



Detection of SARS-CoV-2 Infection in Gargle, Spit, and Sputum Specimens

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ABSTRACT The gold standard for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection diagnosis is reverse transcription (RT)-PCR from a nasopharyngeal swab specimen (NPS). Its collection involves close contact between patients and health care workers, requiring a significant amount of workforce and putting them at risk of infection. We evaluated self-collection of alternative specimens and compared their sensitivity and cycle threshold (C_T) values to those of NPS. We visited acute coronavirus disease 2019 (COVID-19) outpatients to collect concomitant NPS and gargle specimens and had patients self-collect gargle and either sputum or spit specimens the next morning. We included 40 patients and collected 40 concomitant NPS and gargle specimens, as well as 40 gargle, 22 spit, and 16 sputum specimens the next day (2 patients could not produce sputum). All specimens were as sensitive as NPS. Gargle specimens had a sensitivity of 0.97 (95% confidence interval [CI], 0.92 to 1.00), whether collected concomitantly with NPS or the next morning. Next-morning spit and sputum specimens showed sensitivities of 1.00 (95% CI, 1.00 to 1.00) and 0.94 (95% CI, 0.87 to 1.00), respectively. The gargle specimens had significantly higher mean C_T values of 29.89 (standard deviation [SD], 4.63; $P < 0.001$) and 29.25 (SD, 3.99; $P < 0.001$) when collected concomitantly and the next morning, respectively, compared to NPS (22.07 [SD, 4.63]). C_T values obtained with spit (23.51 [SD, 4.57]; $P = 0.11$) and sputum (25.82 [SD, 9.21]; $P = 0.28$) specimens were close to those of NPS. All alternative specimen collection methods were as sensitive as NPS, but spit collection appeared more promising, with a low C_T value and ease of collection. Our findings warrant further investigation.

IMPORTANCE Control of the COVID-19 pandemic relies heavily on a test-trace-isolate strategy. The most commonly used specimen for diagnosis of SARS-CoV-2 infection is a nasopharyngeal swab. However, this method is quite uncomfortable for the patient, requires specific equipment (nose swabs and containers), and requires close proximity to health care workers, putting them at risk of infection. Developing alternative sampling strategies could decrease the burden for health care workers, help overcome potential shortages of equipment, and improve acceptability of testing by reducing patient discomfort.

KEYWORDS alternative testing methods, COVID-19, gargle, nasopharyngeal swab, SARS-CoV-2, spit, sputum


Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection control relies on a test-trace-isolate strategy with early diagnosis and isolation of infected individuals followed by identification of contacts (1). It has led to an initial shortage of

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 Could spit be used to decrease healthcare workers exposure to SARS-CoV-2 and improve testing capacities: Findings from a Finnish exploratory study conducted by @hennamailis and @timodub at @THLorg

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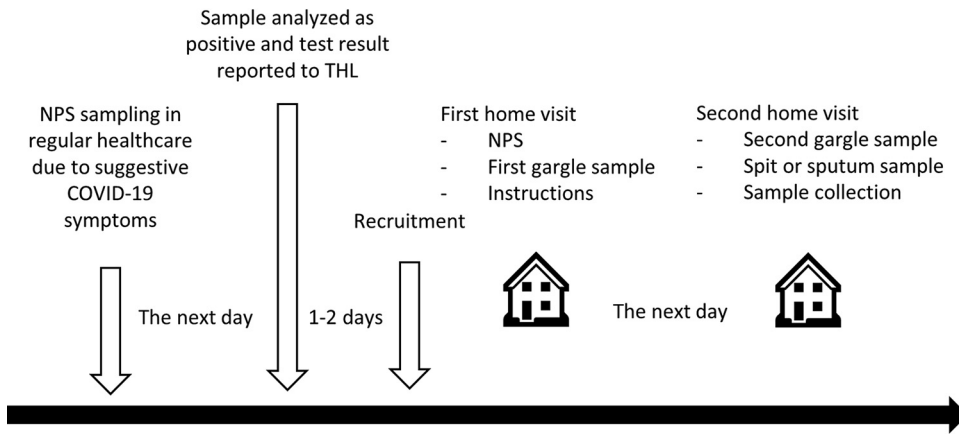


FIG 1 Timeline of the study.

personal protective and sampling equipment, as well as increasing the workload of health care workers (HCWs) (2, 3). In Finland alone (approximately 5,500,000 inhabitants), by 20 March 2021 over 3,700,000 tests had been conducted and analyzed nationwide since the start of the coronavirus disease 2019 (COVID-19) pandemic (4, 5), including 145,000 tests in the first week of March 2021.

Collection of a nasopharyngeal swab specimen (NPS) is the gold standard for SARS-CoV-2 infection diagnosis (6). It is an unpleasant procedure, requiring close contact with a HCW and increasing the HCW's risk of infection (2, 7). The use of an alternative specimen collection method could increase testing capacities, as well as decreasing HCWs' workload and risk of infection (7). There have been several studies evaluating alternative specimen collection methods, but none of the specimen collection methods has yet superseded NPS collection (3, 8–10), even though one Finnish private health care provider now offers asymptomatic patients the possibility to self-collect a gargle specimen as an alternative to NPS collection (11). We evaluated and compared three alternative specimen collection methods that would not require close contact with a HCW and compared their sensitivity and cycle threshold (C_T) values with those for NPS.

RESULTS

We enrolled 40 patients, with a mean age of 38.7 years (standard deviation [SD], 12.6 years), including 21 female patients (53%). Enrollment was performed as soon as the patients received the positive testing results, either 1 day ($n = 27/40$ [67.5%]) or 2 days ($n = 13/40$ [32.5%]) after they had been sampled by NPS for COVID-19 diagnosis (Fig. 1). Thirty-one patients had been symptomatic since disease onset (see Table S1 in the supplemental material), with most prevalent symptoms being fatigue (86%), headache (81%), and cough (79%). At the time of specimen collection, only 24 patients were symptomatic, with the most prevalent symptoms being cough (44%), anosmia (42%), and headache (40%).

We collected 40 concomitant NPS and gargle specimens on the recruitment day, as well as 40 next-morning gargle specimens. Of 22 patients assigned to the spit specimen collection group, all of them gave back specimens; of 18 patients assigned to the sputum group, 2 patients could not produce sputum, and thus only 16 sputum samples were included in the analysis.

All specimens were generally as sensitive as NPS to diagnose SARS-CoV-2 infection. The morning spit specimens showed the highest sensitivity (sensitivity, 1.00 [95% confidence interval [CI], 1.00 to 1.00]), followed by gargle specimens, regardless of when they had been collected (0.97 [95% CI, 0.92 to 1.00]). The sputum specimens had the lowest sensitivity (0.94 [95% CI, 0.87 to 1.00]) (Table 1).

TABLE 1 Sensitivity and specificity of tested sampling methods

Specimen type and result	No. with NPS result of:		Sensitivity (95% CI)	Specificity (95% CI)	Corrected sensitivity (95% CI)	Corrected specificity (95% CI)	Cohen's kappa (95% CI)	AUC (95% CI)
	Positive	Negative						
Gargle 1								
Positive	37	1	0.97 (0.92–1.00)	0.50 (0.00–1.00)	0.97 (0.02–1.00)	0.50 (0.00–1.00)	0.47 (–0.15–1.00)	0.74 (0.47–1.00)
Negative	1	1						
Gargle 2								
Positive	37	1	0.97 (0.92–1.00)	0.50 (0.00–1.00)	1.00 (1.00–1.00)	1.00 (0.87–1.00)	0.47 (–0.15–1.00)	0.74 (0.47–1.00)
Negative	1	1						
Sputum								
Positive	13	1	0.94 (0.83–1.00)	0			–0.09 (–0.22–0.04)	0.54 (0.50–0.61)
Negative	2	0						
Spit								
Positive	21	0	1.00 (1.00–1.00)	1.00 (1.00–1.00)			1.00 (1.00–1.00)	1 (1–1)
Negative	0	1						

TABLE 2 Comparison of C_T values obtained with alternative samples versus NPS

Sample type	No. of samples	C_T (mean \pm SD)	P^a
NPS	40	22.07 \pm 4.63	Reference
Gargle 1	40	29.89 \pm 4.34	<0.001
Gargle 2	40	29.25 \pm 3.99	<0.001
Sputum	16	25.82 \pm 9.21	0.28
Spit	22	23.51 \pm 4.57	0.11

^aPaired t test.

We compared C_T values obtained from alternative specimens to NPS (Table 2). NPS had the lowest C_T value (22.07; SD, 4.63), although it was not significantly lower than values for sputum (25.82 [SD, 9.21]; $P = 0.28$) and spit (23.51 [SD, 4.57]; $P = 0.11$) specimens. Both gargle specimens had statistically significantly higher C_T values, compared to NPS (Table 2). All patients' different specimen results are presented in Fig. 2.

DISCUSSION

We evaluated self-collection of alternative specimens, i.e., gargle, spit, and sputum specimens, and compared their sensitivity and C_T values to those for NPS for SARS-CoV-2 infection diagnosis. All specimen collection methods were comparably sensitive as NPS, with sensitivities exceeding 90%. Compared to NPS, gargle specimens had significantly higher C_T values, likely due to dilution by gargling water, while spit and sputum specimens collected on the following day had higher, although not significantly different, C_T values. Additionally, sputum samples appeared more challenging to collect among patients with milder symptoms. Therefore, spit would be the most suitable alternative specimen to NPS for SARS-CoV-2 infection diagnosis.

Since the beginning of the COVID-19 pandemic, several studies or meta-analyses have investigated the potential use of saliva specimens for SARS-CoV-2 infection diagnosis

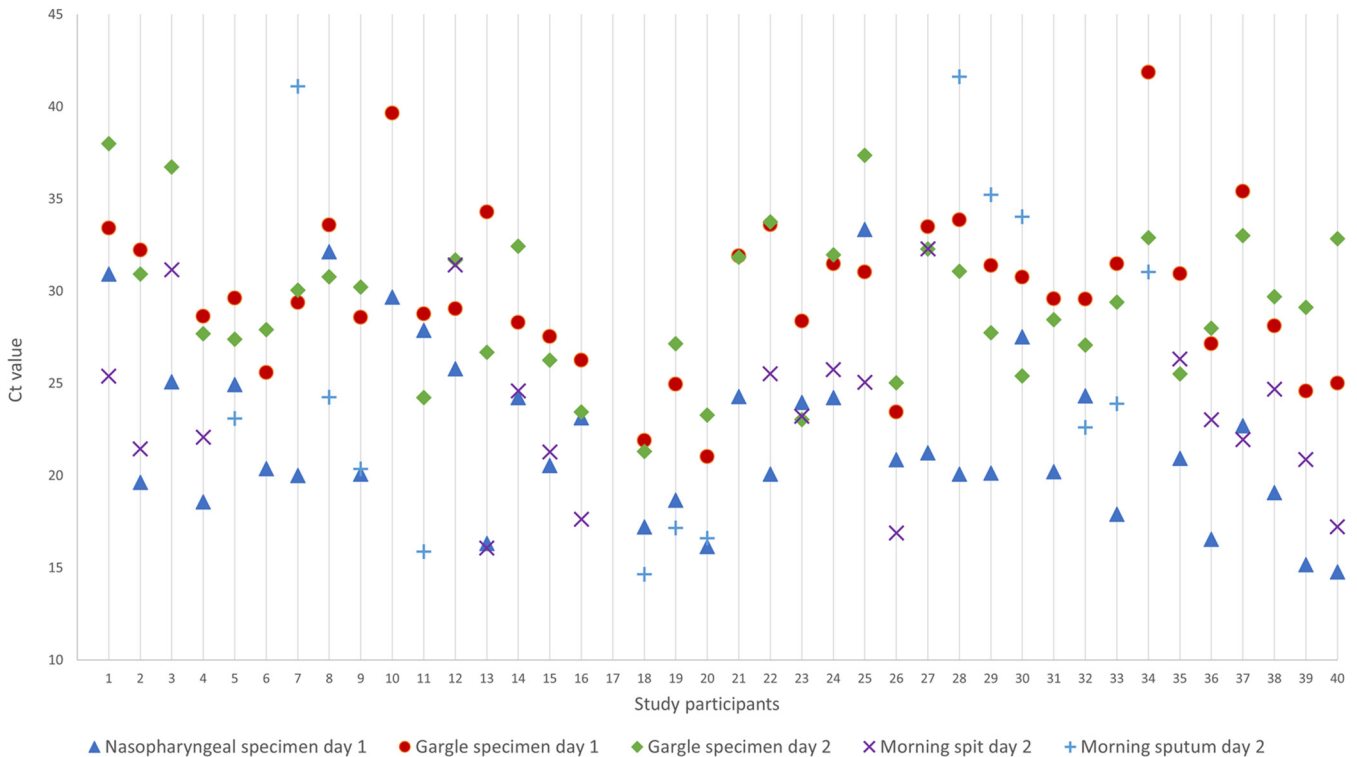


FIG 2 C_T values depending on specimen type and time of collection.

among both symptomatic (2, 7, 8, 12–17) and asymptomatic (2, 7, 8, 12–14, 17, 18) patients. The European Centre for Disease Prevention and Control (ECDC) and the Centers for Disease Control and Prevention (CDC) have approved the use of saliva or sputum samples as diagnostic specimens for COVID-19 testing for patients with a productive cough (19, 20).

There have been mixed results, considering differences in C_T values between NPS and saliva specimens (8, 15, 16, 18). These inconclusive results can be caused by various factors; morning saliva samples might have higher viral loads, compared to those from the rest of the day (14), and levels of SARS-CoV-2 in saliva samples correlate with COVID-19 symptom severity (21). Additionally, the early-morning posterior oropharyngeal spitting technique has been considered to have higher sensitivity for SARS-CoV-2 infection diagnosis, compared to NPS (7).

Previously, gargle specimens were also estimated to be sensitive for SARS-CoV-2 infection diagnosis, compared to NPS, including 50 inpatients (22) and outpatients (3, 23) with confirmed COVID-19. C_T values for gargle specimens were higher than those for NPS in those three studies, including one with 19,620 outpatients that is in line with findings in our study (3, 22, 23). Interestingly, Goldfarb et al., when analyzing 40 COVID-19 outpatients, concluded that gargle specimens had higher sensitivity than saliva specimens (97.5% [95% CI, 86.8 to 99.9%], compared to 78.8% [95% CI, 61.0 to 91.0%]), which is inconsistent with our results. One possible explanation for this could be that patients were instructed to gargle for a longer time in this study (3). Levican-Asenjo et al. and Malczynski et al. showed that diagnostic results and C_T values were comparable between sputum specimen and NPS tests during the first 10 days after COVID-19 diagnosis (24, 25). Our results were also consistent with that study.

The main benefit of alternative specimen collection would be to avoid close contact with a HCW and to avoid an unpleasant procedure. Overall, it would also increase the willingness to apply for SARS-CoV-2 testing and the allocation of current resources for HCWs. However, other practicalities must be considered in developing an alternative sample collection method. Morning spit collection before any food or water intake appears to show the best performance but could not be offered in all situations (appointment for sampling at a later time of day, for example). However, offering this alternative to self-quarantine patients feeling unwell could be a suitable use of the morning spit collection method, assuming that the risk of slightly delayed contact tracing did not outweigh the decreased risk of exposure for HCWs.

The main limitations of our work were small sample size (40 participants in total) and the fact that we did not analyze whether delayed transport or extended storage before analysis could possibly hamper sensitivity; all samples were transported and processed on the same day. Furthermore, the volume and timing of samples (e.g., the second gargle sample after fluid intake) had some variation, which could have an effect on the analysis. Additionally, because the study sample consisted of participants with recent positive results, most patients still tested positive with most methods, meaning that the values obtained regarding specificity should be interpreted with caution. However, we conducted this study as an exploratory assessment of alternative specimen collection methods and focused on patients with the most common clinical picture of COVID-19, with mild symptoms, because they are the ones who would most benefit from noninvasive alternative specimen collection. In addition, we managed to collect all samples within 1 to 2 days after diagnosis, while patients were still in the acute phase of the disease.

CONCLUSION

Among gargle, spit, and sputum specimens, morning collection of spit before any food or water intake or teeth brushing appeared to be the most suitable alternative specimen collection method, with C_T values as low as those obtained with NPS and with ease of collection for patients with mild symptoms. Our findings are promising but warrant further investigations with larger study populations. It is important to note

that the efficiency of different RNA extraction methods can significantly vary among sample materials. For this reason, before changing sample materials, the detection method used must be carefully validated. We will consider offering to patients with mild symptoms who are seeking diagnosis in a pilot testing center the possibility of enrolling in a study assessing whether, in the general population, spit collected on the next morning has the same performance as in our exploratory sample. If so, in the long run, we could offer patients with mild symptoms the choice between at-home self-collection of spit and NPS collection at a testing center. This would not only decrease discomfort but also decrease HCWs' exposure and burden.

MATERIALS AND METHODS

Sample collection. We contacted confirmed COVID-19 outpatients who had been diagnosed with SARS-CoV-2 infection by reverse transcription (RT)-PCR using NPS a few days earlier. Children <2 years of age were not eligible for participation. After calling the patients for recruitment, we visited them on the same day and the following day.

During the first home visit, we gathered informed consent, gave participants a link to an online symptom questionnaire, and collected a NPS and a gargle specimen (gargle 1). We also gave them instructions (see Appendix S1 in the supplemental material) and containers for collection of gargle (gargle 2) and, depending on the recruitment week, sputum or spit specimens the next morning. On the day following the first visit, the patients themselves collected the second gargle (gargle 2) and spit or sputum specimens. We retrieved the self-collected samples later in the day. The timeline of the study is presented in Fig. 1.

All alternative specimens were collected into a 70-ml plastic container. To collect the gargle specimens, the patients were instructed to have a sip of water and gargle it for 5 to 20 s before spitting it in the container. For spit collection, the patients had to spit continuously into the container until they filled one-half its volume; for the sputum specimens, the patients were asked to cough sputum from deep in their lungs and then spit it into the container. Both the spit and sputum specimens were to be collected in the morning before the patients ate, drank, or brushed their teeth, whereas the gargle sample was collected by the participants later in the morning. All specimens were transported at room temperature and analyzed on the same day in the Expert Microbiology Unit at the Finnish Institute for Health and Welfare (THL). The Finnish communicable diseases law and the law on the duties of the THL allowed the implementation of this noninvasive research without seeking further ethical approval (26).

Laboratory methods. RNA extraction from samples was performed using the Chemagic Viral300 DNA/RNA kit H96 (PerkinElmer) according to the manufacturer's instructions. A sample volume of 300 μ l and an elution volume of 50 μ l were used. Highly viscose gargle samples were vortex-mixed with 1 ml phosphate-buffered saline (PBS) before 300 μ l was taken for RNA extraction. Real-time RT-PCR was performed using qScript XLT one-step RT-quantitative PCR (qPCR) ToughMix (Quantabio). SARS-CoV-2 was detected using the E (envelope) gene real-time RT-PCR assay. Primers and probes were based on the Corman E gene primer/probe set (27). The thermal profile for PCR was 55°C for 20 min and 95°C for 3 min, followed by 45 cycles of 95°C for 15 s and 58°C for 1 min, using a CFX thermal cycler (Bio-Rad).

Statistical analysis. We used NPS RT-PCR test results as the reference and C_T values as surrogates for viral load analysis. We used paired *t* tests to compare the measured C_T values between NPS and gargle specimens and between NPS and sputum/spit specimens. Standard methods were used to calculate sensitivity and specificity of the other diagnostic tests (index tests) from saliva and sputum/spit specimens. We used McNemar's exact test to assess the differences in sensitivity and specificity.

We calculated Cohen's kappa coefficient to evaluate the agreement between the reference and the index tests. The area under the receiver operating characteristic curve (AUC) and 95% CI were reported. Because of the imperfect reference test, latent class analysis was used as a correction method to adjust the estimated sensitivity and specificity of the index tests based on the existing sensitivity and specificity of the reference test. Model selection was based on the Bayesian information criterion (BIC). Data analysis was performed using R software (version. 3.6.0).

SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

SUPPLEMENTAL FILE 1, PDF file, 0.4 MB.

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We declare no conflicts of interest.

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L.H., and T.D.; laboratory analysis, N.I., K.L., and the THL virology laboratory; writing and editing, all authors.

REFERENCES

- European Centre for Disease Prevention and Control. 2020. Contact tracing: public health management of persons, including healthcare workers, who have had contact with COVID-19 cases in the European Union: third update. European Centre for Disease Prevention and Control, Solna, Sweden.
- Medeiros da Silva RC, Nogueira Marinho LC, Neto de Araújo Silva D, Costa de Lima K, Pirih FQ, Luz de Aquino Martins AR. 2020. Saliva as a possible tool for the SARS-CoV-2 detection: a review. *Travel Med Infect Dis* 38: 101920. <https://doi.org/10.1016/j.tmaid.2020.101920>.
- Goldfarb DM, Tilley P, Al-Rawahi GN, Srigley JA, Ford G, Pedersen H, Pabbi A, Hannam-Clark S, Charles M, Ditttrick M, Gadkar VJ, Pernica JM, Hoang LMN. 2021. Self-collected saline gargle samples as an alternative to health care worker-collected nasopharyngeal swabs for COVID-19 diagnosis in outpatients. *J Clin Microbiol* 59:e02427-20. <https://doi.org/10.1128/JCM.02427-20>.
- Finnish Institute for Health and Welfare. 2021. COVID-19-epidemiaan hybridistrategian seuranta: tilannearvioraportti 10.3.2021. Finnish Institute for Health and Welfare, Helsinki, Finland.
- Finnish Institute for Health and Welfare. 22 March 2021. Tilannekatsaus koronaviruskesta: infektioaudit ja rokotukset. <https://thl.fi/fi/web/infektioaudit-ja-rokotukset/ajankohtaista/ajankohtaista-koronaviruksesta-covid-19/tilannekatsaus-koronaviruksesta>.
- World Health Organization. 2020. Diagnostic testing for SARS-CoV-2: interim guidance, 11 September 2020. WHO/2019-nCoV/laboratory/2020.6. World Health Organization, Geneva, Switzerland. <https://www.who.int/publications/i/item/diagnostic-testing-for-sars-cov-2>.
- Bastos ML, Perlman-Arrow S, Menzies D, Campbell JR. 2021. The sensitivity and costs of testing for SARS-CoV-2 infection with saliva versus nasopharyngeal swabs: a systematic review and meta-analysis. *Ann Intern Med* 174:501–510. <https://doi.org/10.7326/M20-6569>.
- Wyllie AL, Fournier J, Casanovas-Massana A, Campbell M, Tokuyama M, Vijayakumar P, Warren JL, Geng B, Muenker MC, Moore AJ, Vogels CBF, Petrone ME, Ott IM, Lu P, Venkataraman A, Lu-Culligan A, Klein J, Earnest R, Simonov M, Datta R, Handoko R, Naushad N, Sewanan LR, Valdez J, White EB, Lapidus S, Kalinich CC, Jiang X, Kim DJ, Kudo E, Linehan M, Mao T, Moriayama M, Oh JE, Park A, Silva J, Song E, Takahashi T, Taura M, Weizman O-E, Wong P, Yang Y, Bermejo S, Odio CD, Omer SB, Dela Cruz CS, Farhadian S, Martinello RA, Iwasaki A, Grubaugh ND, Ko AI. 2020. Saliva or nasopharyngeal swab specimens for detection of SARS-CoV-2. *N Engl J Med* 383:1283–1286. <https://doi.org/10.1056/NEJM2016359>.
- Malecki M, Lüsebrink J, Teves S, Wendel AF. 2021. Pharynx gargle samples are suitable for SARS-CoV-2 diagnostic use and save personal protective equipment and swabs. *Infect Control Hosp Epidemiol* 42:248–249. <https://doi.org/10.1017/ice.2020.229>.
- Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W. 2020. Detection of SARS-CoV-2 in different types of clinical specimens. *JAMA* 323:1843–1844. <https://doi.org/10.1001/jama.2020.3786>.
- Terveytalo. 2021. Koronatesti kurlaamalla. <https://www.terveytalo.com/fi/Tietopaketti/Koronatesti-kurlaamalla>.
- Czumbel LM, Kiss S, Farkas N, Mandel I, Hegyi A, Nagy Á, Lohinai Z, Szakács Z, Hegyi P, Steward MC, Varga G. 2020. Saliva as a candidate for COVID-19 diagnostic testing: a meta-analysis. *Front Med (Lausanne)* 7: 465. <https://doi.org/10.3389/fmed.2020.00465>.
- Sagredo-Olivares K, Morales-Gómez C, Aitken-Saavedra J. 2021. Evaluation of saliva as a complementary technique to the diagnosis of COVID-19: a systematic review. *Med Oral Patol Oral Cir Bucal* 26:e526–e532. <https://doi.org/10.4317/medoral.24424>.
- Hung DL-L, Li X, Chiu KH-Y, Yip CC-Y, To KK-W, Chan JF-W, Sridhar S, Chung TW-H, Lung K-C, Liu RW-T, Kwan GS-W, Hung IF-N, Cheng VC-C, Yuen K-Y. 2020. Early-morning vs spot posterior oropharyngeal saliva for diagnosis of SARS-CoV-2 infection: implication of timing of specimen collection for community-wide screening. *Open Forum Infect Dis* 7:ofaa210. <https://doi.org/10.1093/ofid/ofaa210>.
- Williams E, Bond K, Zhang B, Putland M, Williamson DA. 2020. Saliva as a noninvasive specimen for detection of SARS-CoV-2. *J Clin Microbiol* 58: e00776-20. <https://doi.org/10.1128/JCM.00776-20>.
- Yokota I, Hattori T, Shane PY, Konno S, Nagasaka A, Takeyabu K, Fujisawa S, Nishida M, Teshima T. 2021. Equivalent SARS-CoV-2 viral loads by PCR between nasopharyngeal swab and saliva in symptomatic patients. *Sci Rep* 11:4500. <https://doi.org/10.1038/s41598-021-84059-2>.
- Butler-Laporte G, Lawandi A, Schiller I, Yao M, Dendukuri N, McDonald EG, Lee TC. 2021. Comparison of saliva and nasopharyngeal swab nucleic acid amplification testing for detection of SARS-CoV-2: a systematic review and meta-analysis. *JAMA Intern Med* 181:353–360. <https://doi.org/10.1001/jamainternmed.2020.8876>.
- Herrera LA, Hidalgo-Miranda A, Reynoso-Noverón N, Meneses-García AA, Mendoza-Vargas A, Reyes-Grajeda JP, Vadillo-Ortega F, Cedro-Tanda A, Peñaloza F, Frías-Jimenez E, Arriaga-Canon C, Ruiz R, Angulo O, López-Villaseñor I, Amador-Bedolla C, Vilar-Compte D, Cornejo P, Cisneros-Villanueva M, Hurtado-Cordova E, Cendejas-Orozco M, Hernández-Morales JS, Moreno B, Hernández-Cruz IA, Herrera CA, García F, González-Woge MA, Munguía-Garza P, Luna-Maldonado F, Sánchez-Vizcarra A, Osnaya VG, Medina-Molotla N, Alfaro-Mora Y, Cáceres-Gutiérrez RE, Tolentino-García L, Rosas-Escobar P, Román-González SA, Escobar-Arazola MA, Canseco-Méndez JC, Ortiz-Soriano DR, Domínguez-Ortiz J, González-Barrera AD, Aparicio-Bautista DI, Cruz-Rangel A, Alarcón-Zendejas AP, Contreras-Espinosa L, González R, Guerra-Calderas L, Meraz-Rodríguez MA, Montalvo-Casimiro M, Montiel-Manríquez R, Torres-Arciga K, Venegas D, Juárez-González V, Guajardo-Barreto X, Monroy-Martínez V, Guillén D, Fernández J, Herrera J, León-Rodríguez R, Canela-Pérez I, Ruiz-Ordaz BH, Valdez-Vazquez R, Bertin-Montoya J, Niembro-Ortega M, Villegas-Acosta L, López-Castillo D, Soriano-Ríos A, Gastelum-Ramos M, Zamora-Barandas T, Morales-Baez J, García-Rodríguez M, García-Martínez M, Nieto-Patlán E, Quirasco-Baruch M, López-Martínez I, Ramírez-Gonzalez E, Olivera-Díaz H, Escobar-Escamilla N. 2021. Saliva is a reliable and accessible source for the detection of SARS-CoV-2. *Int J Infect Dis* 105:83–90. <https://doi.org/10.1016/j.ijid.2021.02.009>.
- Centers for Disease Control and Prevention. 22 February 2021. Interim guidelines for collecting and handling of clinical specimens for COVID-19 testing. <https://www.cdc.gov/coronavirus/2019-ncov/lab/guidelines-clinical-specimens.html>.
- European Centre for Disease Prevention and Control. 24 March 2021. Diagnostic testing and screening for SARS-CoV-2. <https://www.ecdc.europa.eu/en/covid-19/latest-evidence/diagnostic-testing>.
- Silva J, Lucas C, Sundaram M, Israelow B, Wong P, Klein J, Tokuyama M, Lu P, Venkataraman A, Liu F, Mao T, Oh JE, Park A, Casanovas-Massana A, Vogels CBF, Muenker CM, Zell J, Fournier JB, Campbell M, Chiorazzi M, Ruiz Fuentes E, Petrone M, Kalinich CC, Ott IM, Watkins A, Moore AJ, Nakahata MI, Grubaugh ND, Farhadian S, Dela Cruz C, Ko A, Schulz WL, Ring AM, Ma S, Omer S, Wyllie AL, Iwasaki A. 2021. Saliva viral load is a dynamic unifying correlate of COVID-19 severity and mortality. *medRxiv*. <https://doi.org/10.1101/2021.01.04.21249236>.
- Mittal A, Gupta A, Kumar S, Surjit M, Singh B, Soneja M, Soni KD, Khan AR, Singh K, Naik S, Kumar A, Aggarwal R, Nischal N, Sinha S, Trikha A, Wig N. 2020. Gargle lavage as a viable alternative to swab for detection of SARS-CoV-2. *Indian J Med Res* 152:77–81. https://doi.org/10.4103/ijmr.IJMR_2987_20.
- Kandel CE, Young M, Serbanescu MA, Powis JE, Bulir D, Callahan J, Katz K, McCready J, Racher H, Sheldrake E, Quon D, Vojdani OK, McGeer A, Goneau LW, Vermeiren C. 2021. Detection of severe acute respiratory coronavirus virus 2 (SARS-CoV-2) in outpatients: a multicenter comparison of self-collected saline gargle, oral swab, and combined oral–anterior nasal swab to a provider collected nasopharyngeal swab. *Infect Control Hosp Epidemiol* 1–5. <https://doi.org/10.1017/ice.2021.2>.
- Levicán-Asenjo JE, Almonacid LI, Valenzuela G, García T, Rojas L, Serrano E, Pardo-Roa C, Salinas E, Avendano MJ, Perazzo F, Diaz LA, Valderrama S, Ortega M, Toro A, Montecinos V, Riquelme A, Medina RA. 2020. Viral shedding dynamics reveals sputum as a reliable and cost-saving specimen for SARS-CoV-2 diagnosis within the first 10 days since symptom onset: a prospective cohort study. *medRxiv*. <https://doi.org/10.1101/2020.08.30.20183889>.

25. Malczynski M, Rezaeian S, Rios J, Dirnberger L, Polanco W, Zembower T, Qi C. 2020. Development of a protocol for detection of SARS-CoV-2 in sputum and endotracheal aspirates using Cepheid Xpert Xpress SARS-CoV-2. *Access Microbiol* 2:acmi000176. <https://doi.org/10.1099/acmi.0.000176>.
26. FINLEX. 2016. Säädökset alkuperäisinä: Tartuntatautilaki 1227/2016. <https://finlex.fi/fi/laki/alkup/2016/20161227>.
27. Corman VM, Landt O, Kaiser M, Molenkamp R, Meijer A, Chu DK, Bleicker T, Brünink S, Schneider J, Schmidt ML, Mulders DG, Haagmans BL, van der Veer B, van den Brink S, Wijsman L, Goderski G, Romette J-L, Ellis J, Zambon M, Peiris M, Goossens H, Reusken C, Koopmans MP, Drosten C. 2020. Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR. *Euro Surveill* 25:2000045. <https://doi.org/10.2807/1560-7917.ES.2020.25.3.2000045>.