

ORIGINAL ARTICLE

Biomarker analyses in the phase III ASCENT study of sacituzumab govitecan versus chemotherapy in patients with metastatic triple-negative breast cancer[☆]

A. Bardia¹, S. M. Tolaney², K. Punie³, D. Loirat⁴, M. Oliveira⁵, K. Kalinsky^{6,7}, A. Zelnak⁸, P. Aftimos⁹, F. Dalenc¹⁰, S. Sardesai¹¹, E. Hamilton¹², P. Sharma¹³, S. Recalde¹⁴, E. C. Gil¹⁵, T. Traina¹⁶, J. O'Shaughnessy¹⁷, J. Cortes¹⁸, M. Tsai¹⁹, L. Vahdat²⁰, V. Diéras²¹, L. A. Carey²², H. S. Rugo²³, D. M. Goldenberg^{24,25}, Q. Hong^{24,26}, M. Olivo^{24,26}, L. M. Itri^{24,26} & S. A. Hurvitz^{27*}

¹Massachusetts General Hospital, Harvard Medical School, Boston; ²Medical Oncology, Dana-Farber Cancer Institute, Boston, USA; ³Department of General Medical Oncology and Multidisciplinary Breast Centre, Leuven Cancer Institute, University Hospitals Leuven, Leuven, Belgium; ⁴Medical Oncology Department and D3i, Institut Curie, Paris, France; ⁵Hospital Universitari Vall d'Hebron, Barcelona, Spain; ⁶Columbia University Irving Medical Center, New York; ⁷Winship Cancer Institute, Emory University, Atlanta; ⁸Northside Hospital, Atlanta, USA; ⁹Institut Jules Bordet — Université Libre de Bruxelles, Brussels, Belgium; ¹⁰Institut Claudius Regaud, Toulouse, France; ¹¹The Ohio State University Wexner Medical Center, Columbus; ¹²Sarah Cannon Research Institute/Tennessee Oncology, Nashville; ¹³University of Kansas Medical Center, Westwood, USA; ¹⁴Institut Catala d'Oncologia Hospitalet, Barcelona; ¹⁵Hospital Universitario 12 de Octubre, Madrid, Spain; ¹⁶Memorial Sloan Kettering Cancer Center, New York; ¹⁷Baylor University Medical Center, Texas Oncology, US Oncology, Dallas, USA; ¹⁸International Breast Cancer Center (IBCC), Quiron Group, Madrid & Barcelona, Spain; ¹⁹VPCI Oncology Research, Minneapolis; ²⁰MSK-Norwalk Hospital Partnership, Norwalk, USA; ²¹Centre Eugène Marquis, Rennes, France; ²²University of North Carolina Lineberger Comprehensive Cancer Center, Chapel Hill; ²³University of California San Francisco Comprehensive Cancer Center, San Francisco; ²⁴Immunomedics, Inc., Morris Plains; ²⁵Center for Molecular Medicine and Immunology, Mendham; ²⁶Department of Clinical Development, Gilead Sciences, Inc., Morris Plains; ²⁷Department of Medicine, Division of Hematology/Oncology, David Geffen School of Medicine, University of California, Los Angeles, Jonsson Comprehensive Cancer Center, Los Angeles, USA



Available online 8 June 2021

Background: The pivotal phase III ASCENT trial demonstrated improved survival outcomes associated with sacituzumab govitecan (SG), an anti-trophoblast cell-surface antigen 2 (anti-Trop-2) antibody-drug conjugate linked with the topoisomerase-inhibitor SN-38, over single-agent chemotherapy treatment of physician's choice (TPC) in previously treated metastatic triple-negative breast cancer (mTNBC). This prespecified, exploratory biomarker analysis from the ASCENT trial evaluates the association between tumor Trop-2 expression and germline *BRCA1/2* mutation status with clinical outcomes.

Patients and methods: Patients with mTNBC refractory to or progressing after two or more prior chemotherapies, with one or more in the metastatic setting, were randomized to receive SG (10 mg/kg intravenously days 1 and 8, every 21 days) or TPC (capecitabine, eribulin, vinorelbine, or gemcitabine) until disease progression/unacceptable toxicity. Biopsy or surgical specimens were collected at study entry to determine Trop-2 expression level using a validated immunohistochemistry assay and histochemical scoring. Germline *BRCA1/2* mutation status was collected at baseline.

Results: Of 468 assessable patients, 290 had Trop-2 expression data [64% ($n = 151$ SG) versus 60% ($n = 139$ TPC)] and 292 had known *BRCA1/2* mutation status [63% ($n = 149$ SG) versus 61% ($n = 143$ TPC)]. Median progression-free survival in SG- versus TPC-treated patients was 6.9, 5.6, and 2.7 months versus 2.5, 2.2, and 1.6 months for high, medium, and low Trop-2 expression, respectively. Median overall survival (14.2, 14.9, and 9.3 months versus 6.9, 6.9, and 7.6 months) and objective response rates (44%, 38%, and 22% versus 1%, 11%, and 6%) were numerically higher with SG versus TPC in patients with high, medium, and low Trop-2 expression, respectively. Efficacy outcomes were numerically higher with SG versus TPC in patients with and without germline *BRCA1/2* mutations.

Conclusions: SG benefits patients with previously treated mTNBC expressing high/medium Trop-2 compared with standard-of-care chemotherapy and regardless of germline *BRCA1/2* mutation status. The small number of patients with low Trop-2 expression precludes definitive conclusions on the benefit of SG in this subgroup.

Key words: triple-negative breast cancer, trophoblast cell-surface antigen 2, BRCA

*Correspondence to: Dr Sara A. Hurvitz, Department of Medicine, Division of Hematology/Oncology, David Geffen School of Medicine, University of California, Los Angeles, 10945 Le Conte Ave, PVUB Suite 3360, Los Angeles, CA 90095, USA. Tel: +1-310-998-4747
E-mail: shurvitz@mednet.ucla.edu (S. A. Hurvitz).

[☆]Note: Information in this study was previously presented orally in part at the 43rd San Antonio Breast Cancer Virtual Symposium; 8-11 December 2020; San Antonio, Texas; Abstract GS3-06.

0923-7534/© 2021 The Authors. Published by Elsevier Ltd on behalf of European Society for Medical Oncology. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

INTRODUCTION

Precision medicine has gained momentum in recent years, with increasing attention given to the identification of key biomarkers that predict response to treatment. Triple-negative breast cancer (TNBC) is a heterogeneous disease with distinct molecular subtypes, resulting in unique drug sensitivities.^{1,2} A potential biomarker for response in TNBC is trophoblast cell-surface antigen 2 (Trop-2), a transmembrane glycoprotein calcium signal transducer expressed in human epithelial cells.³ Increased expression of Trop-2 is associated with tumor growth in a variety of solid epithelial tumors, including TNBC and other breast cancer subtypes.³⁻⁵ Elevated levels of membrane Trop-2 are associated with poor prognosis and increased tumor growth in breast cancer, including decreased survival.^{3,5}

Sacituzumab govitecan (SG) is a novel, Trop-2-directed antibody–drug conjugate comprising a humanized anti-Trop-2 IgG₁ kappa antibody coupled to an SN-38 payload, the active metabolite of the topoisomerase 1 inhibitor irinotecan, via a proprietary, hydrolysable linker. SG is distinct from other antibody–drug conjugates due to its high antibody specificity for Trop-2, a high ratio of drug to antibody (7.6: 1),⁶ and delivery of SN-38 in its most active, nonglucuronidated form.⁷ Following SG administration, the anti-Trop-2 monoclonal antibody binds to Trop-2 expressed on the tumor cell surface, allowing internalization and targeted delivery of SN-38 to tumor cells.^{6,8-10} Its proprietary linker also allows SN-38 to be liberated in the tumor microenvironment, enabling antitumor effects (bystander effect) without prerequisite internalization and enzymatic cleavage of SN-38 from the anti-Trop-2 antibody.^{6,8,11}

As *BRCA1* or *BRCA2* (*BRCA1/2*) confer a deficiency in homologous recombination repair of double-stranded DNA breaks, there is interest in *BRCA1/2* as a potential biomarker of response for therapy regimens that target DNA damage, particularly for TNBC.¹² Approximately 15% of patients with TNBC have germline *BRCA* mutations, a higher prevalence compared with other breast cancer subtypes.¹³⁻¹⁵ Topoisomerase I inhibitors like SN-38, the payload in SG, have been shown to increase double-stranded DNA breaks, regardless of *BRCA* mutation status.¹⁴ SG has demonstrated inhibition of tumor growth in translational models of *BRCA*-mutated TNBC and may confer synthetic lethality to TNBC tumors.¹⁴

A phase I/II single-arm basket study (NCT01631552) was conducted of SG in patients with metastatic, epithelial cancers. In the cohort of 108 patients with heavily pre-treated metastatic TNBC (mTNBC), an objective response rate (ORR) of 33%, median progression-free survival (PFS) of 5.5 months, and median overall survival (OS) of 13.0 months were observed.¹⁶ In this cohort of patients with mTNBC, 88% of 48 primary or mTNBC tumors had moderate to strong Trop-2 staining, with the majority expressing Trop-2 in >50% of tumor cells. All responders had moderate to strong Trop-2 staining, demonstrating the

potential for greater benefit of SG in tumors with relatively high Trop-2 expression;¹⁷ however, the number of samples with low or no Trop-2 staining were limited ($n = 6$), highlighting the need for further evaluation of Trop-2 in a larger dataset. The safety and efficacy results from this trial led to an accelerated approval by the United States Food and Drug Administration (FDA), with full approval received based on the results of the phase III ASCENT trial (NCT02574455).

The randomized phase III ASCENT trial of SG versus single-agent chemotherapy treatment of physician's choice (TPC; eribulin, vinorelbine, gemcitabine, or capecitabine) in 468 patients with chemotherapy-pretreated mTNBC confirmed the initial findings from the phase I/II study. Treatment with SG was associated with a significant survival benefit compared with TPC with a median PFS of 5.6 versus 1.7 months {hazard ratio (HR) 0.41 [95% confidence interval (CI) 0.32-0.52]; $P < 0.001$ } and a median OS of 12.1 versus 6.7 months [HR, 0.48 (95% CI 0.38-0.59); $P < 0.001$], along with a tolerable safety profile. Here, we present a pre-specified biomarker assessment of the potential association between tumor membrane Trop-2 expression or germline *BRCA1/2* mutation status on efficacy of SG versus TPC in the phase III ASCENT study.

METHODS

Study design

The study design for ASCENT (NCT02574455) has been described previously.¹⁸ Briefly, ASCENT was an international, multicenter, randomized, phase III study comparing the efficacy and safety of SG versus TPC in patients in the second-line or greater mTNBC setting. Patients were stratified at randomization by the number of prior chemotherapy regimens for advanced disease (2-3 versus >3), presence of known brain metastases at baseline (yes versus no), and geography (North America versus rest of world).

The primary endpoint was PFS (by blinded independent central review) in patients without known baseline brain metastases (measured by computed tomography or magnetic resonance imaging per RECIST version 1.1).¹⁹ Secondary endpoints included investigator-assessed PFS for the full population, including all randomized patients with and without brain metastases, OS, ORR, duration of response, time to response, and safety. Exploratory endpoints included biomarker assessment.

The ASCENT trial was conducted and approved by each investigational site institutional review board/ethics committee before initiation, and in accordance with the Declaration of Helsinki, International Council for Harmonisation Guidelines for Good Clinical Practice, FDA Code of Federal Regulations, national and local drug and data protection laws, and other applicable regulatory requirements. All patients provided written informed consent before enrollment.

Patients

Patients had mTNBC that had progressed following two or more prior standard chemotherapy regimens (no upper limit) for unresectable, locally advanced, or metastatic disease, and included a taxane (any setting). Per protocol, patients were also eligible after only one prior regimen in the metastatic setting if their disease recurred within 12 months of completing (neo)adjuvant therapy. Eligible patients had TNBC according to standard American Society of Clinical Oncology/College of American Pathologists criteria.²⁰ Patients with stable brain metastases for at least 4 weeks before treatment were eligible, but were excluded from evaluation of the primary endpoint and this exploratory analysis.

Sample collection and assay procedure

Primary or metastatic archival biopsy or surgical specimens were requested at study entry to determine tumor Trop-2 expression; however, known Trop-2 expression was not required to determine patient eligibility. Trop-2 expression was determined by using a validated immunohistochemistry (IHC) assay (OptiVIEW DAB detection kit; Roche Diagnostics, Indianapolis, IN) as specified according to manufacturer instructions, including quality control methods and by using Trop-2 mouse monoclonal antibody (ENZ ABS380-0100; ENZO Life Sciences, Farmingdale, NY). IHC was carried out centrally (Laboratory Corporation of America, Research Triangle Park, NC), and interpretation of Trop-2 staining was carried out by a qualified pathologist.

Tumor cell membrane Trop-2 expression was categorized based on a histochemical score (H-score), a numerical value represented by a weighted summation of percent staining which accounts for both the staining intensity and the percentage of cells at that intensity. H-scores were calculated using the following formula: $H\text{-score} = (3 \times \% \text{ cells with strong intensity staining}) + (2 \times \% \text{ cells with moderate intensity staining}) + (1 \times \% \text{ cells with mild intensity staining})$.²¹ The categories were selected based on the distribution of the H-score (range from 0 to 300) to divide the population into low, medium, and high groups. The following Trop-2 expression categories were used: H-score 0 to <100: Trop-2 low; H-score 100-200: Trop-2 medium; H-score >200-300: Trop-2 high. The status of germline *BRCA1/2* mutations was collected at baseline, if known.

intensity staining) + (1 × % cells with mild intensity staining).²¹ The categories were selected based on the distribution of the H-score (range from 0 to 300) to divide the population into low, medium, and high groups. The following Trop-2 expression categories were used: H-score 0 to <100: Trop-2 low; H-score 100-200: Trop-2 medium; H-score >200-300: Trop-2 high. The status of germline *BRCA1/2* mutations was collected at baseline, if known.

Statistical analyses

Subgroup analyses of PFS, OS, and ORR by biomarker were carried out. Efficacy analyses were based on data from the primary study analysis (cutoff date 11 March 2020) of the brain metastases negative patient population.¹⁸ Only patients with known Trop-2 or *BRCA1/2* results are included in the analysis. The analyses were exploratory in nature, with no adjustment for multiple testing and no formal testing of benefit of SG versus TPC subgroups.

PFS was defined as the time from randomization until objective tumor progression or death or censored at the last radiographic assessment for patients without progression or death. PFS and OS were analyzed using the Kaplan–Meier method, with medians and corresponding 95% CIs determined according to the Brookmeyer and Crowley method with log-log transformation. CIs for ORR were calculated by the Clopper-Pearson method.

RESULTS

Patients

As reported previously, 529 patients were enrolled in ASCENT (Figure 1); 61 patients had brain metastases at baseline and 468 patients had no history of brain metastases. Of the patients who were negative for brain metastases (primary efficacy dataset) and included in this analysis, 235 patients were randomized to receive SG, and 233

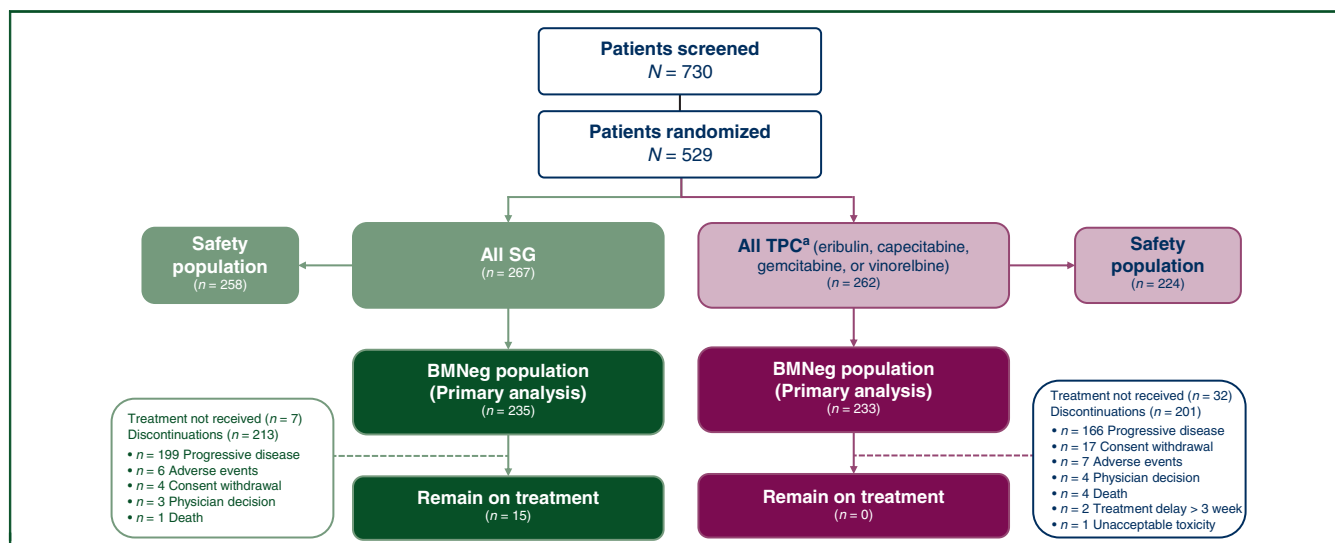


Figure 1. CONSORT diagram: enrollment, intent-to-treat and safety populations, and patient withdrawals.

From *The New England Journal of Medicine*, Bardia A, Hurvitz SA, Tolanev SM, et al: Sacituzumab govitecan in metastatic triple-negative breast cancer. Vol. 384, pp 1529-1541, 2021. Copyright © (2021) Massachusetts Medical Society. <https://doi.org/10.1016/j.jannonc.2021.06.002> article. Reprinted with permission.

BMNeg, brain metastases negative; SG, sacituzumab govitecan; TPC, treatment of physician's choice.

^a Patients in the TPC arm were randomized to eribulin ($n = 139$), vinorelbine ($n = 52$), gemcitabine ($n = 38$), or capecitabine ($n = 33$).

patients were randomized to receive TPC (54% eribulin, 20% vinorelbine, 13% capecitabine, and 12% gemcitabine). Patients in the SG- versus TPC-treated cohorts had a median age of 54 years (range, 29-82 years) and 53 years (range, 27-81 years; Table 1), respectively. The most common prior chemotherapy received in the SG- versus TPC-treated cohorts, respectively, was cyclophosphamide (82% each), carboplatin (63% versus 69%), and capecitabine (63% versus 68%); all patients received a prior taxane. The median number of prior anticancer regimens was 4 (range, 2-17).

Demographics and baseline characteristics of patients with known and unknown Trop-2 expression and those of patients with known or unknown *BRCA1/2* mutational status are presented in Supplementary Tables S1 and S2, respectively, available at <https://doi.org/10.1016/j.annonc.2021.06.002>. Demographics and baseline characteristics of patients included in the biomarker analyses were well-balanced across treatment arms. Patients who were not included in the biomarker analyses due to unknown Trop-2 expression or *BRCA1/2* mutational status generally had similar characteristics as those who were included. Compared with patients who were included in the analyses, minor differences were observed for prior use of checkpoint inhibitors in patients who were not included in the Trop-2 analysis, and for median age and prior use of checkpoint inhibitors in patients who were not included in the *BRCA1/2* analysis. Trop-2 expression levels in patients who did not have TNBC at initial breast cancer diagnosis were broadly similar to those of patients with TNBC at initial breast cancer diagnosis (data not shown).

Among 235 patients who were treated with SG, 151 (64%) had archival tumors evaluated for Trop-2 expression. Of these, the majority of patients had a high H-score [$n = 85$ (56%)], with a medium and low H-score found in 26% ($n = 39$) and 18% ($n = 27$) of patients, respectively. In the SG arm, there were seven patients in the low H-score group who had no Trop-2 expression. A similar distribution of Trop-2 expression was observed in patients who received single-agent chemotherapy with high, medium, and low H-scores of 52%, 25%, and 23%, respectively. In the TPC arm, there were four patients in the low H-score group who had no Trop-2 expression. Data on Trop-2 expression in primary versus metastatic tumors were not collected.

Germline *BRCA1/2* mutational status was known in 149 (63%) patients who received SG and in 143 (61%) of patients who received TPC. Of those with known germline *BRCA1/2* status, there were 16 of 149 patients (11%) who were *BRCA1/2*-positive (germline pathogenic variants in either *BRCA1* or *BRCA2*) and 133 of 149 patients (89%) who were *BRCA1/2*-negative (also considered to be germline *BRCA* wild type) in the SG arm, and 18 of 143 (13%) *BRCA1/2*-positive and 125 of 143 (87%) *BRCA1/2*-negative patients in the TPC arm.

Trop-2 expression and efficacy outcomes

SG-treated patients with high, medium, and low Trop-2 H-scores had median PFS of 6.9 months (95% CI 5.8-7.4

Table 1. Demographics assessed in patients who were negative for brain metastases

	SG (n = 235)	TPC (n = 233)
Median age (range), years	54 (29-82)	53 (27-81)
Female, n (%)	233 (99)	233 (100)
Race or ethnic group, n (%)		
White	188 (80)	181 (78)
Black	28 (12)	28 (12)
Asian	9 (4)	9 (4)
Other or not specified	10 (4)	15 (6)
ECOG PS, n (%)		
0	108 (46)	98 (42)
1	127 (54)	135 (58)
<i>BRCA1/2</i> mutational status, n (%)	149 (63)	143 (61)
Positive	16 (7)	18 (8)
Negative	133 (57)	125 (54)
Trop-2 expression, n (%)	151 (64)	139 (60)
(High) H-score >200-300	85/151 (56)	72/139 (52)
(Medium) H-score 100-200	39/151 (26)	35/139 (25)
(Low) H-score 0 to <100 ^a	27/151 (18)	32/139 (23)
Initial diagnosis of TNBC, ^b n (%)		
Yes	165 (70)	157 (67)
No	70 (30)	76 (33)
Median previous anticancer regimens, ^c n (range)	4 (2-17)	4 (2-14)
Most common prior chemotherapy, n (%)		
Taxane ^d	235 (100)	233 (100)
Cyclophosphamide	192 (82)	192 (82)
Carboplatin	147 (63)	160 (69)
Capecitabine	147 (63)	159 (68)
Previous PARP inhibitor, n (%)	17 (7)	18 (8)
Previous use of checkpoint inhibitors, n (%)	67 (29)	60 (26)
Most common sites of disease, ^e n (%)		
Lung only	108 (46)	97 (42)
Liver	98 (42)	101 (43)
Bone	48 (20)	55 (24)

BRCA, breast cancer gene; ECOG PS, Eastern Cooperative Oncology Group performance status; H-score, histochemical score; PARP, poly (ADP-ribose) polymerase; SG, sacituzumab govitecan; TNBC, triple-negative breast cancer; TPC, treatment of physician's choice; Trop-2, trophoblast cell-surface antigen 2.

^a In this H-score group, seven and four patients in the SG and TPC arms, respectively, had no Trop-2 expression.

^b Patients in study either had TNBC at initial diagnosis or had hormone receptor-positive disease that converted to hormone-negative at time of study entry.

^c Anticancer regimens refer to any treatment regimen that was used to treat breast cancer in any setting.

^d Includes paclitaxel, paclitaxel albumin, and docetaxel.

^e Based on independent central review of target and non-target lesions at baseline.

months), 5.6 months (95% CI 2.9-8.2 months), and 2.7 months (95% CI 1.4-5.8 months), respectively. Compared with SG, TPC-treated patients had numerically lower median PFS across high (2.5 months; 95% CI 1.5-2.9 months), medium (2.2 months; 95% CI 1.4-4.3 months), and low (1.6 months; 95% CI 1.4-2.7 months) Trop-2 H-scores (Figure 2A). Median OS with SG treatment was 14.2 months (95% CI 11.3-17.5 months), 14.9 months (95% CI 6.9 months to not evaluable), and 9.3 months (95% CI 7.5-17.8 months) in patients with high, medium, and low Trop-2 scores, respectively. In patients with who received TPC, median OS was 6.9 months (95% CI 5.3-8.9 months), 6.9 months (95% CI 4.6-10.1 months), and 7.6 months (95% CI 5.0-9.6 months) for high, medium, and low Trop-2 H-scores, respectively; Figure 2B).

The ORR in SG-treated patients with high, medium, and low Trop-2 H-scores was 44%, 38%, and 22%, respectively. In comparison, the ORR in the TPC arm was 1%, 11%, and

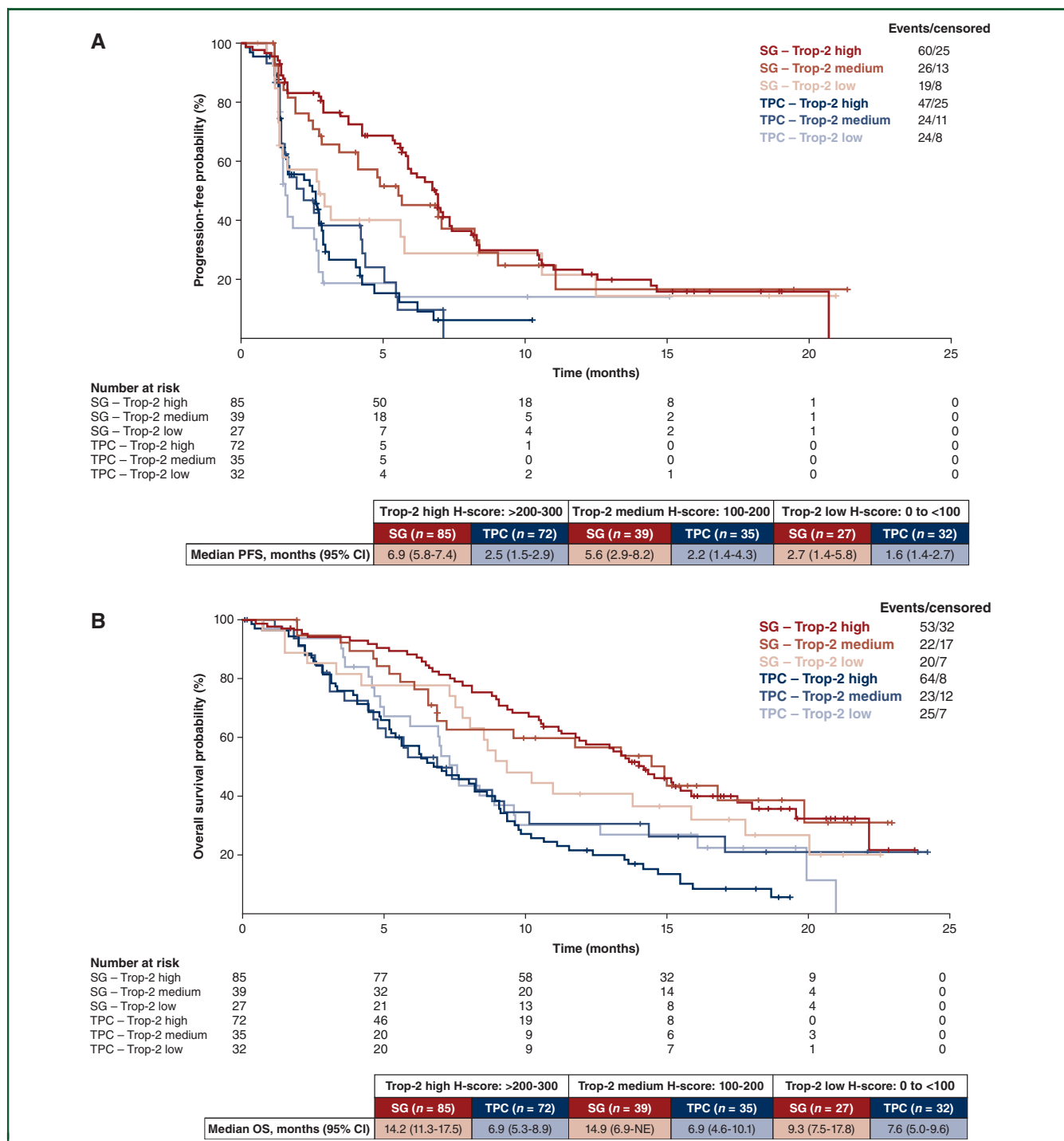


Figure 2. (A) Progression-free survival by Trop-2 expression. (B) Overall survival by trophoblast cell-surface antigen 2 (Trop-2) expression. CI, confidence interval; H-score, histochemical score; OS, overall survival; PFS, progression-free survival; SG, sacituzumab govitecan; TPC, treatment of physician’s choice; Trop-2, trophoblast cell-surface antigen 2.

6% in patients with high, medium, and low Trop-2 H-scores, respectively (Figure 3).

BRCA mutational status and efficacy outcomes

Compared with TPC, treatment with SG resulted in numerically higher median PFS, median OS, and response outcomes, regardless of germline BRCA1/2 mutation status

at study entry (Table 2). In BRCA1/2-positive patients, the median PFS was 4.6 versus 2.5 months in SG-treated versus TPC-treated patients, respectively; median OS was 15.6 versus 4.4 months in SG-treated versus TPC-treated patients, respectively. In BRCA1/2-negative patients, the median PFS was 4.9 and 1.6 months in SG-treated versus TPC-treated patients, respectively; median OS was 10.9 versus 7 months in SG-treated versus TPC-treated patients,

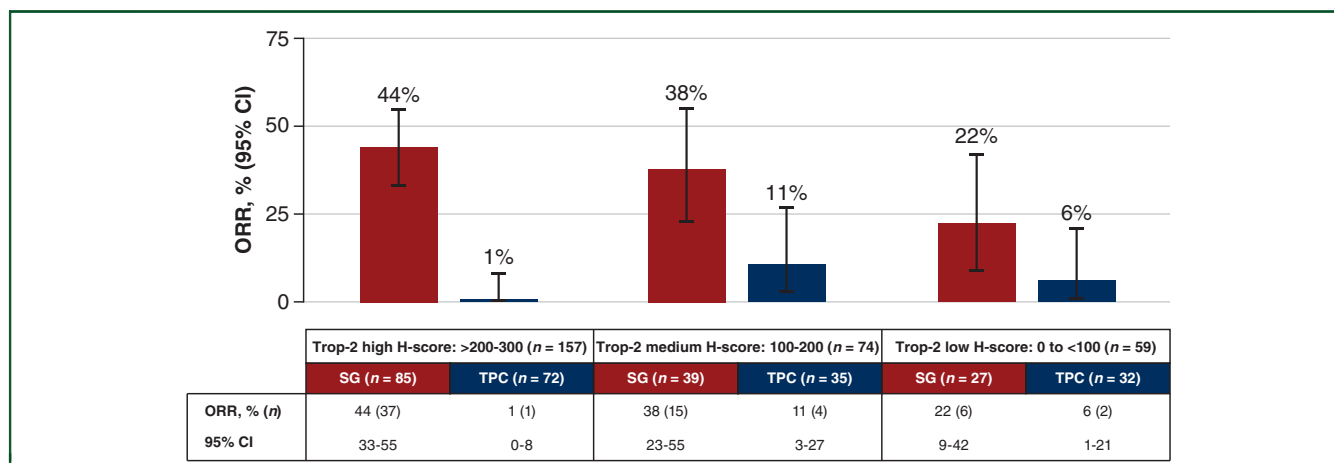


Figure 3. Objective response rate by Trop-2 expression.

CI, confidence interval; H-score, histochemical score; ORR, objective response rate; SG, sacituzumab govitecan; TPC, treatment of physician's choice; Trop-2, trophoblast cell-surface antigen 2.

	Germline BRCA1/2-positive		Germline BRCA1/2-negative	
	SG (n = 16)	TPC (n = 18)	SG (n = 133)	TPC (n = 125)
ORR, n (%)	3 (19)	1 (6)	44 (33)	7 (6)
Odds ratio (95% CI)	3.9 (0.4-42.2)		8.3 (3.6-19.4)	
Median PFS, months (95% CI)	4.6 (1.3-10.3)		4.9 (3.8-5.9)	
HR (95% CI)	0.6 (0.2-1.6)		0.4 (0.3-0.6)	
Median OS, months (95% CI, months)	15.6 (6.2-NE)		10.9 (9.6-13.4)	
HR (95% CI)	0.4 (0.2-0.9)		0.5 (0.4-0.7)	

BRCA, breast cancer gene; CI, confidence interval; HR, hazard ratio; NE, not evaluable; ORR, objective response rate; OS, overall survival; PFS, progression-free survival; SG, sacituzumab govitecan; TPC, treatment of physician's choice.

respectively. In BRCA1/2-positive patients, ORR was 19% versus 6% in SG-treated versus TPC-treated patients, respectively. In BRCA1/2-negative patients, the ORR was 33% versus 6% in SG-treated versus TPC-treated patients, respectively.

DISCUSSION

In the pivotal phase III ASCENT study, SG demonstrated improved outcomes compared with TPC (eribulin, vinorelbine, gemcitabine, or capecitabine), thereby becoming the first Trop-2-directed antibody–drug conjugate to show a significant PFS and OS benefit compared with standard-of-care chemotherapy in the second-line or greater mTNBC setting.¹⁸ These efficacy data, however, were based on a patient population unselected for Trop-2 expression. In this prespecified biomarker analysis, we assessed efficacy outcomes by tumor membrane Trop-2 expression and BRCA1/2 mutational status. Our results show the SG arm had numerically higher efficacy outcomes in high and medium Trop-2 expression subgroups compared with those in the TPC arm. The benefit of SG over TPC was also similar in BRCA1/2-positive and -negative patients.

Although Trop-2 is overexpressed across a variety of epithelial cancers, overexpression is more common in TNBC compared with other breast cancer subtypes.²² Preclinical studies of TNBC demonstrated that expression levels of

Trop-2 and homologous recombination repair deficiency can influence the antitumor activity of SG; SG provided higher activity in preclinical models with higher tumor Trop-2 expression and homologous recombination deficiency.²³ In addition, the presence of intratumoral heterogeneity in Trop-2 expression in clinical breast cancer samples and the possibility for Trop-2 plasticity in the tumor microenvironment highlight the need to assess the clinical impact of SG in TNBC tumors with varying Trop-2 expression.²⁴ The availability of the ASCENT clinical dataset allowed for assessment of SG efficacy in patients with varying tumor Trop-2 expression and the potential for Trop-2 as a biomarker of response in TNBC.

In this study, outcomes among Trop-2 subgroups were numerically higher with SG versus TPC treatment in patients with high and medium Trop-2 expression; SG-treated patients with high/medium Trop-2 expression had similar median PFS, median OS, and response rates as SG-treated patients in the overall primary analysis from ASCENT.¹⁸ As the majority of breast cancers express Trop-2,^{3,4} the number of patients with low Trop-2-expression tumors was small. These results suggest evaluating Trop-2 expression may not be needed to predict which patients are unlikely to derive benefit from SG versus TPC, but the small numbers of patients in the low Trop-2 expression groups and absence of formal testing limit definitive conclusions on the benefit of

SG (or lack thereof) in patients with low Trop-2 expression. Additional studies are needed to address whether higher Trop-2 expression is predictive of better response to SG and if primary tumor Trop-2 expression is predictive of response to SG in mTNBC. Finally, although high Trop-2 expression has previously been associated with poor prognosis in breast cancer,^{3,5} this study did not suggest an adverse prognostic effect of high Trop-2 expression in either treatment arm.

Because the *BRCA1* and *BRCA2* genes encode proteins involved in double-stranded DNA break repair, it is hypothesized that cancers with pathogenic variants in *BRCA1/2* may be more sensitive to chemotherapies that cause DNA damage, such as platinum-based regimens.²⁵⁻²⁷ Due to the strong association of TNBC with germline *BRCA1/2* mutations,^{13,14,28} we assessed whether TNBC with germline pathogenic variants in *BRCA1/2* had increased sensitivity to or was a predictive biomarker for response to SG, which primarily exerts its antitumor activity by eliciting DNA damage. In our analysis, there did not appear to be a difference in the effect size of SG between *BRCA1/2*-positive and -negative patients. Due to the small numbers of patients who were *BRCA1/2*-positive, further comparison of efficacy outcomes with *BRCA1/2*-negative patients was not carried out. However, DNA repair may be altered through other mechanisms beyond germline mutations, and additional research is needed into biomarkers of response and resistance for SG. As we are unable to exclude secondary reversion mutations and due to the advanced setting of the ASCENT trial, these data are not sufficient to draw definitive conclusions on the predictive value of germline *BRCA* mutations for SG in advanced TNBC.

The primary limitations of this study are that ASCENT was not powered to detect predictive effects, and there was no formal testing carried out between treatment arms in this subanalysis. In addition, the number of patients in both the low Trop-2 expression subgroup and the germline *BRCA1/2*-positive subgroup were small, which limited meaningful interpretation of the data when compared with other subgroups. Formal statistical analysis of Trop-2 expression and various baseline patient characteristics to determine a potential correlation was not carried out, due to the variability of patient characteristics and methodological concerns about exploratory non-hypothesis-based *post hoc* analyses. These data also reflect the current measurement of Trop-2 expression by IHC, and future assays may be developed to better quantify its expression. An additional limitation is that these data did not assess somatic *BRCA1/2* mutations in germline *BRCA1/2*-negative patients or mutations in other genes associated with homologous recombination, such as *PALB2*. Finally, differences in Trop-2 expression in primary breast cancers compared with metastatic lesions were not assessed and could be different. Most samples were archival tumor samples, and the biopsies were not from baseline when patients started therapy, so it is uncertain if the Trop-2 expression was reflective of the baseline state when the patient went on study, especially as Trop-2 may be a dynamic biomarker.

Conclusion

The collective results from the current study suggest SG benefits patients with previously treated mTNBC expressing high/medium Trop-2 compared with standard-of-care single-agent chemotherapy and regardless of germline *BRCA1/2* mutation status. Further studies are warranted to fully elucidate outcomes among these patient populations and those with low Trop-2 expression. Currently, studies are ongoing to further evaluate SG in earlier-stage TNBC and human epidermal growth factor receptor 2 (HER2)-negative breast cancer (NeoSTAR, NCT04230109; SASCIA, NCT04595565), in combination with other agents for the treatment of mTNBC (SEASTAR, NCT03992131; MORPHEUS-TNBC, NCT03424005; NCT04468061), and for the treatment of hormone receptor-positive/HER2-negative metastatic breast cancer (TROPiCS-02, NCT03901339). Biomarker data from these studies will provide further insight into the relationship of Trop-2 expression and germline *BRCA1/2* mutation status as a potential predictive biomarker of response to SG in breast cancer.

ACKNOWLEDGEMENTS

We thank the patients and their caregivers for helping us realize the possibilities of this research. We thank the dedicated clinical trial investigators and their devoted team members participating in the ASCENT trial. William A. Wegener, MD, PhD, and Robert M. Sharkey, PhD, former employees of Immunomedics, contributed to the development of the ASCENT trial. We thank Sharon K. Wyhopen, PhD, for critical review of the manuscript and participation in discussions. Medical writing and editorial assistance were provided by Susan O'Donnell, PharmD, and Shala Thomas, PhD, CMPP, at Team 9 Science, and was funded by Immunomedics, Inc. a subsidiary of Gilead Sciences, Inc.

FUNDING

This work was supported by Immunomedics, Inc., a subsidiary of Gilead Sciences, Inc. (no grant number).

DISCLOSURE

AB: consultancy/advisory role with Biotheranostics Inc., Pfizer, Novartis, Genentech, Merck, Radius Health, Immunomedics, Taiho, Sanofi, Daiichi Sanyo/AstraZeneca, Puma, Philips, Eli Lilly, and Foundation Medicine; research funding from Genentech, Novartis, Pfizer, Merck, Sanofi, Radius Health, Immunomedics; travel/accommodations/expenses from Pfizer, Novartis, Genentech, Merck, Radius Health, Immunomedics, Taiho, and Sanofi. SMT: research funding from Bristol Myers Squibb, Eisai, Immunomedics, Genentech/Roche, Pfizer, Novartis, Nektar, Merck, AstraZeneca, Eli Lilly, and Exelixis. KP: consultancy/advisory role with AstraZeneca, Eli Lilly, Novartis, Pfizer, Pierre Fabre, Hoffmann-La Roche, Vifor Pharma, and European Centre for Clinical Research Training; speaker's bureau with Eli Lilly, Mundipharma, Novartis, Pfizer, and Hoffmann-La Roche; research funding from Sanofi; expert testimony for Hoffmann-La Roche; travel/accommodations/expenses

from AstraZeneca, Novartis, Pfizer, PharmaMar, and Hoffmann-La Roche. DL: consultancy/advisory role with Novartis, Merck Sharp & Dohme (MSD), and Roche. MO: research funding from Immunomedics, Roche, Genentech, PUMA Biotechnology, AstraZeneca, Seattle Genetics, Boehringer Ingelheim, and Novartis; expenses from Roche, Genentech, PUMA Biotechnology, AstraZeneca, Seattle Genetics, and Novartis; non-financial support from Roche, Pierre Fabre, Eisai, GP Pharma, Grünenthal, and Novartis. KK: employment (spouse) with Array Biopharma, Pfizer, and Grail; consultancy/advisory role with Immunomedics, Pfizer, Eisai, Eli Lilly, Amgen, and AstraZeneca; institutional research funding from Immunomedics, Novartis, Incyte, Genentech/Roche, Eli Lilly, Pfizer, Calithera Biosciences, Acetylon, Seattle Genetics, Amgen, Zentalis Pharmaceuticals, and CytomX Therapeutics; research funding (spouse) from Array Biopharma, Pfizer, and Grail. AZ: consultancy roles with Novartis and Pfizer. PA: travel/accommodations/expenses from Boehringer Ingelheim, MacroGenics, Amvure, Synthon, Servier, G1 Therapeutics, Roche, Novartis, Amgen, Radius, MSD, and Pfizer. SS: advisory role with Immunomedics, Novartis, and Biotheranostics. EH: consultancy/advisory role with Genentech/Roche, Boehringer Ingelheim, Novartis, Dantari, Eli Lilly, Merck, Puma Biotechnology, Silverback Therapeutics, CytomX, Pfizer, Mersana, Black Diamond, H3 Biomedicine, Daiichi Sankyo, AstraZeneca, Arvinas, Deciphera Pharmaceuticals, Eisai, and SeaGen; institutional research funding from OncoMed, Genentech/Roche, Zymeworks, Rgenix, ArQule, Clovis, Silverback Therapeutics, Millennium, Acerta Pharma, Sermonix Pharmaceuticals, Torque, Black Diamond, Karyopharm, Infinity Pharmaceuticals, Curis, Syndax, Novartis, Boehringer Ingelheim, Immunomedics, FujiFilm, Taiho, Deciphera, Fochon, Molecular Templates, Onconova Therapeutics, Dana-Farber Cancer Hospital, Hutchinson MediPharma, MedImmune, SeaGen, Puma Biotechnology, Compugen, TapImmune, Eli Lilly, Pfizer, H3 Biomedicine, Takeda, Merus, Regeneron, Arvinas, StemCentRx, Vera-stem, eFFECTOR Therapeutics, CytomX, InventisBio, Lycera, Mersana, Radius Health, AbbVie, Nucana, Leap Therapeutics, Zenith Epigenetics, Harpoon, Orinove, AstraZeneca, Tesaro, MacroGenics, EMD Serono, Daiichi Sankyo, Syros, Sutro, G1 Therapeutics, Merck, PharmaMar, Olema, Polyphor, Immunogen, Plexikon, Amgen, Akesobio Australia, and Shattuck Labs. PS: consultancy role with Immunomedics. ECG: consultancy role with Pfizer, Roche, AstraZeneca, Novartis, and Eli Lilly; speaker's bureau with Roche, Pfizer, and Eli Lilly. TT: consultancy/advisory role with Genentech/Roche, Pfizer, AstraZeneca, Merck, Puma Biotechnology, Advaxis, Celgene, Innocrin Pharma, Genomic Health, Bristol Myers Squibb, Samsung, Athenex, Aduro Biotech, Halozyme, Daiichi Sankyo, Ionis, and Seattle Genetics; speaker's bureau with Roche/Genentech; research funding from Eisai, Pfizer, Novartis, Innocrin Pharma, AstraZeneca, Astellas Pharma, Immunomedics, Genentech/Roche, Daiichi Sankyo, Carrick Pharm. JO: consultancy/advisory role with AbbVie, Agendia,

AstraZeneca, Bristol Myers Squibb, Celgene, Eisai, Genentech, Immunomedics, Jounce Therapeutics, Eli Lilly, Merck, Novartis, Pfizer, Puma Biotechnology, Roche, and Seattle Genetics. JC: consultancy/advisory role with Roche, Celgene, Cellestia, AstraZeneca, Biothera Pharmaceuticals, Merus, Seattle Genetics, Daiichi Sankyo, Erytech, Athenex, Polyphor, Eli Lilly, Servier, MSD, GlaxoSmithKline (GSK), Leuko, Bioasis, and Clovis Oncology; speaker's bureau for Roche, Novartis, Celgene, Eisai, Pfizer, Samsung Bioepis, Eli Lilly, MSD, and Daiichi Sankyo; institutional research funding from Roche, Ariad Pharmaceuticals, AstraZeneca, Baxalta GMBH/Servier Affaires, Bayer Healthcare, Eisai, F. Hoffman-La Roche, Guardant Health, MSD, Pfizer, Piquar Therapeutics, Puma C, Queen Mary University of London; ownership interest in MedSIR. LV: consultancy role with Berg Pharma, Seattle Genetics, Polyphor, Immunomedics, and Osmol Therapeutics; speaker's bureau with Eisai; contracted research with Roche, OncoTherapy Science, Arvinas, Genentech, Seattle Genetics, and Immunomedics. VD: consultancy/advisory role with Roche/Genentech, Novartis, Eli Lilly, Pfizer, AstraZeneca, Eisai, AbbVie, MSD, Daiichi Sankyo, and Seattle Genetics; speaker's bureau for Roche, Novartis, Pfizer, Eli Lilly, AstraZeneca, and Daiichi Sankyo; travel/accommodations/expenses from Roche, Novartis, Pfizer, Eli Lilly, AstraZeneca, and Daiichi Sankyo. LAC: royalties (spouse) from Falcon Therapeutics; research funding from Innocrin Pharma, Syndax, Immunomedics, Novartis, and NanoString Technologies; institutional research funding from Sanofi-Aventis, Novartis, G1 Therapeutics, Genentech/Roche, and GSK. HSR: consultancy/advisory role with Samsung and Puma; research funding from Pfizer, Novartis, Eli Lilly, Genentech/Roche, MacroGenics, OBI, Merck, Eisai, Immunomedics, Daiichi, Seattle Genetics, and Odonate; travel/accommodations/expenses from Daiichi, Mylan, Pfizer, Merck, AstraZeneca, Novartis, and MacroGenics. DMG: intellectual property rights/patent holder at Immunomedics; ownership interest with Immunomedics; other interests with Center for Molecular Medicine & Immunology. QH, MO, LMI: employment with Immunomedics. SAH: research funding from Ambrx, Amgen, Arvinas, Bayer, Daiichi Sankyo, Genentech/Roche, GSK, Immunomedics, Eli Lilly, MacroGenics, Novartis, Pfizer, OBI Pharma, Pieris, PUMA, Radius, Sanofi, Seattle Genetics, and Dignitana; ownership interest with NKMax. All other authors have declared no conflicts of interest.

DATA SHARING

Immunomedics, Inc., a subsidiary of Gilead Sciences, Inc. will provide the study protocol and statistical analysis plan with publication of this manuscript as well as post results on clinicaltrials.gov, as required.

REFERENCES

1. Lehmann BD, Bauer JA, Chen X, et al. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J Clin Invest*. 2011;121(7):2750-2767.

2. Lehmann BD, Pietsenpol JA. Identification and use of biomarkers in treatment strategies for triple-negative breast cancer subtypes. *J Pathol*. 2014;232(2):142-150.
3. Ambrogio F, Fornili M, Boracchi P, et al. Trop-2 is a determinant of breast cancer survival. *PLoS One*. 2014;9(5):e96993.
4. Trerotola M, Cantanelli P, Guerra E, et al. Upregulation of Trop-2 quantitatively stimulates human cancer growth. *Oncogene*. 2013;32(2):222-233.
5. Goldenberg DM, Stein R, Sharkey RM. The emergence of trophoblast cell-surface antigen 2 (TROP-2) as a novel cancer target. *Oncotarget*. 2018;9(48):28989-29006.
6. Goldenberg DM, Cardillo TM, Govindan SV, Rossi EA, Sharkey RM. Trop-2 is a novel target for solid cancer therapy with sacituzumab govitecan (IMMU-132), an antibody-drug conjugate (ADC). *Oncotarget*. 2015;6(26):22496-22512.
7. Sharkey RM, McBride WJ, Cardillo TM, et al. Enhanced delivery of SN-38 to human tumor xenografts with an anti-Trop-2-SN-38 antibody conjugate (sacituzumab govitecan). *Clin Cancer Res*. 2015;21(22):5131-5138.
8. Goldenberg DM, Sharkey RM. Sacituzumab govitecan, a novel, third-generation, antibody-drug conjugate (ADC) for cancer therapy. *Expert Opin Biol Ther*. 2020;20(8):871-885.
9. Nagayama A, Vidula N, Ellisen L, Bardia A. Novel antibody-drug conjugates for triple negative breast cancer. *Ther Adv Med Oncol*. 2020;12:1758835920915980.
10. Cardillo TM, Govindan SV, Sharkey RM, et al. Sacituzumab govitecan (IMMU-132), an anti-Trop-2/SN-38 antibody-drug conjugate: characterization and efficacy in pancreatic, gastric, and other cancers. *Bioconjug Chem*. 2015;26(5):919-931.
11. Lopez S, Perrone E, Bellone S, et al. Preclinical activity of sacituzumab govitecan (IMMU-132) in uterine and ovarian carcinosarcomas. *Oncotarget*. 2020;11(5):560-570.
12. Lebert JM, Lester R, Powell E, Seal M, McCarthy J. Advances in the systemic treatment of triple-negative breast cancer. *Curr Oncol*. 2018;25(suppl 1):S142-S150.
13. Jin J, Zhang W, Ji W, Yang F, Guan X. Predictive biomarkers for triple negative breast cancer treated with platinum-based chemotherapy. *Cancer Biol Ther*. 2017;18(6):369-378.
14. Cardillo TM, Sharkey RM, Rossi DL, Arrojo R, Mostafa AA, Goldenberg DM. Synthetic lethality exploitation by an anti-Trop-2-SN-38 antibody-drug conjugate, IMMU-132, plus PARP inhibitors in BRCA1/2-wild-type triple-negative breast cancer. *Clin Cancer Res*. 2017;23(13):3405-3415.
15. Sharma P. Biology and management of patients with triple-negative breast cancer. *Oncologist*. 2016;21(9):1050-1062.
16. Bardia A, Mayer IA, Vahdat LT, et al. Sacituzumab govitecan-hziy in refractory metastatic triple-negative breast cancer. *N Engl J Med*. 2019;380(8):741-751.
17. Bardia A, Mayer IA, Diamond JR, et al. Efficacy and safety of anti-Trop-2 antibody drug conjugate sacituzumab govitecan (IMMU-132) in heavily pretreated patients with metastatic triple-negative breast cancer. *J Clin Oncol*. 2017;35(19):2141-2148.
18. Bardia A, Hurvitz SA, Tolaney SM, et al. Sacituzumab govitecan in metastatic triple-negative breast cancer. *N Engl J Med*. 2021;384:1529-1541.
19. Eisenhauer EA, Therasse P, Bogaerts J, et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur J Cancer*. 2009;45(2):228-247.
20. Hammond ME, Hayes DF, Dowsett M, et al. American Society of Clinical Oncology/College Of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer. *J Clin Oncol*. 2010;28(16):2784-2795.
21. Bychkov A, Sampatanukul P, Shuangshoti S, Keelawat S. TROP-2 immunohistochemistry: a highly accurate method in the differential diagnosis of papillary thyroid carcinoma. *Pathology*. 2016;48(5):425-433.
22. Vidula N, Yau C, Rugo HS. Trop2 gene expression (Trop2e) in primary breast cancer (BC): correlations with clinical and tumor characteristics. *J Clin Oncol*. 2017;35(suppl 15):1075.
23. Cardillo TM, Rossi DL, Zalath MB, et al. Predictive biomarkers for sacituzumab govitecan efficacy in Trop-2-expressing triple-negative breast cancer. *Oncotarget*. 2020;11(43):3849-3862.
24. Remsik J, Bino L, Kahounova Z, et al. Trop-2 plasticity is controlled by epithelial-to-mesenchymal transition. *Carcinogenesis*. 2018;39(11):1411-1418.
25. Bhattacharyya A, Ear US, Koller BH, Weichselbaum RR, Bishop DK. The breast cancer susceptibility gene BRCA1 is required for subnuclear assembly of Rad51 and survival following treatment with the DNA cross-linking agent cisplatin. *J Biol Chem*. 2000;275(31):23899-23903.
26. Kaufman B, Shapira-Frommer R, Schmutzler RK, et al. Olaparib monotherapy in patients with advanced cancer and a germline BRCA1/2 mutation. *J Clin Oncol*. 2015;33(3):244-250.
27. Stoppa-Lyonnet D. The biological effects and clinical implications of BRCA mutations: where do we go from here? *Eur J Hum Genet*. 2016;24(suppl 1):S3-S9.
28. Chen H, Wu J, Zhang Z, et al. Association between BRCA status and triple-negative breast cancer: a meta-analysis. *Front Pharmacol*. 2018;9:909.