize and enhance their effects even under limited bioavailability. Nature has gifted us with numerous natural products, which we consume as food and which are the armamentarium of bioactive substances having diverse activities. Since diseases like cancer are multifactorial phenomenon in which many normal cellular pathways become aberrant, it is highly unlikely that one agent could prove effective against such disorders. In this regard foods, unlike drugs may have the advantage of simultaneously influencing various pathways that go awry in diseases like cancer. Another aspect that needs to be explored is that why the excessive use of fruits and vegetables is not harmful although they are routine part of human diet whereas an isolated compound may show detrimental effects as projected in various studies. Although, it is well understood that dose does determines whether a substance acts as a toxicant or not, it is important to note the significance of synergism among the components that are present together in a particular food [160-165]. In this respect nutrient-nutrient interactions and synergism are required to be studied to augment their beneficial effects or otherwise reduce the side effects. The concept of food as medicine needs to be propagated to ensure healthy food habits. However, for better and mechanism-based understanding of the potential health benefits of dietary polyphenols further studies are warranted. Flavanoids compounds in food play an important role as a health-protecting factor and reduce the risk for various chronic diseases including cancer

and heart diseases. Primary sources of naturally occurring flavanoids are whole grains, fruits and vegetables. Edible plant sourced food antioxidants like vitamin C, vitamin E, carotenes, phenolic acids, phytate and phytoestrogens have been recognized as having the potential to reduce disease risk. Most of the flavanoids in a diet are derived from plant sources and belong to various classes of compounds with a wide variety of physical and chemical properties. Some compounds, such as gallates, have strong antioxidant activity, while others, such as the mono-phenols are weak antioxidants. The main characteristic of an antioxidant is its ability to trap free radicals. Highly reactive free radicals and oxygen species are present in biological systems from a wide variety of sources. These free radicals may oxidize nucleic acids, proteins, lipids or DNA and can initiate degenerative disease. Antioxidant compounds like phenolic acids, polyphenols and flavonoids scavenge free radicals such as peroxide, hydroperoxide or lipid peroxy and thus inhibit the oxidative mechanisms that lead to degenerative diseases [166-170]. There are a number of clinical studies suggesting that the flavanoids are the main factors for the observed efficacy of these foods in reducing the incidence of chronic diseases including heart disease and some cancers.

CONCLUSION

The last two decades have witnessed a major drift in the interests of the scientific community towards explaining better means to containing the health risks of the human race. The century old chemotherapies against various disorders have never been a success, albeit not a total failure. Such therapies have a major drawback of side effects that give rise to unseen disorders that emerge as a new challenge. In this regard, the concept of foodstuffs as natural medicines is very attractive. Epidemiological studies suggest that the vegetarian food habit is associated with reduced risk of cancer, cardiovascular and neurodegenerative disorders. Consistent with this hypothesis is the fact that the incidence of these disorders is least in Asian populations where fruits, vegetables and spices are the major elements in the human diet. Recent research has shown that plant-derived polyphenolic

compounds are promising nutraceuticals for control of various disorders such as cardiovascular, neurological and neoplastic disease. The richness of the polyphenolic contents of green tea and red wine has made them popular

choice for associated anticancer and cardiovascular health benefits. The present article is a brief review of the promises plant polyphenols, bioactive components of our food, hold for the future.

REFERENCES

1. Geissman TA. Flavonoid compounds, tannins and related compounds, In: Florkin, M. and Stotz, E.H., (Ed), Pyrrole Pigments, Isoprenoid Compounds and Phenolic Plant Constituents. Elsevier, New York, 1963: 265.
2. Schultes RE: The kingdom of plants, In: WAR Thomas (Ed), Medicines from the Earth. Mc Graw Hill Book Co, New York 1978: 208.
3. Wollenweber E. Occurrence of flavonoid aglycones in medicinal plants. *Prog Clin Biol Res*, 1988; 280, 45-55.
4. Beretz A, Anton R and Stoclet JC. Flavonoid compounds are potent inhibitors of cyclic AMP phosphodiesterase. *Experimentia*, 1977; 34: 1045-55.
5. Dewick PM (1995). The biosynthesis of shikimate metabolites. *Nat Prod Rep*, 1995; 12, 579-607.
6. Middleton E. The flavonoids. *Trends Pharmacol Sci*, 1985; 5: 335-8.
7. Scalbert A and Williamson G. Dietary intake and bioavailability of polyphenols. *J Nutr*, 2000; 130: 2073-85.
8. Rowland I, Faughnan M L, Hoey K, et al. Bioavailability of phyto-oestrogens. *Br J Nutr*, 2003; 89: 838-52.
9. Lambert DJ, Hong J, Yang G, Lias J, Yand SC. Inhibition of carcinogenesis by polyphenols evidence from laboratory investigations. *AJCN*, 2005; 81: 2845-91.
10. Ireson CR, Orr S, Jones DJL, et al. Characterization of metabolites of the chemopreventive agent curcumin in humans and rat hepatocytes and in rat plasma and evaluation of their ability to inhibit phorbol ester-induced prostaglandin E2 production. *Cancer Res*, 2001; 61: 1058-64.
11. Sugiyama Y, Kawakashi S, Osawa, T. Involvement of the beta-diketone moiety in the antioxidant mechanism of tetrahydrocurcumin. *Biochem Pharmacol*, 1996; 52: 519-25.
12. Mukhopadhyay A, Basu N, Ghatak N, Gujral PK. Antiinflammatory and irritant activities of curcumin analogues in rats. *Agents Actions*, 1982; 12: 508-15.
13. Wenzel E, Somoza V. Metabolism and bioavailability of trans-resveratrol. *Mol Nutr Food Res*, 2005; 49, 472-81.
14. Adlercreutz H, Markkanen H, Watanabe S. Plasma concentration of phyto-oestrogen in Japanese men. *Lancet*, 1993; 342: 1209-10.
15. Mills PK, Beeson WL, Philips RL, Fraser GE. Cohort study of diet, lifestyle and prostate cancer in Adventist men. *Cancer*, 1989; 64: 598-604.
16. Middleton, E. J. R.; Kandaswami, C. Effect of flavonoids on immune and inflammatory cell function. *Biochem. Pharmacol.* 1992; 43(6): 1167-1179.
17. Cooks, N. C.; Samman, S. Flavonoids - Chemistry, metabolism, cardioprotective effects and dietary sources. *J. Nutr. Biochem.*, 1996; 7: 66.
18. Laughton, J. M.; Evans, P. J.; Moroney, M. A.; Hoult, J. R. S. Inhibition of mammalian lipoxygenase and cyclooxygenase by flavonoid and phenolic dietary additives. *Biochem. Pharmacol.* 1991; 18: 1673-1681.
19. Smith T J, Yang CS. Effect of food phytochemical on metabolism and tumorigenesis. *Food Phytochemicals For Cancer Prevention I*. Huang, M. T., Ed.; Am Chem Soc: Washington, DC, 1994; p 48.
20. De Whaley, C. V.; Rankin, S. M.; Hoult, J. R. S.; Jessup, W.; Leake, D. S. Flavonoid inhibit the oxidative modification of low-density lipoprotein by macrophages. *Biochem. Pharmacol.* 1990, 39, 1743-1750.
21. Yoshida, M.; Sakai, T.; Hosokawa, N.; Marui, N.; Matsumoto, K.; Akihiro, F.; Nishino H.; Aoike, A. The effect of quercetin on cell cycle progression and growth of human gastric cancer cells. *FEBS Lett.* 1990; 10-13.
22. Vinson JA, Dabbagh YA, Serry MM, Jang J. Plant flavonoids, especially tea flavonols, are powerful antioxidants using an in vitro oxidation model for heart disease. *J. Agric. Food Chem.* 1995; 43: 2800-2804.

23. Hollman PCH, Van Trijp J MP, Buysman MNCP, Gaag M SV d, Mengelers MJB, De Vries JHM, Katan MB. Relative bioavailability of the antioxidant flavonoid quercetin from various foods in man. *FEBS Lett.* 1997; 418, 152-156.
24. Rhodes MJC, Price KR. Analytical problems in the study of flavonoid compounds in onions. *Food Chem.* 1996; 57(1): 113-117.
25. Hertog MGL. (1995) Flavonoid intake and long-term risk of coronary heart disease and cancer in the 7 countries study. *Arch. Int. Med.* 1995; 155(11); 1184-1195.
26. Rodriguez-Saona LE, Giusti MM, Wrolstad RE. Anthocyanin pigment composition of red-fleshed potatoes. *J. Food Sci.* 1998; 63: 458-465.
27. Satue-Gracia MT, Heinonen M, Frankel EN. Antioxidant activity of anthocyanins in LDL and lecithin liposome systems. *J. Agric. Food Chem.* 1997; 45: 3362-3367.
28. Velioglu YS, Mazza G, Gao L, Oomah B D. Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *J. Agric. Food Chem.* 1998; 46: 4113-4117.
29. White PJ, Xing, Y. Antioxidants from Cereals and Legumes. In *Natural Antioxidants: Chemistry, Health Effects, and Applications*; Shahidi, F., Ed.; AOCS Press: Champaign, IL, 1997.
30. Terao J, Karasawa H, Arai H, Nagao A, Suzuki T, Takama K. Peroxyl radical scavenging activity of caffeic acid and its related phenolic compounds in solution. *Biosci. Biotechnol. Biochem.* 1993; 57: 1204-1205.
31. Dimberg LH, Theander O, Lignert H. Avenanthramides—a group of phenolic antioxidants in oats. *Cereal Chem.* 1993; 70: 637-641.
32. Pereira DM, Patrícia Valentao P, Pereira JA, Andrade PB. Phenolics: From Chemistry to Biology. *Molecules*, 2009; 14: 2202-2211.
33. Hakkinen SH, Mykkanen H M, Karenlampi SO, Torronen AR. Phenolic profiles in Finnish berries. *Cost 916 1st Workshop: Polyphenols in Food, April*; 1997.
34. Chen JH, Ho CT. Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds. *J. Agric. Food Chem.* 1997; 45: 2374-2378.
35. Meyer AS, Donovan JL, Pearson DA, Waterhouse AI, Frankel EN. Fruit hydroxycinnamic acids inhibit human low-density lipoprotein oxidation *in vitro*. *J. Agric. Food Chem.* 1998; 46: 1783-1787.
36. Constantino L, Albasino A, Rastelli G, Benvenuti S. Activity of polyphenolic crude extracts as scavengers of superoxide radicals and inhibitors of xanthine oxidase. *Planta Med.* 1992; 58: 342-344.
37. Chin KW, Sun H L. Analysis of the phenolic compound in betel quid. *J. Chin. Agric. Chem. Soc.* 1990; 31: 623-632.
38. Jong CP, Jung O H, Kun Y P. Antimutagenic effect of flavonoid isolated from *O. javanica*. *J. Korean Soc. Food Sci. Nutr*, 1990; 25(4): 588-592.
39. Kim SY, Lee KH, Chang KS, Bock JY, Jung M. Y. Taste and flavour compound in box thorn (*Lycium chinense miller*) leaves. *Food Chem.*, 1997; 58(4): 297-303.
40. Park WJ. Studies on chemical composition and biological activity of *Lycium chinense miller*. Ph.D. Thesis, 1995; Gunkuk University, Seoul, Korea.
41. Ohshiro M, Kuroyanagi M, Ueno A. Structure of sesquiterpenes from *Curcuma longa*. *Phytochemistry*, 1990; 29(7): 2201-2205.
42. Ong KC, Khoo HE. Biological effects of myricetin. *Gen. Pharmacol.*, 1997; 29: 121-126.
43. Leighton T, Ginther C, Fluss L, Harter WK, Cansado J, Notario V. Quercetin and its glycosides in *Allium* vegetables. In *Phenolic Compounds In Food and Their Effects On Health II*; Ho, C.-T., Lee, C. Y., Huang, M.-T., Eds.; American Chemical Society: Washington, DC, 1992; pp 220-238.
44. Goet RK, Pandey VB, Dwivedi SPD, Rao YV. Antiinflammatory and antiulcer effects of kaempferol, a flavone, isolated from *Rhamnus procumbens*. *Indian J. Exp. Biol.* 1988; 26: 121-124.
45. Brinkworth RI, Chen J, Leuenberger PM, Freiburghaus AU, Follath F. Flavones are inhibitors of HIV proteinase. *Biochem. Biophys. Res. Commun.* 1992; 188: 631-637.
46. Hempel J, Bohm H. Quality and quantity of prevailing flavonoid glycosides of yellow and green French beans. *J. Agric. Food Chem.* 1996; 44: 2114-2116.

47. Park YK, Lee CY. Identification of isorhamnetin 4- β -glucoside in onions. *J. Agric. Food Chem.* 1996; 44: 34-36.
48. Hertog MGL, Hollman PCH, Katan MB, Kromhout D. Intake of potentially anticarcinogenic flavonoid and their determinants in adults in The Netherlands. *Nutr. Cancer*, 1993; 20(1): 21-29.
49. Ferreres F, Tomas-Barberan FA, Gil MI, Tomas-Lorente, F. An HPLC technique for flavonoid analysis in honey. *J. Sci. Food Agric.* 1991; 56: 49-56.
50. Herrmann, K. Flavonols and flavones in food plants: A review. *J. Food Technol.* 1996; 11: 433-448.
51. Mizuno M, Tsuchida H, Kozukue N, Mizuno S. Rapid quantitative analysis and distribution of free quercetin in vegetables and fruits. *J. Japan. Soc. Food Sci. Technol. (Nippon Shokuhin Kogyo Gakkaishi)*, 1992; 39(1), 88-92.
52. Crozier A, Michael EJL, McDonald MS, Black C. Quantitative analysis of the flavonoid content of commercial tomatoes, onions, lettuce, and celery. *J. Agric. Food Chem.* 1997; 45: 590-595.
53. Cao G, Booth SL, Sadowski JA, Prior RL. Increases in human plasma antioxidant capacity after consumption of controlled diets high in fruit and vegetables. *Am. J. Clin. Nutr.* 1998; 68, 1081-1087.
54. Hertog MGL, Hollman PCH, Van de Putte B. Content of potentially anticarcinogenic flavonoids of tea infusions, wines, and fruit juices. *J. Agric. Food Chem.* 1993; 41: 1242-1246.
55. Rice-Evans CA, Miller NJ, Bolwell PG, Bramley PM, Pridham J B. The relative antioxidant activity of plant-derived polyphenolic flavonoid. *Free Radical Res.* 1995; 22: 375-383.
56. Pratt D E. Natural antioxidant from plant material. In *Phenolic Compounds In Food and Their Effects On Health II*. Ho, C.-T., Lee, C. Y., Huang, M.-T., Eds.; American Chemical Society: Washington, DC, 1992; pp 55-71.
57. Gronbaek M, Deis A, Sorensen TIA, Becker U, Schnohr P, Jensen G. Mortality associated with moderate intakes of wine, beer/spirits. *Br. Med. J.*, 1995; 310: 1165-1169.
58. Simonetti P, Pietta P, Testolin G. Polyphenol content and total antioxidant potential of selected Italian wines. *J. Agric. Food Chem.* 1997; 45: 1152-1155.
59. Balestieri F, Flori A, Marini D. Strategies for food quality control and analytical methods in Europe. Proceedings of the 6th European Conference on Food Chemistry. *Lebensmittelchem. Gesellschaft* 1991; 23: 369-378.
60. Fernandez de Simon B, Perez-Illarbe J, Hernandez T, Gomez-Cordoves C, Estrella I. Importance of phenolic compounds for the characterization of fruit juices. *J. Agric. Food Chem.* 1992; 40: 1531-1535.
61. Benavente-Garcia O, Castillo J, Marin FR, Ortuno A, del Rio JA. Uses and properties of citrus flavonoid. *J. Agric. Food Chem.* 1997; 45(12): 4505-4515.
62. Price KR, Bacon JR, Rhodes M JC. Effect of storage and domestic processing on the content and composition of flavonol glucosides in onion (*Allium cepa*) *J. Agric. Food Chem.* 1997; 45: 938-942.
63. Desrosiers RC. Prospects for an AIDS vaccine. *Nature Med*, 2004; 10, 221-3.
64. Joyce CL, Mack SR, Anderson RA, Zaneveld LJD. Effect of hyaluronidase, beta-glucuronidase and beta-N-acetyl glycosaminidase inhibitors on sperm penetration of the mouse oocytes. *Biol Reprod*, 1986; 35: 336-46.
65. Critchfield JW, Butsera ST, Folks TM. Inhibition of HIV activation in latently infected cells by flavonoids compounds. *AIDS Res Hum Retroviruses*, 1996; 12: 39-46.
66. Zhang GH, Wang Q, Chen JJ, et al. The anti-HIV I effect of scutellarin. *Biochem Biophys Res Commun*, 2005; 334: 812-816.
67. Li CJ, Dezube BJ, Biswas DK, Ahlers CM Pardee AB. Inhibition of HIV-1 Transcription. *Trends Microbiol*, 1994; 5: 164-9.
68. Critchfield JW, Coligan JE, Folks TM, Butera ST. Casein kinase II is a selective target of HIV-1 transcriptional inhibitors. *Proc Natl Acad Sci USA*, 1997; 94: 6110-5.
69. Wu Baer F, Lane WS, Gaynor RB. The cellular factor TRP-185 regulates RNA polymerase binding to HIV-1 TAR RNA. *EMBO J*, 1995; 14; 5995-6009.
70. Fauci AS. Host factors and the pathogenesis of HIV induced disease. *Nature*, 1996; 384: 529-34.
71. Kawai K, Tsuno NH, Kitayama J, et al. Epigallocatechin gallate the main component of tea polyphenol binds to CD4 and interferes with

- gp120 binding. *J Allergy Clin Immunol*, 2003; 112: 851-3.
- 72.de Clereq. Natural products for chemotherapy of human immunodeficiency virus (HIV) infection. *Med Res Rep*, 2000; 20: 323-49.
- 73.Verma R, Gupta RB, Singh K, et al. Distribution of CCR5 Delta 32, CCR2-641 and SDF1-3A and plasma levels of SDF-1 in HIV-1 seronegative North Indians. *J Clin Virol*, 2007; 38: 198-203.
- 74.Nair MP, Kandaswami C, Mahajan S, et al. Grape seed extract proanthocyanidins down regulate HIV-1 entry coreceptors, CCR2b, CCR3 and CCR5 gene expression by normal peripheral blood mononuclear cells. *Biol Res*, 2002; 35: 421-1.
- 75.Cowan MM. Plant products as antimicrobial agents. *Clin Microbiol Rev*, 1999; 12: 564-82.
- 76.Tsuchiya H, Sato M, Miyazaki T, et al. Comparative study on the antibacterial activity of phytochemical flavanones against methicillin resistant *Staphylococcus aureus*. *J Ethnopharmacol*, 1996; 50: 27-34.
- 77.Mason TL, Wasserman BP. Inactivation of red beet β glucan synthetase by native and oxidized phenolic compounds. *Phytochem*, 1987; 26: 2197.
- 78.Manjunatha BK. Antibacterial activity of *Pterocarpus santalinus*. *Ind J Pharm Sci*, 200; 68; 115-6.
- 79.Krishnaveni KS, Rao JVS. A new isoflavone glucoside from *Pterocarpus santalinus*. *Asian Nat Prod Res*, 2000; 2: 219.
- 80.Dixon RA, Dey PM, Lamb CJ. Phytoalexins: enzymology and molecular biology. *Adv Enzymol*, 1983; 55: 1-69.
- 81.Toda MS, Okubo R, Ohnishi OR, Shimamura T. Antibacterial and bactericidal activities of Japanese green tea. *Jpn J Bacteriol*, 1989; 45, 561-6.
- 82.Borris RP. Natural products research: perspectives from a major pharmaceutical company. *J Ethnopharmacol*, 1996; 51: 29-38.
- 83.Batista O, Doarte A, Nascimento J, Simoes MF. Structure and antimicrobial activity of diterpenes from the roots of *Plectranthus hereroensis*. *J Nat Prod*, 1994; 57: 858-86.
- 84.Vijaya K, Ananthan S, Nalini R. Antibacterial effect of theaflavin, polyphenol (*Camellia sinensis*) and *Euphorbia hirta* on *Shigella* spp.- a cell culture study. *J Ethnopharmacol*, 1995; 49: 115-8.
- 85.Nakahara K, Kawabata S, Ono H, et al. Inhibitory effects of oolong tea polyphenols on glucosyltransferases of mutans streptococci. *Appl Environ Microbiol*, 1993; 59: 968-73.
- 86.Stern SL, Hagerman AE, Steinberg PD, Mason PK. Phlorotannin-protein interactions. *J Chem Ecol*, 1996; 22: 1887-90.
- 87.Kazmi MH, Malik A, Hameed S, Akhtar N, Ali SN. An anthraquinone derivative from *Cassia italica*. *Phytochem*, 1994; 36: 761-3.
- 88.Criqui MH, Ringel BL. Does diet or alcohol explain the French paradox? *Lancet*, 1994; 344: 1719-23.
- 89.German JB, Walzem R. The health benefits of wine. *Annu Rev Nutr*, 2000; 20: 561-93.
- 90.Li HL, Huang Y, Zhang CN, et al. Epigallocatechin-3 gallate inhibits cardiac hypertrophy through blocking reactive oxidative species-dependent and -independent signal pathways. *Free Radic Biol Med*, 2006; 40: 1756-75.
- 91.Szewczuk LM, Forti L, Stivala LA, Penning TM. Resveratrol is a peroxidase-mediated inactivator of COX-1 but not COX-2: a mechanistic approach to the design of COX-1 selective agents. *J Biol Chem*, 2004; 279: 22727-37.
- 92.Falchi M, Bertelli A, Le Scalzo R, et al. Comparison of cardioprotective abilities between the flesh and skin of grapes. *J Agric Food Chem*, 2006; 54: 6613-22.
- 93.Belaia OL, Fomina IG, Baider LM, Kuropteva ZV, Tiukavkina NA. The influence of the bioflavonoid dicvertin on the antioxidative system ceruloplasmin-transferrin and lipid peroxidation in patients suffering from stable coronary heart disease with dyslipidemia. *Clin Med*, 2006; 84: 46-50.
- 94.Andriambeloson E, Kleschyov AL, Muller B. Nitric oxide production and endothelium-dependent vasorelaxation induced by wine polyphenols in rat aorta. *Br J Pharmacol*, 1997; 20: 1053-8.
- 95.Wallerath T, Deckert G, Ternes T. Resveratrol, a polyphenolic phytoalexin present in red wine, enhances expression and activity of endothelial nitric oxide synthase. *Circulation*, 2002; 106: 1652-8.

96. Kuiper GGM, Lemmen JG, Carlsson B, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor. *Endocrinology*, 1998; 139: 4252-63.
97. Figtree GA, Griffiths H, Lu YQ, et al. Plant-derived estrogens relax coronary arteries in vitro by a calcium antagonistic mechanism. *J Am Coll Cardiol*, 2000; 35: 1977-85.
98. Walker HA, Dean TS, Sanders TA, et al. The phytoestrogen genistein produces acute nitric oxide dependent dilation of human forearm vasculature with similar potency to 17 β -estradiol. *Circulation*, 2001; 103: 258-62.
99. Shen J, White M, Husband AJ, Hambly BD, Bao S. Phytoestrogen derivatives differentially inhibit arterial neointimal proliferation in a mouse model. *Eur J Pharmacol*, 2006; 548: 123-8.
100. Firuzi. Hypochlorite scavenging activity of flavonoids. *J Pharma Pharmacol*, 2004; 56: 801-7.
101. Morris MC, Evans DA, Bienias JL, et al. Dietary intake of antioxidant nutrients and the risk of incident Alzheimer disease in a biracial community study. *JAMA*, 2002; 287: 3230-7.
102. Bauer V, Bauer F. Reactive oxygen species as mediators of tissue protection and injury. *Gen Physiol Biophys*, 1999; 18: 7-14.
103. Perry G, Sayre LM, Atwood CS, et al. The role of iron and copper in the aetiology of neurodegenerative disorders: therapeutic implications. *CNS Drugs*, 2002; 16: 339-52.
104. Ghadge GD, Lee JP, Bindokas VP, et al. Mutant superoxide dismutase-1-linked familial amyotrophic lateral sclerosis, molecular mechanisms of neuronal death and protection. *J Neuro Sci*, 1997; 17: 8756-66.
105. Philips M, Sabas M, Greenberg J. Increased pentane and carbon disulfide in the breath of patients with schizophrenia. *J Clin Pathol*, 1993; 46: 861-864.
106. Wang J, Ho L, Zhao Z, et al. Moderate consumption of Cabernet Sauvignon attenuates Abeta neuropathology in a mouse model of Alzheimer's disease. *FASEB J*, 2006; 20: 2313-20.
107. Dai Q, Borenstein AR, Wu Y, Jackson JC, Larson EB. Fruit and vegetable juices and Alzheimer's disease: the Kame project. *Am J Med*, 2006; 119: 751-9.
108. Bastianetto S, Yao ZX, Papadopoulos V, Quirion R. Neuroprotective effects of green and black teas and their catechin gallate esters against betaamyloid- induced toxicity. *Eur J Neurosci*, 2006; 23: 55-64.
109. Koh SH, Lee SM, Kim HY, et al. The effect of epigallocatechin gallate on expressing diseases progression of ALS model mice. *Neuro Sci Lett*, 2006; 375: 103-7.
110. Xu Z, Chen S, Li X, Luo G, Li L, Le W. Neuroprotective effects of (-)-epigallocatechin-3-gallate in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurochem Res*, 2006; 31: 1263-9.
111. Ramassamy C. Emerging role of polyphenolic compounds in the treatment of neurodegenerative diseases: a review of their intracellular targets. *Eur J Pharmacol*, 2006; 545: 51-64.
112. Shang YZ, Qin BW, Cheng JJ, Miao H. Prevention of oxidative injury by flavonoids from stems and leaves of *Scutellaria baicalensis* georgi in PC12 cells. *Phytother Res*, 2006; 20: 53-7.
113. Kim H, Park BS, Lee KG, et al. Effects of naturally occurring compounds on fibril formation and oxidative stress of beta-amyloid. *J Agric Food Chem*, 2005; 53: 8537-41.
114. Schroeter H, Spencer JP, Evans CR, William RT. Flavonoids protect neuron from toxicologic low-density lipoprotein induced apoptosis. *Biochem J*, 2006; 388: 547-57.
115. Joseph JA, Denisova NA, Arendash G, et al. Blueberry supplementation enhances signaling and prevents behavioral deficits in an Alzheimer disease model. *Nutr Neurosci*, 2003; 6: 153-62.
116. Joseph JA, Shukitt-Hale B, Denisova N, et al. Reversals of age-related declines in neuronal signal transduction, cognitive, and motor behavioral deficits with blueberry, spinach, or strawberry dietary supplementation. *J Neurosci*, 1999; 19: 8114-21.
117. Adlercreutz CH, Goldin BR, Gorbach SL, et al. Soybean phytoestrogen intake and cancer risk. *J Nutr*, 2003; 125: 757-70.
118. Ullah MF. Cancer multidrug resistance (MDR): A major impediment to effective chemotherapy. *Asian Pac J Chem Prev*, 2008; 9: 1-6.
119. Alhasan S, Pietraszkiewicz-Alonso MD, Ensley J, Sarkar FH. Genistein induced cell cycle

- arrest and apoptosis in a head and neck squamous cell carcinoma cell line. *Nutr Cancer*, 1999; 34: 12-9.
- 120.Hillman GG, Forman JD, Kucuk O, et al. Genistein potentiates the radiation effect on prostate carcinoma cells. *Clin Cancer Res*, 2001; 7: 382-90.
- 121.Hillman GG, Wang Y, Kucuk O, et al. Genistein potentiates inhibition of tumor growth by radiation in a prostate cancer orthotopic model. *Mol Cancer Ther*, 2004; 3: 1271-9.
- 122.Hsu JT, Hung HC, Chen CJ, Ying C. Effect of dietary phytoestrogen biochanin A on cell growth in the mammary carcinoma cell line MCF-7. *J Nut BioChem*, 1999; 10, 510-7.
- 123.Fung MC. Effect of biochanin A on the growth and differentiation of myeloid leukemia. *Life Sci*, 1997; 61: 105-15.
- 124.Lyn Cook BP, Stottman HL, Yan Y, Hamous GJ. The effects of phytoestrogen on human pancreatic tumor cells in vitro. *Cancer Lett*, 1999; 142: 111-119.
- 125.Rike L, Samedì VG, Medrans TA, Baker HV. Mechanism of the growth inhibitory effect of the isoflavonoid biochanin A on LNCaP cells and xenograft. *The Prostate*, 2002; 52: 201-12.
- 126.Ulrich A, Schlessinger J. Signal transduction by receptors with tyrosine kinase activity. *Cell*, 1990; 61: 203-212.
- 127.Kyle E, Neckers L, Takimoto C, Curt G, Bergan R. Genistein-induced apoptosis of prostate cancer cells is preceded by a specific decrease in focal adhesion kinase activity. *Mol Pharmacol*, 1997; 5; 193-200.
- 128.Papavassiliou AG. Transcription factors: Structure, function and implication in malignant growth. *Anticancer Res*, 1995; 15: 891-4.
- 129.Davis JN, Kucuk O, Sarkar FH. Genistein inhibits NFkappa B activation in prostate cancer cells. *Nutr Cancer*, 1999; 35: 167-74.
- 130.Tsvetkov P, Asher G, Reiss V, et al. Inhibition of NAD[P]H: Quinone oxido reductase 1 activity and induction of P53 degradation by natural phenolic compound curcumin. *Proc Natl Acad Sci USA*, 2005; 102: 5535-40.
- 131.Maheshwari RK, Singh AK, Gaddipati J, Srimal RC. Multiple biological activities of curcumin: A short review. *Life Sci*, 2006; 78: 2081-87.
- 132.Singh S, Aggarwal BB. Activation of transcription factor NF-kB is suppressed by curcumin (diferulolylmethane). *J Biol Chem*, 1995; 270: 24995-5000.
- 133.Xia Y, Jin L, Zhang B, et al. The potentiation of curcumin on insulin-like growth factor-1 action in MCF-7 human breast carcinoma cells. *Life Sci*, 2007; 80: 2161-9.
- 134.Narayanan BA. Chemopreventive agents alters global gene expression pattern: Predicting their mode of action and targets. *Curr Cancer Drug Targets*, 2006; 6: 711-27.
- 135.Yacoub A, Park JS, Qiao L, Dent P, Hegan MP. MAPK dependence of DNA damage repair: ionizing radiation and the induction of expression of the DNA repair genes XRCC1 and ERCC1 in DU145 human prostate carcinoma cells in MEK K dependent fashion. *Int J Radiat Biol*, 2001; 77: 1067-78.
- 136.Scarlatti F, Sala G, Rici C et al. Resveratrol sensitization of DU145 prostate cancer cells to ionizing radiation is associated to ceramide increase. *Cancer Lett*, 2007; 253: 124-30.
- 137.Casagrande F, Darbon JM. p21CIP1 is dispensable for the G2 arrest caused by genistein in human melanoma cells. *Exp Cell Res*, 2000; 258: 101-8.
- 138.Morley KL, Ferguson PJ, Koropatrik J. Tanageretin and nobiletin induce G1 cell arrest but not apoptosis in human breast and colon cancer cells. *Cancer Lett*, 2007; 251: 168-78.
- 139.Bagli E, Stefaniotou M, Morbidelli L. Luteolin inhibits vascular endothelial growth factor induced angiogenesis; inhibition of endothelial cell survival and proliferation by targeting phosphatidylinositol 3'-kinase activity. *Cancer Res*, 2004; 64: 7936-46.
- 140.Azmi AS, Bhat SH, Hanif S, Hadi SM. Compounds lymphocytes leading to oxidative DNA breakage: Implication for a putative mechanism for anticancer properties. *FEBS Lett*, 2006; 580: 533-8.
- 141.Hadi SM, Asad SF, Singh S, Ahmad A. Putative mechanism for anticancer and apoptosis-inducing properties of plant-derived polyphenolic compounds. *IUBMB Life*, 2000; 50: 1-5.
- 142.Hadi SM, Asfar AS, Bhat SH, et al. Oxidative breakage of cellular DNA by plant polyphenols: A putative mechanism for anticancer properties. *Semin Cancer Biol*, 2007; 17: 370-7.

143. Bors W, Heller W, Michael M. Flavonoids as antioxidants: determination of radical scavenging efficiencies. Edited by CA Rice, E Vans and L Packer, Marcel Dekker, 1998; New York.
144. Gali HV, Perchellet EM, Klish DS, Johnson JM, Perchellet JP. Hydrolyzable tannins: potent inhibitors of hydroperoxide production and tumor promotion in mouse skin treated with 12-O-tetradecanoyl phorbol-13-acetate *in vivo*. *Int J Cancer*, 1992; 51: 425-32.
145. Yoshida D, Ikada Y, Nakayama S. Quantitative analysis of copper, zinc and copper/zinc ratio in selective human brain tumors. *J Neuro Oncol*, 1993; 16: 109-15.
146. Ebadi M, Swanson S. The status of zinc, copper and metallothionein in cancer patients. *Prog Clin Biol Res*, 1988; 259: 161-75.
147. Ullah MF. Cancer therapeutics: Emerging targets and trends. *Curr Cancer Ther Rev*, 2008; 4: 50-6.
148. Shuzhong Z, Xinning Y, Robert AC, Marilyn EM. Structure activity relationships and quantitative structure activity relationships for the flavonoid-mediated inhibition of breast cancer resistance protein. *Biochem Pharmacol*, 2005; 70: 627-39.
149. Chung SY, Sung MK, Kim NH, et al. Inhibition of P-glycoprotein by natural products in human breast cancer cells. *Arch Pharm Res*, 2005; 28: 823-8.
150. Feng Q, Dongzhi W, Qiang Z, Shengli Y. Modulation of P-glycoprotein function and reversal of multidrug resistance by (-)-epigallocatechin gallate in human cancer cells. *Biomed Pharmacother*, 2005; 59: 64-9.
151. Wahl H, Tan L, Griffith K, Choi M, Liu RJ. Curcumin enhances Apo2L/TRAIL-induced apoptosis in chemoresistant ovarian cancer cells. *Gynecol Oncol*, 2007; 105: 104-12.
152. Aggarwal BB, Bhardwaj A, Aggarwal RS, et al. Role of resveratrol in prevention and therapy of cancer: Preclinical and clinical studies. *Anticancer Res*, 2004; 24: 2783-840.
153. Boivin D, Lamy S, Lord-Dufour S, Jackson J, Beaulieu E, Côté M, Moghrabi A, Barrette S, Gingras D, Béliveau R. Antiproliferative and antioxidant activities of common vegetables: A comparative study. *Food Chem*, 2009; 112: 374-380.
154. Brand-Williams W, Cuvelier M, Bersert EC. Use of a free radical method to evaluate antioxidant activity. *LWT-Food Sci. Technol.* 1995; 28: 25-30.
155. Bui ML, Grayer RJ, Veitch NC, Kite GC, Tran H, Nguyen QK. Uncommon 8-oxygenated flavonoids from *Limnophila aromatic* (Scrophulariaceae). *Biochem. Sys. Ecol.* 2004; 32: 943-947.
156. Cao G H, Sofic E, Prior RL. Antioxidant capacity of tea and common vegetables. *J. Agric. Food Chem.* 1996; 44, 3426-3431.
157. Chanwitheesuk A, Teerawutgulrag A, Rakariyatham N. Screening of antioxidant activity and antioxidant compounds of some edible plants of Thailand. *Food Chem.* 2005; 92: 491-497.
158. Eldahshan OA, Ayoub NA, Singab AB and Al-Azizi MM. Potential Superoxide Anion Radical Scavenging Activity of Doum Palm (*Hyphaene thebaica* L.) Leaves Extract. *Rec. Nat. Prod.* 2008; 2(3) 83-93.
159. Finger, A.; Engelhardt, U. H.; Wray, V. Flavonol glycosides in tea-kaempferol and quercetin rhamnoglucosides. *J. Sci. Food Agric.* 1991; 55: 313-321.
160. Hafidh RR, Abdulmir AS, Abu Bakar F, Abas F, Jahanshahi F and Sekawi Z. Antioxidant Research in Asia in the Period from 2000-2008. *Am J Pharmacol & Toxicol*, 2009; 4 (3): 48-66.
161. Huda-Faujan N, Norham A, Norrakiah AS, Babji AS. Antioxidative Activities of Water Extracts of Some Malaysian Herbs. *ASEAN Food Journal* 2007; 14(1): 61-68.
162. Kampa M, Nifli, AP, Notas G, Castanas E. Polyphenols and cancer cell growth. *Rev. Physiol. Biochem. Exp. Pharmacol.* 2007; 159: 79- 113.
163. Lee, Y.; Howard, L. R.; Villalon, B. Flavonoid and antioxidant activity of fresh pepper (*Capsicum annum*) cultivars. *J. Food Sci.*, 1995; 60 (3), 473-476.
164. Marja P. Kahkonen, Anu I. Hopia, Heikki J. Vuorela, Jussi-Pekka Rauha, Kalevi Pihlaja, Tytti S. Kujala, and Marina Heinonen. Antioxidant Activity of Plant Extracts Containing Phenolic Compounds. *J. Agric. Food Chem.*, 1999; 47: 3954-3962.
165. Ullah MF, Khan MW. Food as Medicine: Potential Therapeutic Tendencies of Plant Derived Polyphenolic Compounds. *Asian Pacific J Cancer Prev*, 2009; 9: 187-196.

166.Nanasombat S, Teckchuen N. Antimicrobial, antioxidant and anticancer activities of Thai local vegetables. *J Med Plants Res*, 2009; 3(5): 443-449.

167.Nielson JK, Olsen CE, Petersen MK. Acylated flavonol glycosides from cabbage leaves. *Phytochemistry*, 1993; 34(2): 539-544.

168.Sukrasno N, Yeoman MM. Phenylpropanoid metabolism during growth and development of *Capsicum frutescens* fruits. *Phytochem*, 1993, 32: 839-844.

169.Tsushida T, Suzuki M. Isolation of flavonoid-glycosides in onion and identification by chemical synthesis of the glycosides. *Nipp. Shok. Kaga Koga Kaishi* 1995; 42(2): 100-108.

170.Wang F, Yu F, Fang Y, Xin Z, Yang F, Xu J, Zhao L, Hu Q Li. *In vitro* antioxidant and anticancer activities of ethanolic extract of selenium-enriched green tea. *Food Chem*, 2008; 111: 165-170.