

1 **Enhancement of UV disinfection of urine matrixes by**
2 **electrochemical oxidation**

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11

12 **Abstract**

13 This work focuses on the removal of antibiotic-resistant bacteria (ARB) contained in
14 hospital urines by UV disinfection enhanced by electrochemical oxidation to overcome
15 the limitations of both single processes in the disinfection of this type of effluents. UV
16 disinfection, electrolysis, and photoelectrolysis of synthetic hospital urine intensified
17 with *K. pneumoniae* were studied. The influence of the current density and the anode
18 material was assessed on the disinfection performance of combined processes and the
19 resulting synergies and/or antagonisms of coupling both technologies were also
20 evaluated. Results show that the population of bacteria contained in hospital urine is only
21 reduced by 3 orders of magnitude during UV disinfection. Electrolysis leads to complete
22 disinfection of hospital urine when working at 50 A m⁻² using Boron Doped Diamond
23 (BDD) and Mixed Metal Oxides (MMO) as anodes. The coupling of electrolysis to the
24 UV disinfection process leads to the highest disinfection rates, attaining a complete
25 removal of ARB for all the current densities and anode materials tested. The use of MMO
26 anodes leads to higher synergies than BDD electrodes. Results confirm that UV
27 disinfection can be enhanced by electrolysis for the removal of ARB in urine, considering
28 both technical and economic aspects.

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32 **Keywords**

33 UV disinfection; electrolysis; photoelectrolysis; antibiotic-resistant bacteria; urine

34 **Highlights**

- 35 - UV disinfection decreases *K. pneumoniae* in hospital urine by 3-logs.
36 - Electrolysis ensures a complete bacteria removal with BDD and MMO anodes at
37 50 Am⁻².
38 - The highest disinfection rates are obtained during photoelectrolysis.
39 - Synergies are found when coupling UV disinfection and electrolysis at 5 A m⁻².
40 - Hypochlorite and chloramines are the main responsible species for disinfection.

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54 **1. Introduction**

55 In last years, concern about hospital wastewater treatment has sharply increased in the
56 scientific community, not only related to the presence of a great variety of drugs but also
57 for the high concentration of pathogens, viruses, and fungi [1, 2]. Wastewater generated
58 in hospital facilities may have very different sources (laundry, kitchen, toilets...) and
59 among them, hospital urine can be considered as the main hotspot of both drugs and
60 microorganisms, despite urine only constitutes 2-3 % of the total volume of wastewater
61 generated [3]. In turn, hospital wastewater (including urine) is merged with urban
62 wastewater and treated in conventional wastewater treatment plants (WWTPs) that are
63 not able to efficiently remove the microorganisms owing to the limitations of the
64 technology used [4, 5]. This promotes the occurrence of antibiotic-resistant bacteria
65 (ARB) [6, 7], one of the biggest threats of global health, food security, and development
66 today considered by the World Health Organization (WHO). To prevent the appearance
67 of ARB, hospital urines could be pre-treated before its merge with hospital wastewater
68 and the subsequent discharge to WWTP, using advanced and efficient processes for the
69 removal of microorganisms.

70 To the authors' knowledge, the disinfection of urine has not been studied in-depth and
71 few recent works have been previously reported in the literature [8-10]. Researches
72 focused on the disinfection of urban and hospital wastewater have been more regular,
73 using different Advanced Oxidation Processes (AOPs) among other techniques [11-13].
74 One of the AOPs widely evaluated for the treatment of urban and hospital wastewater is
75 ultraviolet (UV) disinfection [13-15]. The irradiation of UV light to an effluent containing
76 microorganisms can promote bacteria removal by destroying the genetic material [16].
77 Likewise, UV light can penetrate the cell wall, destroying the genetic material. However,
78 the damage caused on bacteria can be repaired after relatively small times, promoting

79 bacteria regrowth [17, 18]. Besides, the presence of suspended solids and colloids in
80 wastewater decreases the process efficiency. This can be an important drawback in the
81 removal of ARB in hospital urines and this should be taken into consideration to ensure
82 efficient and persistent disinfection.

83 At this point, the use of combined treatments appears as a good option to overcome these
84 limitations. Among the different alternatives, electrochemical oxidation is a potential
85 candidate to be considered. Depending on the electrode material and applied current
86 density [19, 20], this technology can promote the formation of hydroxyl radicals from
87 water electrolysis and has been satisfactorily tested for the disinfection of urban
88 wastewater and even hospital urines [9, 21, 22]. Furthermore, during electrolysis, other
89 oxidants can be generated from the oxidation of the ions naturally contained in the
90 effluents that can also contribute to the disinfection process, preventing the bacteria
91 regrowth. At this point, the role of anode material is critical. Thus, the use of anodes based
92 on Mixed Metal Oxides (MMO) such as RuO₂, IrO₂, or Ta₂O₅ promotes the evolution of
93 chlorine during electrolysis [23-25]. On the other hand, Boron Doped Diamond (BDD)
94 anodes favour the formation of a cocktail of oxidants (e.g., hypochlorite, persulfate,
95 peroxodiphosphate, percarbonate, ozone) which seems to be the main responsible for the
96 good performance of this electrode in the treatment of industrial wastewater [26, 27] and
97 wastewater disinfection [28, 29].

98 With this background, the goal of this work is to evaluate the synergies or antagonisms
99 when coupling electrochemical oxidation to UV disinfection for the removal of ARB in
100 hospital urines. To do this, *Klebsiella pneumoniae* (*K. pneumoniae*) has been selected as
101 a model of ARB because it is mainly excreted in urine [30, 31]. It is a gram-negative
102 bacteria that belongs to the Enterobacteriaceae family and presents antibiotic resistance
103 to third-generation cephalosporins and carbapenems [32, 33]. UV disinfection,

104 electrolysis, and the combined process (photoelectrolysis) were carried out using an
105 electrochemical microfluidic flow-through reactor, paying special attention to the
106 influence of the current density (5-50 A m⁻²) and the anode material (BDD and MMO).
107 This will allow us to assess if the UV disinfection process can be enhanced for the
108 removal of ARB in very complex media such as from hospital urines.

109

110 **2. Material and methods**

111 **2.1. Chemicals and microorganisms**

112 Compounds employed for the formulation of synthetic urine (urea, creatinine, uric
113 acid, potassium chloride, magnesium sulfate, calcium phosphate, sodium carbonate, and
114 diammonium hydrogen phosphate) were analytical grade and used as received (Sigma
115 Aldrich, Spain). The bacterial strain used during UV disinfection, electrolysis, and
116 photoelectrolysis was *K. pneumoniae* ATCC 4352 (CECT, Spain). Potassium phosphate
117 monobasic, sodium phosphate dibasic, ethylenediaminetetraacetic acid (EDTA), N,N-
118 Diethyl-p-phenylenediamine sulfate (Sigma Aldrich, Spain), and sulfuric acid (VWR)
119 were used for the determination of chloramines. Acetone, 2,6-pyridinedicarboxylic acid,
120 and nitric acid (Sigma Aldrich, Spain) were used for the determination of ion
121 concentrations. Arsenic trioxide used for the determination of hypochlorite ion was also
122 purchased by Sigma Aldrich, Spain. All chemicals used for analytical measurements were
123 analytical grade and used as received. Double deionized water (Millipore Milli-Q system,
124 resistivity: 18.2 MΩ cm at 25 °C) was used to prepare all solutions.

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127 2.2. Experimental procedure

128 UV disinfection process was carried out in a cylindrical reactor made of borosilicate
129 where a UV germicidal lamp of 5 W ($\lambda = 254$ nm) was located in the middle of the system.
130 Infected synthetic urine (2 dm^3) was recirculated with a peristaltic pump (Percom N-M,
131 JP Selecta) to favor the mass transfer in the photoreactor. Electrolysis experiments were
132 developed in a microfluidic flow-through electrochemical cell equipped with mesh
133 electrodes. Boron Doped Diamond (BDD) and Mixed Metal Oxides (MMO) were used
134 as anodes and Stainless Steel (SS) as the cathode. BDD anode consisted of a thin diamond
135 film supported on a niobium mesh (Diachem®) with a size of $9.5 \times 8 \text{ cm}^2$, supplied by
136 Condias GmbH (Germany). MMO anode consisted of an $\text{IrO}_2/\text{Ta}_2\text{O}_5$ (70:30) coated
137 titanium mesh with a total dimension of $9.5 \times 8 \text{ cm}^2$, supplied by Tianode (India). The
138 surface area was 53 and 50 cm^2 for MMO and BDD, respectively [34]. A Delta
139 Electronika ES030-10 power supply (0-30V, 0-10A) provided the electric current. The
140 effluent was recirculated with a peristaltic pump. Photoelectrolysis was carried out by
141 coupling the photoreactor used for the UV disinfection process to the microfluidic flow-
142 through electrochemical reactor. A detail of the experimental setup used is shown in Fig.
143 S1. All electrochemical and photoelectrochemical experiments were employed under
144 galvanostatic conditions with an initial volume of 2 dm^3 and a flow rate of $40 \text{ dm}^3 \text{ h}^{-1}$.
145 Current densities applied were between 5 and 50 A m^{-2} . These values have been selected
146 considering the two different limit behaviors of electrolysis: direct oxidation which is
147 promoted at low current densities (5 A m^{-2}) and mediated oxidation that is expected to
148 prevail when working at 50 A m^{-2} [35, 36]. All experiments were conducted in triplicate.
149 The simulated hospital urine was prepared with synthetic human urine as supporting
150 electrolyte intensified with *K. pneumoniae* (10^6 CFU mL^{-1}). Table 1 shows the
151 composition of the synthetic urine used in this work [37].

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Table 1. Composition of the synthetic urine.

Compound	Concentration (mg dm ⁻³)
CH ₄ N ₂ O	3333.34
C ₄ H ₇ N ₃ O	166.67
C ₅ H ₄ N ₄ O ₃	50.00
KCl	1000.00
MgSO ₄	170.00
(Ca) ₃ (PO ₄) ₂	28.34
Na ₂ CO ₃	166.67
(NH ₄) ₂ HPO ₄	83.34

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154 **2.3. Analytical methods**

155 Bacteria (*K. pneumoniae*) counts were performed by an indirect impedance method using
 156 a μ -Trac[®] 4200 system. It uses a standard impedance signal (Media Impedance = M-
 157 value) that is registered as the result of microbial metabolism, which causes the change
 158 of the impedance in an AC field. During the growth of microorganisms low or non-
 159 charged nutrient molecules are metabolized. This break down of nutrients generates small
 160 and highly charged metabolites that result in a change of the impedance of the nutrient
 161 media that is recorded by the instrument [38, 39]. The impedance values are related to the
 162 concentration of *K. pneumoniae* in CFU mL⁻¹ through an initial calibration of the
 163 equipment by plate counts.

164 The DPD (N,N-diethyl-phenylenediamine) standard colorimetric method [40], was used
 165 for the determination of inorganic chloramines. This is a selective method that allows us
 166 to determine the presence of mono-, di- and trichloramine without the interferences of

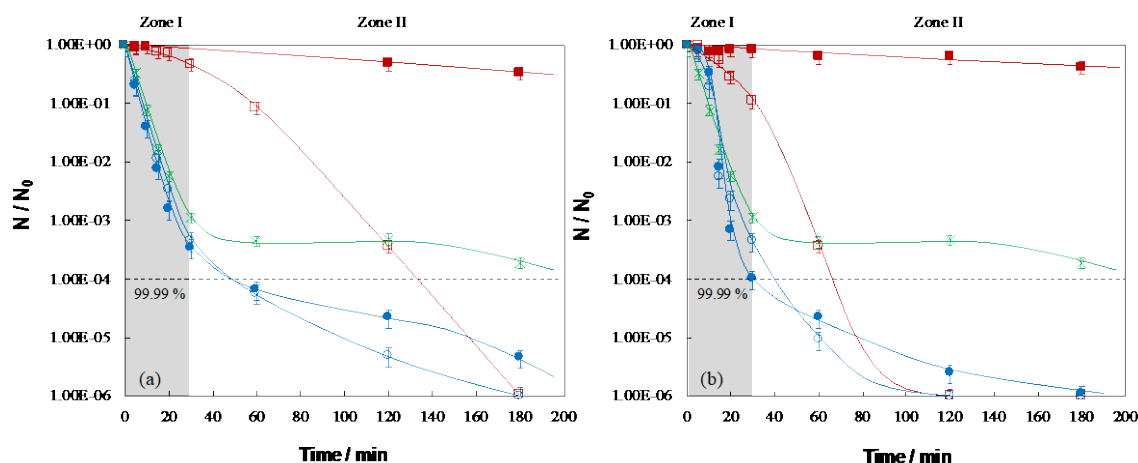
167 other oxidizing species formed during the process, such as, ozone or hydrogen peroxide.
168 Ion chromatography (IC), Metrohm 930 Compact IC Flex coupled to a conductivity
169 detector, was used to analyze the inorganic ions. The columns Metrosep A Supp 7 and
170 Metrosep C6 250 were used to measure anions and cations, respectively. The mobile
171 phase of 85:15 v/v 3.6 mM Na₂CO₃/acetone with a flow rate of 0.8 cm³ min⁻¹ was used
172 to measure anions and a mobile phase of 1.7 mM HNO₃ + 1.7 mM 2,6-
173 pyridinedicarboxylic acid with a flow rate of 0.9 cm³ min⁻¹ was employed for cations.
174 The volume injection of each sample was 20 μL. The peak of the chromatogram for
175 hypochlorite interferes with the chloride peak in the IC technique, therefore, hypochlorite
176 was analyzed by titration with 0.001 M As₂O₃ in 2 M NaOH [41, 42]. This analytical
177 technique is selective in measuring anion hypochlorite. The pH and conductivity were
178 simultaneously measured using a Sension+ MM150 Portable Multi-Parameter Meter
179 from HACH.

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181 **3. Results and discussion**

182 Fig. 1 shows changes in the concentration of *K. pneumoniae* with the operation time
183 during the UV disinfection, electrodisinfection, and photo-electrodisinfection of hospital
184 urines. As can be observed, UV disinfection reduces 3 orders of magnitude the
185 concentration of bacteria in hospital urine after 3 hours of treatment. The removal of
186 microorganisms may be due to the penetration of UV light into the cell, destroying the
187 genetic material [43]. Two zones can be distinguished in which the disinfection rate is
188 very different. Initially, the inactivation of *K. pneumoniae* is very fast, reaching the
189 maximum log removal around 30 min (zone I). After that, the disinfection rate decreases
190 drastically (zone II). This behavior has also been reported in the literature during the

191 application of the UV disinfection process to the treatment of wastewater, although the
 192 occurrence of a plateau zone has not yet been clearly explained [44, 45]. Several
 193 hypotheses have been proposed such as the aggregation of microorganisms or a resistant
 194 subpopulation, but no clear evidences have been reported [17].



196 **Fig. 1.** Time course of the concentration of *K. pneumoniae* during the treatment of
 197 hospital urines. Green symbols (x): UV disinfection; red symbols (■, □): electrolysis; blue
 198 symbols (●, ○): photoelectrolysis; full symbols: 5 A m⁻²; empty symbols: 50 A m⁻²; (a)
 199 BDD anode; (b) MMO anode. *K. pneumoniae*₀ (N₀): 10⁶ CFU mL⁻¹.

200 On the other hand, complete inactivation of *K. pneumoniae* can be attained in hospital
 201 urines using electrolysis with BDD and MMO anodes at 50 A m⁻², while at low current
 202 densities (5 A m⁻²) the population of microorganisms slightly decreases without removing
 203 1 order of magnitude, regardless the anode material used. This reveals that a minimum
 204 current density (higher than 5 A m⁻²) should be applied to ensure complete disinfection.
 205 However, different trends can be seen during electrolysis at higher current densities with
 206 both anodes. Specifically, an operation time of 180 min (which corresponds with 0.42 Ah
 207 dm⁻³ of electric charge) is needed for killing bacteria with BDD anodes whereas a total
 208 inactivation is achieved in 120 min (which corresponds with 0.28 Ah dm⁻³ of electric

209 charge) using MMO electrodes at 50 A m^{-2} . This is an unexpected behavior because
210 electrochemical disinfection with BDD is generally more efficient and faster than with
211 MMO anodes [46]. Nonetheless, it should be noted that the properties of diamond anodes
212 significantly influence the performance of an electrochemical process [47-49] and
213 therefore, general conclusions should not be drawn for all diamond electrodes. The
214 electrode used in the work was made over a niobium substrate, which has shown not to
215 promote the production of large amounts of powerful oxidants during electrolysis, as
216 tantalum and silicon substrates [50]. So, this could explain the lower disinfection rate
217 registered in comparison with MMO anodes. Likewise, it is important to point out that
218 the MMO anodes used are based on $\text{IrO}_2/\text{Ta}_2\text{O}_5$ which favors the evolution of oxygen and
219 chlorine during electrolysis and the subsequent disinfection process [51]. A detailed
220 discussion related to the influence of the anode material on the production of oxidants
221 during electrochemical disinfection is reported later. Nevertheless, at this point, it should
222 be mentioned that the concentration of *K. pneumoniae* was also measured after 24 h to
223 evaluate a possible bacteria regrowth. For this additional test, samples were stored in dark
224 at room temperature when finishing the experiments. The population of microorganisms
225 was similar to that obtained at the end of electrolysis which confirms that the
226 electrochemical process ensures persistent disinfection.

227 Regarding the photoelectrolysis process, results obtained show the highest disinfection
228 rates, reaching a reduction higher than 5 orders of magnitude for all the current densities
229 and anode materials tested. The removal of *K. pneumoniae* follows the same profile as
230 UV disinfection at the beginning of the treatment (Zone I), reaching 3-4 orders of
231 magnitude reduction for both anodes, regardless the current density applied. Then, unlike
232 behavior observed during single photolysis, the population of microorganisms continuous
233 to decrease until the total disinfection (Zone II), being the removal rate higher at 50 A m^{-2}

234 ². Photoelectrolysis is much better than single processes since improves 2-3 orders of
235 magnitude the UV disinfection process and even 5-6 orders of magnitude the
236 electrochemical process at 5 A m⁻². At high current densities, electrochemical and
237 photoelectrochemical processes attain complete disinfection but the last one shows a
238 higher removal rate. Likewise, the use of MMO anodes leads to higher disinfection rates
239 in comparison with BDD anodes due to their different electrocatalytic properties for the
240 production of disinfectants [52]. In this context, the large amounts of disinfectants
241 generated during the electrolysis of hospital urines [9] can be activated by the irradiation
242 of UV light, favoring the production of free radicals which are more reactive than the
243 parent oxidants [53]. Hence, the higher disinfection efficiency can be attributed to the
244 photo-activation of oxidants during the combined treatment. The two different zones
245 observed indicate that the contribution of UV light irradiation is more significant at the
246 beginning of the process (zone I) whereas the combination of photochemical and
247 electrochemical processes leads to remarkable synergies in the disinfection at the end of
248 the treatment (zone II). Likewise, the low electric charges applied within 30 min of the
249 experiments ($Q < 0.1 \text{ Ah dm}^{-3}$) may support the almost negligible contribution of
250 electrochemical oxidation at the beginning of the treatment because of the expected low
251 generation and accumulation of disinfectants in the system.

252 For comparison purposes, experimental data were fitted to a first-order kinetic model and,
253 the resulting kinetic constants, the residual variance (S^2_R), and the correlation coefficient
254 (R^2) are presented in Table 2. Two different values have been calculated for
255 photochemical and photoelectrochemical processes that correspond to the different zones
256 observed in the removal of microorganisms and only one value is reported for the single
257 electrochemical process.

258 Comparing the combined process with UV disinfection, the kinetic constants are a little
 259 higher in the zone I during photoelectrolysis, which confirms that there exists a small
 260 contribution of the electrochemical process to the removal of microorganisms at the
 261 beginning of the treatment. On the contrary, the values registered in zone II are much
 262 higher, being more noticeable at higher current densities where the production of oxidants
 263 is expected to be higher by electrolysis and, hence, the disinfection rate. These results
 264 point out the higher contribution of the electrochemical process at the end of the
 265 treatment. To quantify the improvement of UV disinfection by electrolysis, the synergy
 266 coefficient was calculated according to Eq. 1 [54]. Fig. 2 shows the synergy coefficient
 267 for the combined process and the logarithmic removal increase between UV disinfection
 268 and photoelectrolysis.

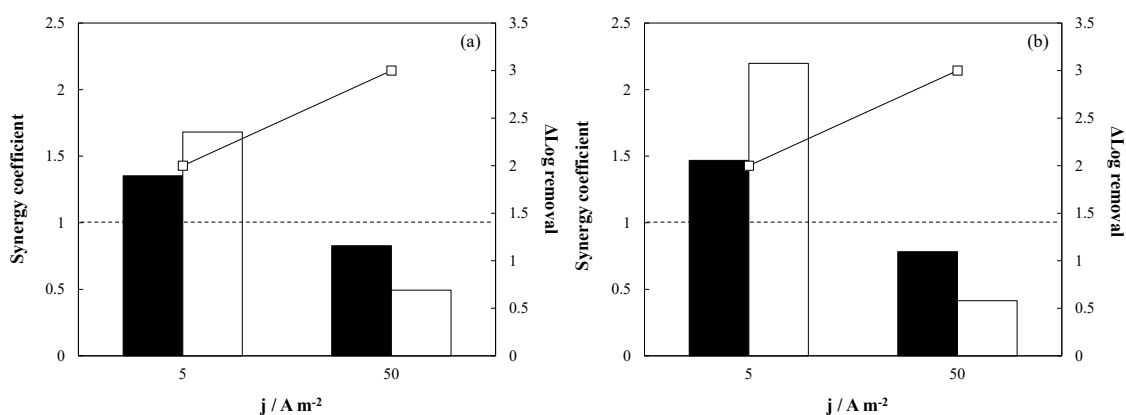
269 **Table 2.** Kinetic constants for the disinfection by UV irradiation, electrolysis, and
 270 photoelectrolysis.

Process	Anode material	Current density (A m ⁻²)	<i>k</i> _{Zone I} (min ⁻¹)	<i>S</i> ² _R (CFU ² /mL ²)	<i>R</i> ²	<i>k</i> _{Zone II} (min ⁻¹)	<i>S</i> ² _R (CFU ² /mL ²)	<i>R</i> ²
UV disinfection	-	-	0.2338	0.1327	0.9795	0.0098	0.1498	0.7141
Electrolysis*	BDD	5	0.0062	0.0002	0.9992	0.0062	0.0002	0.9992
		50	0.0770	0.5861	0.9747	0.0770	0.5861	0.9747
	MMO	5	0.0039	0.0125	0.8193	0.0039	0.0125	0.8193
		50	0.1400	0.4833	0.9848	0.1400	0.4833	0.9848
Photoelectrolysis	BDD	5	0.3247	0.0003	0.9999	0.0269	0.1967	0.9411
		50	0.2570	0.1237	0.9842	0.0428	0.2258	0.9726
	MMO	5	0.3491	1.2676	0.9168	0.0301	0.3666	0.9135
		50	0.2926	0.9813	0.9089	0.062	2.2737	0.7498

271 *only one zone

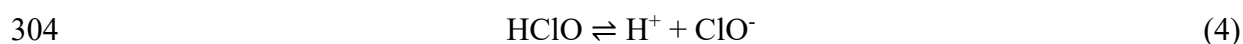
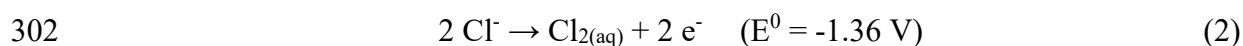
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$$\text{Synergy coefficient} = \frac{k_{\text{photoelectrolysis}}}{k_{\text{UV disinfection}} + k_{\text{electrolysis}}} \quad (1)$$

273 A synergistic effect resulted in the disinfection of urines during photoelectrolysis at 5 A
 274 m⁻², regardless the anode material used. Under these conditions, there is a marked
 275 contribution of UV disinfection at the beginning of the treatment and then, the
 276 electrochemical process has a more significant influence on the removal of ARB. The
 277 highest synergistic effect was obtained in zone II during photoelectrolysis with MMO
 278 anodes where the disinfection by UV irradiation is very limited. These synergies could
 279 be explained by the generation and photoactivation of the disinfectant species formed
 280 during the combined process. More details about the disinfectants produced during the
 281 different processes tested are reported later. At higher current densities, antagonistic
 282 effects (synergy coefficient < 1) can be observed during photoelectrolysis with both
 283 anodes. This can be due to the competitive oxidation between *K. pneumoniae* and
 284 organics contained in the urine matrix (urea, creatinine, and uric acid) under these
 285 operating conditions [55]. These results reveal that photoelectrolysis is an efficient
 286 technology for the disinfection of hospital urines when working at low current densities
 287 since electrolysis can enhance by 2 orders of magnitude the removal of *K. pneumoniae*
 288 attained by UV disinfection. The use of current densities of 50 A m⁻² improves the
 289 removal in 3 orders of magnitude but clear antagonistic effects have been obtained.

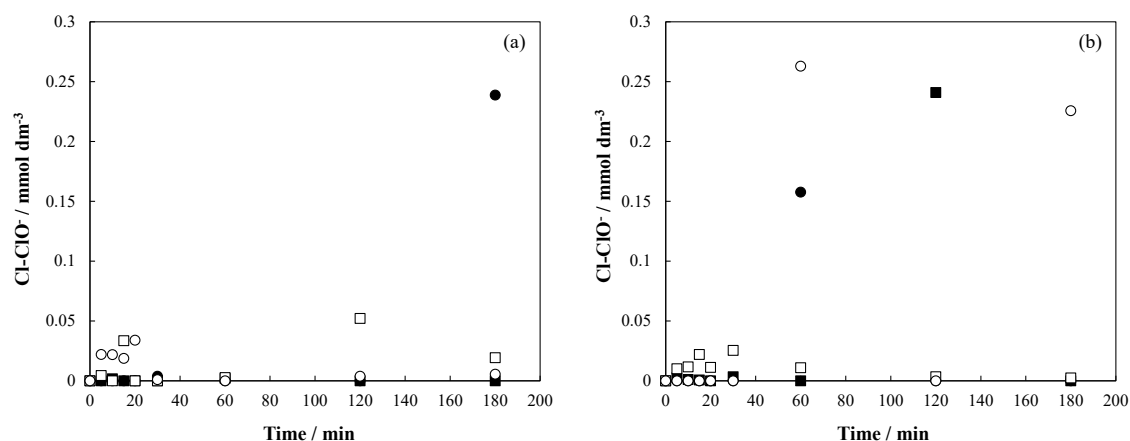


291 **Fig. 2.** Synergistic effect (bars) and ΔLog removal (points) calculated at different current
292 densities. Black bars: zone I; white bars: zone II; (a) BDD anode; (b) MMO anode.

293 The removal of ARB from urine by electrochemical processes could take place by the
294 direct oxidation of microorganisms over the anode surface (electroadsorption) or by
295 mediated oxidation via electrogenerated oxidants. The first option mainly occurs when
296 using porous materials such as carbon felt or cloth as the anode, where bacteria can be
297 adsorbed [56]. On the other hand, the in-situ production of disinfectants and the
298 subsequent attack to microorganisms seems to be the main mechanism for killing bacteria
299 during electrolysis with BDD and MMO anodes [9, 57]. Urine matrix used in this work
300 contains large amounts of chloride in its composition ($475.52 \text{ mg dm}^{-3}$) whose first
301 oxidation product is hypochlorite (Eqs. 2-4).



305 This species is a well-known disinfectant that is used in conventional disinfection
306 processes [58] and, for this reason, its concentration was monitored during the treatment.
307 Fig. 3 shows the hypochlorite electrogenerated as a function of the operation time during
308 the disinfection of hospital urines by electrolysis and photoelectrolysis with BDD and
309 MMO anodes at the current densities tested. Data from UV disinfection is not included
310 in the figure because hypochlorite was not detected during this photochemical process.

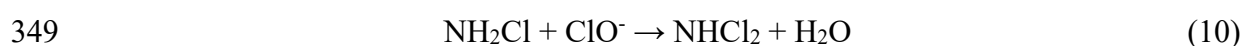
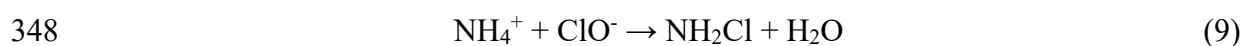
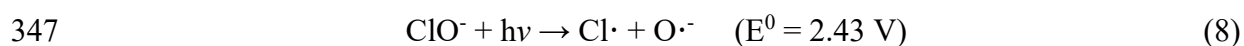
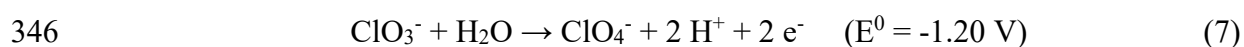
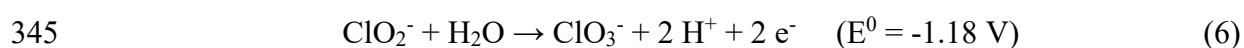
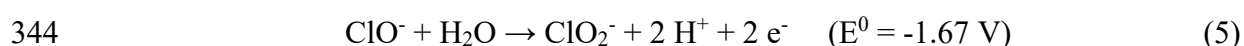


312 **Fig. 3.** Time course of the concentration of hypochlorite during the treatment of hospital
 313 urines. Black symbols: electrolysis; white symbols: photoelectrolysis; (■, □) 5 A m^{-2} ; (●,
 314 ○) 50 A m^{-2} ; (a) BDD anode; (b) MMO anode.

315 The amount of electrogenerated hypochlorite is lower than $0.3 \text{ mmol dm}^{-3} \text{ Cl}$, regardless
 316 the current density and the anode material used. However, it should be pointed out that
 317 the hypochlorite measured is referred to the free concentration that has not reacted in the
 318 effluent and much higher amounts are expected to have been generated during the
 319 treatment. To check this hypothesis, electrolysis of $1000 \text{ mg dm}^{-3} \text{ KCl}$ was carried out
 320 (Fig. S2a) since this compound is contained in the urine matrix in the same concentration.
 321 In these tests, electrogenerated hypochlorite is expected to be easily accumulated because
 322 it cannot react with other species as occur in the urine matrix. Results confirm that large
 323 amounts of hypochlorite are generated at higher current densities (50 A m^{-2}).
 324 Nevertheless, the concentration of this species is very low at 5 A m^{-2} , which implies that
 325 higher values are required to ensure a significant production of hypochlorite during
 326 electrolysis. This also supports the low disinfection rates registered during the treatment
 327 of hospital urines under these conditions (5 A m^{-2}) where it is not possible to achieve 1
 328 order of magnitude removal. Furthermore, it can be observed that the electrochemical
 329 production of hypochlorite is influenced by the anode material used. Specifically,

330 concentrations around 1.8 mmol dm^{-3} Cl are achieved with MMO anodes at 50 A m^{-2}
 331 whereas the use of BDD anodes leads to the generation of concentrations lower than 0.6
 332 mmol dm^{-3} Cl at the end of the process under the same operating conditions. This
 333 confirms the better electrocatalytic properties of the MMO anode tested to produce
 334 chlorine disinfectants and the subsequent removal of *K. pneumoniae* from urine.

335 Hypochlorite concentration does not follow a clear trend during the disinfection tests, and
 336 it could be related to the high reactivity of this species in the urine matrix: 1) hypochlorite
 337 can promote to other chlorine compounds in high oxidation state (i.e., chlorite, chlorate,
 338 and perchlorate) (Eqs. 5-7), 2) the photoactivation of hypochlorite can occur by the
 339 irradiation of UV light, favoring the production of free chlorine radicals (Eq. 8), 3) the
 340 degradation of organics naturally contained in urine (urea, creatinine and uric acid) can
 341 take place by chlorination reaction or 4) hypochlorite can react with ammonium contained
 342 in the effluent, favoring the production of combined chlorine species (chloramines) (Eqs.
 343 9-11). All these reactions can occur simultaneously during the process.

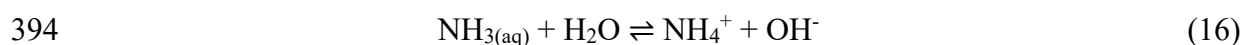
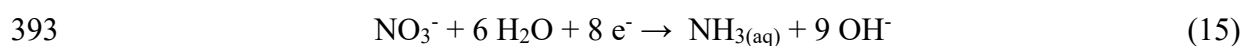
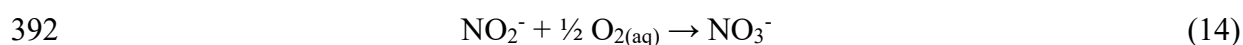
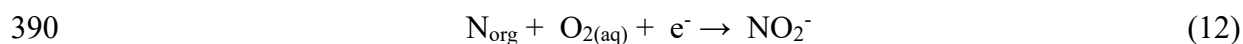


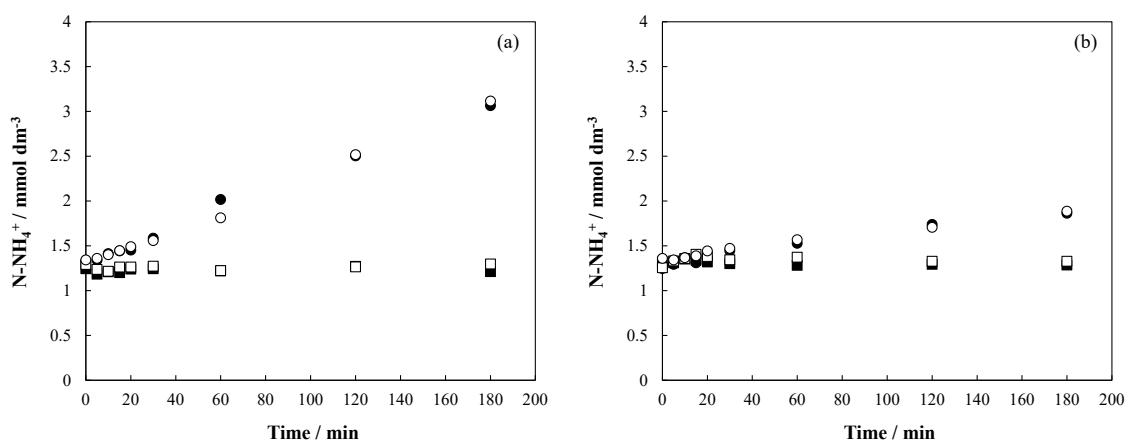
351 The formation of chlorate and perchlorate was not detected by ion chromatography for
352 all the tests carried out with BDD and MMO anodes. The measured concentration was
353 below the detection limit for each species (ClO_3^- : $7 \mu\text{g dm}^{-3}$; ClO_4^- : $7 \mu\text{g dm}^{-3}$). This data
354 is of great importance because it reveals that the removal of ARB from urine by
355 electrolysis and photoelectrolysis could be carried out under the operating conditions
356 tested without the generation of undesirable chlorine by-products.

357 On the other hand, the concentration of free chlorine radicals is difficult to measure owing
358 to their short lifetime. Thus, to evaluate the possible formation of these species during the
359 disinfection of hospital urines by photoelectrolysis, experiments with 1000 mg dm^{-3} were
360 performed (Fig. S2b). As can be observed, the concentration of hypochlorite increases
361 with the operation time at 50 A m^{-2} whereas a negligible concentration was obtained at 5
362 A m^{-2} for both anodes. This behavior is similar to that previously obtained during single
363 electrolysis of KCl solutions (Fig. S2a). However, the concentration achieved is lower
364 during photoelectrolysis which indicates that hypochlorite has been photoactivated,
365 favoring the production of free chlorine radicals (Eq. 8). Therefore, the occurrence of
366 these radicals can be considered as one way for electrogenerated hypochlorite
367 consumption during photoelectrolysis of hospital urines.

368 Regarding the chlorination reaction of organics, urine contains large amounts of urea,
369 creatinine, and uric acid which are susceptible to react with hypochlorite during the
370 electrochemical treatment, decreasing their concentration [9]. The concentration of urea
371 remained constant for all the tests carried out with both anodes. Creatinine concentration
372 decreased between 10 and 25 % at 5 A m^{-2} and, up to 40 % at 50 A m^{-2} with BDD anodes
373 whereas its concentration remained constant when using MMO anodes, regardless the
374 current density applied. Finally, a percentage removal of uric acid around 50 % was
375 achieved during electrolysis and photoelectrolysis with BDD and MMO anodes at 5 and

376 50 A m⁻² (data not shown). These reactions could promote the formation of
 377 organochlorinated compounds in hospital urines which may increase the toxicity of the
 378 resulting effluents. Nonetheless, their generation and accumulation are not expected to be
 379 very high due to the low applied electric charges required for killing microorganisms. The
 380 degradation of urea, creatinine and uric acid releases a higher nitrogen load to the effluent
 381 which can be transformed into other inorganic species [59]. Specifically, nitrite is formed
 382 from the oxidation of organic nitrogen released (Eq. 12, which is rapidly oxidized to
 383 nitrate (Eqs. 13-14). Besides, this last species can be electrochemically reduced, favoring
 384 the generation of ammonium (Eqs. 15-16) [60]. Hence, the concentration of ammonium
 385 could be considered as an indirect measurement of the degradation of organics during the
 386 disinfection of urines. Fig. 4 shows the time course of the ammonium concentration
 387 during the disinfection of hospital urines by electrolysis and photoelectrolysis with BDD
 388 and MMO anodes at different current densities. The initial amount in the urine matrix is
 389 around 1.26 mmol dm⁻³ N.



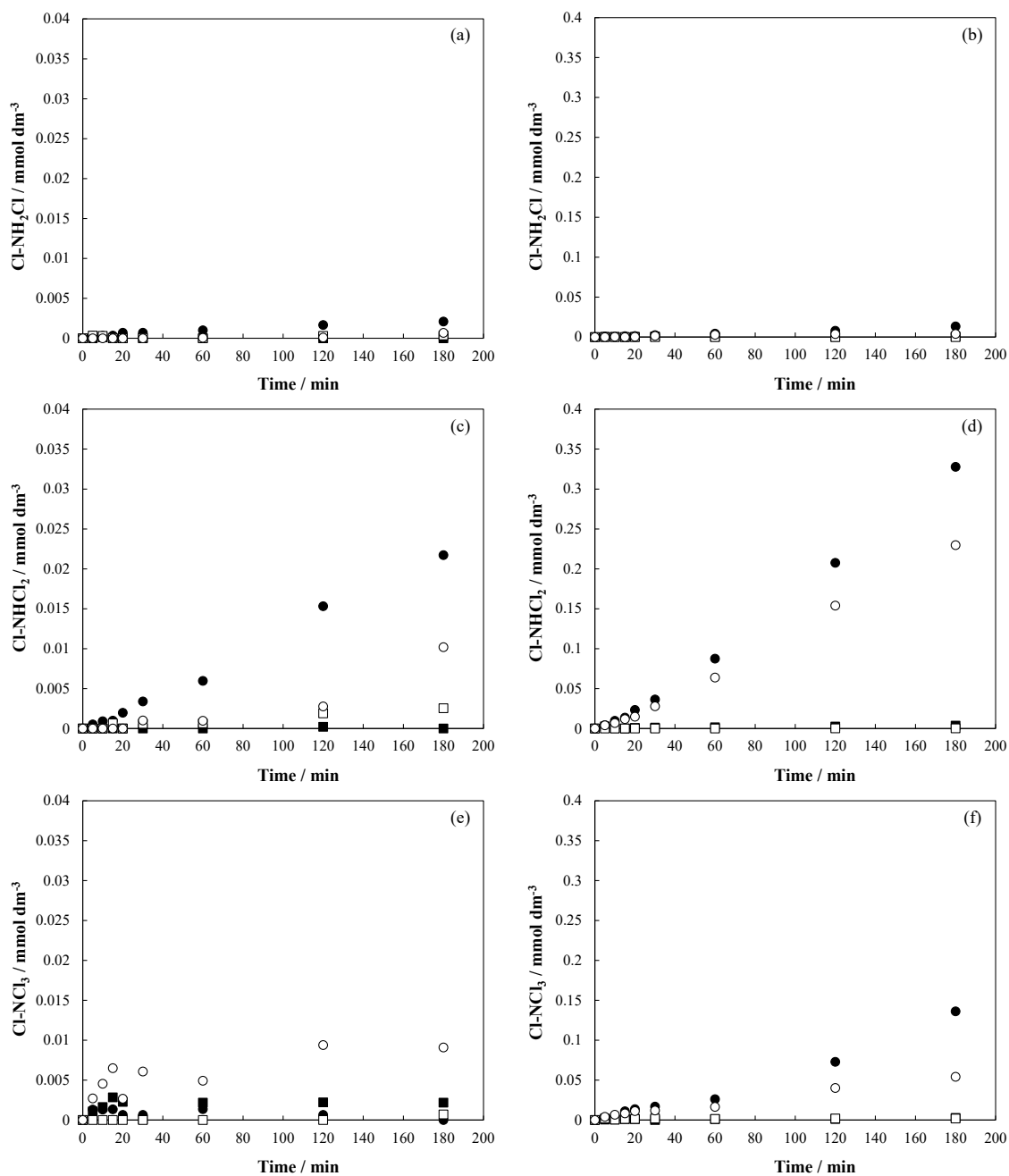


396 **Fig. 4.** Time course of the concentration of ammonium during the treatment of hospital
 397 urines. Black symbols: electrolysis; white symbols: photoelectrolysis; (■, □) 5 A m⁻²; (●,
 398 ○) 50 A m⁻²; (a) BDD anode; (b) MMO anode.

399 As can be observed, the concentration of ammonium remains constant at 5 A m⁻² for both
 400 anodes. This suggests that the organics removal rate is very low, and the chlorination of
 401 organics does not lead to the release of nitro groups but the formation of
 402 organochlorinated compounds. Likewise, this low current density does not favor the
 403 production of other oxidants that could be generated and contribute to the removal of
 404 organics contained in urine. On the contrary, ammonium concentration continuously
 405 increases with the operation time at 50 A m⁻², being higher when working with BDD
 406 anodes. These results suggest that the electrogenerated hypochlorite could also be
 407 consumed in the degradation of organics contained in the urine matrix together with other
 408 oxidants generated during electrolysis and photoelectrolysis under these conditions.
 409 However, the irradiation of UV light does not influence the removal of organics since
 410 similar results are observed in both processes, regardless the anode material tested. The
 411 use of MMO anodes seems to be less effective for the removal of organics than BDD.
 412 This is an unexpected behavior considering the previous disinfection data where BDD
 413 anode showed lower rates for killing bacteria. At this point, it is important to remark that

414 the removal of organics can also take place by direct oxidation over the anode surface and
415 hydroxyl radical-mediated oxidation [36] and these mechanisms seem to be more relevant
416 when using BDD anodes at 50 A m^{-2} . Hence, there could be a marked competitive reaction
417 between the disinfection and the oxidation of organics with this anode.

418 Nonetheless, ammonium released could also be consumed together with the
419 electrogenerated hypochlorite to form chloramines (Eqs. 9-11) during the disinfection of
420 urines. This could explain the low concentration registered with MMO anodes since the
421 electrogeneration of hypochlorite is also expected to be higher with this anode (Fig. S2a).
422 For this reason, the concentration of chloramines was also monitored during the
423 disinfection of urines by electrolysis and photoelectrolysis and, the results obtained as
424 function of the operation time for both anodes at different current densities are plotted in
425 Fig. 5.



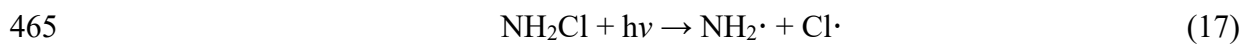
427 **Fig. 5.** Time course of the concentration of chloramines during the treatment of hospital
 428 urines. Black symbols: electrolysis; white symbols: photoelectrolysis; (■, □) 5 A m⁻²; (●,
 429 ○) 50 A m⁻²; (a) Monochloramine-BDD; (b) Monochloramine-MMO; (c) Dichloramine-
 430 BDD; (d) Dichloramine-MMO; (e) Trichloramine-BDD; (f) Trichloramine-MMO.

431 The concentration of chloramines increases with the operation time, being one order of
 432 magnitude higher when using MMO anodes. This can be due to the large amounts of

433 hypochlorite generated with this anode and, it also seems to confirm that the formation of
434 chloramines is the main way of consumption of hypochlorite with MMO anodes whereas
435 the use of BDD anodes promotes the oxidation of organics. Chloramines also contribute
436 to the disinfection of urine although their disinfectant capacity is lower than hypochlorite.
437 Likewise, the presence of chloramines minimizes the production of trihalomethanes
438 (THMs) and trihalogenated haloacetic acids as disinfection by-products [61, 62] and,
439 hence, decreases the accumulation of hazardous organochlorinated compounds in treated
440 effluents. The formation of chloramines follows a sequential reaction that starts with the
441 generation of monochloramine (Eq. 9). This species is the most reactive chloramine and
442 it has been widely used for the disinfection of wastewater [63, 64]. Then,
443 monochloramine can be continuous to react with hypochlorite, favoring the formation of
444 dichloramine (Eq. 10), and, finally, trichloramine is formed by the reaction between
445 dichloramine and hypochlorite (Eq. 11). The presence of one or another chloramine will
446 depend mainly on the ratio Cl/N and the pH of the effluent [65]. In this context, hospital
447 urines used in this work present a pH value around 6, and, according to literature, it favors
448 the presence of dichloramine. This agrees with the results obtained during electrolysis
449 and photoelectrolysis at 50 A m^{-2} where dichloramine is the predominant species followed
450 by trichloramine and, finally, monochloramine. At low current densities, the
451 concentration of chloramines is negligible owing to the low concentration of hypochlorite
452 electrogenerated.

453 Overall, the concentration of chloramines is lower during photoelectrolysis and it can be
454 related to the photoactivation of hypochlorite (Eq. 8) that decreases its free concentration
455 to form chloramines. Likewise, the low concentration of monochloramine (Figs. 5a, 5b)
456 registered for both anodes can be explained by the photoactivation of this species (Eq.
457 17) [66]. In this context, monochloramine can be decomposed in free amine and chlorine

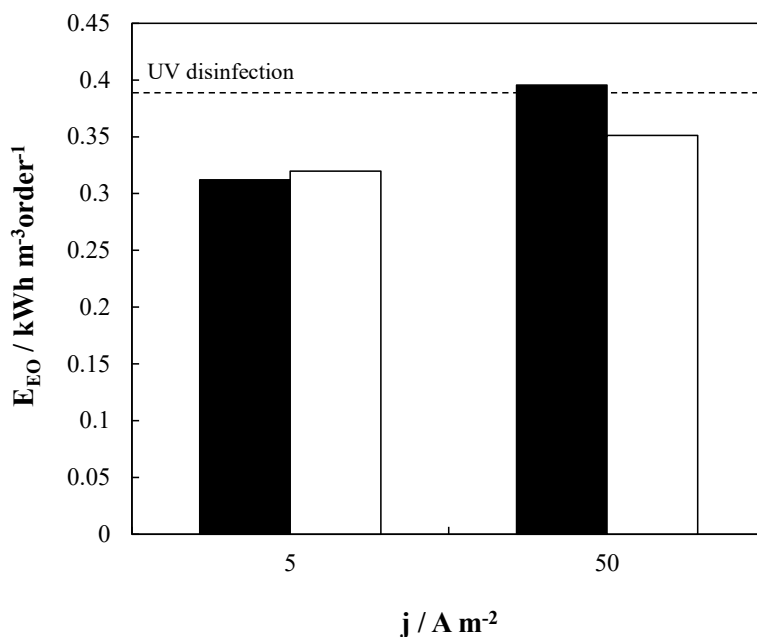
458 radicals by the irradiation of UV light. This also helps to remove ARB from urine since
 459 chlorine radical significantly contributes to the disinfection process. The higher the
 460 concentration of hypochlorite, the higher the concentration of chloramines and, hence,
 461 the higher the concentration of free chlorine radicals during photoelectrolysis. This
 462 supports the disinfection results from Fig. 1 because the concentration of hypochlorite is
 463 expected to be higher with MMO anodes and the removal rates are also higher with this
 464 anode.



466 Finally, to evaluate the energy requirements of the combined process in comparison with
 467 UV disinfection, the specific electric energy (E_{EO}) was calculated by Equations 18 and
 468 19, where E_{cell} is the cell voltage (V), I the current intensity (kA), t the operation time (h),
 469 V the volume (m^3), C_0 and C_f the initial and final bacteria concentration after 1 order of
 470 magnitude removal (CFU ml^{-1}), respectively and W_{lamp} the power of the lamp used (kW).
 471 This parameter determines the electric energy required to decrease the population of *K.*
 472 *pneumoniae* by one order of magnitude in a unit volume [67]. The results obtained are
 473 plotted in Fig. 6.

474
$$E_{\text{EO}} = \frac{E_{\text{cell}} \cdot I \cdot t}{V \cdot \text{Log} \frac{C_0}{C_f}} \quad (18)$$

475
$$E_{\text{EO}} = \frac{W_{\text{lamp}} \cdot t}{V \cdot \text{Log} \frac{C_0}{C_f}} \quad (19)$$



477 **Fig. 6.** Specific electric energy during UV disinfection and photoelectrolysis of hospital
 478 urines. Line: UV disinfection; black bars: BDD anode; white bars: MMO anode.

479 As can be observed, the lowest electric energy resulted in the combined process at low
 480 current densities using both BDD and MMO anodes. These results are in line with the
 481 synergistic effects found previously, where the highest values resulted when applying 5
 482 A m^{-2} . At 50 A m^{-2} , the electric energy for the process with BDD anodes is slightly higher
 483 than that required for the UV disinfection process (0.395 vs. $0.389 \text{ kWh m}^{-3} \text{ order}^{-1}$).
 484 Nevertheless, the use of MMO anodes at higher current densities leads to lower electric
 485 energy.

486 This reveals that electrolysis improves the efficiencies obtained during the UV
 487 disinfection process, decreasing, in turn, the energy requirements. This good performance
 488 of the combined process could be explained in terms of the promotion of free chlorine
 489 radicals. However, the use of BDD anodes is limited to their application at lower current
 490 densities, whereas MMO anodes can be used for all the current densities studied.

491 4. Conclusions

492 UV disinfection allows to decrease the population of ARB in synthetic hospital urine, but
493 it cannot reach complete disinfection. The irradiation of UV light to urine polluted with
494 *K. pneumoniae* could be limited by the occurrence of bacteria agglomeration or a resistant
495 subpopulation. On the other hand, electrochemical oxidation with BDD and MMO anodes
496 leads to complete removal of ARB from urine when applying 50 A m⁻². At 5 A m⁻²,
497 bacteria removal is negligible because the concentration of disinfectants produced is
498 rather low. The disinfection rate is higher when working with MMO anodes and it can be
499 related to competitive oxidation between bacteria and the organics contained in urine,
500 which seems to be favored using BDD anodes. Finally, photoelectrolysis enhances single
501 UV disinfection and electrolysis performances for all the current densities tested with
502 BDD and MMO anodes. A more remarkable contribution of UV disinfection is observed
503 at the beginning of the process whereas the electrochemical process has more influence
504 at the end of the treatment. A marked synergistic effect was found when UV disinfection
505 was enhanced by electrolysis at 5 A m⁻² with BDD and MMO anodes. The use of BDD
506 anodes at 50 A m⁻² led to antagonistic effects due to the possible competitive degradation
507 between bacteria and organics under these conditions. Regarding energy requirements for
508 single and combined processes, the specific electric energy required for photoelectrolysis
509 at low current densities is lower than that required for UV disinfection. At high current
510 densities, MMO shows lower electric energy whereas the use of BDD anodes leads to a
511 slightly higher value than UV disinfection.

512

513

514

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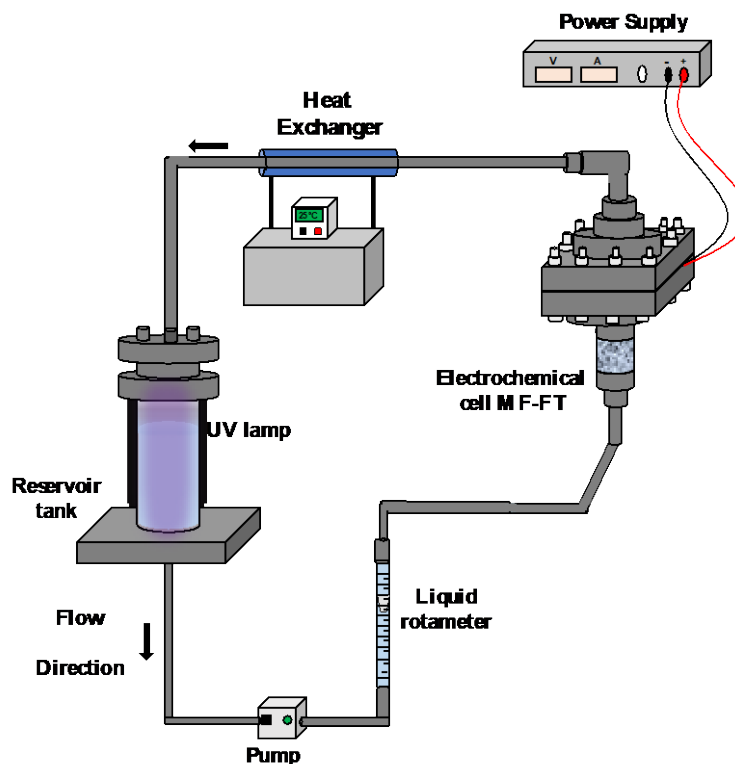
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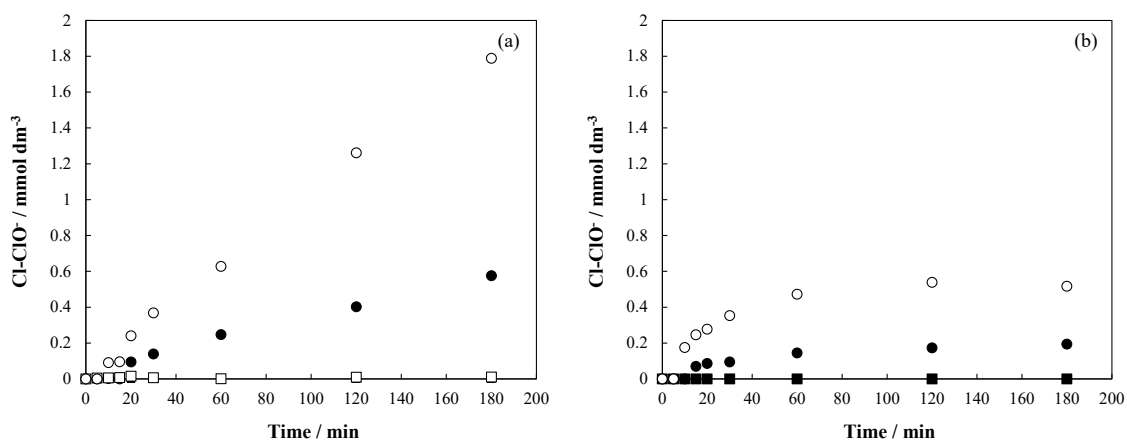
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Fig. S1. Experimental setup.

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730 **Fig. S2.** Time course of the concentration of hypochlorite during the electrolysis of 1000
 731 mg dm^{-3} KCl. Black symbols: BDD anode; white symbols: MMO anode; (■, □) 5 A m^{-2} ;
 732 (●, ○) 50 A m^{-2} ; (a) electrolysis; (b) photoelectrolysis.

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