

Biological activity, molecular docking, and ADME predictions of amphibine analogues of *Ziziphus spina-christi* towards SARS-CoV-2 M^{pro}

Taufik Muhammad Fakhil^{1*}, Dwi Syah Fitra Ramadhan², Fitrianti Darusman¹

¹Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Universitas Islam Bandung

Jl. Ranga Gading No. 8 Bandung, West Java, Indonesia. 40116

*Email: taufikmuhammadf@gmail.com

²Department of Pharmacy, Universitas Mandala Waluya

Jl. Jend. AH. Nasution Blok G-37 Kendari, Southeast Sulawesi, Indonesia. 93561

ABSTRACT. The main protease of the SARS-CoV-2 virus, SARS-CoV-2 M^{pro}, can be discovered as a promising target to treat the COVID-19 pandemic. The peptide-based inhibitors may present better options than small molecules to inhibit SARS-CoV-2 M^{pro}. *Ziziphus spina-christi* species reported have a peptide-based of alkaloids group, i.e., amphibine whose analogues can be identified the potential as an inhibitor of SARS-CoV-2 M^{pro}. The compound structure was drawn and optimized using semi-empirical AM-1 method using Quantum ESPRESSO v.6.6, while the biological activity using PASS. Prediction server and molecular docking simulation using MGLTools 1.5.6 with AutoDock 4.2 were performed. Afterward, the ADME profiles were predicted using the SWISS-ADME server. PASS server was predicting amphibine B-F and H showed potency both as antiviral and as a protease inhibitor. The molecular docking simulation of amphibine analogues showed lower binding energy than the native ligand. The binding energy of the native ligand was -7.69 Kcal/mol compared to the lowest binding energy of amphibine analogues was -10.10 Kcal/mol (amphibine-F). The ADME prediction showed that amphibine-F has the best bioavailability as an oral drug, amphibine-B, C, and D have good bioavailability, and amphibian-E and H have poor bioavailability. Concluded, amphibine B-F and H of amphibine analogues showed potency as COVID-19 treatment targeting SARS-CoV-2 M^{pro}.

Keywords: Christ's thorn jujube; COVID-19 therapy; *in silico* study; SARS-CoV-2 M^{pro}; semi-empirical AM-1

Article History: Received 25 April 2021; Received in revised form 6 May 2021; Accepted 30 May 2021; Available online 30 June 2021

How to Cite This Article: Fakhil TM, Dwi Syah Fitra Ramadhan DSF, Darusman F. 2021. Biological activity, molecular docking, and ADME predictions of amphibine analogues of *Ziziphus spina-christi* towards SARS-CoV-2 M^{pro}. *Biogenesis: Jurnal Ilmiah Biologi*. vol 9(1): 109-117. doi: <https://doi.org/10.24252/bio.v9i1.21335>.

INTRODUCTION

The Coronavirus disease 2019 has spread worldwide and still become a health problem that needs attention (Thompson, 2020; Zhu *et al.*, 2020). Released on March 2021 in the present situational report from WHO, 3.8 million COVID-19 new cases, and 64000 recent deaths were reported globally. (WHO, 2021). Clinically, COVID-19 can lead to severe respiratory complications and death with fever and respiratory symptoms (Calcagno *et al.*, 2020).

SARS-CoV-2 main protease (M^{pro}), which can be crystallized along with its inhibitors, was one of the most promising targets for COVID-19 drug discovery (Jin *et al.*, 2020; Khaerunnisa *et al.*, 2020; Mirza & Froeyen, 2020; Reiner *et al.*, 2020), and become the key enzyme of viral polyprotein maturation, replication and transcription cycle (Dai *et al.*, 2020; Fu *et al.*, 2020; Goyal & Goyal, 2020). SARS-CoV M^{pro}

is resistant to peptide-like anti-HIV-1 drugs, hence SARS-CoV and SARS-CoV-2 SARS-CoV-2 is closely related to, but distinct from the SARS-CoV branch on phylogenetic relationship (Choudhury & Mukherjee, 2020; Dong *et al.*, 2020). Both virus rely on main protease associated with N3 inhibition (Griffin, 2020). Small compounds may not be as effective as peptide-based inhibitors in the treatment of COVID-19 (Gentile *et al.*, 2020; Han & Král, 2020; Maiti, 2020; Murdocca *et al.*, 2021).

The natural product compounds based on peptide-like from medicinal plants become our orientation research. Hence, they have not been explored intensively in drug discovery, especially those that can inhibit COVID-19 (Dias *et al.*, 2012; Harvey *et al.*, 2015; Benarba & Pandiella, 2020; Lakshmi *et al.*, 2020). *Z. spina-christi* as important medicinal plant (El Maaiden *et al.*, 2019) is a deciduous tree that

generally comes from warm and subtropical climates, such as North Africa, South Europe, Mediterranean, tropical America, South and East of Asia, and others, including Indonesia (Kwape *et al.*, 2013; Moossavi *et al.*, 2017). There are many names for *Z. spina-christi*, known as Christ's thorn jujube, belongs to the Rhamnaceae family with large shade tree (Baghazadeh-Daryaii *et al.*, 2017; Gorai *et al.*, 2019).

The previous studies have reported that *Z. spina-christi* provided a variety of pharmacological activities, including antibacterial, antifungal, antioxidant, antihyperglycemic, and anti-diabetic (Kalayou *et al.*, 2012; Ads *et al.*, 2017; Al-Ghamdi *et al.*, 2017; Alotibi *et al.*, 2020). According to our previous studies, the main phytochemicals were discovered in this plant include alkaloids, flavonoids, and saponins (Darusman & Fakih, 2021). Tuentner *et al.* (2017) and Sakna *et al.* (2019) stated cyclopeptide alkaloids can be found in their stem-bark.

To date, the need for a vaccine and antiviral development is increasing, especially those targeting SARS-CoV-2 M^{pro}. Computational approaches were demonstrated to predict the affinity and molecular behavior of amphibine analogues from the plant compound. We are interested in investigating the potency of the amphibine analogues (cyclopeptide alkaloids) from *Z. spina-christi*, as a promising future treatment for COVID-19 targeting SARS-CoV-2 M^{pro}.

MATERIALS AND METHODS

Ligand preparation. The ligands chosen for this research were peptide alkaloids, amphibine analogues in *Ziziphus spina-christi*, i.e., amphibine A-H compounds. The 3D ligand structures were drawn and optimized based semi-empirical AM-1 method using Quantum ESPRESSO v.6.6 (Giannozzi *et al.*, 2020). The research protocols were following our previous studies (Fakih *et al.*, 2021).

Receptor preparation. The 3D structure of SARS-CoV-2 M^{pro} was obtained from the Protein Data Bank (PDB) (<http://www.rcsb.org/pdb/>). The high resolution of the SARS-CoV-2 M^{pro} receptor

(2.15 Å) with PDB ID: 6WTT was chosen (Ma *et al.*, 2020). The receptor was complexed with boceprevir, an HCV protease inhibitor as a native ligand. Afterward, all the unique ligands and water molecules were removed from the receptor, and then the polar hydrogen and a charge (Kollman charge) were added to the protein structure. The protein preparation procedures were executed using MGLTools 1.5.6 with AutoDock 4.2 (Tanbin *et al.*, 2021).

Biological activity prediction. The biological activity spectra of amphibine analogues were assessed using the PASS prediction web server (<http://www.way2drug.com/PASSOnline/predict.php>) (Lagunin *et al.*, 2000). The predicted spectrum was estimated as probable activity (Pa) and probable inactivity (Pi), based on structure-activity relationship analysis of the training set containing more than 205000 compounds exhibiting more than 3750 kinds of biological activities. Pa and Pi values vary from 0.000 to 1.000 since they are probabilities. The PASS prediction was interpreted and used flexibly, according to Anzali *et al.* (2001).

Molecular docking simulation. The molecular docking simulation method was validated using RMSD calculation by redocked the crystallographic native ligand. The best conformation of docked native ligand was taken and superimposed with the native ligand before docked, and the Root-Mean-Square Deviation (RMSD) was calculated. The acceptable RMSD value must be less than 2.0 Å (Bell & Zhang, 2019). Afterward, the amphibine analogues were docked into the binding pocket of the SARS-CoV-2 M^{pro}. The grid box was set with coordinates 5.499, 27.197, and -11.76 (x, y, and z, respectively), and the dimensions of the grid box were 64, 60, and 60 (x, y, and z), and numbers of GA run was 100 (Atilgan & Hu, 2011).

ADME prediction. We analyzed the adsorption, distribution, metabolism, and excretion (ADME) profile of the amphibine analogues, which could be used as a drug. We used the SWISS-ADME web server to predict the ADME profile (<https://www.swissadme.ch>), which allows the user to draw or input their molecules data and

provides the parameters such as lipophilicity, water-solubility, pharmacokinetics, drug-likeness rules, and medicinal chemistry (Mahanthesh *et al.*, 2020).

RESULTS AND DISCUSSION

Biological activity prediction. The PASS prediction web server's biological activity was carried out on amphibine analogues (A-H) compounds to see the level as a COVID-19 main protease inhibitor (Table 1).

The PASS web server predicts various biological activities of Amphibine analogues, but the focus of the research here is on the prediction of antiviral and protease inhibitor agents. All amphibine A-H were predicted to have an activity as antiviral agents. As protease enzyme inhibitors, amphibine B, C, D, E, F, and H showed activity, whereas amphibine A and G showed no activity. Amphibine-C showed the best-predicted activity spectrum of 0.298 for antiviral activity and 0.151 for protease inhibitor activity. Overall, in line with Abdelli *et al.* (2021) and Shah *et al.* (2021), amphibine B, C, D, E, F, and H compounds showed potency both as antiviral activity and protease inhibitors.

Table 1. *In silico* prediction of activity spectra for substances (PASS) results.

Compounds	Activities prediction	Pa	Pi
Amphibine-A	Antiviral	0.162	0.145
	Protease Inhibitor	–	–
Amphibine-B	Antiviral	0.278	0.005
	Protease inhibitor	0.136	0.057
Amphibine-C	Antiviral	0.298	0.037
	Protease Inhibitor	0.151	0.050
Amphibine-D	Antiviral	0.278	0.045
	Protease Inhibitor	0.112	0.075
Amphibine-E	Antiviral	0.200	0.096
	Protease Inhibitor	0.102	0.085
Amphibine-F	Antiviral	0.290	0.040
	Protease Inhibitor	0.098	0.090
Amphibine-G	Antiviral	0.195	0.015
	Protease Inhibitor	–	–
Amphibine-H	Antiviral	0.263	0.052
	Protease Inhibitor	0.118	0.070

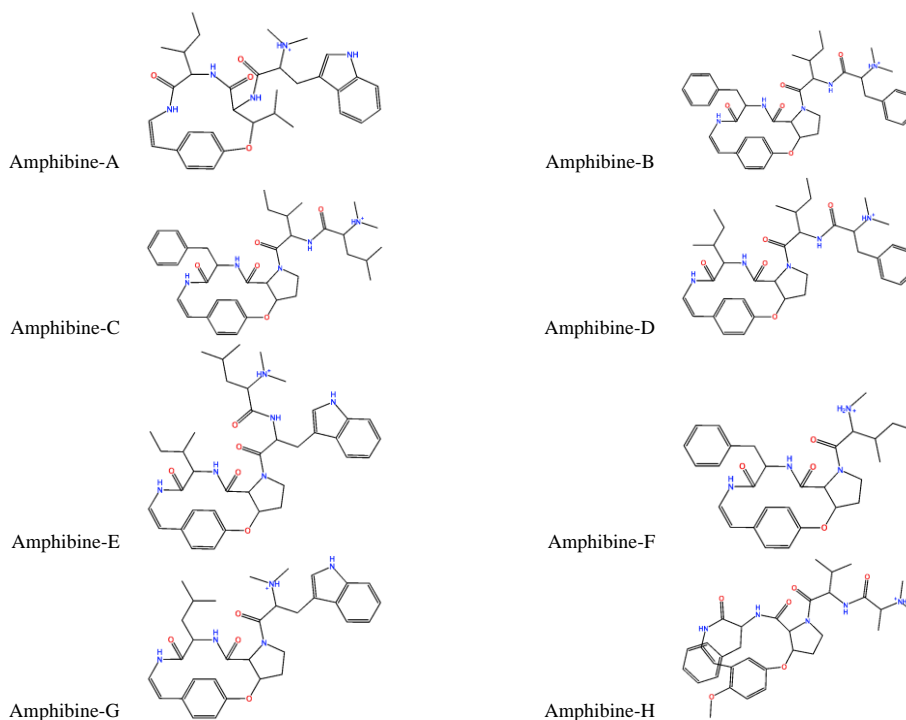


Fig. 1. Chemical structure of amphibine A-H.

Molecular docking simulation.

Molecular docking simulation study was performed on the crystal structure of SARS CoV-2 M^{pro} to assess the binding affinity potency of amphibine analogues (amphibine B, C, D, E, F, and H) that were previously predicted using the PASS website. The docking methods were validated to see the strength of binding mode prediction through re-docking the native ligand. The RMSD value of the native ligand obtained was 1.30 Å, which shows that the molecular docking method was valid. The amphibine analogues were then docked into the binding site of the crystal structure of SARS-CoV-2 M^{pro}. All of the docking results of amphibine A-H showed high binding energy and Ki compared to the native ligand (Table 2). The negative sign or the lowest binding energy is considered to be a stable binding affinity to the receptor. The binding energy of the native ligand was -7.69 Kcal/mol. The amphibine analogues binding energy sort by lowest to highest were -10.10 Kcal/mol (amphibine-F), -9.22 Kcal/mol (amphibine-E), -9.07 Kcal/mol (amphibine-B), -8.83 Kcal/mol (amphibine-H), -8.71 Kcal/mol (amphibine-D), and -8.07 Kcal/mol (amphibine-C).

Table 2. The binding energy and Ki of the amphibine of *Ziziphus spina-christi* to the SARS-CoV-2 M^{pro} receptor.

No	Compounds	Binding energy (Kcal/mol)	Ki (nM)
1	Native ligand	- 7.69	2300
2	Amphibine-B	- 9.07	224.22
3	Amphibine-C	- 8.07	1220
4	Amphibine-D	- 8.71	411.85
5	Amphibine-E	- 9.22	173.95
6	Amphibine-F	- 10.10	39.51
7	Amphibine-H	- 8.83	335.11

Molecular interactions. The ligand-receptor interactions of the best binding mode of the Amphibine analogues were analyzed and compared to reference native ligand binding mode toward the SARS-CoV-2 M^{pro}. The tabulation data of the amino acid interactions were provided in Table 3, and the 2D

interaction was provided in Fig. 2. Native ligand in its interactions, form hydrogen bonds with amino acids Glu166, His164, Phe140, Gln189, Cys145, and other types of interaction with residue Cys145 (unfavorable bump), His163 (unfavorable acceptor-acceptor), Pro168 (Pi-alkyl), Met165 (alkyl), His41 (alkyl & carbon-hydrogen bond), and His172 (carbon-hydrogen bond). Amphibine-B showed hydrogen bond interaction with three amino acids, i.e., Glu166, Ser144, Asn142, and other types of interaction with amino acids Glu166 (Pi-Anion), Pro168 (Pi-sigma), His41 (alkyl), Cys145 (Pi-alkyl), and His163 (unfavorable acceptor-acceptor). Amphibine-C showed hydrogen bond interaction with two amino acids, including Glu166 and Gln189. Other types of interaction with amino acids Gln189 (carbon-hydrogen bond), Glu166 (carbon-hydrogen bond), Pro168 (Pi-sigma), Met49 (Pi-sulfur), His41 (Pi-alkyl & Pi-pi stacked), Leu167 (Pi-alkyl), Leu141 (alkyl). Amphibine-D showed hydrogen bond interaction with one amino acid (Glu166) and other types of interaction with amino acids Met165 (alkyl & Pi-sulfur), His41 (carbon-hydrogen bond), Thr24 (carbon-hydrogen bond), Cys145 (Pi-hydrogen bond). Amphibine-E showed hydrogen bond interaction with three amino acids, including Glu166, Gln192, and Gln189, and other types of interaction with amino acids Ala191 (Pi-alkyl), Pro168 (Pi-alkyl), His41 (Pi-sigma). Amphibine-F showed hydrogen bond interaction with three amino acids, including His41, Glu166, and Gln189, and other types of interaction with amino acids His41 (Pi-sigma & unfavorable acceptor-Acceptor), Glu166 (Pi-anion), Gln189 (carbon-hydrogen bond), Arg188 (carbon-hydrogen bond), Met165 (alkyl). Amphibine-H showed hydrogen bond interaction with three amino acids, including Glu166, Gln192, and Thr190, and other types of interaction with amino acids Ala191 (Pi-alkyl), Met165 (alkyl), Glu166 (carbon-hydrogen bond), and Phe140 (carbon-hydrogen bond).

Table 3. Tabulation data of amino acid interactions of reference ligand (native ligand) compared to cyclopeptide alkaloids in SARS-CoV-2 M^{Pro}.

Ligands		H-bond interactions	Other type of interactions
Reference	native ligand	Glu166, His164, Phe140, Gln189, Cys145	Cys145k, His163l, Pro168d, Met165c, His41c,f, His172f
Amphibine-B		Glu166, Ser144, Asn142	Glu166a, Pro168b, His41c, Cys145d, His163e
Amphibine-C		Glu166, Gln189	Gln189f, Glu166f, Pro168b, Met49g, His41d,h, Leu167d, Leu141c
Amphibine-D		Glu166	Met165c,g, His41f, Thr24f, Cys145i
Amphibine-E		Glu166, Gln192, Gln189	Ala191d, Pro168d, His41b
Amphibine-F		His41, Glu166, Gln 189	His41b,e, Glu166j, Gln189f, Arg188f, Met165c
Amphibine-H		Glu166, Gln192, Thr190	Ala191d, Met165c, Glu166f, Phe140f

Notes: a= Pi-anion; b= Pi-sigma; c= alkyl; d= Pi-alkyl; e= unfavorable acceptor-acceptor; f= carbon-hydrogen bond; g= Pi-sulphur; h= Pi-pi stacked; i= Pi-hydrogen bond; j= Pi-anion; k= unfavorable bump; l= unfavorable acceptor-acceptor.

The hydrogen bond is an attractive interaction between a hydrogen atom from fragment X–H, and enhance receptor-ligand interactions (Arunan *et al.*, 2011; Chen *et al.*, 2016). Native ligands binding mode still showed the highest intensity of hydrogen bonding (five hydrogen bonds), followed by amphibine-B, amphibine-E, amphibine-F, and amphibine-H with three numbers of the hydrogen bond, then amphibine-C (two hydrogen bond), and Amphibine-D (1 hydrogen bond). The similarity of amino acid interaction types between the native ligand as a reference and amphibine analogues showed in

the amphibine-B, amphibine-C, amphibine-D, and amphibine-H provided one type of hydrogen bond interaction similar (Glu166). The amphibine-E and amphibine-F compounds showed two similar hydrogen bond interactions to the reference ligand (Glu166 & Gln189). The other interactions, i.e., Pi-sigma, Pi-alkyl, and Pi-sulphur, mostly involve charge transfer assisting in intercalating the drug at the receptor-binding site. The highest number of amino acid interactions that form those other interactions were dominated by amphibine-C, amphibine-B, amphibine-F, amphibine-H, amphibine-E, and amphibine-D, respectively.

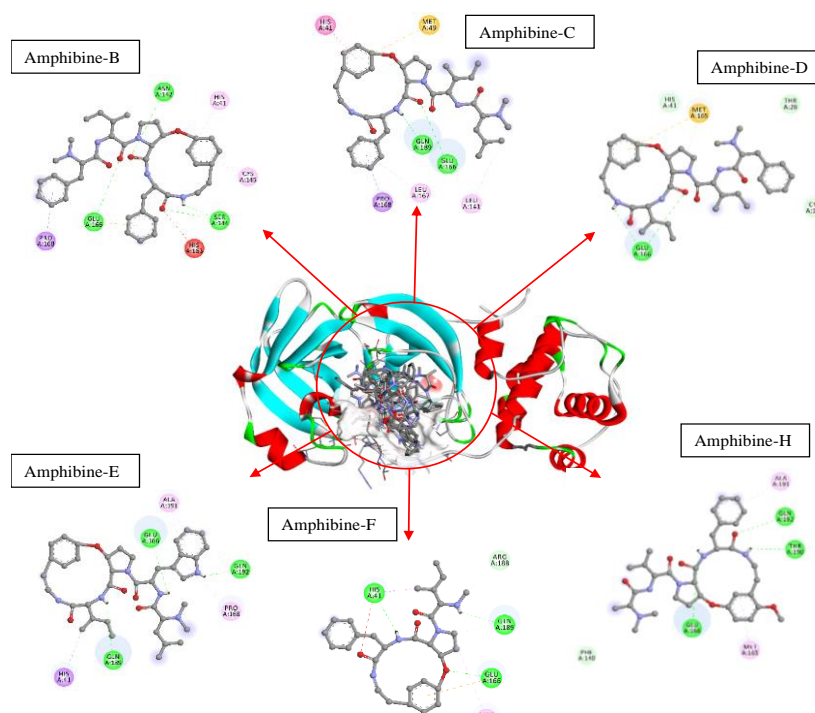


Fig. 2. Molecular interaction of native ligand and amphibine B, C, D, E, F, and H

ADME prediction. The amphibine analogues predicted before (B, C, D, E, F, and H) have been analyzed by ADME profile using SWISS-ADME (Fig. 3). The ADME profile was provided with radar that shown six predicted ADME parameters that are closely related to the oral bioavailability of a compound, including LIPO (lipophilicity), SIZE (size), POLAR (polarity), INSOLU (insolubility), INSATU (insaturation), and FLEX (flexibility). The colored zone was a physical chemistry area that is suitable for oral bioavailability. Analysis of ADME profiles performed by radar showed Amphibine-B,

Amphibine-C, Amphibine-D, and Amphibine-F have suitable in polarity and insaturation following by oral drug bioavailability criteria, except for the lipophilicity, size, insolubility, and flexibility parameter. Amphibine-E compound radar is suitable for insaturation parameters. Amphibine-F compound radar shows a suitable in all parameters, i.e., lipophilicity, size, polarity, insolubility, insaturation, and flexibility. Amphibine-H compound radar shows a suitable in polarity, insolubility, insaturation, and lipophilicity criteria.

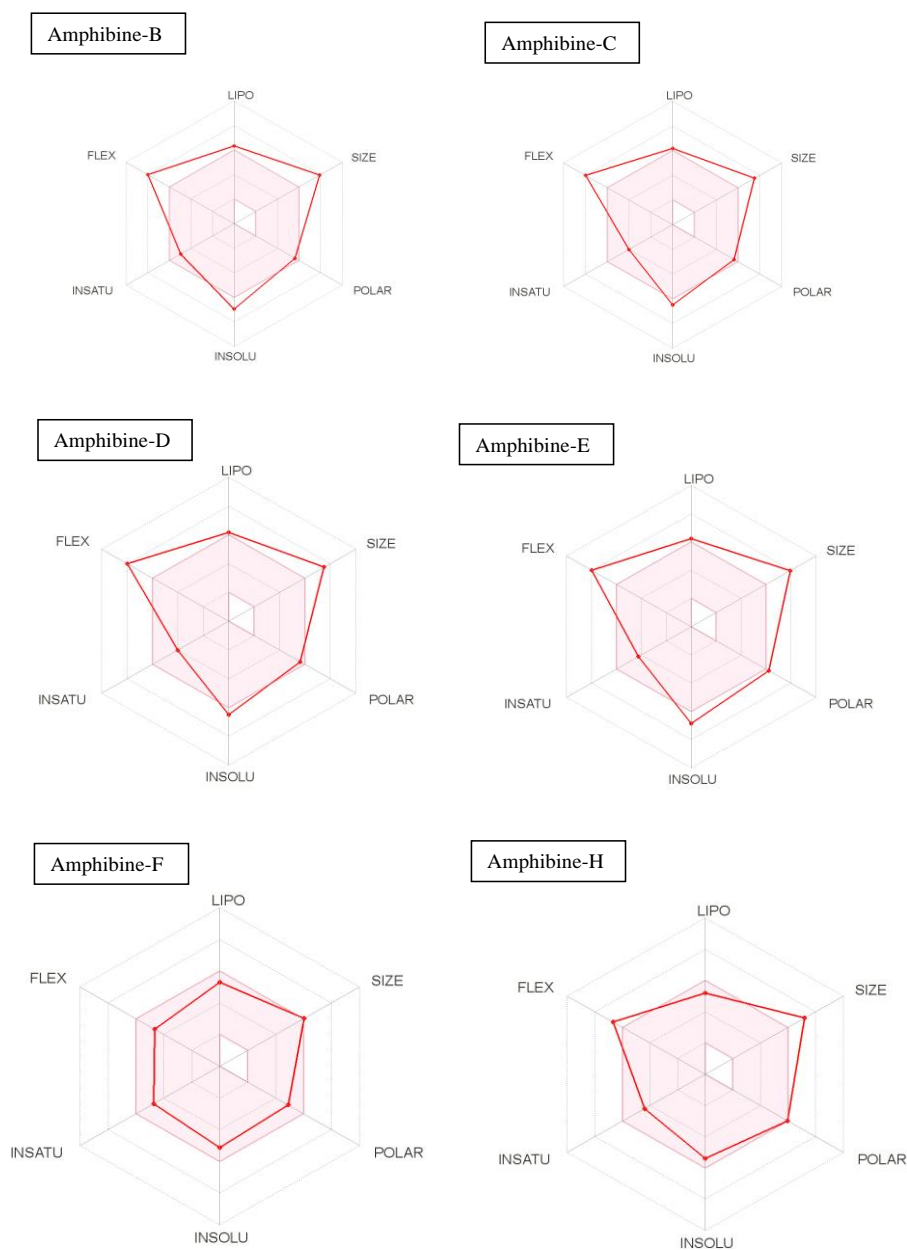


Fig. 3. Administration, distribution, metabolism, and elimination (ADME) parameters for amphibine B, C, D, E, F, and H that were evaluated by SWISS-ADME.

Furthermore, lipinski analyses were performed to look for drug similarities or determine whether a chemical compound has specific pharmacological or biological activity has chemical and physical properties that make it pharmacokinetically effective in the human body, including ADME. In Lipinski drug-likeness analysis, amphibine-B provided one violation (MW >500), amphibine-C provided one violation (MW >500), amphibine-D provided one violation (MW >500), amphibine-E provided two violation (MW >500), and (NorO > 10), amphibine-F provided one violation (MW>500), Amphibine-H provided two violations (MW>500, NorO>10). The drug-likeness results showed amphibine-E and amphibine-H provided more than one violation, indicates poor bioavailability as oral drugs. Amphibine-B, C, D, and F showed one violation for drug-likeness criteria, indicates good bioavailability. In general, the amphibine-F compounds showed the best bioavailability as an oral drug, amphibine-B, C, and D showed good bioavailability as an oral drug, and amphibine-E and H showed poor bioavailability as oral drugs.

CONCLUSION

The amphibine analogues from *Ziziphus spina-christi* species analyzed by biological activity, molecular docking, and ADME predictions were showed as potentially inhibitor candidates for the SARS-CoV-2 M^{pro} receptor. The biological activity prediction by PASS web server of amphibine analogues (A-H) showed amphibine-B, C, D, E, F, and H have potential as antiviral and protease inhibitor agents. The molecular docking results of the amphibine-B, C, D, E, F, and H showed better binding affinity against SARS-CoV-2 M^{pro} compared to the native ligand as a reference inhibitor. These compounds also form interactions that are similar in some residues with the native ligand. The ADME prediction showed amphibine-F has the best bioavailability as an oral drug, amphibine-B, C, and D have good bioavailability as an oral drug from drug-likeness criteria, while amphibine-E and H show poor bioavailability as an oral drug. Concluded the Amphibine-B, C, D, E, F, and H

have potential as a treatment of COVID-19 through inhibits the protease enzyme of SARS-CoV-2 M^{pro}, and some compounds can be formulated as oral administration (amphibine-B, C, D, and F), and some in other administration (amphibine-E and H).

ACKNOWLEDGEMENTS

The authors thank to the Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Universitas Islam Bandung, for the research financially supported by the Independent Research Grant Program 2020, No. 03/PEN-PKM/I/2021.

REFERENCES

- Ads EN, Rajendrasozhan S, Hassan SI, Sharawy SMS, Humaidi JR. 2017. Phytochemical, antimicrobial and cytotoxic evaluation of *Ziziphus spina-christi* (L.) stem bark. *Biomedical Research*. vol 28(15): 6646–6653.
- Al-Ghamdi AA, Shahat AA. 2017. Antioxidant, hypoglycemic and anti-diabetic activities of *Ziziphus spina-christi* (L) Willd (Rhamnaceae) leaf extract. *Tropical Journal of Pharmaceutical Research*. vol 16(11): 2601–2610. doi: <https://doi.org/10.4314/tjpr.v16i11.5>.
- Alotibi FO, Ashour EH, Al-Basher G. 2020. Evaluation of the antifungal activity of *Rumex vesicarius* L. and *Ziziphus spina-christi* (L) Desf. Aqueous extracts and assessment of the morphological changes induced to certain myco-phytopathogens. *Saudi Journal of Biological Sciences*. vol 27(10): 2818–2828. doi: <https://doi.org/10.1016/j.sjbs.2020.06.051>.
- Anzali S, Barnickel G, Cezanne B, Krug M, Filimonov D, Poroikov V. 2001. Discriminating between drugs and nondrugs by prediction of activity spectra for substances (PASS). *Journal of Medicinal Chemistry*. vol 44(15): 2432–2437. doi: <https://doi.org/10.1021/jm0010670>.
- Atilgan E, Hu J. 2011. Improving Protein Docking Using Sustainable Genetic Algorithms. *International Journal of Computer Information Systems and Industrial Management Applications*. vol 3(2011): 248–255.
- Baghzadeh-Daryaii L, Sharifi-Sirchi GR, Samsampoor D. 2017. Morphological, phytochemical and genetic diversity of *Ziziphus spina-christi* (L) Des. in South and Southeastern of Iran. *Journal of Applied Research on Medicinal and Aromatic Plants*. vol 7: 99–107. doi: <https://doi.org/10.1016/j.jarmap.2017.06.006>.
- Bell EW, Zhang Y. 2019. DockRMSD: An open-source tool for atom mapping and RMSD calculation of symmetric molecules through graph isomorphism. *Journal of Cheminformatics*. vol 11(1): 1–9. doi:

- <https://doi.org/10.1186/s13321-019-0362-7>.
- Benarba B, Pandiella A. 2020. Medicinal plants as sources of active molecules against COVID-19. *Frontiers in Pharmacology*. vol 11: 1–16. doi: <https://doi.org/10.3389/fphar.2020.01189>.
- Calcagno N, Colombo E, Maranzano A, Pasquini J, Sarmiento IJK, Trogu F, Silani V. 2020. Rising evidence for neurological involvement in COVID-19 pandemic. *Neurological Sciences*. vol 12: 1–3. doi: <https://dx.doi.org/10.1007%2Fs10072-020-04447-w>.
- Chen D, Oezguen N, Urvil P, Ferguson C, Dann SM, Savidge TC. 2016. Regulation of protein-ligand binding affinity by hydrogen bond pairing. *Science Advances*. vol 2(3): 1–16. doi: <https://doi.org/10.1126/sciadv.1501240>.
- Choudhury A, Mukherjee S. 2020. *In silico* studies on the comparative characterization of the interactions of SARS-CoV-2 spike glycoprotein with ACE-2 receptor homologs and human TLRs. *Journal of Medical Virology*. vol 92(10): 2105–2113. doi: <https://doi.org/10.1002/jmv.25987>.
- Dai W, Zhang B, Jiang XM, Su H, Li J, Zhao Y, Xie X, Jin Z, Peng J, Liu F, Li C, Li Y, Bai F, Wang H, Cheng X, Cen X, Hu S, Yang X, Wang J, Liu X, Xiao G, Jiang H, Rao Z, Zhang LK, Xu Y, Yang H, Liu H. 2020. Structure-based design of antiviral drug candidates targeting the SARS-CoV-2 main protease. *Science*. vol 368(6497): 1331–1335. doi: <https://doi.org/10.1126/science.abb4489>.
- Darusman F, Fakih TM. 2021. Comprehensive *in silico* analysis of chitinase molecular behaviour from *Ziziphus spina-christi* leaves on *Propionibacterium acnes*. *Pharmaceutical Sciences and Research*. vol 8(1): 55–64. doi: <https://doi.org/10.7454/psr.v8i1.1112>.
- Dias DA, Urban S, Roessner U. 2012. A historical overview of natural products in drug discovery. *Metabolites*. vol 2(2): 303–336. doi: <https://doi.org/10.3390/metabo2020303>.
- Dong R, Pei S, Yin C, He RL, Yau SST. 2020. Analysis of the hosts and transmission paths of SARS-CoV-2 in the COVID-19 outbreak. *Genes*. vol 11(6): 1–16. doi: <https://doi.org/10.3390/genes11060637>
- El Maaiden E, El Kharrassi Y, Moustaid K, Essamadi AK, Nasser B. 2019. Comparative study of phytochemical profile between *Ziziphus spina-christi* and *Ziziphus lotus* from Morocco. *Journal of Food Measurement and Characterization*. vol 13(1): 121–130. doi: <https://doi.org/10.1007/s11694-018-9925-y>.
- Fakih TM, Ramadhan DSF, Darusman F. 2021. *In silico* activity identification of cyclo peptide alkaloids from *Ziziphus spina-christi* species against Sars-Cov-2 main protease. *Jurnal Biodjati*. vol 6(1): 71–81. doi: <https://doi.org/10.15575/biodjati.v6i1.10603>.
- Fu L, Ye F, Feng Y, Yu F, Wang Q, Wu Y, Zhao C, Sun H, Huang B, Niu P, Song H, Shi Y, Li X, Tan W, Qi J, Gao GF. 2020. Both Boceprevir and GC376 efficaciously inhibit SARS-CoV-2 by targeting its main protease. *Nature Communications*. vol 11(1): 1–8. doi: <https://doi.org/10.1038/s41467-020-18233-x>.
- Gentile D, Patamia V, Scala A, Sciortino MT, Piperno A, Rescifina A. 2020. Putative inhibitors of SARS-COV-2 main protease from a library of marine natural products: A virtual screening and molecular modeling study. *Marine Drugs*. vol 18(4): 1–16. doi: <https://doi.org/10.3390/md18040225>.
- Giannozzi P, Baseggio O, Bonfà P, Brunato D, Car R, Carnimeo I, Cavazzoni C, De Gironcoli S, Delugas P, Ruffino FF, Ferretti A, Marzari N, Timrov I, Urru A, Baroni S. 2020. Quantum ESPRESSO toward the exascale. *Journal of Chemical Physics*. vol 152(15): 1–11. doi: <https://doi.org/10.1063/5.0005082>.
- Gorai M, Romdhane R, Maraghi M, Neffati M. 2019. Relationship between leaf gas-exchange characteristics and the performance of *Ziziphus spina-christi* (L.) Willd. seedlings subjected to salt stress. *Photosynthetica*. vol 57(3): 897–903. doi: <https://doi.org/10.32615/ps.2019.093>.
- Goyal B, Goyal D. 2020. Targeting the dimerization of the main protease of coronaviruses: a potential broad-spectrum therapeutic strategy. *ACS Combinatorial Science*. vol 22(6): 297–305. doi: <https://doi.org/10.1021/acscmbosci.0c00058>.
- Griffin JW. 2020. SARS-CoV and SARS-CoV-2 main protease residue interaction networks change when bound to inhibitor N3. *Journal of Structural Biology*. vol 211(3): 1–9. doi: <https://doi.org/10.1016/j.jsb.2020.107575>.
- Han Y, Král P. 2020. Computational design of ACE2-based peptide inhibitors of SARS-CoV-2. *ACS Nano*. vol 14(4): 5143–5147. doi: <https://doi.org/10.1021/acsnano.0c02857>.
- Harvey AL, Edrada-Ebel R, Quinn RJ. 2015. The re-emergence of natural products for drug discovery in the genomics era. *Nature Reviews Drug Discovery*. vol 14(2): 111–129. doi: <https://doi.org/10.1038/nrd4510>.
- Jin Z, Du X, Xu Y, Deng Y, Liu M, Zhao Y, Zhang B, Li X, Zhang L, Peng C, Duan Y, Yu J, Wang L, Yang K, Liu F, Jiang R, Yang X, You T, Liu X, Yang X, Bai F, Liu H, Liu X, Guddat LW, Xu W, Xiao G, Qin C, Shi Z, Jiang H, Rao Z, Yang H. 2020. Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors. *Nature*. vol 582(7811): 289–293. doi: <https://doi.org/10.1038/s41586-020-2223-y>.
- Kalayou S, Haileselassie M, Gebre-egziabher G, Tiku'e T, Sahle S, Taddele H, Ghezu M. 2012. *In-vitro* antimicrobial activity screening of some ethnoveterinary medicinal plants traditionally used against mastitis, wound and gastrointestinal tract complication in Tigray Region, Ethiopia. *Asian Pacific Journal of Tropical Biomedicine*. vol 2(7): 516–522. doi: [https://doi.org/10.1016/S2221-1691\(12\)60088-4](https://doi.org/10.1016/S2221-1691(12)60088-4).

- Khaerunnisa S, Kurniawan H, Awaluddin R, Suhartati S, Soetjipto S. 2020. Potential inhibitor of COVID-19 main protease (Mpro) from several medicinal plant compounds by molecular docking study. *Preprints*. vol 2020: 1–14. doi: <https://doi.org/10.20944/preprints202003.0226.v1>.
- Kwape TE, Chaturvedi P, Kamau JM, George S. 2013. Hepato-protective potential of methanol extract of leaf of *Ziziphus mucronata* (ZMLM) against dimethoate toxicity: biochemical and histological approach. *Ghana Medical Journal*. vol 47(3): 112–120.
- Lagunin A, Stepanchikova A, Filimonov D, Poroikov V. 2000. PASS: Prediction of activity spectra for biologically active substances. *Bioinformatics*. vol 16(8): 747–748. doi: <https://doi.org/10.1093/bioinformatics/16.8.747>.
- Lakshmi SA, Shafreen RMB, Priya A, Shunmugiah KP. 2020. Ethnomedicines of Indian origin for combating COVID-19 infection by hampering the viral replication: using structure-based drug discovery approach. *Journal of Biomolecular Structure and Dynamics*. vol 39(13): 4594–4609. doi: <https://doi.org/10.1080/07391102.2020.1778537>.
- Ma C, Sacco MD, Hurst B, Townsend JA, Hu Y, Szeto T, Zhang X, Tarbet B, Marty MT, Chen Y, Wang J. 2020. Boceprevir, GC-376, and calpain inhibitors II, XII inhibit SARS-CoV-2 viral replication by targeting the viral main protease. *Cell Research*. vol 30(8): 678–692. doi: <https://doi.org/10.1038/s41422-020-0356-z>.
- Mahanthesh MT, Ranjith D, Yaligar R, Jyothi R, Narappa G, Ravi MV. 2020. Swiss ADME prediction of phytochemicals present in *Butea monosperma* (Lam.) Taub. *Journal of Pharmacognosy and Phytochemistry*. vol 9(3): 1799–1809.
- Maiti BK. 2020. Potential role of peptide-based antiviral therapy against SARS-CoV-2 infection. *ACS Pharmacology & Translational Science*. vol 3(4): 783–785. doi: <https://doi.org/10.1021/acsptsci.0c00081>.
- Mirza MU, Froeyen M. 2020. Structural elucidation of SARS-CoV-2 vital proteins: Computational methods reveal potential drug candidates against main protease, Nsp12 polymerase and Nsp13 helicase. *Journal of Pharmaceutical Analysis*. vol 10(4): 320–328. doi: <https://doi.org/10.1016/j.jpha.2020.04.008>.
- Moossavi M, Hoshyar R, Hemmati M, Farahi A, Javdani H. 2017. An invivo study on the hepato-protective effects of *Crocus sativus*, *Ziziphus jujuba* and *Berberis vulgaris* against acute acetaminophen and rifampicin-induced hepatotoxicity. *Clinical Phytoscience*. vol 2(1): 1–7. doi: <https://doi.org/10.1186/s40816-016-0030-7>.
- Murdocca M, Citro G, Romeo I, Lupia A, Miersch S, Amadio B, Bonomo A, Rossi A, Sidhu SS, Pandolfi PP, Alcaro S, Sanguolo FC, Novelli G. 2021. Peptide platform as a powerful tool in the fight against COVID-19. *Viruses*. vol 13(8): 1–21. doi: <https://doi.org/10.3390/v13081667>.
- Reiner Z, Hatamipour M, Banach M, Pirro M, Al-Rasadi K, Jamialahmadi T, Radenkovic D, Montecucco F, Sahebkar A. 2020. Statins and the Covid-19 main protease: *In silico* evidence on direct interaction. *Archives of Medical Science*. vol 16(3): 490–496. doi: <https://dx.doi.org/10.5114%2Faoms.2020.94655>.
- Sakna ST, Mocan A, Sultani HN, El-Fiky NM, Wessjohann LA, Farag MA. 2019. Metabolites profiling of *Ziziphus* leaf taxa via UHPLC/PDA/ESI-MS in relation to their biological activities. *Food Chemistry*. vol 293: 233–246. doi: <https://doi.org/10.1016/j.foodchem.2019.04.097>.
- Tanbin S, Fuad FAA, Hamid AAA. 2021. Virtual screening for potential inhibitors of human hexokinase II for the development of anti-dengue therapeutics. *Biotech*. vol 10(1): 1–28. doi: <https://doi.org/10.3390/biotech10010001>.
- Thompson RN. 2020. Novel coronavirus outbreak in Wuhan, China, 2020: intense surveillance is vital for preventing sustained transmission in new locations. *Journal of Clinical Medicine*. vol 9(2): 1–7. doi: <https://doi.org/10.3390/jcm9020498>.
- WHO. 2021. Weekly operational update on COVID-19 – 1 March 2021. Geneva: World Health Organization. <https://www.who.int>.
- Zhu H, Wei L, Niu P. 2020. The novel coronavirus outbreak in Wuhan, China. *Global Health Research and Policy*. vol 5(1): 1–3. doi: <https://doi.org/10.1186/s41256-020-00135-6>.