



FACTORS INVOLVED IN IMMUNITY TO NEMATOSPIROIDES DUBIUS  
INFECTIONS IN MICE

by

Varunee Desakorn

B.Sc.Med.Tech. (Thailand)

M.P.H. (Philippines)

A thesis submitted for the degree of  
Master of Science

in

The Department of Microbiology and Immunology  
The University of Adelaide,  
Adelaide,  
South Australia

December, 1983

## ABSTRACT

The present investigation examined the interaction between mouse peritoneal macrophages with various developmental forms of Nematospiroides dubius (Heligmosomoides polygyrus) a natural nematode parasite of the mouse and the ability of serum from mice immune to this infection to damage exsheathed L<sub>3</sub> in the absence of phagocytic cells.

Initial studies examined the adherence of peritoneal exudate cells to the surface of the parasite and the factors involved in promoting this adhesion. Peritoneal exudate cells from normal mice did not adhere to the cuticle of exsheathed L<sub>3</sub> in the absence of serum, but did so when the larvae had been sensitised with normal mouse serum. In this case the cells bound to the larvae via their C3 receptors and confirmed previous observations that the parasite activated complement via the alternative pathway. It was also found that serum from mice immune to this infection contained antibodies directed against the cuticle of all larval stages as measured by the adhesion of cells to larvae that had been sensitised with heat inactivated serum. The cells in this instance adhered to the larvae via their Fc receptors. The factors involved in promoting adhesion of peritoneal exudate cells from normal mice and those from mice resistant to this infection were similar. The peritoneal exudate cells adhering to the larvae were found to be predominantly macrophages.

The cell adhesion studies demonstrated that all of the developing post-infective and the adult form of the parasite were capable of activating complement by the alternative pathway with the marked exception of larvae collected 96 hours post-infection. This

indicated a possible change in the antigenic structure of the cuticle.

Experiments were carried out to determine the ability of various immunoglobulin isotypes from mouse immune to reinfection to mediate cell binding to all developmental forms and adult worms of the parasite. It was found that the ability of IgM to promote the binding of cells to the larval stages increased dramatically from the pre-infective stages including sheathed and exsheathed L<sub>3</sub> up to 96 hours post-infective larvae, but promoted little binding to adult worms. The IgG<sub>1</sub> promoted cell binding in a similar pattern to that of IgM, but binding of the cells to 96 hours post-infective larvae was considerably reduced. Both IgG<sub>2a</sub> and IgG<sub>2b</sub> enhanced the binding of cells to pre-infective larvae, but were not active in promoting cell binding to post-infective larvae. However at a physiological level only IgG<sub>1</sub> and IgM were active in promoting cell adhesion.

An in vitro assay to determine the effect of cell binding upon the infectivity of exsheathed L<sub>3</sub> indicated that peritoneal exudate cells from mice infected with 2 doses of L<sub>3</sub> were capable of damaging the larvae (as measured by a loss in infectivity) providing the larvae had been sensitised with antibody and complement or complement alone, but had no effect if the larvae had been sensitised with antibody alone even though antibody mediated cell adherence. Peritoneal exudate cells from normal mice were unable to damage the larvae even in the presence of antibody and complement.

Further in vitro studies showed that fresh untreated serum from mice infected with 4 doses of L<sub>3</sub>, IMS(4) was able to damage exsheathed L<sub>3</sub> as measured by a reduction in their infectivity.

This effect was not apparent when fresh untreated serum from mice given 2 immunising doses, IMS(2) was used. The larvicidal activity was found only in long-term N. dubius infected mice since serum from mice infected with Mesocostoides corti an unrelated helminth parasite did not reduce the infectivity of the larvae. The larvicidal activity of IMS(4) appeared to be dependent on specific antibodies of the IgM class reacting with the cuticle of the larvae and binding complement.

Studies in vivo showed that fresh IMS(4) given intravenously to naive mice protected them against a subsequent intravenous challenge with exsheathed L<sub>3</sub>, but not against a challenge with sheathed L<sub>3</sub>. It was also found that serum from immune mice from which the greater proportion of the immunoglobulins had been removed was unable to transfer immunity passively to naive mice. The relative importance of these studies to other investigations dealing with nematode parasites is discussed.

STATEMENT

This thesis contains no material previously submitted by me for a degree in any university, and to the best of my knowledge and belief it contains no material previously published or written by another person, except where reference is made in the text.

Varunee Desakorn

December, 1983

ACKNOWLEDGEMENTS

I am deeply indebted to Dr. C.R. Jenkin for his supervision and encouragement throughout the course of this investigation.

I am most grateful to Dr. P.L. Ey for his understanding, advice criticism and enthusiasm, and to the other members of the Department of Microbiology and Immunology for their comments of this work.

Thanks are due to Mrs. A. Hallett for the culture of parasite larvae, to Mr. N. Shead for his help with the preparation of graphs and photographs, and to Mrs. J. Fallon for the typing of this thesis. I should also acknowledge the receipt of an Australian Government Award for SEAMEO centre staff development programme.

Finally, I would like to thank my parents and friends for their kindness and moral support.

ABBREVIATIONS USED IN THIS THESIS

BSA	Bovine serum albumin
c.a.	approximately
EDTA	ethylenediaminetetraacetate
EGTA	ethyleneglycol-bis ( $\beta$ -aminoethyl) N,N'-tetraacetate
ELISA	Enzyme-linked immuno-sorbent assay
Ig	Immunoglobulin
IMS	immune mouse serum
IMS(2)	serum from 2 times <u>N. dubius</u> infected mice
IMS(4)	serum from 4 times <u>N. dubius</u> infected mice
L <sub>3</sub>	Third-stage (infective) <u>N. dubius</u> larvae
N.D.	not done
NMS	normal mouse serum
PBS	Phosphate-buffered saline
tris	Tris(hydroxymethyl)aminomethane
VBS	Ca <sup>++</sup> , Mg <sup>++</sup> - supplemented veronal-buffered saline

TABLE OF CONTENTS

	<u>page</u>
Title	i
Abstract	ii
Statement	v
Acknowledgements	vi
Abbreviations used in this thesis	vii
Table of Contents	viii
 <u>Chapter 1 : Introduction</u>	
1.1. General Introduction	1
1.2. Vaccination of hosts against nematode infection	2
1.3. Mechanisms of immunity to <u>Nippostrongylus brasiliensis</u>	4
1.4. Mechanisms of immunity to <u>Trichinella spiralis</u>	14
1.5. <u>Nematospiroides dubius</u> a natural nematode parasite of the mouse	22
1.5.1. Life Cycle	22
1.5.2. Pathology of the infection	23
1.5.3. Induction of immunity in mice to infection with <u>N. dubius</u>	25
1.5.4. Changes in immunoglobulin levels during infection	29
1.5.5. Changes in cell levels during infection	31
1.5.6. The role of phagocytic cells and antibody in immunity to <u>N. dubius</u> infection	32
 <u>Chapter 2 : Materials and Methods</u>	
2.1. Incubation media and diluents	35
2.1.1. Incubation media	35
2.1.2. Diluents	35
2.2. Mice	35
2.3. Maintenance of the parasite <u>Nematospiroides dubius</u> ( <u>Heligmosomoides polygyrus</u> )	36



2.4.	Collection of exsheathed larvae of <u>N. dubius</u>	37
2.5.	Collection of post-infective larvae	37
2.6.	Method for immunising mice against infection with <u>N. dubius</u>	39
2.7.	Assay of resistance to infection with <u>N. dubius</u>	39
2.8.	Collection of serum	39
2.9.	Inactivation of complement by methylamine	40
2.10.	Collection of peritoneal exudate cells	40
2.11.	Cell counting and viability	40
2.12.	Preparation of siliconised glass tubes	41
2.13.	Treatment of mice with <u>Salmonella enteritidis</u> 11 RX for the collection of activated peritoneal exudate cells	41
2.14.	Measurement of hemolytic complement activity	42
2.15.	Concentration of serum or immunoglobulin fractions by ultrafiltration	42
2.16.	Ammonium sulphate precipitation of immunoglobulins	42
2.17.	Preparation of purified mouse immunoglobulins	43
2.17.1.	Sephadex G-200 chromatography	43
2.17.2	Protein A-Sepharose chromatography	43
2.18.	Preparation of purified mouse IgG and IgM immunoglobulins	44
2.19.	Radial immunodiffusion analyses (Ouchterlony test)	44
2.20.	Enzyme linked immunosorbent assays (ELISA)	45
2.21.	Adsorption of immunoglobulin from immune mouse serum by affinity chromatography	46
2.22.	Adsorption of complement component C3 from mouse serum by affinity chromatography	47
2.23.	Statistics	47

Chapter 3 : Factors involved in the adherence of peritoneal exudate cells to the larval and adult stages of N. dubius

3.1.	Introduction	48
3.2.	Method	
	Assay for the adherence of peritoneal exudate cells to different developmental forms of <u>N. dubius</u>	48
3.3.	Results	49
3.3.1.	Factors involved in the adherence of peritoneal exudate cells from normal mice to exsheathed L <sub>3</sub>	49
3.3.2.	The ability of serum from normal mice or mice resistant to re-infection to promote the adherence of normal peritoneal exudate cells to different developmental forms of <u>N. dubius</u>	51

Chapter 4 : The ability of different immunoglobulin isotypes to promote the adhesion of normal peritoneal exudate cells to N. dubius

4.1.	Introduction	54
4.2.	Results	54
4.2.1.	The efficacy of different immunoglobulin isotypes in promoting the adherence of normal peritoneal exudate cells to exsheathed L <sub>3</sub>	54
4.2.2.	The ability of different immunoglobulin isotypes to promote the adherence of normal peritoneal exudate cells to different developmental forms of <u>N. dubius</u>	55
4.2.3.	Comparison between the ability of killed L <sub>3</sub> and living L <sub>3</sub> to provide the production of antibodies enhancing adhesion of normal peritoneal exudate cells to exsheathed L <sub>3</sub>	56
4.2.4.	The inability of killed larvae to protect mice against infection with <u>N. dubius</u>	58

Chapter 5 : The effect in vitro of peritoneal exudate cells  
 from immune mice on the infectivity of  $L_3$  of  
*N. dubius* sensitised with either specific antibody  
 and/or complement

- 5.1. Introduction 59
- 5.2. Method  
In vitro assay for the effect of peritoneal exudate  
 cells on the infectivity of exsheathed  $L_3$  59
- 5.3. Results
- 5.3.1. The effect of incubating peritoneal exudate cells  
 from resistant mice for different periods of time  
 with exsheathed  $L_3$  on their infectivity after  
 sensitisation with IMS(2) 60
- 5.3.2. The effect in vitro of peritoneal exudate cells  
 from mice given 2 immunising doses on the  
 infectivity of exsheathed  $L_3$  sensitised with  
 NMS or IMS(2) treated or untreated at  $56^{\circ}$  61
- 5.3.3. Comparison between the effect in vitro of  
 peritoneal exudate cells from mice given 2  
 immunising doses of  $L_3$  and cells from mice  
 given 4 immunising doses of  $L_3$  on the infectivity  
 of exsheathed  $L_3$  sensitised with either serum  
 from mice given 2 immunising doses of  $L_3$ , IMS(2);  
 or serum from mice given 4 immunising doses of  
 $L_3$ , IMS(4) 63
- 5.3.4. The effect in vitro of peritoneal exudate cells  
 from mice given living *Salmonella enteritidis* 11 RX  
 on the infectivity of exsheathed  $L_3$  65
- 5.4. Conclusion 67

Chapter 6 : The effect of serum on the infectivity of  $L_3$   
 following incubation in vitro

- 6.1. Introduction 68

6.2.	Method	
	<u>In vitro</u> assay for the effect of a cell-free serum system on the infectivity of exsheathed L <sub>3</sub>	68
6.3.	Results	69
6.3.1.	The optimal incubation time required for serum from mice infected with 4 immunising doses to damage L <sub>3</sub>	69
6.3.2.	Length of time required for IMS(4) to impair the infectivity of L <sub>3</sub>	70
6.3.3.	The larvicidal activity of IMS(4) on varying numbers of exsheathed L <sub>3</sub>	71
6.3.4.	The effect of the further addition of IMS(4) during the course of incubation on the infectivity of exsheathed L <sub>3</sub>	72
6.3.5.	Titration of the larvicidal activity of serum taken at various times during the course of infection from mice infected with varying number of doses of 200 <u>N. dubius</u> L <sub>3</sub>	73
6.3.6.	Comparison between the effects of serum from mice infected with 4 doses of <u>N. dubius</u> L <sub>3</sub> and serum from mice infected with <u>Mesocestoides corti</u> on the infectivity of exsheathed L <sub>3</sub>	74
6.3.7.	The effect of storing IMS(4) on ice for varying periods of time on its larvicidal activity	76
6.3.8.	The effect of storing IMS(4) at -20° on its ability to damage L <sub>3</sub> <u>in vitro</u>	77
6.3.9.	The effect of heat treatment at different temperatures on the larvicidal activity of IMS(4)	77
6.3.10.	The effect of dialysis on the larvicidal activity of IMS(4)	78
6.3.11.	The requirement for complement in mediating damage to exsheathed L <sub>3</sub> <u>in vitro</u> by serum from mice infected with 4 immunising doses of <u>N. dubius</u> L <sub>3</sub>	79

6.3.12.	The ability of IMS(4) after removal of the immunoglobulins to damage exsheathed $L_3$	80
6.4.	Conclusions	81
<u>Chapter 7 : Protection afforded to normal mice to <i>N. dubius</i></u>		
<u>infection following the passive transfer of serum from mice resistant to re-infection</u>		
7.1.	Introduction	84
7.2.	Results	84
7.2.1.	Passive transfer of immunity to naive mice given IMS(4) intraperitoneally	84
7.2.2.	Passive transfer of immunity to naive mice by the intravenous injection of IMS(4)	85
7.2.3.	Passive transfer of immunity to naive mice by the intravenous injection of IMS(4) followed by a challenge infection of either exsheathed $L_3$ or sheathed $L_3$	85
7.2.4.	The ability of various dilutions of IMS(4) to transfer immunity passively to naive mice	86
7.2.5.	The effect of freezing and thawing on the ability of IMS(4) to transfer protection to naive mice	87
7.2.6.	Comparison between the <u>in vivo</u> and <u>in vitro</u> larvicidal effect of IMS(4) after chromatography on either an anti-mouse C3 or an anti-mouse $F(ab')_2$ column	88
7.2.7.	Further evidence that complement is involved in the larvicidal activity of IMS(4)	90
7.2.8.	The effect of an immunoglobulin preparation or an IgG and IgM fraction prepared from IMS(4) on the infectivity of exsheathed $L_3$ in the presence and absence of NMS	92
7.2.9.	The effect of purified IgM prepared from IMS(4) on the infectivity of exsheathed $L_3$ in the presence of NMS	94

7.3. Conclusions	96
<u>Chapter 8 : Discussion</u>	97
Bibliography	111