Hourly and seasonal variation in photosynthesis and stomatal conductance of soybean grown at future CO₂ and ozone concentrations for 3 years under fully open-air field conditions

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ABSTRACT

It is anticipated that enrichment of the atmosphere with CO₂ will increase photosynthetic carbon assimilation in C3 plants. Analysis of controlled environment studies conducted to date indicates that plant growth at concentrations of carbon dioxide ([CO₂]) anticipated for 2050 (~ 550 μ mol mol⁻¹) will stimulate leaf photosynthetic carbon assimilation (A) by 20 to 40%. Simultaneously, concentrations of tropospheric ozone ([O₃]) are expected to increase by 2050, and growth in controlled environments at elevated [O₃] significantly reduces A. However, the simultaneous effects of both increases on a major crop under open-air conditions have never been tested. Over three consecutive growing seasons > 4700 individual measurements of A, photosynthetic electron transport (J_{PSII}) and stomatal conductance (g_s) were measured on Glycine max (L.) Merr. (soybean). Experimental treatments used free-air gas concentration enrichment (FACE) technology in a fully replicated, factorial complete block design. The mean A in the control plots was 14.5 μ mol m⁻² s⁻¹. At elevated [CO₂], mean A was 24% higher and the treatment effect was statistically significant on 80% of days. There was a strong positive correlation between daytime maximum temperatures and mean daily integrated A at elevated [CO₂], which accounted for much of the variation in CO₂ effect among days. The effect of elevated [CO₂] on photosynthesis also tended to be greater under water stress conditions. The

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*Present address: Department of Environmental Sciences, Brookhaven National Laboratory, Upton, NY 11973-5000, USA. [†]Present address: Plant Science Research Unit, Agricultural Research Service, United States Department of Agriculture, 3908 Inwood Rd, Raleigh, NC 27603, USA. elevated $[O_3]$ treatment had no statistically significant effect on mean A, g_s or J_{PSII} on newly expanded leaves. Combined elevation of $[CO_2]$ and $[O_3]$ resulted in a slightly smaller increase in average A than when $[CO_2]$ alone was elevated, and was significantly greater than the control on 67% of days. Thus, the change in atmospheric composition predicted for the middle of this century will, based on the results of a 3 year open-air field experiment, have smaller effects on photosynthesis, g_s and whole chain electron transport through photosystem II than predicted by the substantial literature on relevant controlled environment studies on soybean and likely most other C3 plants.

Key-words: air pollution; chlorophyll fluorescence; climate change; free-air gas concentration enrichment (FACE); global change.

INTRODUCTION

The response of soybean [Glycine max (L.) Merr.] to anthropogenically driven perturbations in atmospheric trace gases has been widely studied in recognition of its potential for economic impact. Two meta-analyses have summarized the responses of soybean to growth in elevated atmospheric concentrations of CO₂ ([CO₂]; Ainsworth et al. 2002), elevated tropospheric ozone concentrations ($[O_3]$) and a combination of the two gases in enclosed environments (Morgan, Ainsworth & Long 2003). Leaf-level gas exchange measurements show soybean photosynthetic carbon assimilation (A) generally increases with growth in elevated [CO2] (Ainsworth et al. 2002) and decreases when grown in elevated $[O_3]$ (Morgan *et al.* 2003). Lower stomatal conductance (g_s) is associated with growth under higher concentrations of both gases (Ainsworth et al. 2002; Morgan et al. 2003). Despite these generalized responses, there are no studies that have explored full diurnal and

seasonal responses of soybean, or any other plant, to growth in elevated $[O_3]$ over the full life cycle and under fully open-air conditions. Such data have only been provided for elevated $[CO_2]$ previously for wheat (Garcia *et al.* 1998) and soybean (Rogers *et al.* 2004), and in each case only for a single year.

Evidence clearly demonstrates different directional responses of instantaneous leaf-level carbon assimilation (A) for soybean grown in elevated $[CO_2]$ versus $[O_3]$ (e.g. Ainsworth et al. 2002; Morgan et al. 2003). The mechanisms driving the responses in A differ substantially between $[CO_2]$ and $[O_3]$. Growth in elevated $[CO_2]$ stimulates A through an increase in the primary substrate, CO₂, and a decrease in photorespiration. As a result of this, a 37% increase in light-saturated steady-state photosynthesis and 11% increase in light-limited photosynthesis are predicted when $[CO_2]$ is increased from 370 to 550 μ mol mol⁻¹ at 25 °C (Farquhar, Caemmerer & Berry 1980; Long 1991; Long et al. 2004; Rogers et al. 2004). This predicts that stimulation of A at elevated [CO₂] should be greatest during sunny weather and in the middle of the day, and least around dawn and dusk. Enhancement of A by elevated [CO₂] in soybean grown in enclosures has often equalled theoretical expectations (e.g. Ainsworth et al. 2002). However, smaller than predicted stimulation was observed for soybean, and other crops, grown in elevated [CO₂] under field conditions using FACE technology (Garcia et al. 1998; Ainsworth et al. 2003; Rogers et al. 2004; Bernacchi et al. 2005). Increases in A with elevated $[CO_2]$ may be diminished by numerous factors including (1) decreased maximum rate of carboxylation at ribulose-1, 5-bisphosphate carboxylase/oxygenase (RUBISCO) ($V_{c,max}$); (2) decreased maximum whole chain rate of photosynthetic electron transport (J_{max}) , which is proportional to the rate of regeneration of RuBP (Xu, Gifford & Chow 1994; Vu et al. 1997; Luo, Sims & Griffin 1998; Ainsworth et al. 2002; Ainsworth et al. 2004); and (3) decreased g_s (Lawlor & Mitchell 1991; Allen et al. 1998; Ainsworth et al. 2002; Polley 2002; Rogers et al. 2004; Bernacchi et al. 2005). In the field, with a continually fluctuating environment, photosynthesis will rarely achieve a true steady state (e.g. Leakey et al. 2002).

Much less is known regarding the mechanism of response to growth in elevated $[O_3]$, although a decrease in A is typically observed via damage to photosynthetic machinery; caused primarily by a loss of the amount and activity of Rubisco (Pell, Eckardt & Enyedi 1992; Mckee, Farage & Long 1995; Reid, Fiscus & Burkey 1998; Long & Naidu 2002). Ozone affects plants by diffusing through the stomata into the leaf and reacting with the water of the exposed mesophyll apoplast and its solutes to form a wide range of reactive oxygen species (ROS). These in turn interact with plasma membranes and initiate a damage response. It is not possible to calculate a theoretical reduction in A at a given $[O_3]$, though substantial decreases in A with elevated [O₃] is generally observed regardless of the enclosure type in which the treatment is applied. However, the youngest fully expanded leaves of soybean grown in elevated [O₃] under field conditions do not show decreases in A (Morgan *et al.* 2004). Only after prolonged exposure in full sunlight does A begin to decrease with elevated $[O_3]$ (Morgan *et al.* 2004). These results were observed based on gas exchange measurements made on detached leaves measured in controlled environments. Whether this lack of response is observed over diurnal, seasonal and interannual time courses has yet to be determined.

True representation of responses to elevated $[CO_2]$ and [O₃] should include variable conditions over short- and long-term timescales as well as over repeated complete life cycles. The influence of rising temperature (Long 1991) or water stress (Allen et al. 1994) on A has also been predicted to interact with the effect of elevated [CO₂]; however, few studies have the temporal resolution to measure this interaction under field conditions. Rogers et al. (2004) demonstrated variability in the response of a soybean cultivar to elevated [CO₂] over seven measurement dates within one growing season. A mean 25% increase in diurnal integrated leaf-level photosynthesis was observed across the growing season; however daily responses ranged from a 50% increase to no effect (Rogers et al. 2004). From the analysis of a single year it is not possible to know if these responses are characteristic or instead reflect weather or environmental conditions unique to that year. Rogers et al. (2004) did not investigate the effects of elevated $[O_3]$. All previous field studies have used open-top chambers which alter the coupling of the leaf canopy to the ozone-polluted air, which could exaggerate the effect of ozone on leaf photosynthesis and gs relative to natural coupling (Elagoz & Manning 2005).

In this field study on soybean, we test the hypotheses that (1) under fully open-air conditions, the response of photosynthesis to elevated [CO₂] will be less than previously reported; (2) the responses will be affected by both plant developmental stage and time of day; (3) that the magnitude of response of elevated [CO2] on photosynthesis will vary with temperature and water stress; and (4) that no effect of elevated $[O_3]$ on photosynthesis or g_s is observable in newly expanded leaves when compared with control or to elevated $[CO_2]$ treatment. In addition to measuring A and g_s , whole chain electron transport through photosystem II (J_{PSII}) was determined by simultaneous modulated chlorophyll fluorescence to aid interpretation of observed responses. In total, > 4700 individual measurements each of A, g_s and J_{PSII} were made over full diurnal cycles during three complete growing seasons and life cycles. This study used the Soybean Free Air gas Concentration Enrichment (SoyFACE) facility at Champaign, IL, USA.

METHODS

Site description

The SoyFACE facility is contained within a 32 ha (80 acre) field of the South Farms, University of Illinois at Urbana-Champaign (40°03'21.3"N, 88°12'3.4"W, 230 m a.s.l.; www.soyface.uiuc.edu). The soil is a Drummer-Flanagan soil series (fine-silty, mixed, mesic Typic Endoaquoll)

typically very deep and formed from loess and silt parent material deposited on the glacial till and outwash plain. This site has been in continuous cultivation of arable crops for over 100 years. Soybean [*Glycine max* (L.) Merr. cv. Pioneer 93B15] and maize (*Zea mays*) each occupied one half of the field and follow an annual rotation. After the autumn maize harvest, the stubble was chopped with a mower followed by tillage with a rip chisel and conventional single-pass cultivator tillage was used in the spring. Soybean row spacing was 0.38 m and cultivation followed typical Illinois agricultural practices (Ainsworth *et al.* 2004; Rogers *et al.* 2004).

Four blocks were nested within the 16 ha of soybean, each containing one control, one elevated $[CO_2]$, one elevated $[O_3]$ and one elevated $[CO_2] + [O_3]$ treatment plot. The plots were octagonal with a mean diameter of 20 m, defined by horizontal pipes which provide the controlled release of O_3 , CO_2 or both gases. These pipes and associated equipment are assembled immediately after sowing, so that treatment was applied at emergence and continued through to final harvest. The plots were separated by at least 100 m to avoid cross-contamination. Treatments were arranged in a randomized complete block design (*n* of 4 for each treatment) to control for topographic and soil variation across the field.

The target $[CO_2]$ of 550 μ mol mol⁻¹ and $[O_3]$ of $1.23 \times \text{current}$ concentration was based on projected future mean global tropospheric concentrations for 2050 (IPCC 2001; Prather et al. 2001, 2003). Seasonal average [CO₂] in the elevated $[CO_2]$ and elevated $[CO_2] + [O_3]$ treatments was 552 μ mol mol⁻¹ in 2002 and 2003, and 550 μ mol mol⁻¹ in 2004. This compared to an ambient $[CO_2]$ of ~ $375 \,\mu$ mol mol⁻¹. Seasonal average [O₃] in the elevated $[O_3]$ and elevated $[CO_2] + [O_3]$ treatments was greater than ambient [O₃] by 21% in 2002, and 25% in 2003 and 2004 (Fig. 1). Mean 8 h maximum [O₃] throughout the season and seasonal AOT40 and SUM06 values (Mauzerall & Wang 2001) were calculated from ozone concentrations measured on site (model 49CO3 analyser; Thermo Environmental Instruments, Franklin, MA, USA; calibration US EPA Equivalent Method EQQA-0880-047, range 0 to $0.05-1.0 \,\mu\text{mol mol}^{-1}$) from control and elevated [O₃] plots (Fig. 1). Elevation of $[CO_2]$ and $[O_3]$ was based on the method of Miglietta et al. (2001) modified as described previously for CO₂ (Ainsworth et al. 2004) and O₃ (Morgan et al. 2004). All 3 years of measurement included the control, elevated $[CO_2]$ and elevated $[O_3]$ treatments while the elevated $[CO_2] + [O_3]$ treatment was added for 2003 and 2004.

Palmer crop moisture index (PCMI) is an estimate of short-term moisture conditions based on temperature, precipitation and modelled soil water content (Palmer 1968). As PCMI decreases below zero, it indicates progressively greater drought stress conditions. PCMI from 1973 to 2004 for East Central Illinois was provided by the Climate Operation Branch of the National Oceanic and Atmospheric Administration (NOAA) (http://www.usda.gov/oce/waob/ jawf/). Meteorological measurements of precipitation, temperature, humidity and solar radiation were collected in 15 min averages from a weather station located at SoyFACE. Thirty year mean averages (1971–2000) of meteorological variables were determined from recorded data of weather stations located within 3 km of the SoyFACE field through the Midwestern Climate Information System (MICIS) operated by the Midwestern Regional Climate Center, Illinois State Water Survey (http://mrcc.sws.uiuc.edu/).

Gas exchange and chlorophyll fluorometry measurements

Measurements were made from predawn to post-dusk on 15 d over three complete growing seasons (Table 1). A total of three teams in 2002 and four teams in 2003 and 2004 measured leaf level gas exchange each using a portable open gas exchange systems incorporating infrared CO₂ and water vapor analysers (Li-Cor 6400; Li-Cor, Inc., Lincoln, NE, USA) coupled with an integrated portable chlorophyll fluorometer (LI-6400-40 leaf chamber fluorometer; Li-Cor, Inc.). Prior to measurements, the infrared CO_2 and water vapor analysers were zeroed using anhydrous calcium carbonate (Drierite, W.A. Hammond Drierite Company, Ltd, Xenia, OH, USA) to remove water, and NaOH/Ca(OH)₂ (soda lime granules, Morgan Medical, Ltd, Kent, UK) to remove CO_2 , from the air entering the cuvettes. The gas exchange systems were calibrated every 30-45 d throughout the season using gas of a known, certified, CO2 concentration with 21% oxygen and nitrogen as balance (S.J. Smith, Champaign, IL, USA), and known water vapor concentrations generated with a controlled humidification system (LI-610 Portable Dew Point Generator; Li-Cor, Inc.).

Measurements were made on the youngest fully expanded leaves from plants within a 4×2 m area in each plot reserved for this study. Measurements were initiated prior to sunrise in 2002 and 2004, and immediately after sunrise in 2003. Early morning measurements often coincided with dew present on the leaves; in which case, leaves were blotted with paper towels immediately prior to enclosure into the leaf chamber. g_s was not calculated at these times, because residual surface moisture likely increased apparent water vapor fluxes from the leaf. Three leaves on separate plants in each plot were measured once approximately every 2 h. Each of the teams sampled one complete block for each time point. Each team started their cycle of measurements in a different block and cycled through the blocks over the day. This avoided confounding time within the approximately 45 min to 1 h that it took to complete all measurements with treatment and avoided confounding any differences between teams with a block.

Immediately prior to the start of a measurement cycle, the red-blue light-emitting diode (LED) light source integrated into each gas exchange system was set to the incident photosynthetically active photon flux (*I*) that was determined using a photometric sensor mounted just above the plant canopy (LI-210, Li-Cor, Inc.). Similarly, the measurement temperature of the gas exchange cuvette block was



Figure 1. Meteorological conditions measured during the 2002, 2003 and 2004 growing seasons. Panels (a, b & c) show the daily maximum and minimum temperatures throughout each growing season (solid lines) and the daily maximum and minimum temperatures averaged for the period 1971–2000 (grey broken line). Also shown is the mean temperature for each of the three growing seasons and the 30 year mean (1971–2000, d). Daily total precipitation is shown for each of the growing seasons (e, f & g) along with mean May–September Palmer crop moisture index (PCMI) values for Illinois Climate Division 5 by the Climate Operation Branch of the National Oceanic and Atmospheric Administration (NOAA) (http://www.ncdc.noaa.gov/oa/ncdc.html; h). Total daily integral of solar radiation for each day of the growing season (i, j, k) is shown along with mean daily solar radiation for each of the growing seasons compared with the 30 year mean value. The dates on which diurnal photosynthesis (*A*), stomatal conductance (g_s) and whole chain electron transport through photosystem II (J_{PSII}) were measured are indicated by the open triangles. Mean 8 h maximum ozone concentrations are shown for each of the three growing seasons (m, n & o) along with ozone exposure calculations using the AOT40 (white bars) and SUM06 (grey bars) for the control (open bars) and elevated [O₃] (striped bars, p).

set to the open-air temperature recorded at the start of the measurement period using an aspirated shielded type T thermocouple (copper-constantan) positioned 2 m above ground and attached to a datalogger (CR7 700X Measurement and Control System; Campbell Scientific, Logan, UT, USA). Humidity in the gas exchange cuvette was not controlled except to avoid condensation inside the gas exchange system during early morning and evening measurements. The light and temperature conditions within the chamber were held constant for the duration of the measurement period, regardless of short-term fluctuations in light and air temperature. [CO₂] in the cuvettes was set to growth [CO₂] for each treatment, with the control and elevated [O₃] plots having a set point of 370 μ mol mol⁻¹, and

the elevated $[CO_2]$ and elevated $[CO_2] + [O_3]$ plots having a set point of 550 μ mol mol⁻¹. Conditions were allowed to stabilize for 45–60 s after enclosing 2 cm² of an attached leaflet in the gas exchange cuvette. The time required for one complete measurement cycle of all 16 plots ranged from 45 min to 1 h.

Calculations of A and g_s followed the equations of von Caemmerer & Farquhar (1981) and for J_{PSII} Genty, Briantais & Baker (1989). Daily integrated rates of carbon assimilation (A') and whole chain electron transport (J') were calculated from the instantaneous measurements made for each day of the growing season as described previously (Rogers *et al.* 2004). In 2003 when predawn measurements were not made, the leaves were assumed to have a net

Table 1. Calendar date, day of year (DOY), diurnal start and finish times of experimental measurements and estimated crop ontogenetic developmental stage (defined in Ritchie *et al.* 1989) for *Glycine max* grown at Soybean Free Air Concentration Enrichment (SoyFACE) in Champaign, IL

Date	DOY	First measurement (h)	Last measurement (h)	Growth stage
1 July 2002	182	4:30	20:30	Vegetative, fourth trifoliate
17 July 2002	198	4:30	20:30	Full bloom, flower in top 2 nodes
31 July 2002	212	4:30	20:30	Beginning pod in top 4 nodes
15 August 2002	227	4:30	20:30	3.175 mm seed in top 4 nodes
27 August 2002	239	5:00	20:00	3.175 mm seed in top 4 nodes
10 September 2002	253	5:30	19:00	Full size seed in top 4 nodes
25 June 2003	176	7:00	21:00	Vegetative, second trifoliate
16 July 2003	197	7:00	21:00	Beginning bloom, first flower
7 August 2003	219	7:30	21:30	Beginning pod in top 4 nodes
26 August 2003	238	7:00	19:30	3.175 mm seed in top 4 nodes
30 June 2004	182	4:30	20:30	Vegetative, fourth trifoliate
14 July 2004	196	4:30	20:30	Beginning bloom, first flower
4 August 2004	217	4:30	19:45	Full pod, three-fourth inch pod in top 4 nodes
24 August 2004	237	4:30	20:00	3.175 mm seed in top 4 nodes
9 September 2004	253	5:00	19:00	Full size seed in top 4 nodes

carbon assimilation of zero at sunrise. A polynomial was fitted to the data and the area under the curve integrated individually for each plot throughout each day. The order of the polynomial was increased stepwise until no significant increase in r^2 was achieved. The resulting polynomials ranged from third to fifth order. Calculations for A' and J' were limited to the period of the day when $A \ge 0$; that is, net photosynthesis was positive.

Statistical analysis

Statistical analyses were conducted using the models developed previously for SoyFACE (Rogers et al. 2004). The plot was identified as the sample unit, with this value being the mean of the three subsamples in a plot at a single time point. The diurnal measurements were analysed separately for each day of year (DOY) using a mixed model complete block repeated measures analysis of variance (ANOVA) (PROC MIXED, SAS v9.1, SAS Institute, Cary, NC, USA), with time of day, treatment and time of day-by-treatment as fixed effects. A priori determined pairwise comparisons between the treatments and the controls were performed for each measurement day. The best-fit variance/covariance matrices were chosen for each variable using Akaike's information criterion to correct for inequality of variance between the sampling time periods (Keselman et al. 1998; Littell, Henry & Ammerman 1998; Littell, Pendergast & Natarajan 2000). Integrated calculation of A' and J' was also analysed as above only with DOY, treatment and DOY-by-treatment as fixed effects. Comparison between years was conducted using a complete block ANOVA with year and treatment as main effects. Because the experiment plots were rotated to different ends of the field each year, repeated measures analysis was not appropriate. Comparisons between the percentage stimulation in photosynthesis and daily maximum temperatures or PCMI were made using a linear regression ANOVA for a first-order fit to the data (SigmaPlot, Systat Software, Inc, Richmond, CA, USA). Given the variability associated with field studies, statistical significance is reported at P < 0.1 as the risk of a Type II error would otherwise be high.

RESULTS

The measured diurnal courses were spaced throughout the growing seasons representing a range of developmental stages (Table 1), ozone concentrations and meteorological conditions (Fig. 1). Of the 3 years, 2002 had the lowest PCMI but all 3 years were higher than the 1973-2001 average (Fig. 1). Daily maximum and minimum temperatures varied substantially throughout the duration of this experiment. Mean temperature was above average in 2002, average in 2003 and below average in 2004 (Fig. 1). Total solar radiation was generally similar among the three growing seasons, with only a slight decrease in solar radiation during the 2004 growing season relative to the previous two seasons (Fig. 1). In addition to variation in temperature, measurements throughout the three growing seasons included days during dry spells (e.g. DOY 238, 2003), rain (e.g. DOY 227, 2002), clear skies (e.g. DOY 182, 2004) and overcast skies (e.g. DOY 227, 2002). Additionally, the crop experienced a hailstorm that destroyed > 50% of the plant canopy on 14 July 2003 (Morgan et al. 2005). Background ambient [O₃] varied within and among seasons with 2002 experiencing the highest and 2004 the lowest concentrations (Fig. 1).

Over the duration of this experiment, ca 320–430 individual leaves were measured on each of the 15 d, spread across the three growing seasons. Averaged across the daylight hours of the 15 d, mean A in the control plots was 14.5 μ mol m⁻² s⁻¹ and 24% higher in the elevated [CO₂] plots at 17.9 μ mol m⁻² s⁻¹. Photosynthesis in the elevated [CO₂] plots was significantly greater on 80% of the measurement days. The mean A for the [O₃] plots was identical to that of the controls, and plots with simultaneous elevated [CO₂] and [O₃] treatment were 19% higher than the controls and significantly higher for 67% of days (Table 2).

Year	Day of year	Main effects			Pairwise comparisons			Means			
		Т	Н	$T \times H$	$\overline{CO_2}$	O ₃	$CO_2 + O_3$	Control	CO ₂	O ₃	$CO_2 + O_3$
2002	182	< 0.001	< 0.001	< 0.001	< 0.001	ns	_	14.7 ± 0.7	19.3 ± 0.7	14.3 ± 0.7	_
	198	< 0.01	< 0.001	< 0.001	< 0.01	ns	_	14.7 ± 0.5	17.5 ± 0.5	14.4 ± 0.5	_
	212	< 0.01	< 0.001	< 0.001	< 0.01	ns	_	16.4 ± 0.9	20.4 ± 0.9	16.9 ± 0.9	_
	227	< 0.001	< 0.001	< 0.001	< 0.001	ns	_	13.7 ± 0.6	18.0 ± 0.6	14.1 ± 0.6	_
	239	ns	< 0.001	< 0.1	ns	ns	_	16.7 ± 0.9	17.7 ± 0.9	15.7 ± 0.9	_
	253	< 0.1	< 0.001	< 0.001	< 0.1	ns	_	11.2 ± 0.9	14.2 ± 0.9	9.7 ± 0.9	-
2003	176	< 0.01	< 0.001	< 0.01	< 0.01	ns	< 0.01	14.0 ± 1.0	18.2 ± 1.0	13.8 ± 1.0	18.5 ± 1.0
	197	< 0.01	< 0.001	< 0.1	< 0.001	ns	< 0.01	16.8 ± 1.0	20.5 ± 1.0	17.9 ± 1.0	20.1 ± 1.0
	219	< 0.1	< 0.001	ns	ns	ns	ns	15.5 ± 0.9	17.4 ± 0.9	15.2 ± 0.9	17.4 ± 0.9
	238	< 0.1	< 0.001	< 0.001	< 0.01	ns	ns	16.8 ± 1.3	22.3 ± 1.3	17.6 ± 1.3	19.7 ± 1.3
2004	182	< 0.01	< 0.001	< 0.01	< 0.001	ns	< 0.01	15.6 ± 1.2	21.0 ± 1.2	15.9 ± 1.2	19.3 ± 1.2
	196	< 0.01	< 0.001	< 0.1	< 0.1	ns	< 0.1	13.7 ± 0.6	15.8 ± 0.6	12.8 ± 0.6	15.0 ± 0.6
	217	< 0.1	< 0.001	ns	ns	< 0.1	ns	14.7 ± 0.8	15.0 ± 0.8	13.4 ± 0.8	15.3 ± 0.8
	237	< 0.01	< 0.001	< 0.001	< 0.001	ns	< 0.01	13.3 ± 0.8	16.9 ± 0.8	13.7 ± 0.8	16.3 ± 0.8
	253	< 0.1	< 0.001	ns	< 0.1	ns	< 0.1	8.8 ± 1.4	14.0 ± 1.4	11.9 ± 1.4	13.8 ± 1.4

Table 2. Statistic alanalysis of the complete-block repeated measures analysis of variance for instantaneous diurnal measurements of leaf-level photosynthesis (*A*)

The main effects are treatment (T), hour (H) and $T \times H$, with H as a repeated term. Pairwise comparisons of the three treatments versus control were determined *a priori*. Significance of *F* values is given in the table where non-statistical significance (ns) > 0.1. Also shown are mean daily values of *A* for each treatment on each date (± one SE of the mean).

 $J_{\rm PSII}$ averaged across all days and times averaged 132 μ mol $m^{-2} s^{-1}$ in the control plots and was 9% higher at 143 μ mol $m^{-2} s^{-1}$ in the elevated [CO₂] treatment. J_{PSII} was also greater in the elevated $[CO_2] + [O_3]$ treatment than the control, but only by 3%, while there was no effect of $[O_3]$ alone. Given the many measurements, only representative figures showing the diurnal course of A and J_{PSII} from each growing seasons are included (Fig. 2; all data collected throughout this experiment may be viewed in the online supplementary information). Over all three growing seasons, elevated $[CO_2]$ and elevated $[CO_2] + [O_3]$ grown leaves show higher A, but the stimulation was only apparent in the middle of the day (Fig. 2). This accounts for the significant treatment by time effect seen most days (Table 2). The pattern of response observed for J_{PSU} was similar to A, with elevated $[CO_2]$ grown plants showing higher rates than the control (Fig. 2). However the proportional increase, relative to control was only about half that observed for A and only significant on 40% of days (Table 3; Fig. 2). When plants were grown in elevated $[O_3]$ and $[CO_2]$, a significant increase in J_{PSII} was only apparent on 33% of days, and when grown in elevated $[O_3]$ alone a significant decrease in J_{PSII} was observed on 1 d (Table 3). Two of the measurement dates in which a statistically significant mean effect of elevated [CO₂] was lacking showed a significant time by treatment response. Further investigation of these days shows certain time periods throughout the day when J_{PSII} is statistically higher for elevated [CO₂] than control grown plants.

Representative plots of diurnal g_s are shown for each of the 3 years (Fig. 3). Averaged across the 15 d, the mean g_s in the control plots was 0.46 mol m⁻² s⁻¹ and 16% lower in the elevated [CO₂] plots at 0.38 mol m⁻² s⁻¹. Decreases in g_s were statistically significant on 53% of measurement days as a result of growth in elevated $[CO_2]$ (Table 4). The average g_s for plants grown under elevated $[O_3]$ was only 1% lower than of the controls and not significantly different, except on 2 d in 2003. With the exception of 1 d the elevated $[CO_2] + [O_3]$ grown plants mirrored the responses of the elevated $[CO_2]$ grown plants (Table 4).

Daily integral of A(A') was typically higher for the plants grown at elevated $[CO_2]$ and elevated $[CO_2] + [O_3]$ over the duration of the experiment (Fig. 4). Increases in seasonal A' ranged from 18% in 2004 to 25% in 2002 for the elevated $[CO_2]$ grown plants and from 16% in 2004 to 20.5% in 2003 for the elevated $[CO_2] + [O_3]$ grown plants (Table 5). There was a significant treatment-by-day interaction in 2002 and 2004, suggesting a variable treatment response over the duration of these growing seasons (Fig. 4; Table 5).

A comparison of A' between control and elevated $[CO_2]$ grown plants for the 3 years showed differences among years and treatments (P < 0.001) but no significant interaction. Analysing this further with pairwise comparisons showed that there was no difference between 2002 and 2003 but A' was significantly lower in 2004 (Fig. 4). Per cent stimulation of A' by elevated $[CO_2]$ rose with daily maximum temperature ($r^2 = 0.51$; Fig. 5). Stimulation of A' declined with increasing PCMI, which is a measure of decreasing water stress ($r^2 = 0.25$, P = 0.0675; Fig. 5). However, the relationship between PCMI and percentage stimulation of photosynthesis is only statistically significant when DOY 217 in 2004 is excluded from the analysis (Fig. 5).

Daily integrated rates of whole chain electron transport through photosystem II (J') were also estimated from the diurnal responses (Fig. 6). The response of J' to elevated [CO₂] is variable, as expected from the diurnal responses;



Figure 2. Representative measurements of *A* (solid lines) and J_{PSII} (dotted lines) over the diurnal time course from one day in 2002 (first column), 2003 (second column) and 2004 (third column). Measurements were made on day of year 182 for the 2002 and 2004 growing season, and day of year 176 during 2003. The top three panels compare control and elevated [CO₂], the middle three panels control versus elevated [O₃] and the bottom two panels control versus elevated [CO₂] + [O₃]. Open symbols represent ambient [O₃], closed symbols represent elevated [O₃], circles represent ambient [CO₂] and triangles represent elevated [CO₂]. Error bars represent one SE around the mean.

Table 3. Statistical analysis of the complete-block repeated measures analysis of variance for instantaneous diurnal measurements of whole chain electron transport through photosystem II (J_{PSII})

Year	Day of year	Main effects			Pairwise comparisons			Means			
		Т	Н	$T \times H$	$\overline{CO_2}$	O ₃	$CO_2 + O_3$	Control	CO ₂	O ₃	$CO_2 + O_3$
2002	182	< 0.05	< 0.001	ns	< 0.05	ns	_	159.1 ± 3.9	168.3 ± 4.1	157.8 ± 4.2	_
	198	ns	< 0.001	ns	ns	ns	_	147.6 ± 3.0	150.9 ± 3.0	147.6 ± 3.0	_
	212	ns	< 0.001	ns	ns	ns	_	153.0 ± 5.1	164.1 ± 5.1	155.5 ± 5.1	_
	227	< 0.05	< 0.001	ns	< 0.05	ns	_	127.7 ± 2.9	134.5 ± 2.9	124.8 ± 2.9	_
	239	ns	< 0.001	ns	ns	ns	_	132.8 ± 4.5	132.0 ± 4.5	125.8 ± 4.5	_
	253	< 0.05	< 0.001	ns	ns	< 0.05	-	127.6 ± 4.0	132.0 ± 4.0	115.1 ± 4.0	-
2003	176	ns	< 0.001	< 0.1	ns	ns	ns	167.4 ± 3.8	173.2 ± 3.8	164.6 ± 3.8	168.4 ± 3.8
	197	ns	< 0.001	ns	< 0.05	ns	ns	114.5 ± 4.5	123.2 ± 4.5	118.9 ± 4.5	119.0 ± 4.5
	219	ns	< 0.001	ns	ns	ns	ns	117.1 ± 3.7	118.1 ± 3.7	118.1 ± 3.7	109.9 ± 3.7
	238	< 0.1	< 0.001	ns	< 0.01	ns	ns	155.6 ± 4.3	170.9 ± 4.3	162.4 ± 4.3	162.6 ± 4.3
2004	182	< 0.01	< 0.001	< 0.05	< 0.05	ns	< 0.001	136.1 ± 4.2	142.4 ± 4.2	137.4 ± 4.2	147.5 ± 4.2
	196	< 0.05	< 0.001	ns	< 0.05	ns	ns	100.0 ± 3.9	107.6 ± 3.9	98.1 ± 3.9	103.9 ± 3.9
	217	< 0.01	ns	< 0.0001	ns	ns	ns	113.0 ± 4.0	108.4 ± 4.0	106.5 ± 4.0	110.9 ± 4.0
	237	< 0.1	< 0.001	ns	< 0.1	ns	< 0.1	125.8 ± 3.4	130.8 ± 3.4	125.0 ± 3.4	131.8 ± 3.5
	253	< 0.1	< 0.01	ns	< 0.05	ns	< 0.05	98.1 ± 4.9	109.2 ± 4.9	105.1 ± 4.9	109.5 ± 4.9

The main effects are treatment (T), hour (H) and T × H, with H as a repeated term. Pairwise comparisons of the three treatments versus control were determined *a priori*. Significance of *F* values is given in the table where non-statistical significance (ns) > 0.1. Also shown are mean daily values of J_{FSII} for each treatment on each date (± one SE of the mean).



Figure 3. Representative measurements of stomatal conductance (g_s) over the diurnal time course from the days represented in Fig. 2. The top three panels compare control and elevated [CO₂], the middle three panels control versus elevated [O₃] and the bottom two panels control versus elevated [CO₂] + [O₃]. Open symbols represent ambient [O₃], closed symbols represent elevated [O₃], circles represent ambient [CO₂] and triangles represent elevated [CO₂]. Error bars represent one SE around the mean.

Table 4. Statistical analysis of the complete-block repeated measures analysis of variance for instantaneous diurnal measurements of stomatal conductance (g_s)

Year	Day of year	Main effects			Pairwise comparisons			Means			
		Т	Н	$T \times H$	CO_2	O ₃	$CO_2 + O_3$	Control	CO_2	O ₃	$CO_2 + O_3$
2002	182	< 0.1	< 0.001	ns	< 0.05	ns	_	0.40 ± 0.03	0.29 ± 0.03	0.38 ± 0.03	_
	198	< 0.01	< 0.001	< 0.05	< 0.001	ns	_	0.46 ± 0.02	0.33 ± 0.02	0.43 ± 0.02	_
	212	ns	< 0.01	< 0.05	ns	ns	_	0.57 ± 0.05	0.54 ± 0.05	0.53 ± 0.05	_
	227	ns	< 0.001	ns	ns	ns	_	0.57 ± 0.04	0.50 ± 0.04	0.57 ± 0.04	_
	239	< 0.05	< 0.001	ns	< 0.05	ns	_	0.59 ± 0.03	0.50 ± 0.03	0.62 ± 0.03	_
	253	ns	< 0.001	ns	ns	ns	-	0.40 ± 0.03	0.40 ± 0.03	0.38 ± 0.03	_
2003	176 197 219 238	< 0.05 < 0.001 < 0.01 ns	< 0.001 < 0.001 < 0.001 < 0.001	ns ns ns ns	< 0.01 < 0.05 < 0.01 ns	ns < 0.05 < 0.05 ns	ns < 0.05 < 0.01 ns	$\begin{array}{c} 0.22 \pm 0.01 \\ 0.49 \pm 0.03 \\ 0.65 \pm 0.03 \\ 0.36 \pm 0.03 \end{array}$	$\begin{array}{c} 0.18 \pm 0.01 \\ 0.43 \pm 0.03 \\ 0.48 \pm 0.03 \\ 0.36 \pm 0.03 \end{array}$	$\begin{array}{c} 0.22 \pm 0.01 \\ 0.55 \pm 0.03 \\ 0.56 \pm 0.03 \\ 0.37 \pm 0.03 \end{array}$	$\begin{array}{c} 0.22 \pm 0.01 \\ 0.42 \pm 0.03 \\ 0.50 \pm 0.03 \\ 0.30 \pm 0.03 \end{array}$
2004	182 196 217 237 253	ns < 0.01 < 0.001 ns ns	< 0.001 < 0.001 < 0.001 < 0.1 < 0.001	ns ns < 0.05 ns ns	ns < 0.01 < 0.001 ns ns	ns ns ns ns ns	ns < 0.01 < 0.001 ns ns	$\begin{array}{c} 0.33 \pm 0.03 \\ 0.42 \pm 0.03 \\ 0.41 \pm 0.02 \\ 0.63 \pm 0.08 \\ 0.38 \pm 0.05 \end{array}$	$\begin{array}{c} 0.30 \pm 0.03 \\ 0.29 \pm 0.03 \\ 0.25 \pm 0.02 \\ 0.57 \pm 0.08 \\ 0.34 \pm 0.04 \end{array}$	$\begin{array}{c} 0.36 \pm 0.03 \\ 0.41 \pm 0.03 \\ 0.42 \pm 0.02 \\ 0.66 \pm 0.08 \\ 0.31 \pm 0.04 \end{array}$	$\begin{array}{c} 0.31 \pm 0.03 \\ 0.29 \pm 0.03 \\ 0.30 \pm 0.02 \\ 0.58 \pm 0.08 \\ 0.40 \pm 0.04 \end{array}$

The main effects are treatment (T), hour (H) and T × H, with H as a repeated term. Pairwise comparisons of the three treatments versus control were determined *a priori*. Significance of *F* values is given in the table where non-statistical significance (ns) > 0.1. Also shown are mean daily values of g_s for each treatment on each date (± one SE of the mean).



Figure 4. Daily integral of carbon assimilation (A') for all measurement dates for all three growing seasons: 2002, 2003 and 2004. Seasonal means of A' for each treatment in each year are also presented as bars to the right in each graph. Open symbols represent ambient [O₃], closed symbols represent elevated [O₃], circles represent ambient [CO₂] and triangles represent elevated [CO₂]. Error bars represent one SE around the mean.

however, there was a general and significant increase in J' averaging 5% across the 3 years (Fig. 6, Table 5). No treatment effect of elevated [O₃] was observed for any of the 3 years. Seasonal average J' was 4.4% greater in the elevated [CO₂] + [O₃] treatment than the control treatment in 2004.

To more clearly assess the impacts of the various treatments on A, J_{PSII} and g_s , and to allow for a more direct comparison of these with previously published reports, midday values were extracted from each diurnal data set (Fig. 7). The stimulation of midday rates of A and J_{PSII} was slightly higher than A' and J', respectively, for all 3 years (Table 5). A decrease in midday g_s with elevated [CO₂] and elevated [CO₂] + [O₃] occurred on a majority of the measurement days (Fig. 7) and resulted in statistically significant decreases throughout all growing seasons (Table 5). However, the responses of midday g_s over the three seasons to elevated [CO₂] appear to differ, as 2002 and 2003 showed a 20 and 23% decrease in midday g_s , respectively, whereas 2004 resulted in a 35% decrease (Fig. 7). In parallel to the diurnal response, no effect of elevated [O₃] on midday g_s was apparent.

DISCUSSION

We tested the hypotheses that (1) the response of leaf photosynthesis (A), g_s and whole chain photosystem II electron transport (J_{PSII}) to elevated [CO₂] will be less under fully open-air conditions than previous reports; (2) that responses to [CO₂] will be affected by both date and time of day; (3) stimulation of photosynthesis by elevated [CO₂] will be greater with higher temperatures and water stress; and (4) no effect of elevated [O₃] will be apparent on A, g_s and J_{PSII} in newly expanded leaves when compared with control or to elevated [CO₂] treatment. Hypothesis 1 was partially supported and hypotheses 2, 3 and 4 were fully supported by the results.



Figure 5. The relationship between per cent stimulation in A' in elevated [CO₂] compared with control plotted as a function of (a) daily maximum temperature and (b) Palmer crop moisture index (PCMI) for all 3 years combined. The lines represent first-order linear regressions fitted through the data ($r^2 = 0.51$, P = 0.003 for panel a and $r^2 = 0.25$, P = 0.0675 for panel b). One measurement date, day of year 217 in 2004, was excluded from the linear regression analysis for panel b and is represented by a triangle symbol.

Table 5. Statistical analysis of the main effects and pairwise comparisons for midday rates of carbon uptake (*A*), daily integral of *A* (*A'*), midday rates of whole chain electron transport through photosystem II (J_{PSII}), daily integrated J_{PSII} (J') and midday values of stomatal conductance (g_s)

2002	Main effec	ets		Control ve	rsus		% Deviation from control		
	T	D	$T \times D$	$\overline{CO_2}$	O ₃	$CO_2 + O_3$	CO ₂	O ₃	$CO_2 + O_3$
Midday A	< 0.001	< 0.01	ns	< 0.01	ns	_	24.2	5.5	
Α'	< 0.001	< 0.001	< 0.05	< 0.001	ns	-	22.6	-1.9	_
Midday J_{PSII}	< 0.01	ns	ns	< 0.1	ns	-	5.1	1.7	
J'	< 0.01	< 0.001	ns	< 0.05	ns	_	4.6	-1.7	_
Midday g_s	< 0.1	< 0.001	ns	< 0.1	ns	_	-23.7	-2.4	-
2003									
Midday A	< 0.05	< 0.001	ns	< 0.001	ns	< 0.001	30.6	1.8	29.9
A'	< 0.001	< 0.01	ns	< 0.001	ns	< 0.001	25.4	2.6	20.5
Midday J_{PSII}	< 0.01	< 0.05	ns	< 0.05	ns	< 0.1	7.0	-1.2	4.9
J'	< 0.1	< 0.001	ns	< 0.05	ns	ns	6.7	1.9	0.3
Midday g_s	< 0.01	< 0.01	ns	< 0.01	ns	< 0.05	-20.5	4.1	-17.3
2004									
Midday A	< 0.01	0.0003	ns	< 0.001	ns	< 0.01	22.9	-1.8	20.0
Α'	< 0.001	< 0.001	< 0.01	< 0.001	ns	< 0.001	18.4	-1.6	16.4
Midday J_{PSII}	< 0.001	< 0.001	ns	< 0.01	ns	< 0.001	5.7	0.0	8.4
J'	< 0.01	< 0.001	ns	< 0.05	ns	< 0.05	3.7	-0.5	4.4
Midday g_s	< 0.001	< 0.05	ns	< 0.001	ns	< 0.001	-36.7	-4.1	-30.6

The data were analysed using a repeated measures complete-block analysis of variance with treatment (T), hour (H) and $T \times H$ as the main effects. *A priori* determined pairwise comparisons of the three treatments versus control and the per cent deviation of the various treatments from control are also shown. Non-statistical significance is denoted by *ns.* Means for the control and treatments are shown in Figs 4, 6 and 7.

Meta-analyses of 111 prior studies of soybean grown in enclosed environments show an increase in A of 39% and a decrease in g_s of 40% at [CO₂] elevated on average to 689 µmol mol⁻¹ (Ainsworth *et al.* 2002). Assuming a linear response of these variables to [CO₂], this would translate to an increase in A of 20% and a decrease in g_s of 20% with elevation of CO₂ to 550 µmol mol⁻¹. In this open-air experiment in which CO₂ was elevated to 550 µmol mol⁻¹, A was increased by 24% and g_s decreased by 16%. These suggest similar responses in the two environments. However, assuming a linear response to [CO₂] likely underestimates the true response indicating that the open-air responses might still be slightly less than in the enclosure studies.

Elevated $[CO_2]$ consistently resulted in slightly higher J_{PSII} throughout the 3 years of measurements (Fig. 6, Tables 3 & 5). Rogers *et al.* (2004), studying a different soybean cultivar in 2001, noted a possible increase in J_{PSII} , but this was not significant. Here a statistically significant increase in J_{PSII} is observed. In theory J_{PSII} could increase, decrease, or remain constant with growth in elevated $[CO_2]$ depending on whether photosynthesis is limited by Rubisco, RuBP or triose phosphate export from the chloroplast, respectively (Hymus, Baker & Long 2001). We have previously shown that the photosynthetic operating point for control plants grown at SoyFACE is near, or slightly below, the inflection point where photosynthesis is co-limited by Rubisco and RuBP (Bernacchi *et al.* 2005). Despite the slight down-regulation of Rubisco with

elevated $[CO_2]$, the shift in the operating point of photosynthesis from Rubisco- to RuBP-limiting without a decrease in photosynthetic potential (Bernacchi *et al.* 2005) supports a slight increase in J_{PSII} , but one smaller than the increase in A (Hymus *et al.* 2001) as reported here.

The mean increase in A with elevated [CO₂] varied depending on the timescale under consideration. Percentage increases in A for instantaneous measurements ranged from slight decreases to increases of over 50%. However, when integrated over the day, the stimulation of A' by elevated [CO₂] ranged from 4 to 37% (Fig. 5). Little or no increase in A with plant growth in elevated $[CO_2]$ was apparent at the low-light fluxes of early morning and dusk (Fig. 2) accounting for the significant interaction of time of day and $[CO_2]$ (Table 2). This is consistent with the expectation from Rubisco kinetics that A at 25 °C would be stimulated by 37% at light saturation, but only 11% when photosynthesis is light-limited (Long et al. 2004). This is because at light saturation, elevated [CO₂] both increases the velocity of carboxylation and partially inhibits oxygenation at Rubisco. At low light, CO₂ is not limiting the rate of carboxylation but does inhibit oxygenation. As a result, the proportion of RuBP that is oxygenated decreases as $[CO_2]$ rises, resulting in a net increase in CO_2 uptake (Long et al. 2004), This is also consistent with the observed effect of elevated $[CO_2]$ on J_{PSII} where increase is evident only around midday when photosynthesis may be limited by the amount of active Rubisco and increased velocity of



Figure 6. Daily integral of electron transport rates (J') for all measurement dates for all three growing seasons: 2002, 2003 and 2004. Seasonal means of J' for each treatment in each year are also presented as bars to the right in each graph. Open symbols represent ambient [O₃], closed symbols represent elevated [O₃], circles represent ambient [CO₂] and triangles represent elevated [CO₂]. Error bars represent one SE around the mean.

carboxylation at elevated [CO₂] will increase the electron transport requirement. In low light when RuBP is limiting, the electron transport requirement will be unaffected by a change in the ratio of oxygenation/carboxylation (Long & Bernacchi 2003). Elevated $[CO_2]$ significantly decreased g_s and the diurnal pattern of decrease was more variable than for the increase in A. On all dates, midday g_s was significantly less at elevated [CO₂] and not significantly different in elevated $[O_3]$ (Table 5). By contrast, g_s measured over the diurnal period was significantly less on only eight of the 15 measurement dates for elevated [CO₂] and on two dates for elevated [O₃]. Inconsistency between diurnal versus midday g_s was likely caused by smaller or no response in the early morning and around dusk, and also potentially compromised by dew on the leaves in the morning of several of the measurement dates. However, variation in the CO₂ effect on g_s, during the middle of the diurnal period, was predictable for dates in 2003 through use of the Ball, Woodrow & Berry (1987) model (Leakey *et al.*, in press). The absence of an elevated $[CO_2]$ effect on g_s on specific dates could be explained if it corresponds to periods when soil moisture depletion is reducing g_s in ambient $[CO_2]$, but at elevated $[CO_2]$ lower whole plant transpiration is conserving soil water and allowing plants to avoid water stress and maintain high g_s .

Because both the solubility of CO₂ in water and specificity of Rubisco for CO₂ decline relative to O₂ with rising temperature, the stimulatory effect on A of rising $[CO_2]$ should be greater as temperature rises (Long 1991). In practice this theoretical expectation has not been realized in various field studies (e.g. Delgado et al. 1994; Naumburg, Loik & Smith 2004). However, here a strong positive correlation between temperature and per cent stimulation of A with growth in elevated $[CO_2]$ was observed (Fig. 5). Possibly this was achieved by the statistical sensitivity provided by the very large number of measurements (>4700)contributing to this analysis. Drought has similarly been expected to result in an increased response to elevated $[CO_2]$. Decreased g_s at elevated $[CO_2]$ may conserve soil moisture delaying or avoiding any decrease in assimilation due to decreased leaf and plant water status (Rogers et al. 1984; Allen et al. 1994, 1998). The PCMI provides an indicator of potential crop water stress for a specific region. Because the reported PCMI values represent a large portion of East Central Illinois, they may not reflect the actual crop moisture status of the SoyFACE field. Nevertheless, the response of A to elevated $[CO_2]$ significantly increased during periods of greater water stress (Fig. 5). This corresponds with the observation at SoyFACE of enhanced A in maize during periods of drought stress (Leakey et al. 2004). C_4 photosynthesis of maize is saturated at ambient [CO₂], but lower g_s leads to reduced soil water depletion and the potential for amelioration of drought stress and indirect enhancement of carbon gain (Leakey et al. 2006). The same mechanism may well occur in soybean, but is difficult to dissect from the concurrent direct stimulation of A by elevated [CO₂].

A meta-analysis of 53 prior chamber studies of soybean in which $[O_3]$ was elevated to 70 nmol mol⁻¹ (daytime average) showed highly significant reductions in A (20%) and g_s (17%) relative to clean air (< 30 nmol mol⁻¹; Morgan *et al.* 2003). Although this first open-air study of soybean under elevated [O₃] covers a smaller increase in concentration, not only was there no significant effect of elevation but no change in either A or g_s was indicated. The difference between elevated $[O_3]$ effects on A and g_s in this study versus previous experiments may be driven by many factors. Firstly, the meta-analysis focuses on comparisons between $[O_3]$ free air and elevated [O₃] treatments. Fumigation at SoyFACE is a fixed percentage above the background levels, which represents a smaller differential than the majority of previous studies (Morgan et al. 2003). Secondly, it is possible that previous experiments showing an effect of elevated $[O_3]$ on plants may overestimate the responses because of forced coupling between the atmosphere and the canopy. The



Figure 7. Midday measurement of A, J_{PSII} and g_s for each measurement period for all three growing seasons: 2002, 2003 and 2004. Data from the measurement period closest to solar noon were used in the analysis. Seasonal mean midday values for A, J_{PSII} and g_s are also presented to the right of each graph. Open symbols represent ambient [O₃], closed symbols represent elevated [O₃], circles represent ambient [CO₂] and triangles represent elevated [CO₂]. Error bars represent one SE around the mean.

transfer of water from the soil to the bulk atmosphere via transpiration is influenced by at least nine major resistances (e.g. Bazzaz & Sombroek 1996) which are substantially altered in artificial growth environments. The substantial forced mixing that occurs in artificial enclosures reduces many of the canopy resistance pathways, which results in a higher degree of ozone exposure at the leaf level, which would accentuate the effects of ozone. A third possibility may exist in the selection of the youngest fully expanded leaves. As we have previously demonstrated, leaves grown in elevated [O₃] show damage only after prolonged exposure (Morgan et al. 2004). By selecting only the recently expanded leaves for this experiment, older leaves in which greater cumulative ozone uptake may have induced damage were avoided. Moreover, it should be noted that despite no obvious effects of elevated $[O_3]$ on photosynthesis, g_s or J_{PSII} , significant and consistent decreases in leaf area index (Dermody, Long & DeLucia 2006), total plant biomass and yields (Morgan *et al.* 2005) for soybean grown in elevated $[O_3]$ have been observed in the same plots at SoyFACE. Although photosynthesis was also increased significantly by elevated $[CO_2]$ even when $[O_3]$ was also elevated, the increase was 19%, 5% less than when $[CO_2]$ was elevated without elevation of $[O_3]$. This resulted from a significant interaction of elevation of $[CO_2]$ and $[O_3]$ on six out of the 15 d of measurements across all three growing seasons.

In conclusion, photosynthesis, g_s and whole chain electron transport through photosystem II all responded to growth in elevated [CO₂] in a predictable manner, although the percentage changes are less than predicted from enclosure studies. Temperature and soil moisture both strongly affect the response of photosynthesis to elevated [CO₂]. Elevated [O₃], did not have any effect on recently expanded leaves, although it did slightly diminish the response of photosynthesis to elevated [CO₂].

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SUPPLEMENTARY MATERIAL

The following supplementary material is available for this article:

Figure S1. Meteorological data for each of the 15 measurement days during the 3 year experiment. Data for air temperature (open circles), VPD (ticks) and photosynthetically active radiation flux (PAR, lines) are presented in 10 min averages of 10 s measurements from a weather station located at SoyFACE.

Figure S2. Measurements of leaf-level photosynthesis (*A*) as a function of time of day over the complete diurnal cycle for all 15 measurement days over the 3 year experiment. Each panel represents one measurement day. Each point is the mean of four replicate plots with three subsamples per plot. Open symbols represent ambient and closed symbols elevated [O₃] and circles represent control [CO₂] and triangles elevated $[CO_2]$. Error bars are one SD around the mean. Figure S3. Measurements of leaf-level whole chain electron transport through photosystem II (J_{PSII}) as a function of time of day over the complete diurnal cycle for all 15 measurement days over the 3 year experiment. Each panel represents one measurement day. Each point is the mean of four replicate plots with three subsamples per plot. Open symbols represent ambient and closed symbols elevated [O₃] and circles represent control [CO₂] and triangles elevated $[CO_2]$. Error bars are one SD around the mean.

Figure S4. Measurements of stomatal conductance (g_s) as a function of time of day over the complete diurnal cycle for all 15 measurement days over the 3 year experiment. Each panel represents one measurement day. Each point is the mean of four replicate plots with three subsamples per plot. Open symbols represent ambient and closed symbols elevated $[O_3]$ and circles represent control $[CO_2]$ and triangles elevated $[CO_2]$. Error bars are one SD around the mean.

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