# International Journal of Pharmacy and Pharmaceutical Sciences

ISSN- 0975-1491

Vol 7, Issue 11, 2015

**Original Article** 

# ANTI-ANEMIC ACTIVITY OF SPROUTS OF VIGNA RADIATA L. IN MALE ALBINO RATS

# MANIKANDASELVI S.<sup>1</sup>\*, DAVID RAJ C.<sup>3</sup>, ARAVIND S.<sup>2</sup>, RAVIKUMAR R.<sup>4</sup>, THINAGARBABU R.<sup>2</sup>, NANDHINI S.<sup>1</sup>

<sup>1</sup>PG and Research Department of Biochemistry, S. T. E. T Women's College, Mannargudi, Tamilnadu, India, <sup>2</sup>Department of Chemistry, School of Chemical and Bio-Technology, SASTRA, University, Thanjavur, Tamilnadu, India, <sup>3</sup>Central Animal Facility, SASTRA University, Thanjavur, Tamilnadu, India, <sup>4</sup>Department of Chemistry, Vivekanandha College of Arts and Sciences for Women, Tiruchengode

Email: selvithinagar@gmail.com

# Received: 22 May 2015 Revised and Accepted: 03 Oct 2015

# ABSTRACT

**Objective:** To evaluate the anti-anemic activity of sprouts of *Vigna radiata* L. against phenyl hydrazine induced anemic rats.

**Methods:** Rats were divided into 4 groups of 6 each. Group 1 was given normal saline and served as control and all other groups were given 40 mg/kg b. w of phenyl hydrazine for 2 d to induce anemia. Group 3 was treated with Bioferon (230 mg/kg) and served as the standard. Group 4 was treated with sprouted *Vigna radiata* L. (600 mg/kg bw). All the treatments were given orally. On completion of the experimental period, all the test substance/vehicle-treated rats were sacrificed and the plasma separated was used for estimating various biochemical as well as hematological parameters as per standard procedures.

**Results:** The experimental rats treated with sprouted *Vigna radiata* L. at the dose level 600 mg/kg bw for 13 d revealed significant changes in biochemical and hematological parameters compared to phenyl hydrazine induced anemic rats.

Conclusion: The present study concluded that the sprouted Vigna radiata L. inhibits anemia induced by phenyl hydrazine in male albino rats.

Keywords: Anemia, Phenyl hydrazine, Vigna radiata L., Bioferon.

### INTRODUCTION

Anemia is described as a reduction in Red Blood Cell (RBC) mass or blood hemoglobin (Hb) concentration resulting in a decrease in the oxygen-carrying capacity of the blood. The prevalence of anemia in the developing countries tends to be 3 to 4 times higher than in the developed countries. Anemia affects the physical and mental development of an individual leading to decreased working capacity, which in turn affects the development of the country. Since the technological advancement and economic development of a nation depend heavily on its trained human resources, the behavioral effects of anemia are highly relevant. Consequently, if anemia is highly prevalent in a country, it can substantially affect its intellectual and economic potential.

The World Health Organization (WHO) estimates the number of anemic people worldwide to be a staggering 2 billion (about 30 % of the world's population) and that approximately 50 % of all anemias can be attributed to iron deficiency and in resource-poor areas, this is frequently exacerbated by infectious diseases, malaria, worm infestation and HIV/AIDS [1]. Even though anemia is associated with nutritional deficiencies, acute or chronic disease, drug use or physiological states such as pregnancy, blood loss, impaired erythropoiesis and abnormal erythrocyte destruction are implicated [2, 3].

Plant and plant products are being utilized as a source of medicine since long. Plant extracts are used as phytotherapeutics and are still a large source of natural antioxidants. Natural antioxidants strengthen the endogenous antioxidant defense from ROS ravage and restored the optimal balance by neutralizing the reactive species [4]. Particularly, flavonoids and phenolics are considered as potential therapeutic agents. A wide range of ailments and is widely distributed in the plant kingdom and, therefore, an integral part of the diet, with the significant amount reported in vegetables, fruits and beverages [5].

*Vigna radiata* L. has been consumed as a common food in China for more than 2,000 y. It is well known for its detoxification activities and is used to refresh mentality, alleviate heat stroke, and reduce swelling in the summer. In the book Ben Cao Qiu Zhen, the *Vigna* 

radiata L. was recorded to be beneficial in the regulation of gastrointestinal upset and to moisturize the skin [6]. The seeds and sprouts of Vigna radiata L. are also widely used as a fresh salad vegetable or common food in India, Bangladesh, South East Asia, and western countries [7]. Sprouted Vigna radiata L. contains: Water, 88.8; protein, 3.8; fat, 0.2; crude fiber, 0.7; total carbohydrates, 6.6; and ash, 0.6 g/100g; mineral constituents: Ca, 19; P, 64; Fe, 1.3; Na, 5; and K, 223 mg/kg. Yang and Tsou reported that the available iron in sprouts tested with in vitro dialysis is increased due to the increased ascorbic acid and reduced phytic acid content in Vigna radiata L. during sprouting [8-10]. High levels of proteins, amino acids, oligosaccharides, and polyphenols in Vigna radiata L. are thought to be the main contributors to the antioxidant activity [11-14], antimicrobial activity [15-19], anti-inflammatory activity [20-23], antidiabetic effects [24-25], antihypertensive effects [26], antitumor effects [27-29] and antisepsis effects [30-31].

*Vigna radiata* L. is said to be a traditional source cures paralysis, rheumatism, coughs, fever, liver ailments and for weight reduction. It is employed as a light diet during fever and is considered as a cooling and astringent effect. The pulse is prescribed for vertigo. A decoction of seeds is used an effective treatment for beriberi [32].

In recent years, studies have shown that the sprouts of *Vigna radiata L*. after germination have more obvious biological activities and more plentiful secondary metabolites since relevant bio-synthetic enzymes are activated during the initial stages of germination. Thus, germination is thought to improve the nutritional and medicinal qualities of *Vigna radiata L* [33]. Highly efficient use of *Vigna radiata L*. according to evidence demonstrated from scientific experiments will be beneficial to the application of *Vigna radiata L*. as a health food, medicine, and cosmetic [34].

In the tropical area between 10 to 20% of the population presents less than 10g/dl of hemoglobin. Due to the high prevalence and the possibility of an even further increase, there is the need to prevent it or seek for more cost effective and better treatment strategies. By keeping the above points, the present study has been designed to evaluate the anti-anemic activity of sprouted *Vigna radiata* L. on phenyl hydrazine induced hemolytic anemia in male albino rats.

# MATERIALS AND METHODS

# Animals

Wistar strain male albino rats, weighing 100-120g were selected for the study. The animals were housed individually in polypropylene cages under hygienic and standard environmental conditions (22±3 °C, humidity 30-70%, 12 h light/dark cycle). The animals were allowed to have standard feed and water *adlibtum*. They were acclimated to the environment for one week prior to experimental use. The study protocol was carried out as per the rules and regulations of the Institutional Animal Ethical Committee (IAEC).

#### Chemicals

All chemicals and reagents were used of analytical grade, purchased from SRL Chemie Pvt. Ltd. and from Hi Media Laboratories Pvt. Limited.

### Plant material and preparation of drug

The seeds of *Vigna radiata* L. were purchased from a departmental store during the month of January 2014. The seeds were germinated by soaking them in water for four hours. Collected sprouts were open-air-dried under the shade, pulverized into a moderately coarse powder. The powder was mixed with distilled water just before oral administration.

#### **Experimental design**

Anemia was induced by intra peritoneal injection of phenyl hydrazine at 40 mg/kg for 2 d, as described by Diallo *et al.* [35]. Following the injections, rats were divided into four groups of six rats each.

Group I-Control rats received saline

Group II-Phenyl hydrazine treated rats (40 mg/kg per day for 2 d)

Group III-Phenyl hydrazine treated rats with 40 mg/kg per day for 2 d and sprouted *Vigna radiata* L. single dose (600 mg/kg) per day for 13 d.

Group IV-Phenyl hydrazine treated rats with standard Bioferon treated rats-a single dose (230 mg/kg) per day for 13 d.

On completion of the experimental period, animals were anesthetized with thiopentone sodium (50 mg/kg, ip). The blood was collected with and without EDTA as an anticoagulant. Plasma was separated by centrifugation. Plasma was used for the estimation of various biochemical parameters. Hemoglobin was estimated by cyanomethaemoglobin method [36]. WBC, RBC and platelet were counted by Ochei method, 2000 [37].

Hematocrit or Packed Cell Volume (PCV)

$$PCV \% = \frac{Packed RBC Column height}{Total blood Column height}$$

Mean Corpuscular Hemoglobin (MCH)

This indicates the weight of hemoglobin in a single red blood cell and is expressed in picograms (pg)  $(1pg=10\cdot[12]g)$ .

$$MCH = \frac{\text{Hemoglobin (g/dL)}}{\text{RBC count Million per cu. mm}}$$

Mean corpuscular Hemoglobin concentration (MCHC)

This denotes the hemoglobin concentration per 100 ml of packed red blood cells and is related to the color of the red cells. This is expressed as the percentage of packed cells.

$$MCHC = \frac{\text{Hemoglobin (g/dL)}}{\text{PCV \%}} X100$$

This is expressed as the volume in cubic microns or femtoliters of an average red blood cell.

$$MCV = \frac{PCV \%}{RBC \text{ count Million per cu. mm}} X10$$

Malondialdehyde was estimated by the thiobarbituric acid assay method of Beuge and Aust [38]; reduced glutathione was estimated by the method of Moron *et al.*, [39] and serum GOT was estimated by the method of Reitman and Frankel [40].

### Statistical analysis

Values were expressed as mean±SD and statistically significant differences between mean values were determined by one-way analysis of variance (ANOVA) followed by the Tukey's test for multiple comparisons [41]. Statistical analysis carried out by MS-Windows based graph pad Instat software (Graph Pad software, San Diego, CA, USA) 3 version was used. A value of P<0.05 was considered statistically significant.

# RESULTS

The present study was carried out to assess the anti-anemic activity of sprouted *Vigna radiata* L. The observations made on different groups of experimental and control animals were compared as follows.

Table I represent the effect of sprouted *Vigna radiata* L. on Hemoglobin, WBC, RBC, and platelet content of male albino rats. Group II phenyl hydrazine intoxicated rats showed a significant (8.22±0.87) decrease in the content of Hb when compared to Group I rats. Group III phenyl hydrazine intoxicated rats treated with sprouted *Vigna radiata* L. significantly (14.47±0.93) increase in the level of Hb when compared to group II. Group II phenylhydrazine induced rats showed a significant (2350±236, 2.30±0.18 and 18833.3±2016) decrease in WBC, RBC and platelet levels respectively when compared to Group I rats. Group III phenyl hydrazine induced rats treated with sprouted *Vigna radiata* L. significant (3450±278, 3.89±0.04 and 219000±1870) increase in WBC, RBC and platelet levels respectively when compared to group II.

Table 1: Effect of sprouted Vigna radiata L. on Hemoglobin, WBC, RBC and Platelet count

Parameters	Group I	Group II	Group III	Group IV
Hemoglobin(g/dl)	11.96±1.04	8.22±0.87	$14.47\pm0.93^*$	12.0±0.02
WBC (Cells x 10 <sup>9</sup> /l)	4075±245	2350±236	3450±278 <sup>*</sup>	3949±146
RBC (Cells x 10[12]/l)	4.9±0.39	2.30±0.18	3.89±0.04*	4.2±0.17
Platelet(Cells x $10^9$ /l)	223583±1744	18833.3±2016	$219000 \pm 1870^*$	203410±1080

Values were expressed as mean±SD for six rats in each group, \*Significantly different from Group II P<0.05.

Table 2 represents the effect of sprouted *Vigna radiata* L. on PCV, MCH, MCHC and MCV count in male albino rats. Group II phenylhydrazine intoxicated rats showed a significant (16±0.01) decrease in PCV when compared to Group I rats. Group III rats treated with germinated *Vigna radiata* L. significantly (29±0.11) increase in PCV when compared to group II. Group II phenyl hydrazine induced rats showed a significant increase in MCH, MCHC, MCV when compared to Group I rats. Group III rats treated with germinated *Vigna radiata* L. significantly (33±0.35, 38±0.24 and

 $48{\pm}0.97)$  decreases in MCH, MCHC, and MCV levels respectively when compared to group II.

Table 3 represents the levels of MDA, GSH, and SGOT in male albino rats. Group II phenyl hydrazine intoxicated rats showed a substantial (7.4±1.24) increase in the level of MDA when compared to Group I rats. Group III phenyl hydrazine intoxicated rats treated with germinated *Vigna radiata* L. significantly (4.2±1.02) a decrease in the level of MDA when compared to group II. Group II phenyl hydrazine intoxicated rats showed a significant (2.1±0.54) decrease in the level of GSH when compared to Group I rats. Group III phenyl hydrazine intoxicated rats treated with sprouted *Vigna radiata* L. significantly (4.0±0.67) increase in the level of GSH as compared to group II. Group II phenyl hydrazine intoxicated rats showed a significant increase (205.3 $\pm$ 72.5) in the activity of SGOT when compared to Group I rats. Group III phenyl hydrazine intoxicated rats treated with germinated *Vigna radiata* L. significantly (79.2 $\pm$ 22.8) a decrease in the activity of SGOT when compared to group II.

Parameters	Group I	Group II	Group III	Group IV	
PCV (%)	23±0.13	16±0.01	29±0.11*	25±0.02	
MCH (pg/cell)	24±0.32	50±0.41	33±0.35*	30±0.58	
MCHC (%)	32±0.25	53±0.42	38±0.24*	34±0.03	
MCV (cubic micron)	46±0.85	86±1.58	48±0.97*	48±0.02	

Values were expressed as mean±SD for six rats in each group, \*Significantly different from Group II P<0.05.

Table 3: Effect of s	nrouted <i>Viana radiat</i>	a Lon MDA level	GSH and SGOT activity
Table 5. Effect of 5	pi outcu rignu ruunut	u h. on Mibri ievel,	ush and sub r activity

Parameters	Group I	Group II	Group III	Group IV	
MDA(nmole/l)	3.78±0.16	7.4±1.24	4.2±1.02*	3.8±0.6	
GSH (mg/dl)	4.63±0.05	2.1±0.54	4.0±0.67*	4.61±0.4	
SGOT (IU/l)	49.1±16.6	205.3±72.5	79.2±22.8*	60.3±16.2	

Values were expressed as mean±SD for six rats in each group, \*Significantly different from group II P<0.05.

#### DISCUSSION

Anemia is a disease characterized by a reduction in the concentration of hemoglobin, circulating red blood cell and pack cell volume per unit of the peripheral blood below the normal for the age and sex of the patient. The present study aimed to evaluate the antianemic activity of sprouts of *Vigna radiata* L. against phenyl hydrazine induced anemic rats. Phenyl hydrazine is recognized for its capacity to cause hemolysis both *in-vitro* and *in-vivo* by the formation of aryl and hydroxyl radicals, which have been demonstrated to be associated with its interaction with erythrocytes [42].

Oxidative stress in erythrocytes is considered as an important mechanism of hemolysis. Disruption of membrane integrity arises from fragility, dehydration as well as increased production of reactive oxygen species. Chronic hemolysis leads to loss of hemoglobin. These metabolic changes lead to the depletion of essential nutrients and micronutrients which are required for proper cell function [43]. The accumulation of hydrogen peroxide in addition to the detoxifying capacity of the red cell may lead to the oxidation of essential cellular constituents including membrane phospholipids. Such alterations presumably contribute to the eventual hemolysis of affected cells.

The intoxication of rats with phenyl hydrazine (4 mg/kg for 2 d) resulted in a marked hemolytic anemia characterized by decreased RBC, hemoglobin and PCV [44]. Similar results were obtained in our study when experimental rats were administered phenyl hydrazine in order to induce anemia. Phenyl hydrazine altered the function of RBC by hemolysis characterized by decreased levels of RBC, hemoglobin and PCV. In addition, Ferrali *et al.* [45], observed an increased reticulocytosis, methaemoglobinemia and hemocatheresis in phenyl hydrazine intoxicated rats.

This study is intended to evaluate the effect of sprouted *Vigna radiata* L. on the hemolytic anemia induced by phenyl hydrazine. It has been demonstrated previously that intraperitoneal administration of phenyl hydrazine decreases the hemoglobin concentration, red blood cell number, and hematocrit in rats [46]. This anemia which resulted from the early lysis of the red blood cells was naturally reversed 12 d later by the regeneration of those blood cells due to the increase of the reticulocytes.

The germinated *Vigna radiata* L. could stimulate erythropoiesis process. The increase in the number of young red blood cells (reticulocytes) explains the strong osmotic resistance of the red blood cells in rats treated with the extract. The number of circulating

reticulocytes coincided with the increase in MCV, thus suggesting that erythrocyte precursors become enucleated at a more differentiated stage of erythropoiesis. On the other hand, the increase in MCH observed during the experimental period could be indicative of a certain degree of intravascular hemolysis [47].

Indicators of anemia are reduced hemoglobin concentration (Hb), red blood cell count (RBC), WBC and PCV. Animals are similar to humans in that reduction in Hb, RBC and PCV are indicative of anemia. Mean Corpuscular Hemoglobin Concentration (MCHC-the amount of hemoglobin per unit erythrocyte volume) often reduced in hemolytic anemia or increased in the case of massive intravascular hemolysis. Mean Corpuscular Volume (MCV-average volume of the erythrocyte) is often increased in hemolytic anemia as the result of reticulocytosis. Mean corpuscular hemoglobin (MCHthe average amount of hemoglobin per cell) often increased in hemolytic anemia [48]. In the present study, phenyl hydrazine intoxicated rats decrease hemoglobin levels, RBC, WBC, platelet count and PCV whereas; it induces an increase in MCV, MCH, and MCHC. Our results with earlier reports [49], supplementation of germinated Vigna radiata L. to phenyl hydrazine intoxicated rats restored the altered hematological parameters.

Lipid peroxidation and the resultant perturbation of the structural integrity of the plasma membrane have long been considered to be capable of initiating the hemolytic response [50], though how the generalized destruction of membrane lipids could stimulate a selective macrophage response was not clear. The most recent reports that lipid peroxidation in nucleated cells correlates with the accumulation of Phosphatidylserine (PS) on the outer leaflet of the lipid bilayer. ROS production was associated with extensive binding of oxidized and denatured hemoglobin to the membrane cvtoskeleton. Thus, phenyl hydrazine induced hemolytic injury seems to be derived from oxidative alterations to red blood cell membrane lipids [51]. In the present study, increased lipid peroxidation products, as MDA were observed on phenyl hydrazine, intoxicated rats. Supplementations of germinated Vigna radiata L. restored the MDA content suggested that reduced the oxidative damage.

In the present study, a marked decrease in the concentration of GSH was observed in phenyl hydrazine intoxicated rats when compared to control rats. Administration of sprouted *Vigna radiata* L. significantly increases the levels of GSH in phenyl hydrazine intoxicated rats. Enzymes catalyze specific biochemical reactions in the body. Changes in their levels and of cellular damage, the intracellular concentration of the enzymes and the mass properties

alter the functional ability of an organism. The diagnosis of organ disease/damage is aided by measurement of a number of nonfunctional plasma enzymes characteristic of that tissue or organ. The amount of enzyme released depends on the degree of the affected tissue. The concentration of the enzymes released reflects the severity of the damage. SGOT and SGPT are enzymes normally present in the liver, heart, muscles and blood cells. They are basically located within hepatocytes. So when liver cells are damaged or die transaminases is released into the bloodstream, where they can be measured they are therefore the index of liver injury [52]. The hepatocellular damage indicated by increased activity of SGOT in serum was observed in this study. Supplementation of sprouted *Vigna radiata* L. to phenyl hydrazine intoxicated rats restored the SGOT activity.

Herbal medicine is increasingly gaining greater recognition from the public and medical profession due to greater advances in the understanding of the mechanisms by which herbs positively influence health and quality of life [53]. Several plant products are known to exhibit creditable medicinal properties for the treatment of various ailments and need to be explored to identify their potential application in prevention and therapy of human ailments. Keeping in view the present study has evaluated the anti-anemic activity of sprouted *Vigna radiata* L. Phenyl hydrazine, an alkyl hydrazine was chosen to induce hemolytic anemia. Phenyl hydrazine induces the destruction of red blood cells by oxidation stress and many changes in cellular levels resulting in hemolytic anemia. Supplementation of germinated *Vigna radiata* L. to phenyl hydrazine intoxicated rats shows the following results.

- Improved the Hb content.
- Restored the WBC, RBC and platelet count.

> Secondary parameters of erythrocytes such as PCV, MCH, MCHC, and MCV were restored.

 $\succ$  Reduced oxidative damage confirmed by the decreased MDA content.

 $\succ$  Improved the detoxification mechanism by increased GSH content.

Normalize liver functions as evidenced by SGOT activity.

In developing countries, anemia is one of the major health problems and in India, Lauha bhasma, an iron-based herbo-metallic preparation, is prescribed for treating anemia [54, 55].

#### CONCLUSION

The results of the present study accomplished that sprouted *Vigna radiata* L. inhibits anemia induced by phenyl hydrazine model similar to those induced by parasites such as *Plasmodium falciparum*. This result supports at least partially the traditional use of sprouted *Vigna radiata* L. in the treatment of anemia. Further investigations are needed to understand the mechanism involved in the anti-anemic action of sprouted *Vigna radiata* L.

### **CONFLICT OF INTERESTS**

Declared None

#### REFERENCES

- Penninx BW, Guralnik JM, Onder G, Ferrucci L, Wallace RB, Pahor M. Anemia and decline in physical performance among older persons. Am J Med 2003;11:104–10.
- Baker SJ, DeMaeyer EM. Nutritional anemia: its understanding and control with special reference to the work of the world health organization. Am J Clin Nutr 1979;3:368-417.
- 3. Tulchisky TH, Varavikova. New Public Health. 2<sup>nd</sup>edn. Elsevier Academic press: London. UK; 2009. p. 193-6.
- Herifinda ET, Gourley DR. Textbook of therapeutics, drug and disease management. 6<sup>th</sup>edn. William and Wilkins, Baltimore, USA; 1996. p. 198-226.
- 5. Ravi U Thaker, Bhavin A Vyas, Shrikant V Joshi, Paras K Patel, Dinesh R Shah. Effect of dehydrated water extract of fruits of

*Opuntia ficus indica* on experimentally induced hemolytic anemia in rats. Int J Pharm Res Dev 2012;4:185-91.

- Min L. Research advance in chemical composition and pharmacological action of mung bean. Shanghai J Trad Chin Med 2001;5:18.
- 7. Fery RL. The cowpea: production, utilization, and research in the United States. Hortic Rev 1990;12:197–222.
- Soobratte MA, Neergheen VS, Luximon-Ramma A. Phenolic as potential antioxidant therapeutic agents: mechanism and actions. Mutat Res 2005;579:200-13.
- Nair RM, Yang RY, Easdown WJ, Thavarajah D, Thavarajah P, Hughes Jd, *et al.* Biofortification of mungbean (*Vigna radiata*) as a whole food to enhance human health. J Sci Food Agric 2013;8:1805-13.
- 10. Yang RY, Tsou SCS. Mungbean as a potential iron source in south Asian diets. In 35 International consultation workshop on mungbean. AVRDC: Shanhua; 1998. p. 152-8.
- 11. Wongekalak LSP, Jirasripongpun K, Hongsprabhas P. Potential use of antioxidativemungbean protein hydrolysate as an anticancer asiatic acid carrier. Food Res Int 2011;44:812–7.
- Lai F, Wen Q, Li L, Wu H, Li X. Antioxidant activities of watersoluble polysaccharide extracted from mung bean (*Vigna radiata* L.) hull with ultrasonic assisted treatment. Carbohydr Polym 2010;81:323–9.
- Lee JH, Jeon JK, Kim SG, Kim SH, Chun T, Imm JY. Comparative analyses of total phenols, flavonoids, saponins and antioxidant activity in yellow soy beans and mung beans. Int J Food Sci Tech 2011;46:2513-9.
- 14. Kim JH, Lee BC, Kim JH, Sim GS, Lee DH, Lee KE, *et al*. The isolation and antioxidative effects of vitexin from acerpalmatum. Arch Pharm Res 2005;28:195–202.
- Wang S, Rao P, Ye X. Isolation and biochemical characterization of a novel leguminous defense peptide with antifungal and antiproliferative potency. Appl Microbiol Biotechnol 2009;82:79–86.
- Wang S, Shao B, Fu H, Rao P. Isolation of a thermostable legume chitinase and study on the antifungal activity. Appl Microbiol Biotechnol 2009;85:313–21.
- Wang PF, Ye SY, Rao XY. Research progress on the biological activities and functions of mung beans. J Chin Inst Food Sci Technol 2004;1:26.
- Ye XY, Ng TB. Mungin, a novel cyclophilin-like antifungal protein from the mung bean. Biochem Biophys Res Commun 2000;273:1111–5.
- Wang S, Wu J, Rao P, Ng TB, Ye X. A chitinase with antifungal activity from the mung bean. Protein Expression Purif 2005;40:230–6.
- Lee SJ, Lee JH, Lee HH, Lee S, Kim SH, Chun T, *et al.* Effect of mung bean ethanol extract on pro-inflammtory cytokines in LPS stimulated macrophages. Food Sci Biotechnol 2011;20:519–24.
- Yeap SK, AliN M, Yusof HM, Noorjahan BA, Boon KB, Wan YH, et al. Antihyperglycemic effects of fermented and nonfermentedmung bean extracts on alloxan-induced-diabetic mice. BioMed Res Int 2012;1–7. doi.org/10.1155/2012/ 285430. [Article in Press]
- 22. Bellik Y, Hammoudi S, Abdellah F, Iguer-Ouada M, Boukraa L. Phytochemicals to prevent inflammation and allergy. Recent Pat Inflammation Allergy Drug Discovery 2012;6:147–58.
- 23. Cherng JM, Chiang W, Chiang LC. Immunomodulatory activities of edible beans and related constituents from soybean. Food Chem 2007;104:613–8.
- 24. Yao Y, Chen F, Wang M, Wang J, Ren G. Antidiabetic activity of mung bean extracts in diabetic KK-Ay mice. J Agric Food Chem 2008;56:8869–73.
- Randhir R, Shetty K. Mung beans processed by solid-state bioconversion improves phenolic content and functionality relevant for diabetes and ulcer management. Innovative Food Sci Emerging Technol 2007;8:197–204.
- Hsu GSW, Lu YF, Chang SH, Hsu SY. Antihypertensive effect of mung bean sprout extracts in spontaneously hypertensive rats. J Food Biochem 2011;35:278–88.
- 27. Matousek J, Podzimek T, Pouckova P, Stehlik J, Skvor J, Soucek J, *et al.* Antitumor effects and cytotoxicity of recombinant plant nucleases. Oncol Res 2009;18:163–71.

- Xu B, Chang SK. Comparative study on antiproliferation properties and cellular antioxidant activities of commonly consumed food legumes against nine human cancer cells. Food Chem 2012;134:1287–96.
- 29. Zhao YR, Li ZW, Zhao C, Fu R, Wang XH, Li ZY. Effects of recombinant mung bean trypsin inhibitor fragments on migration of colon cancer cell SW480. J Shanxi Univ Nat Sci 2012;1:29.
- Zhu S, Li W, Li JH, Arvin J, Andrew ES, Wang HC. It is not just folklore: the aqueous extract of mung bean coat is protective against sepsis. J Evidence-Based Complementary Altern Med 2012:1–10. doi.org/10.1155/2012/498467. [Article in Press]
- Lee CH, Yoon SJ, Lee SM. Chlorogenic acid attenuates high mobility group Box 1 (HMGB1) and enhances host defense mechanisms in murine sepsis. Mol Med 2012;18:1437–48.
- Kushwah A, Rajawat P, Kushwah HS. Nutritional evaluation of extruded Faba bean (Vicia *faba* L.) as a protein supplement in cereals based diet in rats. J Exp Biol 2002;40:49–52.
- El-Adawy T, Rahma E, El-Bedawey A, El-Beltagy A. Nutritional potential and functional properties of germinated mung bean, pea and lentil seeds. Plant Foods Hum Nutr 2003;58:1–13.
- Golob P. The use of spices and medicinals as bioactive protectants for grains. Rome: FAO Agricultural Sciences Bulletin; 1999. p. 137.
- Diallo A, Gbeassor M, Ahoefa Vovor B, Kwashie Eklu-Gadegbeku A, Kodjo Aklikokou A, Amegnona Agbonon A, *et al.* Effect of *Tectona grandis* on phenylhydrazine-induced anemia in rats. Fitoterapia 2008;79:332-6.
- Dacie JV, Lewis SM. Practical Hematology. 4<sup>th</sup> edition. Churchhill, UK; 1968. p. 37.
- OcheiJ, Kolhatkar A. Medical laboratory science theory and practice. Tata McGraw-Hill Publishing Company Limited, New Delhi; 2000. p. 281-7.
- Beuge JA, Aust SD. The Thiobarbituric acid assay. Methods in enzymology 1978;52:306-7.
- 39. Moron MS, Dsepierre JW, Manerwik KB. Levels of glutathione, glutathione reductase and glutathione-S-transferase activities in rat lung and liver. Biophysica Acta 1979;582:67-8.
- 40. Reitman S, Frankel SA. Colorimetric method for the determination of serum Glutamic oxaloacetic and glutamic pyruvic transaminase. Am J Clin Pathol 1957;25:56-63.
- 41. Harvey J, Paige SM. The In stat guide to choosing and interpreting statistical tests: a manual for graph pad Instat San Diego, CA USA; 1998.

- Akyüz M. Nutritive value, flavonoid content and radical scavenging activity of the truffle. J Soil Sci Plant Nutr 2013;13:143-51.
- 43. Gannet PM, Lawson TS, Kolar C, Toth B. Aryl radical formation during the metabolism of aryl hydrazine by microsomes. Chem Res Toxicol 1997;10:1372-7.
- Prasad AS. Zinc and trace minerals. Workshop on nutrient metabolism in genetic anemia, Bethesoda, USA, NHLBI; 1999.
- 45. Ferrali M, Signorini C, Pompella A, Lodovic M, Caciotti B, Ciccoli L, *et al*. Release of free redox-active iron in the liver and DNA oxidative damage following phenyl hydrazine intoxication. Biochem Pharmacol 1997;53:1743-51.
- 46. Criswell KA, Sulkanen AP, Hochbaum AF, Bleavins MR. Effects of phenylhydrazine or phlebotomy on peripheral blood, bone marrow and Erythropoietin in Wistar rats. J Appl Toxicol 2000;20:25-34.
- 47. Andre Muller, Helene Jacobsen, Edel Healy, Sinead McMickan, FréderiqueIstace, Marie-Noëlle Blaude, *et al.* Hazard classification of chemicals inducing hemolytic anemia: an EU regulatory perspective. Regul Toxicol Pharmacol 2006;45:229-41.
- Shukla S, Mehta A, John J, Singh S, Mehta P, Vyas S. Clinical manifestations of human parvovirus B19 in adults. Arch Int Med 2012;149:11532-6.
- Tyurina YY, Shvedova AA, Kanai K, Tyurin VA, Kommineni C, Quinn PJ, *et al.* Phospholipid signaling in apoptosis. Peroxidation and externalization of Phosphatidylserine. Toxicology 2000;148:93–101.
- Hochstein P. Perspectives on hydrogen peroxide and druginduced hemolytic anemia in glucose-6-phosphate dehydrogenase deficiency. Free Radical Biol Med 1988;5:387–92.
- McMillan DC, Powell CL, Bowman ZS, Morrow JD, Jollow DJ. Lipids versus proteins as major targets of per-oxidant, directacting hemolytic agents. Toxicol Sci 2005;88:274-83.
- Haram K, Nilsen ST, Ulvik RJ. Iron supplementation in pregnancy--evidence and controversies. Acta Obstet Gynecol Scand 2001;80:683-8.
- Vyshtakaliuk AB, Zobov VV. Antianemic activity of water-soluble Na, Ca, Fe-polygalacturonate. Bull Exp Biol Med 2010;150:45-7.
- Mittal MB, Abhay RS, Yadunath MJ, Vilasrao JK. An intervention on iron deficiency anemia and change in dietary behavior among adolescent girls. Int J Pharm Pharm Sci 2011;3:40-2.
- 55. Balaji K, Brindha P, Sridharan K, Uma Maheshwari K, Swaminathan S, Rajan KS. Elucidation of a core-shell model for Lauhabhasma through physico-chemical characterization. Int J Pharm Pharm Sci 2012;4:644-9.