



Draft Genome Sequence of a Novel “*Candidatus Liberibacter*” Species Detected in a *Zanthoxylum* Species from Bhutan

Grant A. Chambers,^a Nerida J. Donovan,^a Daniel R. Bogema,^a Namgay Om,^{b,c} George A. C. Beattie,^b Jennifer L. Morrow,^d Paul Holford^b

^aNSW Department of Primary Industries, Elizabeth Macarthur Agricultural Institute, Menangle, NSW, Australia

^bWestern Sydney University, School of Science, Penrith, NSW, Australia

^cNational Plant Protection Centre, Department of Agriculture, Ministry of Agriculture & Forests, Thimphu, Bhutan

^dWestern Sydney University, Hawkesbury Institute for the Environment, Penrith, NSW, Australia

ABSTRACT The draft genome sequence of a novel “*Candidatus Liberibacter*” species detected in an unidentified species of *Zanthoxylum* (Rutaceae) collected in Bhutan is reported. The total length is 1,408,989 bp with 1,169 coding sequences in 96 contigs, a GC content of 37.3%, and 76 to 77% average nucleotide identity with several other “*Ca. Liberibacter*” species.

Members of the genus *Liberibacter* are plant pathogens or nonpathogenic endophytes that are transmitted among their hosts by various psyllids. Because psyllids can harbor plant pathogens, a study of the microbiomes of psyllids associated with Rutaceae in Bhutan was performed using high-throughput sequencing (1). This revealed a novel “*Candidatus Liberibacter*” species in *Cornopsylla rotundiconis* and an uncharacterized species of *Cacopsylla*. Preliminary phylogenetic analysis revealed that the bacterium is closely related to “*Candidatus Liberibacter solanacearum*” and “*Candidatus Liberibacter caribbeanus*.” Here, we report the draft genome sequence of the “*Ca. Liberibacter*” isolate sampled in July 2014 from an uncharacterized *Zanthoxylum* species, a host of the psyllids, from Tsirang, Bhutan. The plant sampled exhibited blotchy mottled symptoms; however, the status of the bacterium as a pathogen or endophyte is unknown. “*Candidatus Liberibacter africanus*” was previously found in *Zanthoxylum* spp. (2).

Total DNA was extracted from petioles and midrib tissue using an Isolate II plant DNA kit (Bioline). The bacterial DNA extract was enriched using a NEBNext microbiome DNA enrichment kit (New England Biolabs) and submitted to the Ramaciotti Centre for Genomics (University of New South Wales [UNSW] Sydney, Australia) for library preparation and sequencing. A library was prepared using a Nextera DNA Flex library prep kit (Illumina), and the 2 × 150-bp paired-end library was sequenced using an Illumina NextSeq 500 platform. A total of 3.27 × 10⁸ reads with a mean length of 147 bp were generated.

The reads were analyzed with FastQC v0.11.8 and passed all relevant quality checks, were trimmed using BBDuk (3) using the parameters ktrim=r, k=23, mink=11, and hdist=1, and were assembled with SPAdes v3.13 (4) using k-mer sizes of 21, 41, 71, 101, and 127 and the --careful option. A total of 955,328 scaffolds were generated, with the largest being 122,841 bp long. Scaffolds were analyzed with BLASTn against the nucleotide database (downloaded 3 July 2020), and results were used in further analysis with BlobTools v1.1.1 (5). A BlobPlot based on the BLAST results, GC content, length, and average coverage depth was generated, and a clear cluster of scaffolds with GC content consistent with those of “*Ca. Liberibacter*” spp. (31.1 to 36.4%) and relatively high coverage depth was observed in a plot limited to the *Proteobacteria* phylum. Using this cluster as a guide, *Proteobacteria* and “no-hit” scaffolds with >20× coverage, a

Citation Chambers GA, Donovan NJ, Bogema DR, Om N, Beattie GAC, Morrow JL, Holford P. 2020. Draft genome sequence of a novel “*Candidatus Liberibacter*” species detected in a *Zanthoxylum* species from Bhutan. *Microbiol Resour Announc* 9:e00897-20. <https://doi.org/10.1128/MRA.00897-20>.

Editor Irene L. G. Newton, Indiana University, Bloomington

Copyright © 2020 Chambers et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Grant A. Chambers, grant.chambers@dpi.nsw.gov.au.

Received 3 August 2020

Accepted 8 September 2020

Published 1 October 2020

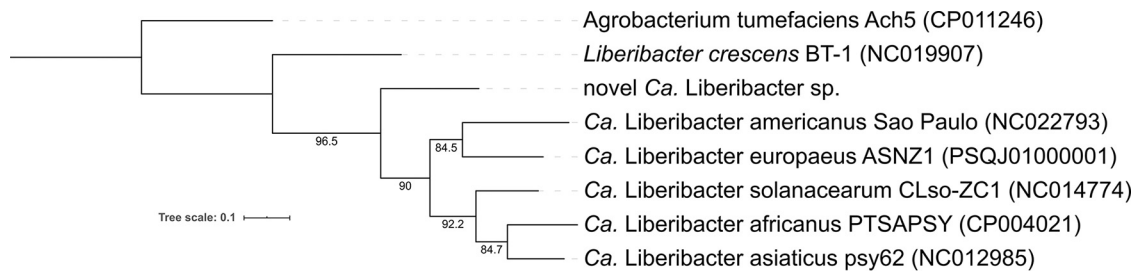


FIG 1 Species tree of *Liberibacter* spp. generated with 511 single-copy orthologs identified with OrthoFinder v2.4.0 (10) using default settings; individual protein sequences were aligned with MAFFT v7.471 (11) using the `--auto` argument. Phylogenetic trees and gene concordance factors (branch labels) were inferred with IQ-TREE v2.0.3 (12–15) with default settings. Branch support values were calculated using the ultrafast bootstrap method within IQ-TREE utilizing 1,000 replicates and showed 100% support for each branch within the tree.

>200-bp length, and 35 to 50% GC content were subsequently used for individual BLASTn nucleotide searches using standard parameters and limited to the proteobacterium database. A total of 96 contigs (N_{50} , 25,705 bp) showed similarity to *Liberibacter* species, with average GC contents of 37.3% and average coverage of $\sim 70.5\times$, forming the draft genome. The draft genome was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (6, 7). The total length of the assembly was 1,408,989 bp, and the genome was predicted to have 1,169 coding sequences and 47 RNA genes. Average nucleotide identity analysis was performed using FastANI v1.0 (8) against reference sequences of “*Ca. Liberibacter africanus*” (GenBank accession number CP004021), “*Candidatus Liberibacter americanus*” (NC022793), “*Candidatus Liberibacter asiaticus*” (NC012985), and “*Ca. Liberibacter solanacearum*” (CP002371) and determined identities of 76 to 77% to each. Analysis with BUSCO v4.1.2 (9), using the proteobacteria_obd10 data set with 219 BUSCOs and default settings, showed 88% complete, 4% fragmented, 7.7% missing, and 0% duplicated BUSCOs; this low level of duplication indicates that the reads have come from a single isolate. Phylogenetic analysis (Fig. 1) shows a very high level of gene concordance and provides strong evidence that a single isolate is present. The discovery of this novel “*Ca. Liberibacter*” species is relevant to the phylogeny and origins of liberibacters and their potential impacts.

Data availability. The sequence data have been deposited in DDBJ/ENA/GenBank under the accession number JACETX000000000. The version described in this paper is the first version, JACETX010000000. The raw reads are deposited in the NCBI Sequence Read Archive (SRA) under the BioProject PRJNA645749.

ACKNOWLEDGMENTS

N.O. was supported by a John Allwright fellowship awarded by the Australian Centre for International Agricultural Research. J.L.M. was supported by the Australian Research Council Industrial Transformation Training Centre (ARC-ITTC).

REFERENCES

- Morrow JL, Om N, Beattie GAC, Chambers GA, Donovan NJ, Liefting LW, Riegler M, Holford P. 2020. Characterization of the bacterial communities of psyllids associated with Rutaceae in Bhutan by high throughput sequencing. *BMC Microbiol* 20:215. <https://doi.org/10.1186/s12866-020-01895-4>.
- Roberts R, Steenkamp ET, Pietersen G. 2015. Three novel lineages of “*Candidatus Liberibacter africanus*” associated with native rutaceous hosts of *Trioxa erythraea* in South Africa. *Int J Syst Evol Microbiol* 65: 723–731. <https://doi.org/10.1099/ijs.0.069864-0>.
- Bushnell B. 2017. BMAP. <http://sourceforge.net/projects/bmap/>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Laetsch DR, Blaxter ML. 2017. BlobTools: interrogation of genome assemblies [version 1; peer review: 2 approved with reservations]. *F1000Res* 6:1287. <https://doi.org/10.12688/f1000research.12232.1>.
- Tatusova T, DiCuccio M, Badretdin A, Chetverin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Haft DH, DiCuccio M, Badretdin A, Brover V, Chetverin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. *Nucleic Acids Res* 46:851–860. <https://doi.org/10.1093/nar/gkx1068>.
- Jain C, Rodriguez RL, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>.

9. Seppey M, Manni M, Zdobnov EM. 2019. BUSCO: assessing genome assembly and annotation completeness. *Methods Mol Biol* 1962: 227–245. https://doi.org/10.1007/978-1-4939-9173-0_14.
10. Emms DM, Kelly S. 2019. OrthoFinder: phylogenetic orthology inference for comparative genomics. *Genome Biol* 20:238. <https://doi.org/10.1186/s13059-019-1832-y>.
11. Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol* 30:772–780. <https://doi.org/10.1093/molbev/mst010>.
12. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. 2015. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol* 32:268–274. <https://doi.org/10.1093/molbev/msu300>.
13. Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermini LS. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat Methods* 14:587–589. <https://doi.org/10.1038/nmeth.4285>.
14. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: improving the ultrafast bootstrap approximation. *Mol Biol Evol* 35:518–522. <https://doi.org/10.1093/molbev/msx281>.
15. Minh BQ, Hahn MW, Lanfear R. 2020. New methods to calculate concordance factors for phylogenomic datasets. *Mol Biol Evol* 37:2727–2733. <https://doi.org/10.1093/molbev/msaa106>.