

PUBLISHED

Sonam Wangchuk, Sonam Pelden, Tenzin Dorji, Sangay Tenzin, Binay Thapa, Sangay Zangmo, Ratna Gurung, Kinzang Dukpa, Tenzin Tenzin

Crimean-congo hemorrhagic fever virus IgG in goats, Bhutan

Emerging Infectious Diseases, 2016; 22(5):919-920

Emerging Infectious Diseases is an open access journal in the public domain. All content is freely available without charge to the user or his/her institution.

Originally published at: <http://doi.org/10.3201/eid2205.151777>

PERMISSIONS

<https://wwwnc.cdc.gov/eid/page/copyright-and-disclaimers>

Open Access and Usage

Emerging Infectious Diseases is an open access journal in the public domain. All content is freely available without charge to the user or his/her institution. In accordance with the [Budapest Open Access Initiative](#) definition of Open Access, users are allowed to read, download, copy, distribute, print, search, or link to the full texts of the articles, or use them for any other lawful purpose, without asking prior permission from the publisher or the author. Because the journal is in the public domain, its usage policy also conforms to conditions set for by [Creative Commons](#).

Emerging Infectious Diseases does request a proper citation be included for its content and that any user indicate clearly if changes have been made.

Emerging Infectious Diseases grants authors permission to self-archive their articles without fees or permission and grants institutions permission to preserve a second copy of articles published by their researchers in the institutional repository.

16 April 2019

<http://hdl.handle.net/2440/118562>

can be mistaken for signs of tuberculosis. Considering the poor hygienic conditions among refugees, LBRF has become an important differential diagnosis in Europe in times of increasing migration.

Acknowledgments

We thank the technician teams of the Institutes of Virology and Laboratory Medicine at the University Hospital Marburg for excellent technical assistance.

References

1. Parola P, Diatta G, Socolovschi C, Mediannikov O, Tall A, Bassene H, et al. Tick-borne relapsing fever borreliosis, rural Senegal. *Emerg Infect Dis*. 2011;17:883–5.
2. Elbir H, Raoult D, Drancourt M. Relapsing fever borreliae in Africa. *Am J Trop Med Hyg*. 2013;89:288–92. <http://dx.doi.org/10.4269/ajtmh.12-0691>
3. Wilting KR, Stienstra Y, Sinha B, Braks M, Cornish D, Grundmann H. Louse-borne relapsing fever (*Borrelia recurrentis*) in asylum seekers from Eritrea, the Netherlands, July 2015. *Euro Surveill*. 2015;20:pii: 21196. <http://dx.doi.org/10.2807/1560-7917.ES2015.20.30.21196>
4. Lucchini A, Lipani F, Costa C, Scarvaglieri M, Balbiano R, Carosella S, et al. Louseborne relapsing fever among East African refugees, Italy, 2015. *Emerg Infect Dis*. 2016;22:298–301. <http://dx.doi.org/10.3201/eid2202.151768>
5. Naddaf SR, Ghazinezhad B, Bahramali G, Cutler SJ. Phylogenetic analysis of the spirochete *Borrelia microti*, a potential agent of relapsing fever in Iran. *J Clin Microbiol*. 2012;50:2873–6. <http://dx.doi.org/10.1128/JCM.00801-12>
6. Schwan TG, Battisti JM, Porcella SF, Raffel SJ, Schrupf ME, Fischer ER, et al. Glycerol-3-phosphate acquisition in spirochetes: distribution and biological activity of glycerophosphodiester phosphodiesterase (GlpQ) among *Borrelia* species. *J Bacteriol*. 2003;185:1346–56. <http://dx.doi.org/10.1128/JB.185.4.1346-1356.2003>
7. Lescot M, Audic S, Robert C, Nguyen TT, Blanc G, Cutler SJ, et al. The genome of *Borrelia recurrentis*, the agent of deadly louse-borne relapsing fever, is a degraded subset of tick-borne *Borrelia duttonii*. *PLoS Genet*. 2008;4:e1000185. <http://dx.doi.org/10.1371/journal.pgen.1000185>
8. Cadavid D, Barbour AG. Neuroborreliosis during relapsing fever: review of the clinical manifestations, pathology, and treatment of infections in humans and experimental animals. *Clin Infect Dis*. 1998;26:151–64. <http://dx.doi.org/10.1086/516276>
9. Guerrier G, Doherty T. Comparison of antibiotic regimens for treating louse-borne relapsing fever: a meta-analysis. *Trans R Soc Trop Med Hyg*. 2011;105:483–90. <http://dx.doi.org/10.1016/j.trstmh.2011.04.004>
10. Bryceson AD, Parry EH, Perine PL, Warrell DA, Vukotich D, Leithead CS. Louse-borne relapsing fever. *Q J Med*. 1970;39:129–70.

Address for correspondence: Christian Keller, Institute of Virology, University Hospital Marburg, Hans-Meerwein-Str 2, 35043 Marburg, Germany; email: christian.keller@staff.uni-marburg.de

Crimean-Congo Hemorrhagic Fever Virus IgG in Goats, Bhutan

Sonam Wangchuk, Sonam Pelden, Tenzin Dorji, Sangay Tenzin, Binay Thapa, Sangay Zangmo, Ratna Gurung, Kinzang Dukpa, Tenzin Tenzin

Author affiliations: Ministry of Health Public Health Laboratory, Thimphu, Bhutan (S. Wangchuk, S. Pelden, T. Dorji, B. Thapa, S. Zangmo); Ministry of Agriculture and Forests National Centre for Animal Health, Thimphu (S. Tenzin, R. Gurung, K. Dukpa, T. Tenzin)

DOI: <http://dx.doi.org/10.3201/eid2205.151777>

To The Editor: Crimean-Congo hemorrhagic fever (CCHF) is a highly infectious tickborne disease caused by a high-risk group of viruses belonging to the family *Bunyaviridae* (1,2). In humans, the overall case-fatality rate of CCHF is ≈30%, but in severe and hospitalized patients, fatalities may be up to 80% (3,4). CCHF is widespread in various countries in Africa, Asia, and Europe; the virus had been identified in humans in China, Pakistan, and Afghanistan and has been recently reported for the first time in humans in India (4–7). Humans can be infected by bites from infected ticks, mainly of the *Hyalomma* genus; by unprotected contact with blood or tissue of viremic patients; or during slaughtering of infected animals. In addition, nosocomial infections are found in humans (1,4,8,9).

Fatal cases of CCHF in humans were confirmed in Ahmadabad in India in 2011, but a recent serosurvey in livestock showed that this disease has widespread seroprevalence in domestic animals across India (7–10). Bhutan shares a long, porous border with India, and animals and humans frequently cross the border. Comprehensive surveillance was needed to determine the presence of CCHF virus (CCHFV) in livestock in Bhutan and to assess risk for zoonotic infection in humans.

During October 2015, in collaboration with the National Centre for Animal Health Bhutan, we retrospectively tested serum samples collected during April–May 2015 from 81 goats and 92 cattle for CCHFV-specific IgG by using ELISA kits (Sheep/goat anti-CCHFV IgG ELISA kit and Cattle anti-CCHFV IgG ELISA kit; National Institute of Virology, Pune, India), as described (10). CCHFV IgG was detected in 31 (38.2%) goats; no cattle had positive results. The samples from goats, which were collected in early 2015 as part of surveillance of peste des petits ruminants, originated from the southern district of Sarpang, which shares a porous border with the state of Assam in India (Figure). The samples from cattle were collected



Figure. Locations in Bhutan where serum samples were collected from goats (triangles) and cattle (square and circle) and tested for Crimean-Congo hemorrhagic fever virus. The shaded area shows the boundaries of Sarpang district and subdistricts, where samples from goats were collected.

from the National Nublang Breeding Center (Trashigang district) and the National Jersey Breeding Center (Samtse district) (Figure). Findings indicated that all goats that tested positive for CCHFV were reported to have been either bred within households that kept goat herds or procured from other villages within the district. Exact sources of those seropositive goats could not be ascertained. However, in a few instances in the past, breeding goats (male and female) were procured from India by the Bhutan government and distributed to farmers for breed improvement. We also believe that cross-border movement of animals and unofficial imports of goats by farmers along the porous borders of southern Bhutan likely occurred. Furthermore, a large number of dairy cattle are imported annually from India for enhancing milk production and breeding purposes. Not all imported animals (both cattle and goats) were tested for CCHF because of a lack of diagnostic facilities and the negligible occurrence of the disease in livestock.

Our findings indicate that the risk of importing emerging infectious diseases along with live animals poses a serious risk to public health. Consequently, detailed risk-based surveillance is necessary to understand the complete scenario of CCHFV prevalence in livestock in Bhutan because *Hyalomma* tick species, the primary vectors of CCHF, are present on animals here. In addition, a survey among at-risk human populations is also needed. Findings from these surveillance activities would help institute more diagnostic facilities and risk-based surveillance and assist in developing a preparedness plan at the human-animal interface. Although our study has limitations because of the low number of serum samples tested from limited animal species from only 3 areas, the study provides evidence that CCHFV is circulating in goats in Bhutan.

Acknowledgments

We gratefully acknowledge the encouragement and support extended by the Secretary of the Ministry of Health in Bhutan. We also extend our thanks for the support that the World Health Organization Country Office in Bhutan provided to carry out this study. We further acknowledge the director and technical experts at the National Institute of Virology, Pune, India, for their technical assistance and guidance for this study.

Mr. Wangchuk is Laboratory Chief of the Public Health Laboratory, Ministry of Health, Thimphu, Bhutan. His main research interest is infectious and zoonotic diseases.

References

- Ergönül O. Crimean-Congo hemorrhagic fever. *Lancet Infect Dis*. 2006;6:203–14. [http://dx.doi.org/10.1016/S1473-3099\(06\)70435-2](http://dx.doi.org/10.1016/S1473-3099(06)70435-2)
- Bente DA, Forrester NL, Watts DM, McAuley AJ, Whitehouse CA, Bray M. Crimean-Congo hemorrhagic fever: history, epidemiology, pathogenesis, clinical syndrome and genetic diversity. *Antiviral Res*. 2013;100:159–89. <http://dx.doi.org/10.1016/j.antiviral.2013.07.006>
- Whitehouse CA. Crimean-Congo hemorrhagic fever. *Antiviral Res*. 2004;64:145–60. [http://dx.doi.org/10.1016/S0166-3542\(04\)00163-9](http://dx.doi.org/10.1016/S0166-3542(04)00163-9)
- Hoogstraal H. The epidemiology of tick-borne Crimean-Congo hemorrhagic fever in Asia, Europe, and Africa. *J Med Entomol*. 1979;15:307–417. <http://dx.doi.org/10.1093/jmedent/15.4.307>
- Sargianou M, Papa A. Epidemiological and behavioral factors associated with Crimean-Congo hemorrhagic fever virus infections in humans. *Expert Rev Anti Infect Ther*. 2013;11:897–908. <http://dx.doi.org/10.1586/14787210.2013.827890>
- Ince Y, Yasa C, Metin M, Sonmez M, Meram E, Benkli B, et al. Crimean-Congo hemorrhagic fever infections reported by ProMED. *Int J Infect Dis*. 2014;26:44–6. <http://dx.doi.org/10.1016/j.ijid.2014.04.005>
- Mourya DT, Yadav PD, Patil DY, Bhatia R. Highly infectious tick-borne viral diseases: Kyasanur forest disease and Crimean Congo hemorrhagic fever in India. *WHO South East Asia J Public Health*. 2014;3:8–21. <http://dx.doi.org/10.4103/2224-3151.115828>
- Yadav PD, Raut CG, Patil DY, Majumdar TD, Mourya DT. Crimean-Congo hemorrhagic fever: current scenario in India. *Proc Indian Natl Sci Acad B Biol Sci*. 2014;84:9–18. <http://dx.doi.org/10.1007/s40011-013-0197-3>
- Mourya DT, Yadav PD, Shete AM, Gurav YK, Raut CG, Jadi RS, et al. Detection, isolation, and confirmation of Crimean-Congo hemorrhagic fever virus in human, ticks and animals in Ahmadabad, India, 2010–2011. *PLoS Negl Trop Dis*. 2012;6:e1653.
- Mourya DT, Yadav PD, Shete AM, Sathe PS, Sarkale PC, Pattnaik B, et al. Cross-sectional serosurvey of Crimean Congo Hemorrhagic fever virus IgG antibody in domestic animals in India. *Emerg Infect Dis*. 2015;21:1837–9. <http://dx.doi.org/10.3201/eid2110.141961>

Address for correspondence: Sonam Wangchuk, Laboratory Chief, Public Health Laboratory, Thimphu, Bhutan; email: swangchuk@health.gov.bt